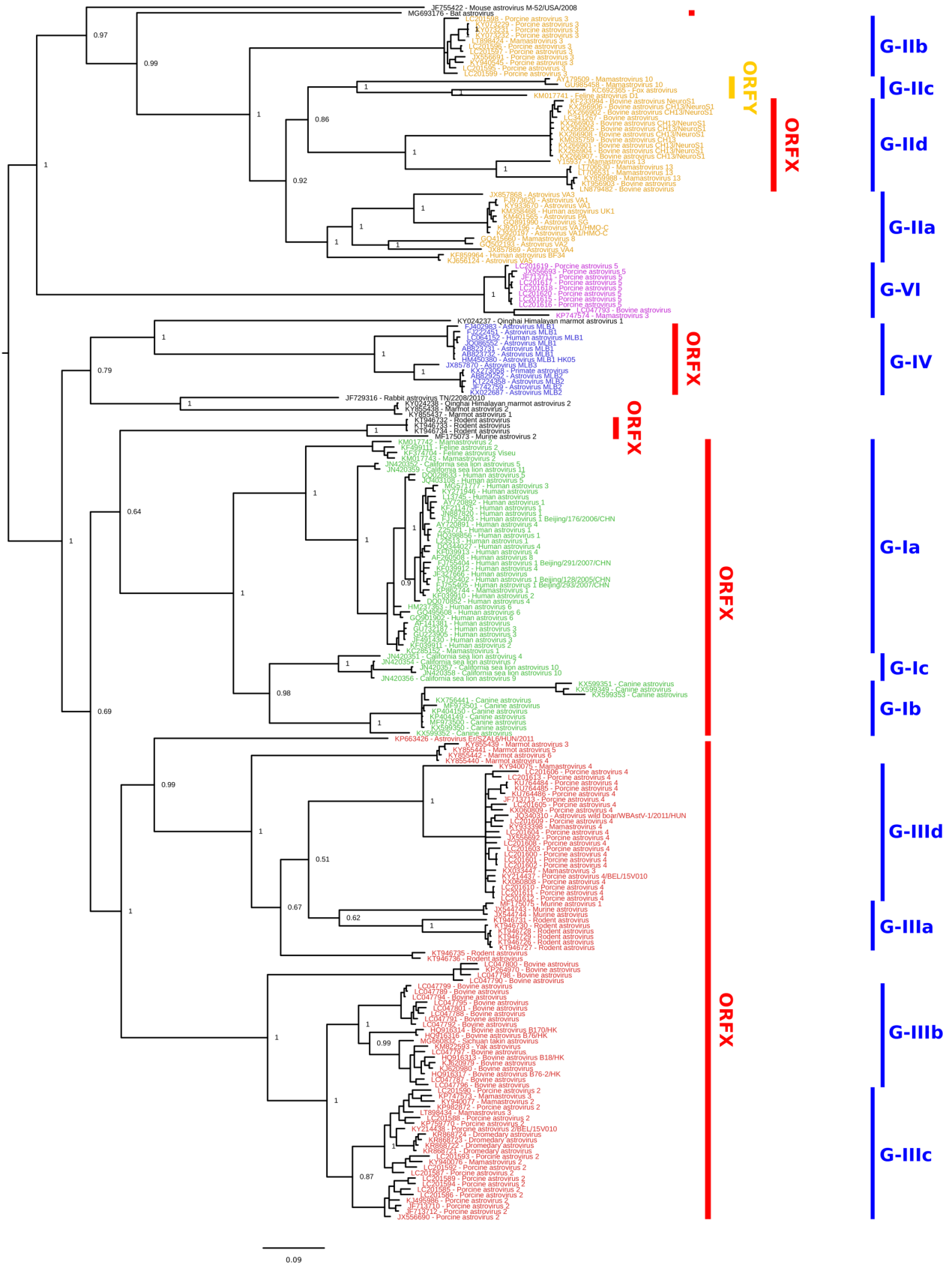


# **A hidden gene in astroviruses encodes a viroporin**

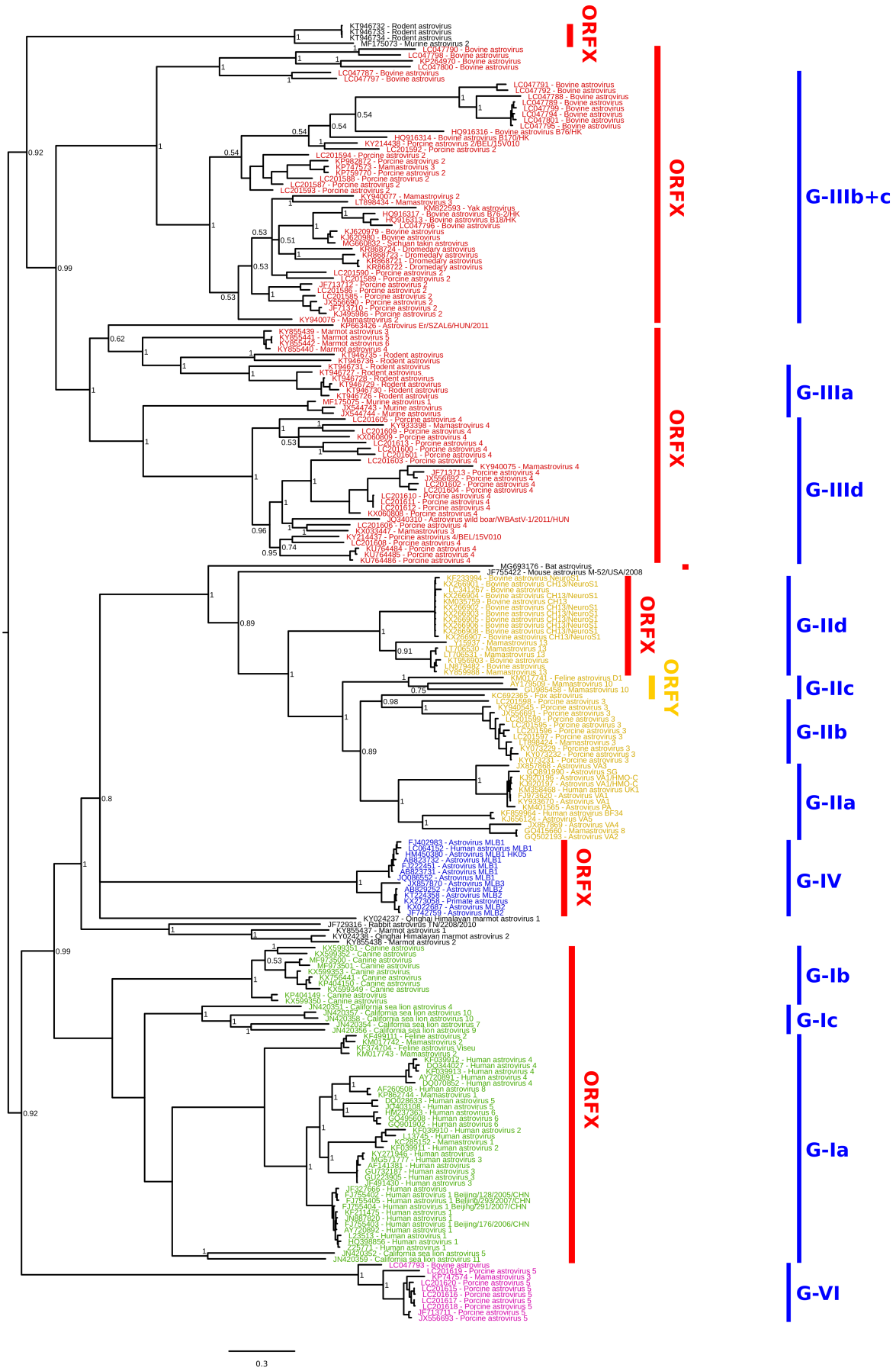
**Lulla *et al.***

**Supplementary information**



**Supplementary Figure 1 | Phylogenetic tree of mammalian astrovirus RdRp sequences.** All full-length mammalian astrovirus genome sequences were obtained from NCBI on 26 July 2018 and the ORF1b amino acid sequences were extracted and aligned with MUSCLE<sup>1</sup>. A maximum likelihood phylogenetic tree was estimated using the Bayesian Markov chain Monte Carlo method implemented in MrBayes version 3.2.3<sup>2</sup>, sampling across the default set of fixed amino acid rate matrices, with one

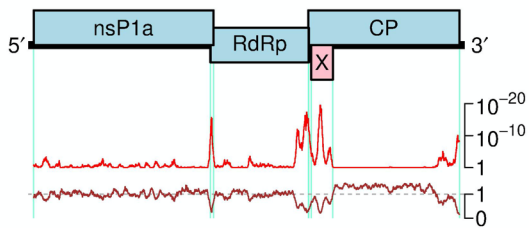
million generations, discarding the first 25% as burn-in. The tree was visualized with FigTree (<http://tree.bio.ed.ac.uk/software/figtree/>). Genogroups (based on the scheme of Yokoyama et al<sup>3</sup>) are indicated with coloured text: green – genogroup I, yellow – genogroup II, red – genogroup III, blue – genogroup IV, magenta – genogroup VI, unclassified sequences – black. Subgroups of sequences, defined on the basis of RdRp phylogeny for the purposes of this study only, are indicated in blue at right (G-Ia, G-Ib, etc). Taxa that have a putative ORFX (ORFY) are indicated in red (yellow) at right. The tree is midpoint rooted and deeper nodes are labelled with posterior probability values.



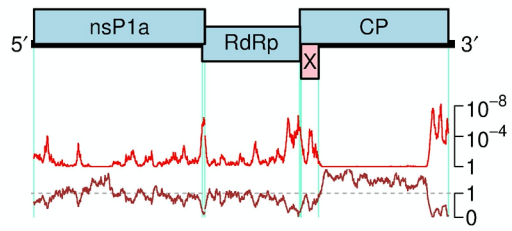
**Supplementary Figure 2 | Phylogenetic tree of mammalian astrovirus CP sequences.** All full-length mammalian astrovirus genome sequences were obtained from NCBI on 26 July 2018 and the

ORF2 amino acid sequences were extracted and aligned with MUSCLE<sup>1</sup>. A maximum likelihood phylogenetic tree was estimated using the Bayesian Markov chain Monte Carlo method implemented in MrBayes version 3.2.3<sup>2</sup>, sampling across the default set of fixed amino acid rate matrices, with one million generations, discarding the first 25% as burn-in. The tree was visualized with FigTree (<http://tree.bio.ed.ac.uk/software/figtree/>). Genogroups (based on the scheme of Yokoyama et al<sup>3</sup>) are indicated with coloured text: green – genogroup I, yellow – genogroup II, red – genogroup III, blue – genogroup IV, magenta – genogroup VI, unclassified sequences – black. Subgroups of sequences, defined on the basis of RdRp phylogeny (Supplementary Fig. 1) for the purposes of this study only, are indicated in blue at right (G-Ia, G-Ib, etc). Taxa that have a putative ORFX (ORFY) are indicated in red (yellow) at right. The tree is midpoint rooted and deeper nodes are labelled with posterior probability values.

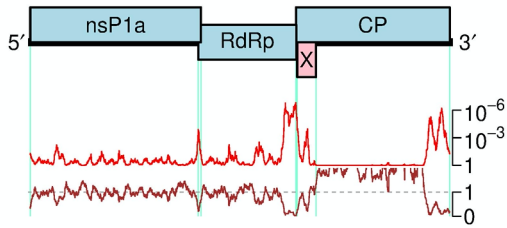
**a** Z25771, human astrovirus 1, G-Ia



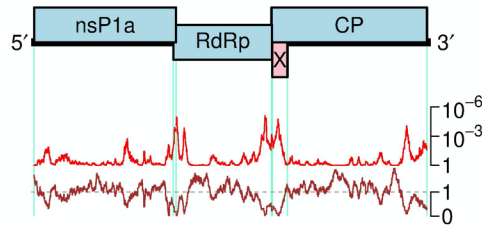
**b** KP404149, canine astrovirus, G-Ib



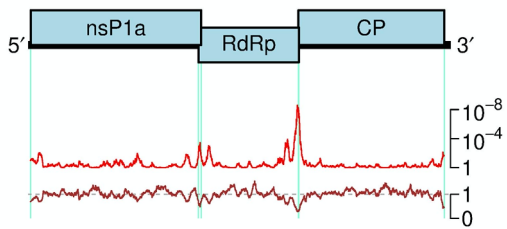
**c** JN420356, California sea lion astrovirus 9, G-Ic



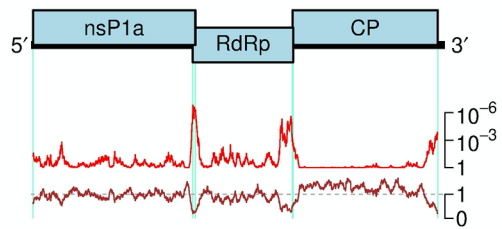
**d** KT946734, rodent astrovirus



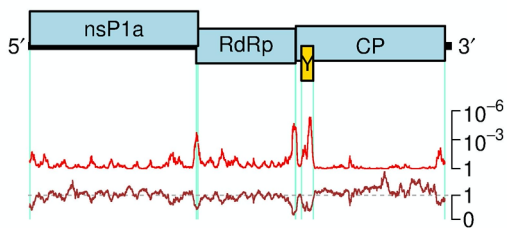
**e** FJ973620, astrovirus VA1, G-IIa



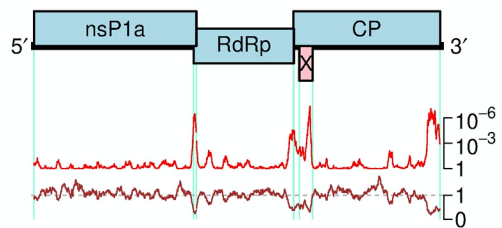
**f** JX556691, porcine astrovirus 3, G-IIb



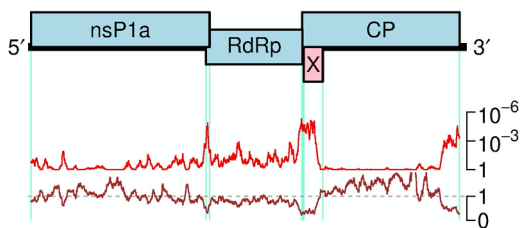
**g** AY179509, mamastrovirus 10, G-IIc



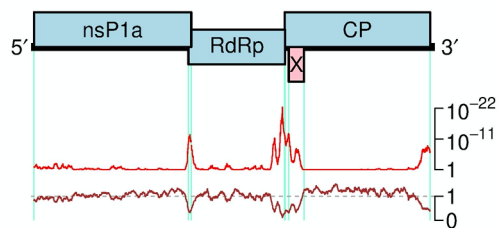
**h** Y15937, mamastrovirus 13, G-IId

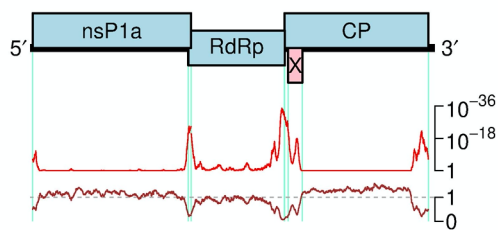
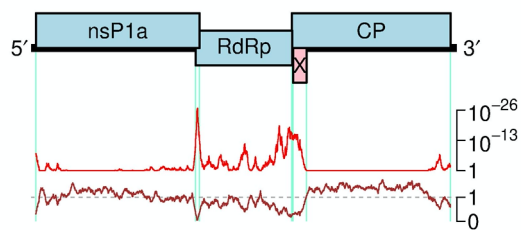
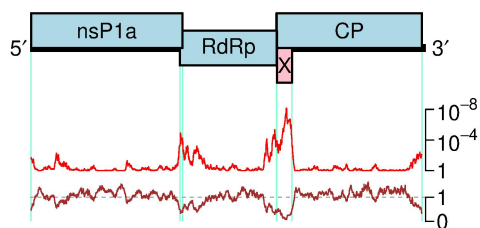
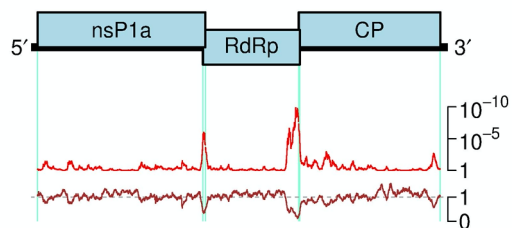


**i** JX544743, murine astrovirus, G-IIIa

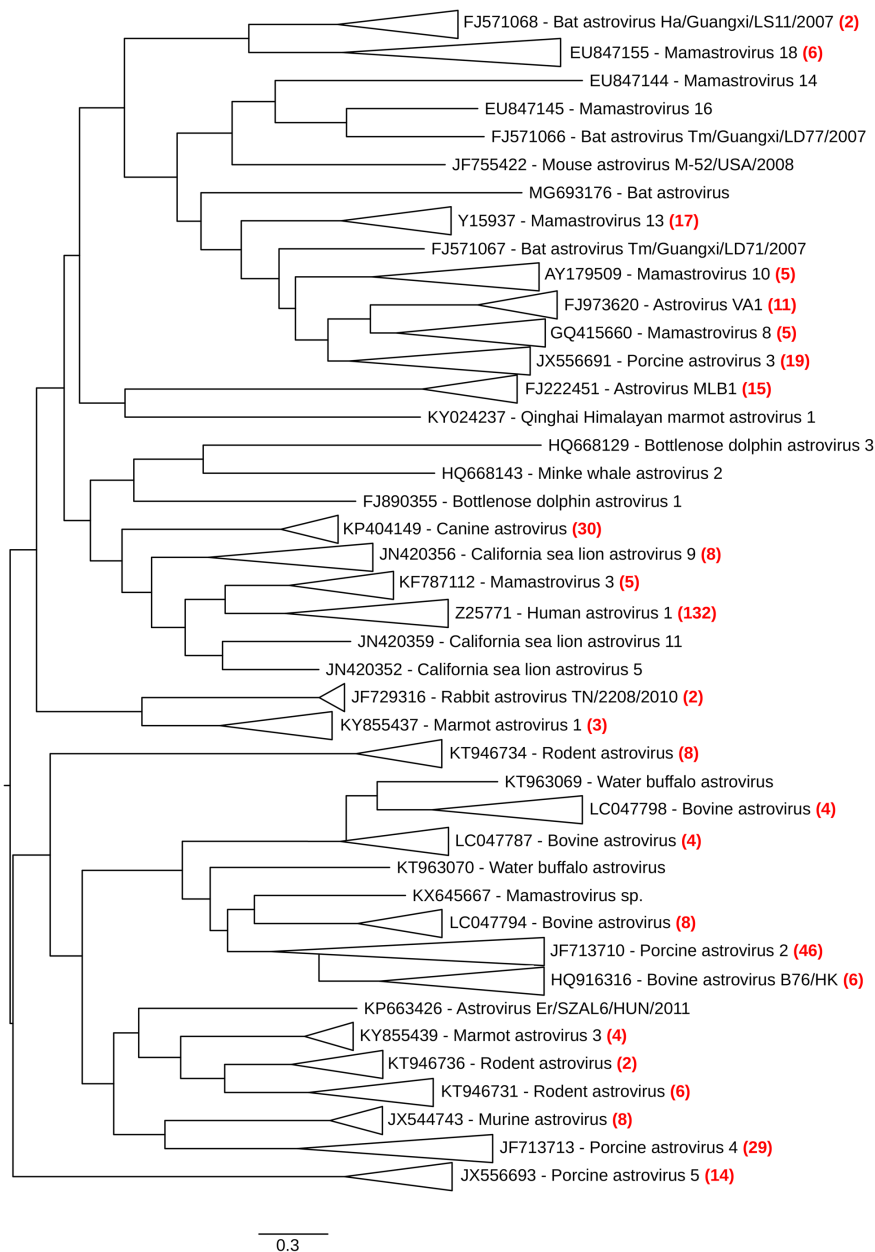


**j** HQ916313, bovine astrovirus B18/HK, G-IIIb



**k** JF713710, porcine astrovirus 2, G-IIIc**l** JF713713, porcine astrovirus 4, G-IIIId**m** FJ222451, astrovirus MLB1, G-IV**n** JX556693, porcine astrovirus 5, G-VI

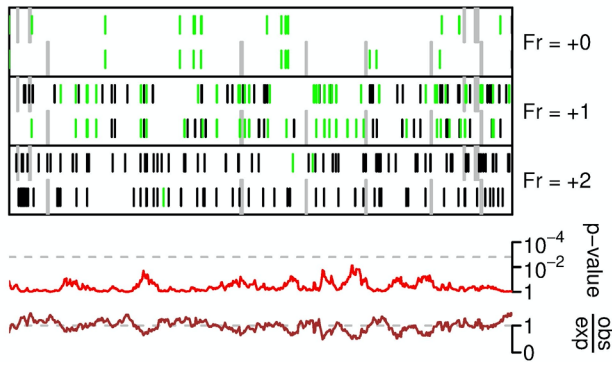
**Supplementary Figure 3 | Synonymous site conservation analysis of astroviruses.** In each subfigure, a genome map is shown at top, indicating the nsP1a, RdRp and CP ORFs (blue) and the putative additional ORF where present (pink – ORFX; yellow – ORFY). Below, is shown the analysis of conservation at synonymous sites in the nsP1a, RdRp and CP ORFs. The red line shows the probability that the observed conservation could occur under a null model of neutral evolution at synonymous sites, whereas the brown line depicts the ratio of the observed number of substitutions to the number expected under the null model. Peaks in synonymous site conservation may indicate functionally important overlapping elements such as the  $-1$  PRF signal between the nsP1a and RdRp ORFs, sgRNA promoter sequences, and overlapping coding sequences (i.e. the putative X and Y ORFs). Each synonymous site conservation analysis is based on an alignment of virus sequences in the indicated clade (Ia, Ib, Ic, etc) using the genome coordinate system of the indicated reference sequence / isolate name. The subgroup designations are defined in Fig. 1b and the sequences used in each alignment are as shown in Supplementary Fig. 9.



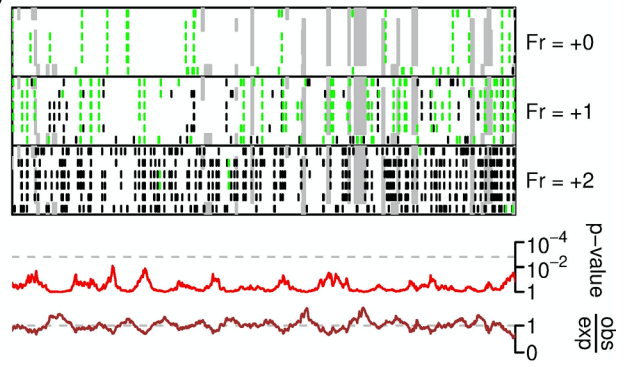
**Supplementary Figure 4 | Phylogenetic tree of mammalian astrovirus CP sequences.** Mammalian astrovirus sequences with complete or nearly complete coverage of ORF2 were obtained from NCBI on 26 July 2018, and the ORF2 amino acid sequences were extracted and aligned with MUSCLE<sup>1</sup>. A maximum likelihood phylogenetic tree was estimated using the Bayesian Markov chain Monte Carlo method implemented in MrBayes version 3.2.3<sup>2</sup>, sampling across the default set of fixed amino acid rate matrices, with five million generations, discarding the first 25% as burn-in. The tree was midpoint rooted and visualized with FigTree (<http://tree.bio.ed.ac.uk/software/figtree/>). Related groups of sequences (indicated by isosceles triangles) have been replaced in the figure by a single representative accession number and virus name; the total number of sequences in each group is shown in red. The 26 groups and 16 singletons correspond to those used in Supplementary Fig. 5 and Supplementary Fig. 7, respectively. The complete list of 415 sequences is shown in Supplementary Table 2.



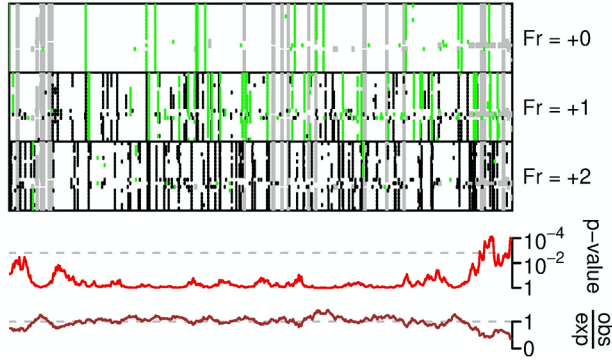
**a** FJ571068 – Bat astrovirus



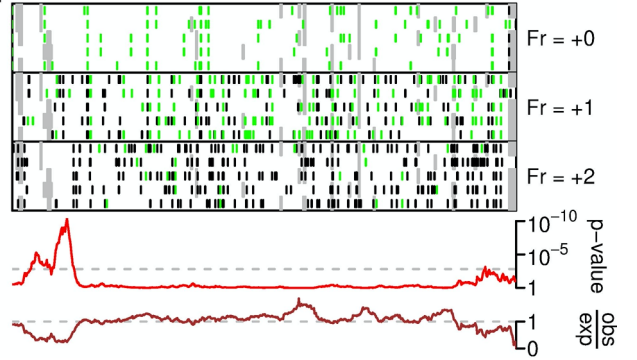
**b** no ORFX EU847155 – Mamastrovirus 18



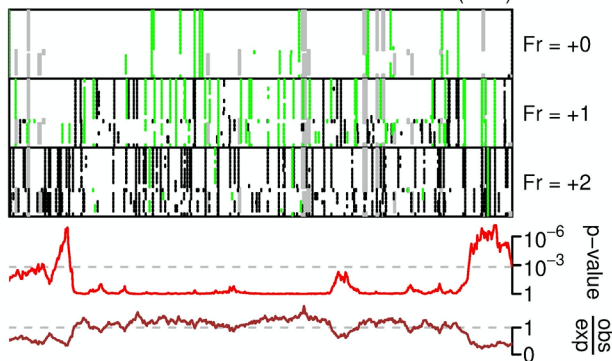
**c** no ORFX JX556691 – Porcine astrovirus 3 (G-II)



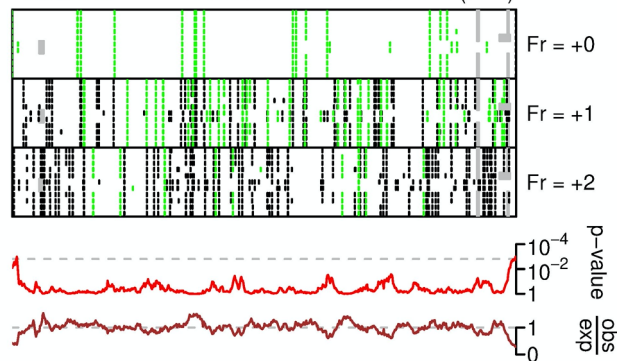
**d** ORFY AY179509 – Mamastrovirus 10 (G-II)



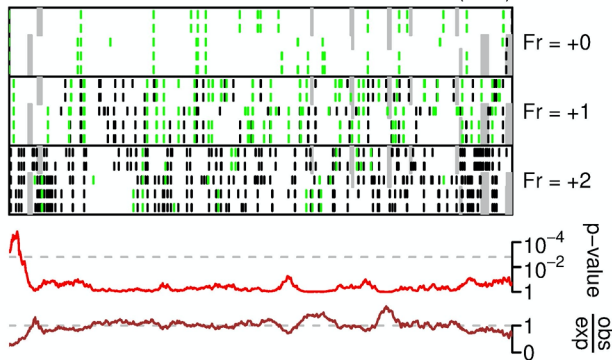
**e** ORFX Y15937 – Mamastrovirus 13 (G-II)



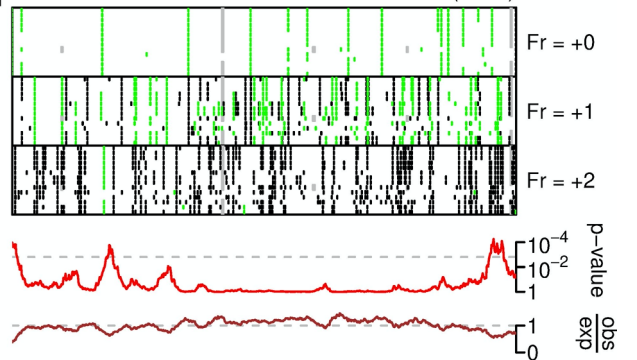
**f** no ORFX FJ973620 – Astrovirus VA1 (G-II)



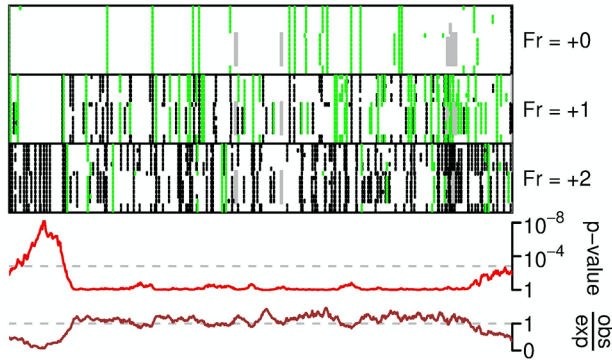
**g** no ORFX GQ415660 – Mamastrovirus 8 (G-II)



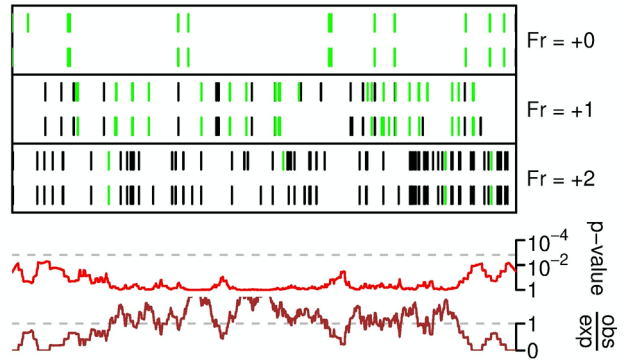
**h** no ORFX JX556693 – Porcine astrovirus 5 (G-VI)



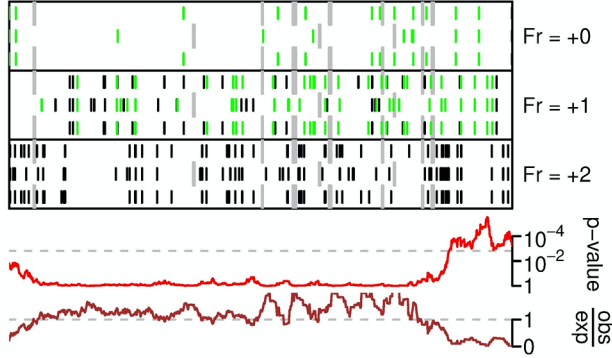
**i** ORFX FJ222451 – Astrovirus MLB1 (G–IV)



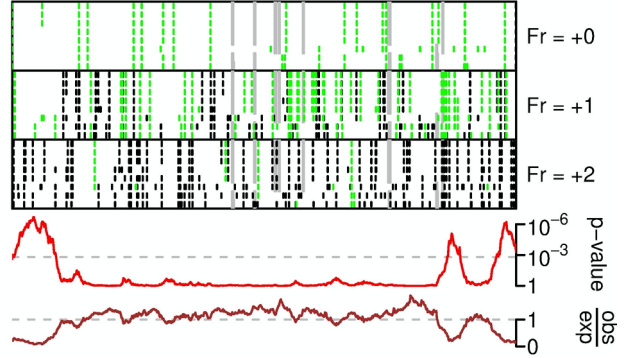
**j** no ORFX JF729316 – Rabbit astrovirus



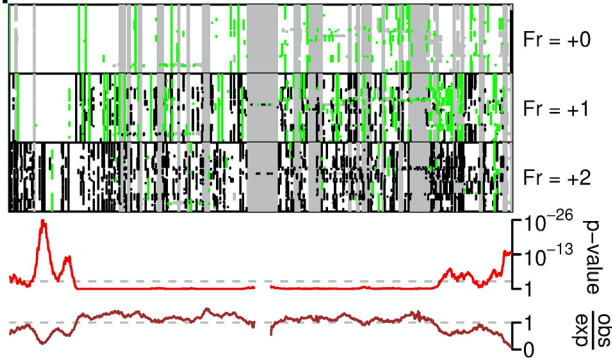
**k** no ORFX KY855437 – Marmot astrovirus 1



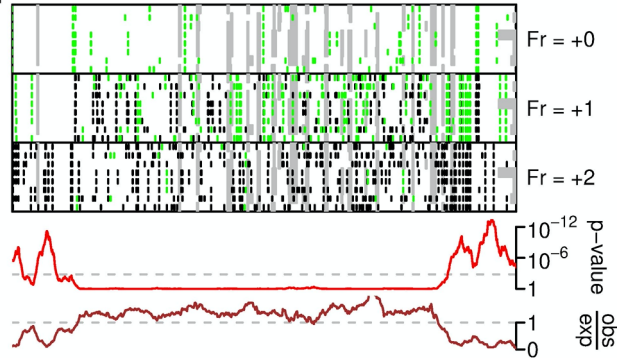
**l** ORFX KT946734 – Rodent astrovirus



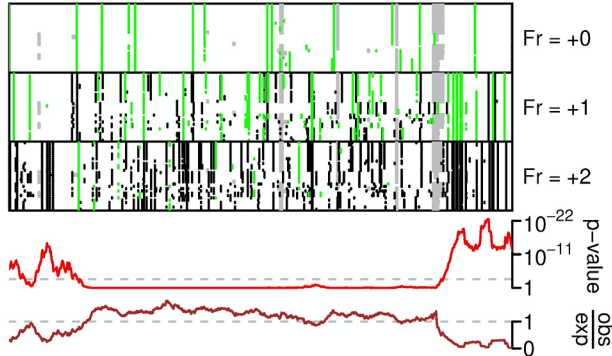
**m** ORFX Z25771 – Human astrovirus 1 (G–I)



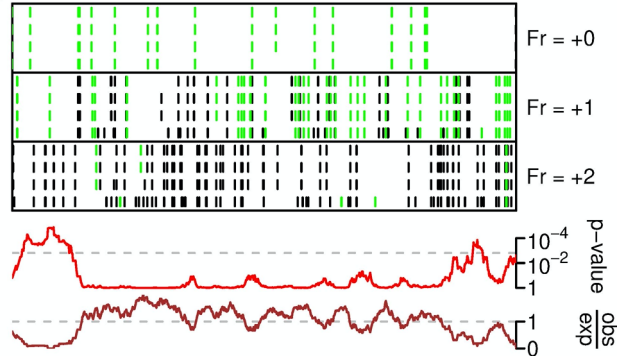
**n** ORFX JN420356 – California sea lion AstV 9 (G–I)



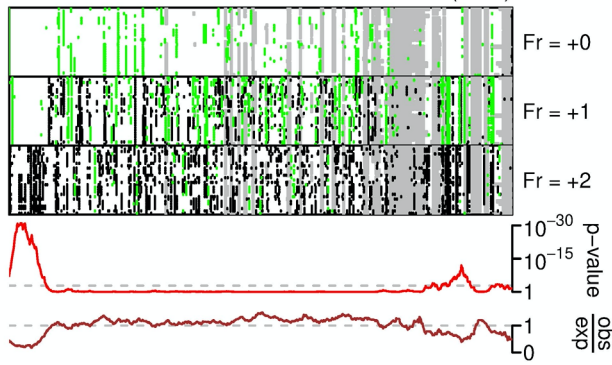
**o** ORFX KP404149 – Canine astrovirus (G–I)



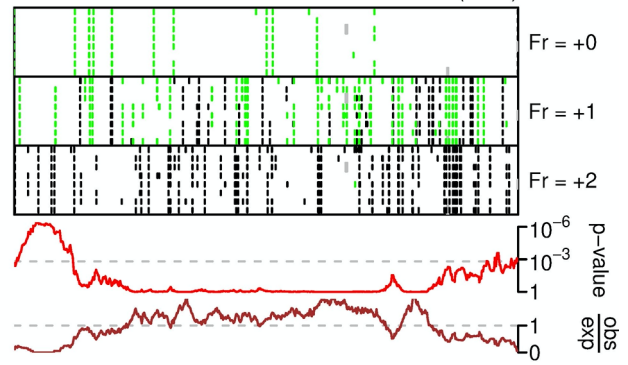
**p** ORFX KY855439 – Marmot astrovirus 3 (G–III)



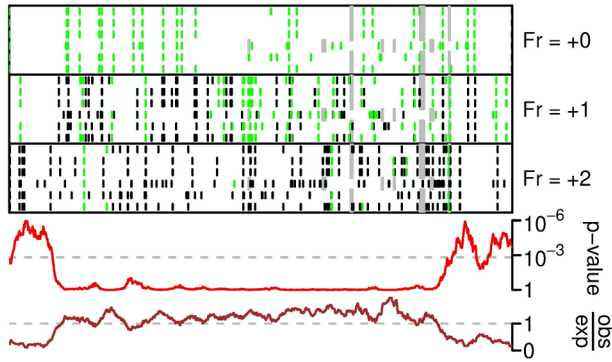
**q** ORFX JF713713 – Porcine astrovirus 4 (G-III)



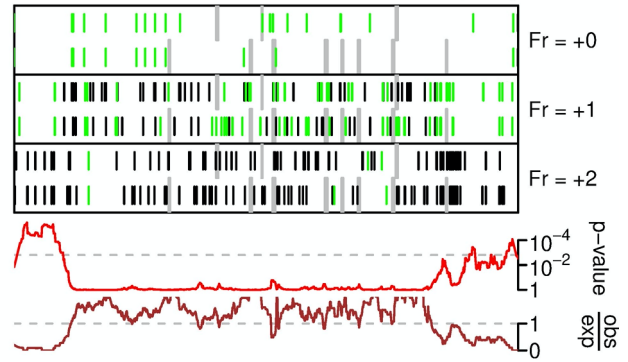
**r** ORFX JX544743 – Murine astrovirus (G-III)



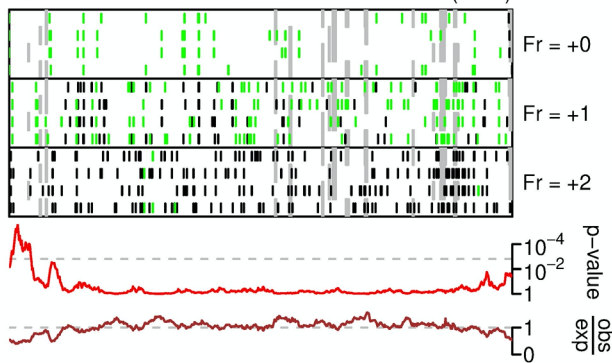
**s** ORFX KT946731 – Rodent astrovirus (G-III)



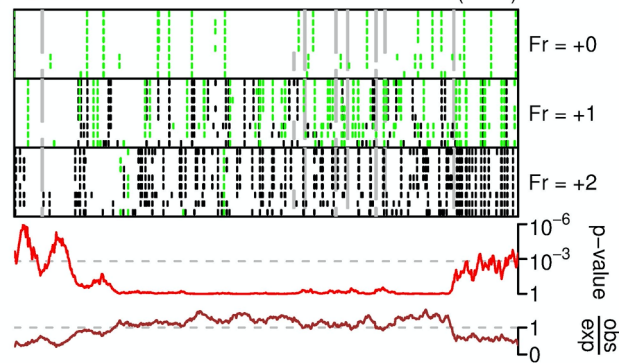
**t** ORFX KT946736 – Rodent astrovirus (G-III)



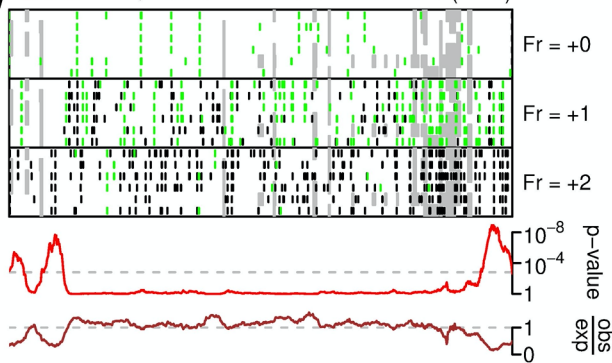
**u** ORFX LC047798 – Bovine astrovirus (G-III)



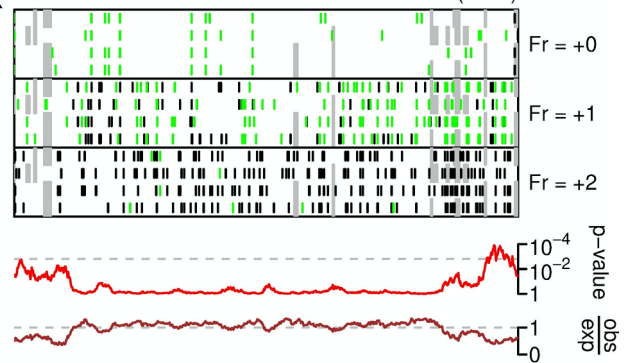
**v** ORFX LC047794 – Bovine astrovirus (G-III)

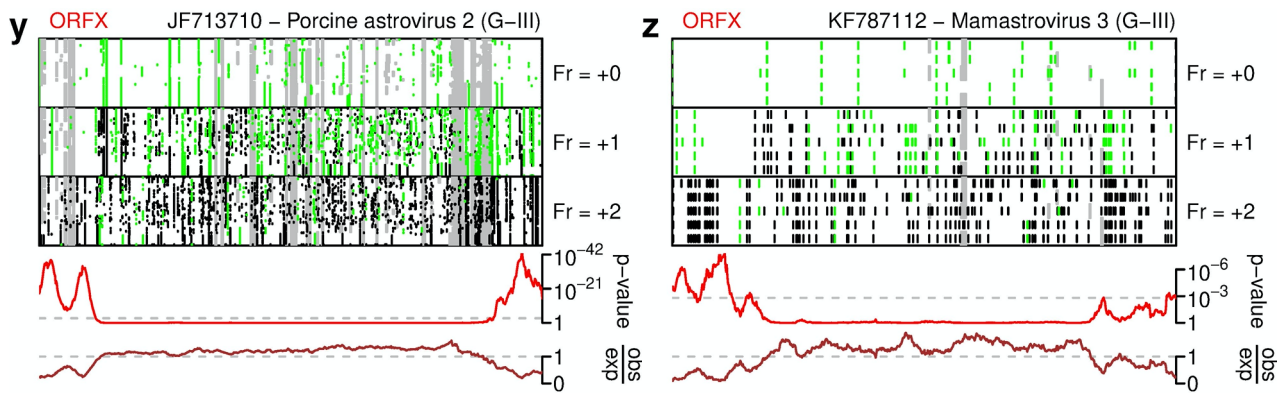


**w** ORFX HQ916316 – Bovine astrovirus B76 (G-III)

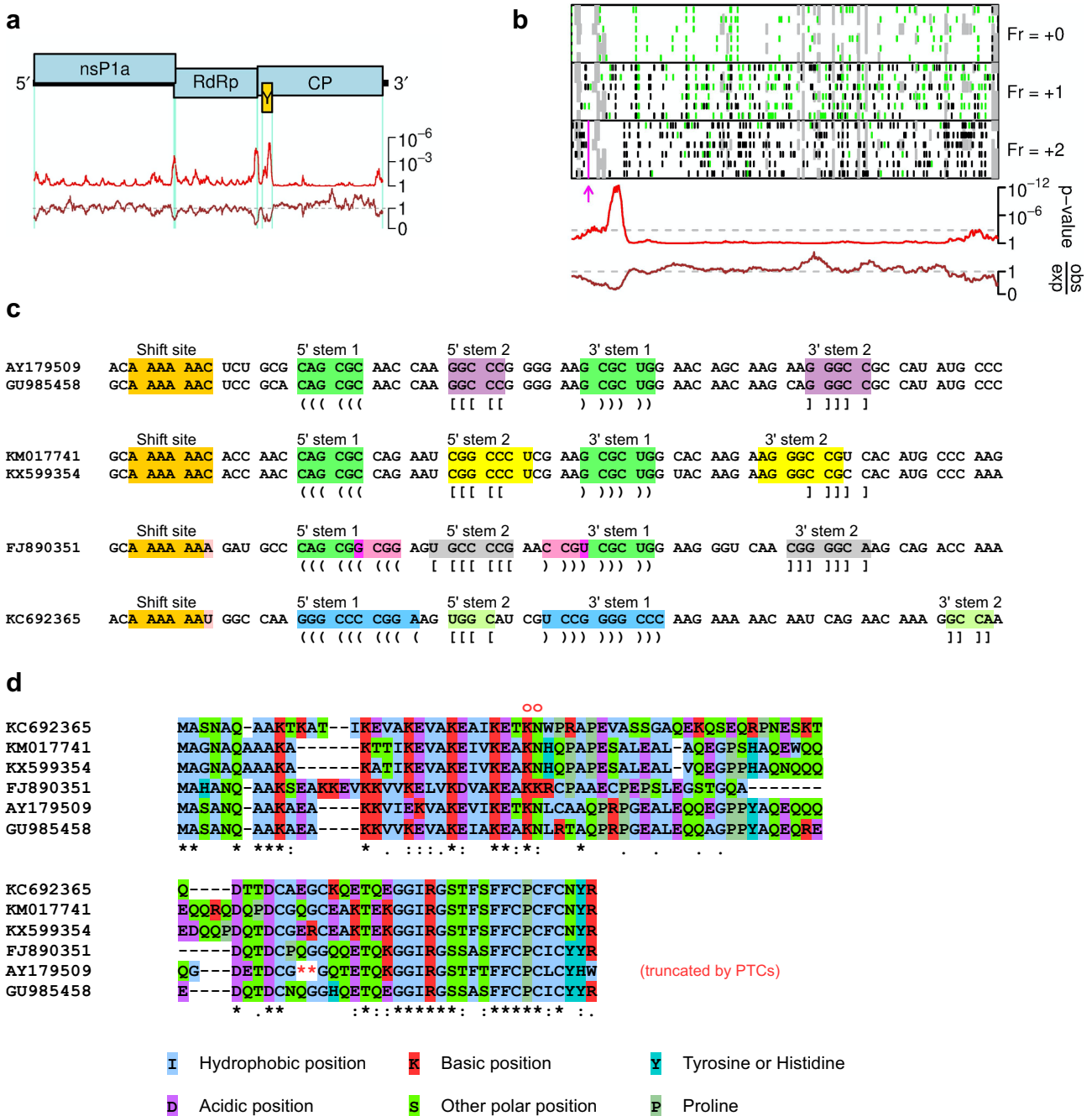


**x** ORFX LC047787 – Bovine astrovirus (G-III)





**Supplementary Figure 5 | Comparative genomic analysis of CP ORF alignments.** Sequences with coverage of the capsid coding region of mammalian astroviruses were obtained from NCBI, CP ORF sequences were extracted, and clustered into 26 multi-sequence and 16 singleton groups based on CP amino acid identity (see Methods; Supplementary Fig. 4). For each multi-sequence group, duplicate sequences were removed and remaining sequences were aligned with MUSCLE<sup>1</sup> and analysed with synplot2<sup>4</sup>. The header of each plot indicates one sequence (accession number and virus name) from the group, genogroup (G-I, G-II etc; where defined), and whether the group is predicted to contain ORFX, ORFY or neither. The upper three panels show the positions of alignment gaps (grey), stop codons (black) and AUG codons (green) in each of the three reading frames in each sequence in the alignment. Below, is shown the analysis of conservation at synonymous sites. The red line shows the probability that the observed conservation could occur under a null model of neutral evolution at synonymous sites, whereas the brown line depicts the ratio of the observed number of substitutions to the number expected under the null model. Peaks in synonymous site conservation may indicate functionally important overlapping elements such as overlapping coding sequences (i.e. the putative X and Y ORFs) besides regulatory elements (e.g. functional RNA structures). See Supplementary Fig. 7 for the singleton groups.



**Supplementary Figure 6 | A putative -1 PRF overlapping ORF in subgroup IIc astroviruses.** (a) Comparative genomic analysis of subgroup IIb full-length sequences AY179509, GU985458, KC692365 and KM017741. Top – genome map indicating the nsP1a, RdRp and CP ORFs (blue) and the putative additional Y ORF (yellow). Bottom – analysis of conservation at synonymous sites in the nsP1a, RdRp and CP ORFs. The red line shows the probability that the observed conservation could occur under a null model of neutral evolution at synonymous sites, whereas the brown line depicts the ratio of the observed number of substitutions to the number expected under the null model. Peaks in synonymous site conservation may indicate functionally important overlapping elements such as the -1 ribosomal frameshifting signal between the nsP1a and RdRp ORFs, sgRNA promoter sequences, and overlapping coding sequences (i.e. the putative Y ORF). (b) Synonymous site conservation plot for the CP ORF for AY179509, GU985458, KC692365, KM017741 and two additional partial-genome sequences with CP ORF coverage, FJ890351 and KX599354 (lower two panels). In the upper three panels, alignment gaps (grey), stop codons (black) and AUG codons (green) are indicated in each of the three reading frames. The location of the putative A<sub>-1</sub>AAA<sub>-1</sub>AAZ<sub>-1</sub> ribosomal frameshifting site is indicated in pink. (c) Predicted -1 PRF shift sites and 3'-adjacent

RNA secondary structures in each of the six sequences. Predicted base-pairings are indicated with matching highlights and “()”s (stem 1) or “[ ]”s (stem 2). **(d)** Alignment of the predicted -1 PRF product. The location of the A\_AAA\_AAN shift site is indicated with two red circles. The upstream sequence is encoded by the 5' end of the CP ORF whereas the downstream sequence is encoded by the overlapping -1 frame Y ORF. Amino acids are colour-coded according to their physicochemical properties. In AY179509, the Y ORF is truncated by two tandem premature termination codons (red asterisks).

**EU847145 - Mamastrovirus 16**

MRLEINLTKRISTRVSKSMLGRRLRRKRGRMVMVDLNLNLLMLLLLLLGLLEAVMSLI  
????????????????????

**FJ890355 - Bottlenose dolphin astrovirus 1**

MLALRLKPPARREARVVFVQGLEEEHPLSKSOLIPKOKDLPEDQVDALLEIKIIVSNKLEINSRNKVSQGPQRLSRPRLLALLDQILAMMQRGRFPSI  
????????????????

**HQ668129 - Bottlenose dolphin astrovirus 3**

MAAKSQLKSSPEPELRIVKAITRELVEEKEVIRTIKRMLANNQEIITRLGVSEQRVHVQLTAWAYADPNQIKSSSLQCLVPSVETHQEGSKWRRWFSLTFCSLRKSGQITT  
LDHCK

**HQ668143 - Minke whale astrovirus 2**

MPHNLKRNASVSGVTQEELELLEILTAILESHDEVIVDVINRMEGRLELRLNAWSEGLSQSHRSLRPRWAPLVPMRRLFLRLKGFFTSTPHSLRRSPGVWLDLRFPP

**JN420352 - California sea lion astrovirus 5**

MAATGANLVPAFRVAGNQMSRSQSIQNPEEQVELDAVNVNLIITVSVNLSNNSLTSVLVQDSQPSDKKPLLRLLALSTLIRLIRRSLLR  
????????????????

**JN420359 - California sea lion astrovirus 11**

MKNPTGAILAAAHNPEATRQSRSPSIQRSQEVETDAVNVNLSVSVQLSINNSGNRVSDQKQKQNSGQQQHSEPLEPTPLGSQSLRRASFVTRFSLRTVQVLLAQFRC

**KT963069 - Water buffalo astrovirus**

MAMGMPRAWFRPLRVNRRRTKVDVGDGARLRSMSTSQTRQDHQEDQDNNNLRDNNVNFQCGATAALRAVKLRFSTIRYARR  
????????????????

**KT963070 - Water buffalo astrovirus**

MDQHPQEQVDDVDGVEPNPENKLLCCPLRKFRRVSGPAVRASITMWFGRSPPPLEQLEPTPTGK

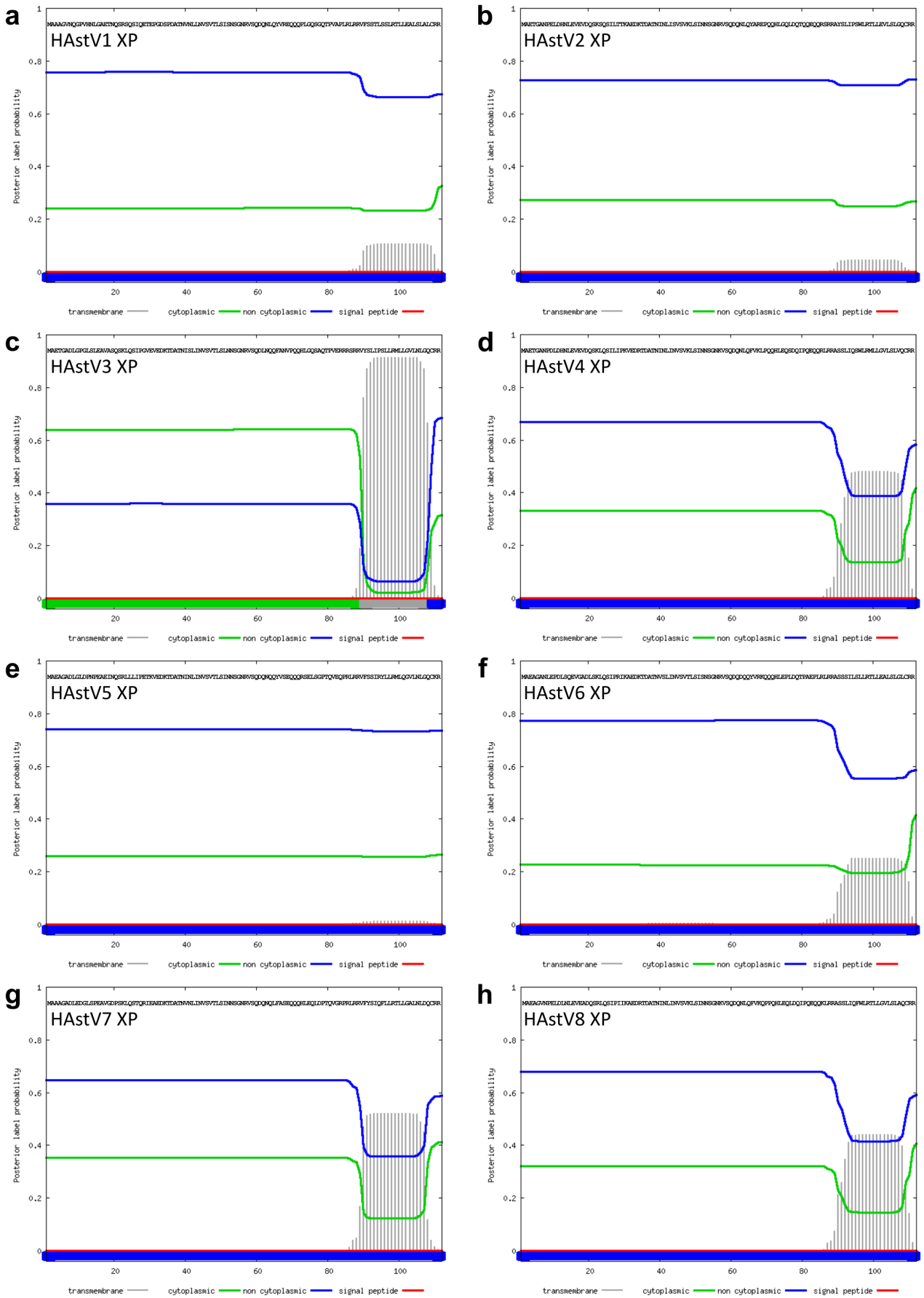
**KX645667 - Mamastrovirus sp.**

MEDQHNPPNPSRDETDGRDIMRRLRLLYALYKELKNVADQILQEDVVEGGLYCRKLOORSEQLDQMLQKQSKTSSQSSSIPPP

**MG693176 - Bat astrovirus**

MPSLRTRRLRLRSRLNLIISNILRNTNRNRPRARVLEQSLRRLRRLRRSVLRGRNRGQSVSRPLSASLTGPPRRARCWLLVQIFILLPRNLKVALLSALWLRRRPSLANGA  
FLGLLSVLPWLALLLLALLSLAFFPSIQQAGPFO

**Supplementary Figure 7 | Potential XP sequences in unclustered astroviruses.** Sequences with coverage of the capsid coding region of mammalian astroviruses were obtained from NCBI, CP ORF sequences were extracted, and clustered into 26 multi-sequence and 16 singleton groups based on CP amino acid identity (see Methods). Potential XP peptides encoded within 10 of the 16 unclustered sequences are shown. Amino acids are colour-coded according to their physicochemical properties. Transmembrane regions predicted by Phobius<sup>5</sup> are indicated with pink bars. Potential transmembrane regions (hydrophobic regions scored below threshold by Phobius) are indicated with pink question marks. See Supplementary Fig. 5 for the multi-sequence groups.



**Supplementary Figure 8 | Transmembrane domain predictions for HAstV XPs.** Transmembrane domains for representative sequences (see Fig. 1f) were predicted with Phobius<sup>5</sup>. An above-threshold TM was predicted only for HAstV3.





Genogroup Ic - California sea lion astrovirus etc

Genetic sequence for Genogroup Ic, California sea lion astrovirus etc. Includes accession numbers (e.g., JN420351) and amino acid sequences with color-coded residues.

Genogroup Ib - canine astrovirus etc

Genetic sequence for Genogroup Ib, canine astrovirus etc. Includes accession numbers (e.g., KX599351) and amino acid sequences with color-coded residues.

Genogroup III - marmot astrovirus

Genetic sequence for Genogroup III, marmot astrovirus. Includes accession numbers (e.g., KY855439) and amino acid sequences with color-coded residues.

Genogroup IIIc - porcine astrovirus 4 etc

Genetic sequence for Genogroup IIIc, porcine astrovirus 4 etc. Includes accession numbers (e.g., KY940075) and amino acid sequences with color-coded residues.

Genogroup IIIa - murine astrovirus etc

Genetic sequence for Genogroup IIIa, murine astrovirus etc. Includes accession numbers (e.g., MF175075) and amino acid sequences with color-coded residues.

\* KT946731 lacks the ORFX AUG codon - possibly a defective sequence

Genogroup III - Rodent astrovirus

Genetic sequence for Genogroup III, Rodent astrovirus. Includes accession numbers (e.g., KT946735) and amino acid sequences with color-coded residues.

Genogroup III - Bovine astrovirus

Genetic sequence for Genogroup III, Bovine astrovirus. Includes accession numbers (e.g., LC047800) and amino acid sequences with color-coded residues.

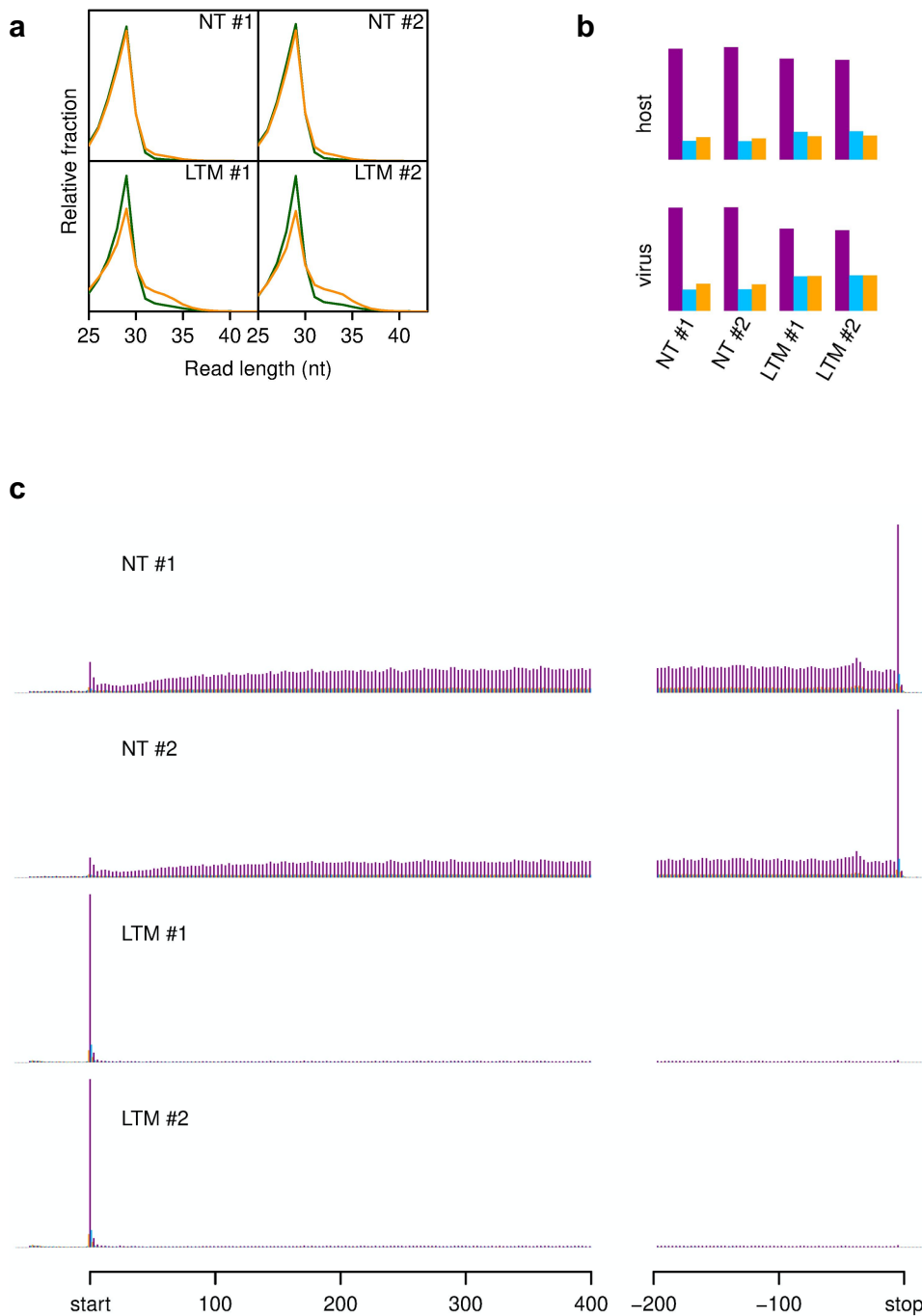
Genogroup IIIb - bovine astrovirus etc

LC047799 MDQLLHKVQEIFSSSSRG-RDEPDGGDSKIK---LGCLPFRLRLRVPGV-VSVR--VRALSTRRLSOHLAQWARIIRRC-----  
 LC047789 MDQLLHKVQEIFNSSSRG-RDEPDGGDSKIK---LGCLPFRLRLRVPGV-VSVR--VRALSTRRLSOHLAQWARIIRRC-----  
 LC047794 MDQLLHKVQEIFNSSSRG-RDEPDGGDSKIK---LGCLPFRLRLRVPGV-VSVR--VRALSTRRLSOHLAQWARIIRRC-----  
 LC047795 MDQLLHKVQQAENSSSRG-KDEPDGGDSKIR---LGCLPFRLRLRVPGV-VSVR--VRALSTRRLSOHLAQWARIIRRC-----  
 LC047801 MDQLPHVQQAENSSSRG-KDEPDGGDSKIR---LGCLPFRLRLRVPGV-VSVR--VRALSTRRLSOHLAQWARIIRRC-----  
 LC047788 MDQLPQVGRPNSSSNRE-GDEPDGGDSRIR---LGCLPFRLRLRVPGV-VSVR--VRALSTRRLSOHLAQWARIIRRC-----  
 MDQPPHQVGRPNSSSRREGEDELDDGGKLC---LGCHPFRLRLRVPGV-ASVR--IRELSTSGSSRRLGRLAQIILNKLIN---  
 LC047792 MDQLPHQAGQPNSSSSREGEDELDDGGKLC---LGCLPFRLRLRVPGV-ASVR--IRELSTSGLSQRSERLAQIILNKLIN---  
 HQ916314 MDQLPPKVEAASQDSNN---SSVVDGAEQDHRPRS CGCFPTKLRVA-GGG-LGFHGSITVWWSRRLSLSVRSAPMAQAQ-----  
 HQ916316 MDQLPPKVEAASQDSNN---SSAVDGAQCGRPRWCGCYPSKIRVA-GGG-FGLRGSVTVWFRRLSLSAR-----  
 MG660832 MEQLPTRQELSEDG---VDGGTGETAKRRRSTCGCYPTKLRVS-AGI-LAARASVTVWFSRRSTQPSVOLDPTEVNRNLNASLRA  
 KM822593 MDQLPHVQQAENSSSRG-RDEPDGGDSKIR---LGCLPFRLRLRVPGV-ASVR--IRELSTSELSQLRGLARIQTK-----  
 LC047797 MDQLPPKAEGLKLVCH---ADAVDGCKCRKCH-RGCLPIG-RIRLSGAFLAARMSITVVFIRRSRRSGP-----  
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 KJ620979 MEQLPTRQELSEDG---VDGGTRGTGRRRSTCGCYPTKLRVS-AGV-LAARASVTVWFSRRSTQPSVOLDPTEVNRNLNASRA  
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 LC047787 MEQLPQKVEGSDSSKIKH-ADDVDGCKCRKCH-RGCLPVR-RIRLAGVFLAARMSITVVFIRRSRRSGP-----  
 LC047796 MEQLPTRQELLEDS---ADGGAGGITRPRRSTCGCYPTKLRFS-AGF-LAARASVTVWFSRRSIQPSVOLDPTEVNRNLNASLRA

Genogroup IIIc - porcine astrovirus 2 etc

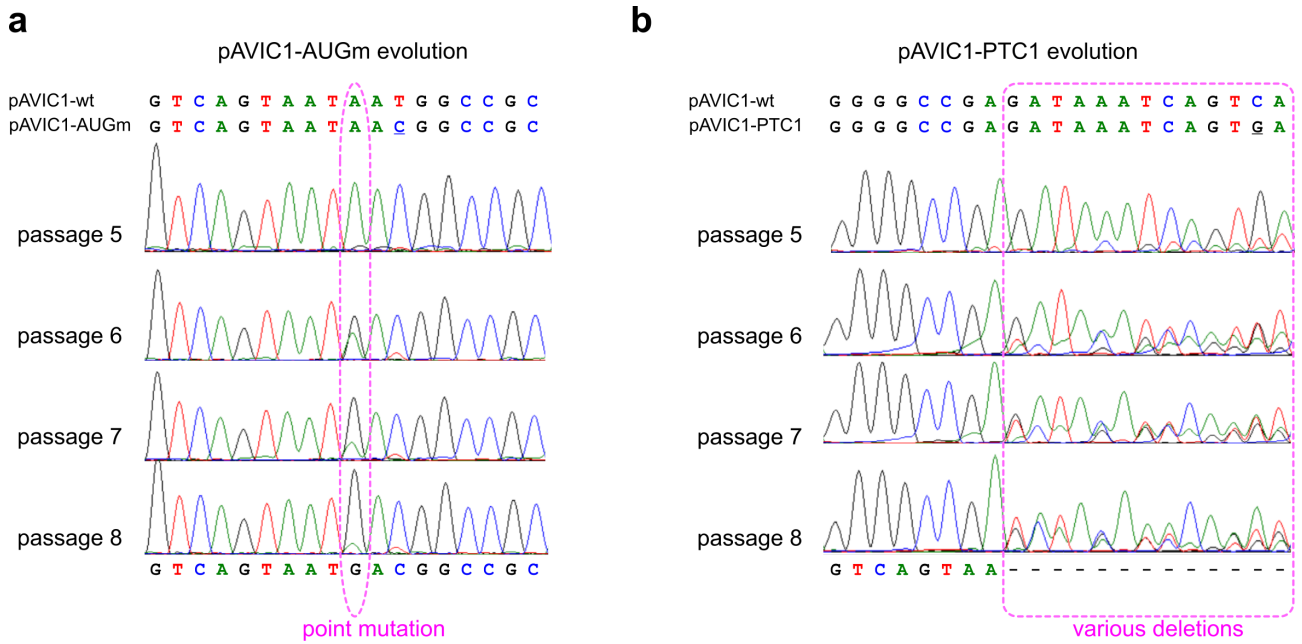
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 KY940077 --MAPARIRRALPELNQEGEET-GENH---RLTSGCLPIKIRLSAGVLAARVSVVWFSRRSRRLPAQSDQMVVSKSSVK-----  
 KP982872 --MAPARIRRALPEPNRQEGEET-GENH---RPWCGCYPTKIRVAGAVLGLRASVTVWLSRRLSRLSEQLDPTARDRSRQSSLS CSTPAQ  
 LT898434 --MAPARIRRALPEHNRQEGEET-GENH---RSTSGCWVFKIRLSAGVLAARVSVVWFSRRSRQPSAQSDQMVVTRSSVR-----  
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 KP759770 --MAPPRIRRALPEQIQEGEET-GENHN---RPWCGCYPTKRVVAGAILGLFRASVTVWLSRRLSLSQSDQEMGVGOSRNSLSSTPAQ  
 KY214438 --MAPPRIRRALPEIQEGEET-EGNHN---RPLCGCYPTKRVVAGAVLGLRASVTVWLSRRLSRLSARSGRMCAQ-----  
 KR868724 --MAPPOIRRALPROSRQEGEV-EGNH---RLTSGCCPIKVRLSASFLAARASVTVWFSRRSIRPLEQLDQMAVNR-----  
 KR868723 --MAPPRIRRALPRORQEGEET-GENH---RSTSGCCPVKIRLSAGVLAARVSVVWFSRRSIRPLGQLDPMVAVNKS SVR  
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 KR868721 --MAPPRIRRALPRORQEGEET-EGNH---RLTSGCCPIKIRLSARFLAARVSVVWFSRRSIRPLGQLGPMVAVSKSSVR-----  
 LC201593 --MAPPRIRRALPEPNRQEGEET-REDHS---OSTCGCLEPTKRVVAGAVLGLFRASVTVWVHRLSRLSRSRVE TDPGT  
 KY940076 --MAPARIRRALPEPNRQEGEETQGDRE---GSTCGCFPIKVRVAGAVLGLRASVTVWFSRRSIRPLAQLGRMRAERLSVN-----  
 LC201592 --MAPPRIRRALPALGRDAGGCT-GENNNORSTCGCYPTKIRVAGAVLGLRASVTVWWSRRLSRLSRLGPTGLEK-----  
 LC201587 --MAPARIRRAHPGPSRQEGEV-GENHS---RPWCGCYPTKIRVAGAILGLRASVTVWLSRRLSRPSEQSGFMEDRSRONSLS CSIPVO  
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 LC201585 MEQLPPR-RGRILRQQA-EGGTEQGDSP---RSMCGCYPLKLRVAAGVFLGARVSVVWFSRRSIRRSQAQARMGVNKK-----  
 LC201586 MEQLPTR-RGRILRQATEGEGIEQGDRE---RSTCGCFVVKIRVAGAVLGLRASVTVWFSRRSIRLLAQSDQEMGVNKK-----  
 KJ495986 MEQLPPR-RGRILRQQA-VGGEIEQGDNE---KSTCGCYPLKLRVAAGVFLGARVSVVWFSRRSIRRSQAQARMGVNKK-----  
 JF713710 MEQLPPR-RGRILRQQA-VGGEIEQGDNE---RSTCGCYPLKLRVAAGVFLGARVSVVWFSRRSIRRSQAQARMGVNKK-----  
 JF713712 MEQLPPR-RGRILRQQA-VGGEIEQGDNE---NSTCGCFVVKIRVAGAVLGLRASVTVWFSRRSIRRSQAQARMGVNKK-----  
 JX556690 MEQLPPR-RGRILRQQA-VGGEIEQGDNE---RSMCGCYPLKLRVAAGVFLGARVSVVWFSRRSIRRSQAQARMGVNKK-----

**Supplementary Figure 9 | Sequences of putative XP proteins encoded in diverse astroviruses.** Amino acid alignments of putative XP proteins in different astrovirus groups. Sequences were aligned with MUSCLE<sup>1</sup>. Sequences are listed in the same order as in the ORF1b phylogenetic tree (Supplementary Fig. 1). Amino acids are colour-coded according to their physicochemical properties. Transmembrane regions predicted by Phobius<sup>5</sup> are indicated with pink bars above the alignment. Potential transmembrane regions (hydrophobic regions scored below threshold by Phobius) are indicated with pink question marks. Signal peptides predicted by Phobius are indicated with pink “s”s. In all cases, predictions are based on the first sequence in each alignment.

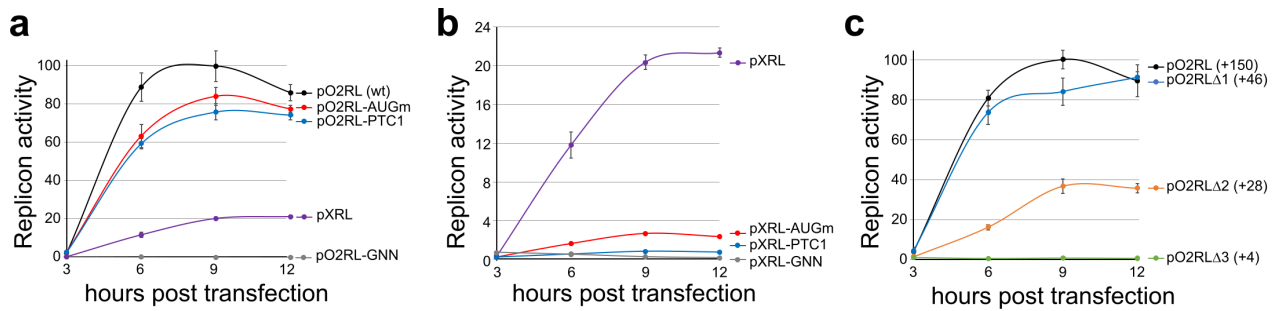


**Supplementary Figure 10 | Assessment of ribosome profiling quality.** Cells were harvested at 12 hpi and either flash frozen with no pre-treatment (NT), or pre-treated with lactimidomycin for 30 min followed by flash freezing (LTM). **(a)** Relative length distributions for Ribo-Seq reads mapping to virus (orange) and host (green) mRNA coding regions. **(b)** Phasing of 5' ends of RPFs ( $\geq 25$  nt) that map to the viral ORFs (excluding dual coding regions) or host mRNA coding regions. **(c)** Histograms of approximate P-site positions of RPFs ( $\geq 25$  nt) relative to annotated initiation and termination sites summed over all host mRNAs. See Supplementary Table 4 for read counts. Source data are provided as a Source Data file.





**Supplementary Figure 12 | Evolution of XP knockout viruses.** Sanger sequencing chromatograms of RT-PCR products obtained from passaged **(a)** pAVIC1-AUGm and **(b)** pAVIC1-PTC1. Evolution occurred via a pseudo-reversion (pAVIC1-AUGm) or 5- or 8-codon deletions (pAVIC1-PTC1). See Fig. 3c for individually sequenced cloned virus RT-PCR products.

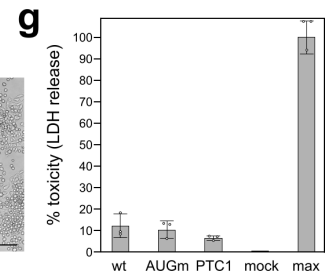
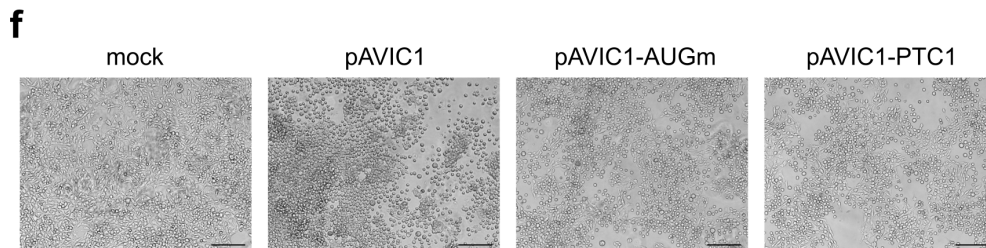


**d** Maximum replicon activity in BSR cells

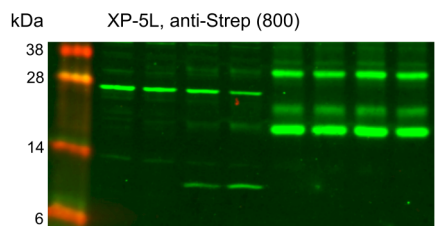
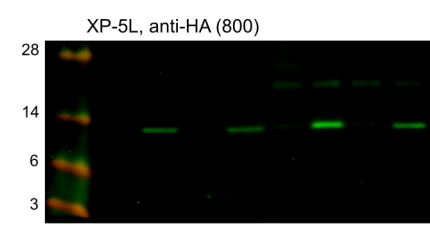
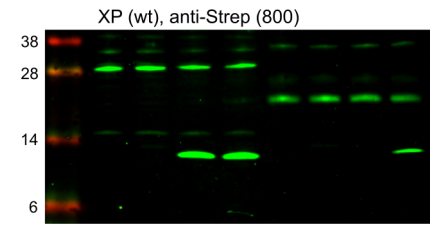
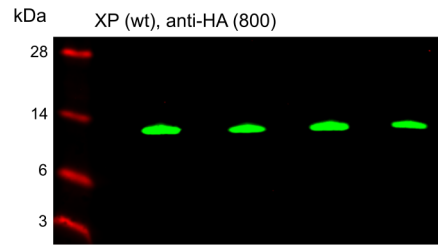
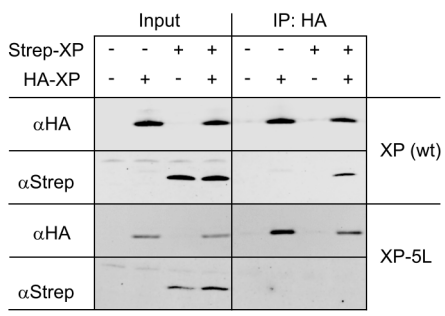
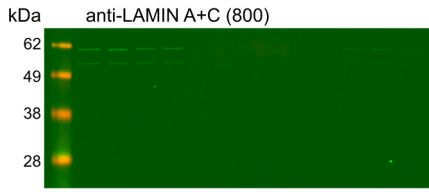
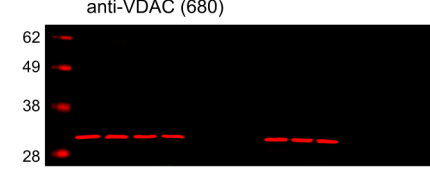
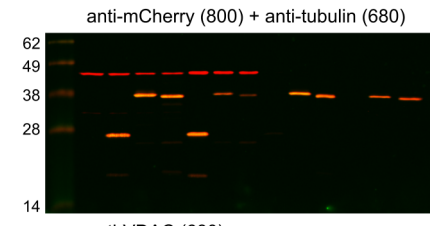
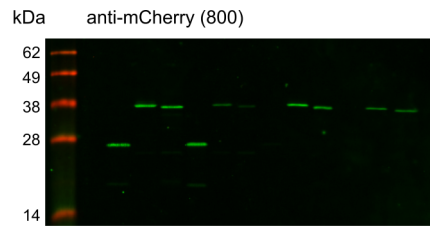
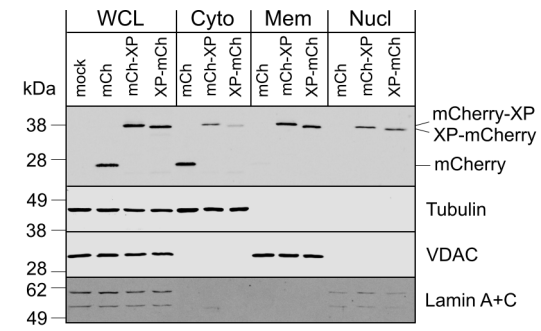
replicon	wt	GNN	AUGm	PTC1	$\Delta 1 (+46)$	$\Delta 2 (+28)$	$\Delta 3 (+4)$
pO2RL	100 ± 3.0%	0.048 ± 0.002%	92.6 ± 3.6%	85.4 ± 2.6%	92.2 ± 1.9%	12.5 ± 0.1%	0.645 ± 0.027%
pXRL	13.7 ± 0.2%	0.111 ± 0.001%	2.45 ± 0.01%	0.747 ± 0.027%			

**e** Maximum replicon activity in Huh7.5.1 cells

replicon	wt	GNN	AUGm	PTC1	$\Delta 1 (+46)$	$\Delta 2 (+28)$	$\Delta 3 (+4)$
pO2RL	100 ± 8.1%	0.108 ± 0.004%	84.2 ± 4.6%	76.1 ± 4.3%	91.1 ± 2.9%	36.6 ± 3.6%	0.620 ± 0.104%
pXRL	21.4 ± 0.5%	0.274 ± 0.007%	2.68 ± 0.02%	0.828 ± 0.049%			

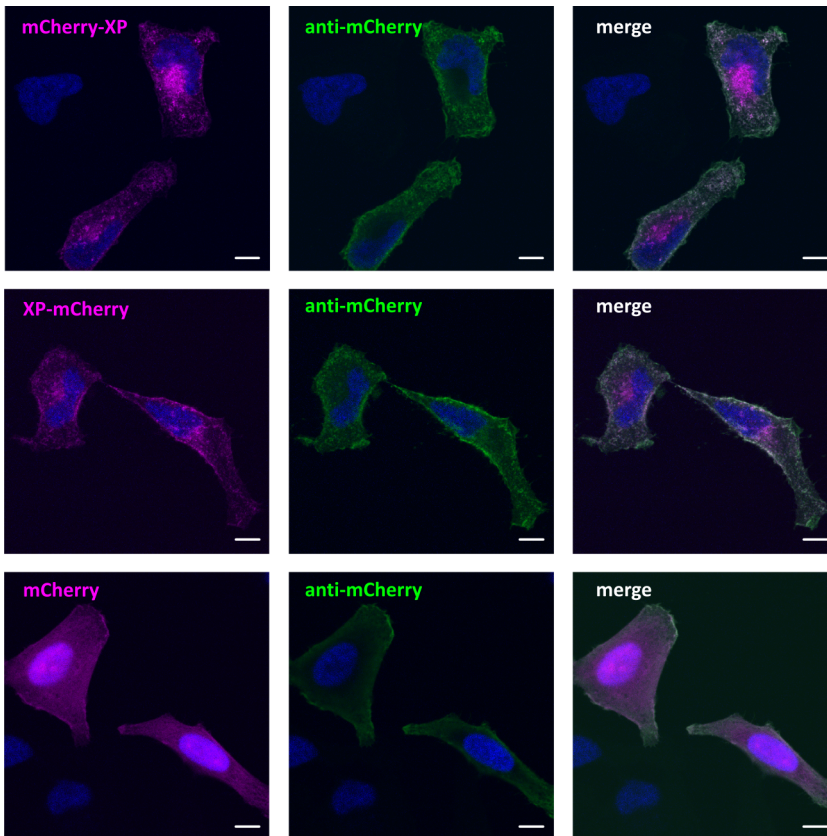


**Supplementary Figure 13 | Supporting data for Figure 4. (a-c)** Relative replicon luciferase activities representing translated product associated with activity of the subgenomic promoter, measured after RNA transfection of Huh7.5.1 cells (mean ± s.d.;  $n = 3$  biologically independent experiments). Values are normalized so that the maximum wt value for ORF2-frame translation is 100%. **(d)** Maximum replicon activities obtained in the assays presented in Fig. 4c, d, f (BSR cells). **(e)** Maximum replicon activities obtained in the assays presented in Supplementary Fig. 13a, b, c (Huh7.5.1 cells). **(f)** Representative bright field images at 48 hpe of BSR cells electroporated with mock, pAVIC1, pAVIC1-AUGm or pAVIC1-PTC1 RNAs. Scale bars represent 100  $\mu\text{m}$ . **(g)** Cell viability at 48 hpe for BSR cells electroporated with pAVIC1 or the indicated mutant RNAs, calculated as the ratio of released to total lactate dehydrogenase (LDH) activity (i.e.  $([\text{LDH}]_{\text{test}} - [\text{LDH}]_{\text{negative}}) / ([\text{LDH}]_{\text{max}} - [\text{LDH}]_{\text{negative}})$ ) (mean ± s.d.;  $n = 3$  biologically independent experiments). Max = maximum LDH measured for fully lysed cells. Source data are provided as a Source Data file.

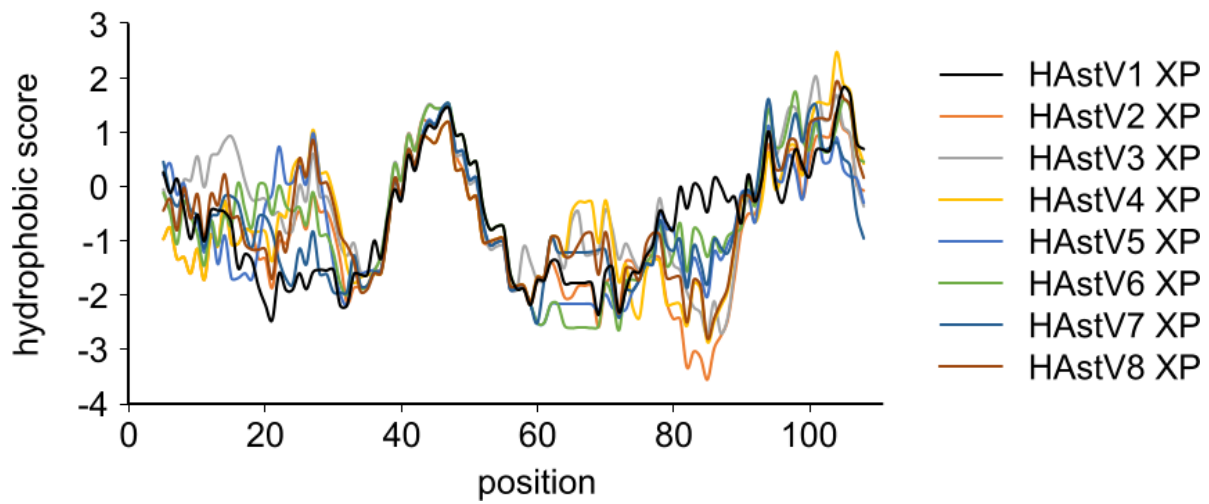


Supplementary Figure 14 | Original western blot scans for Fig. 5b,h.

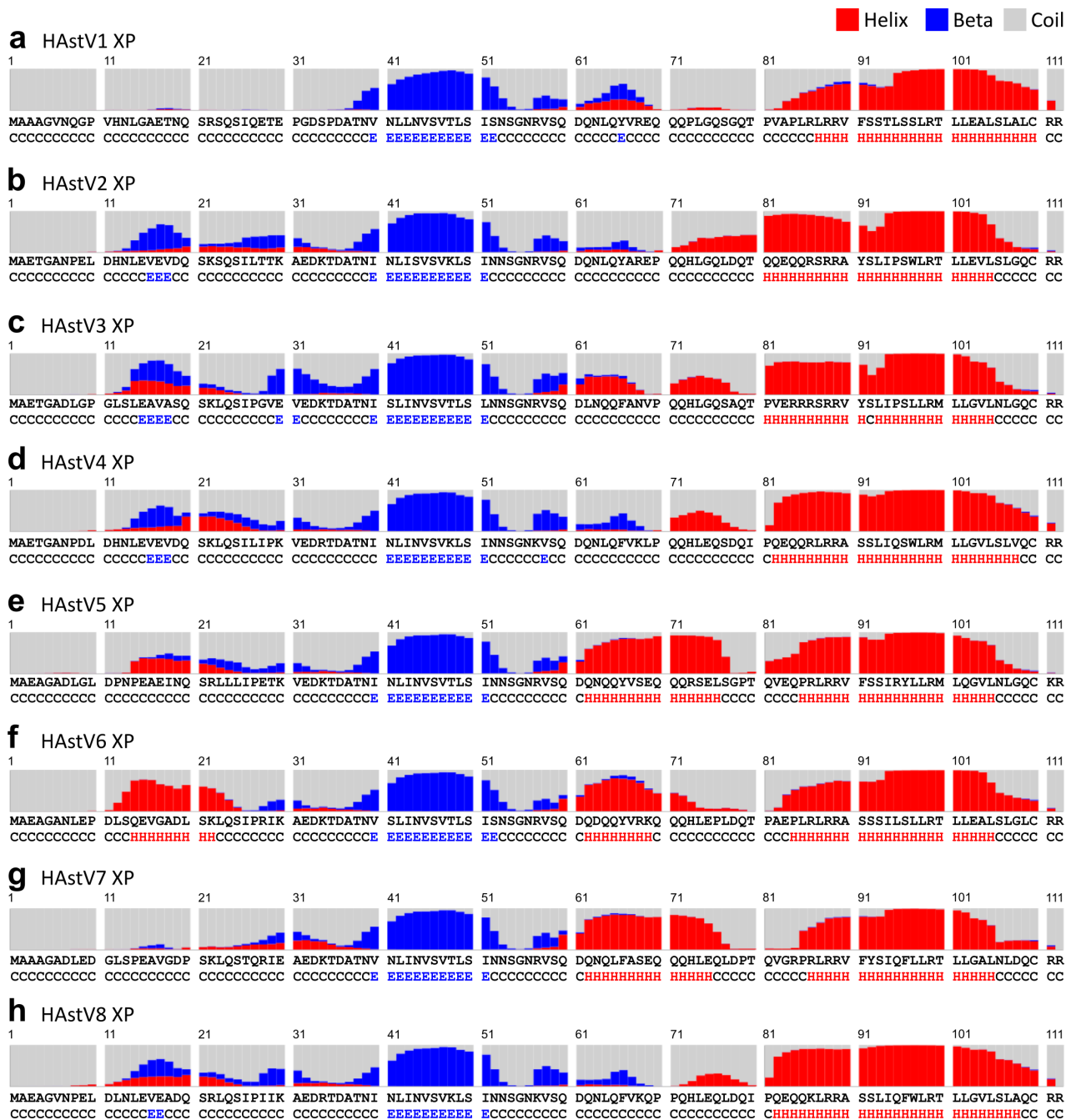




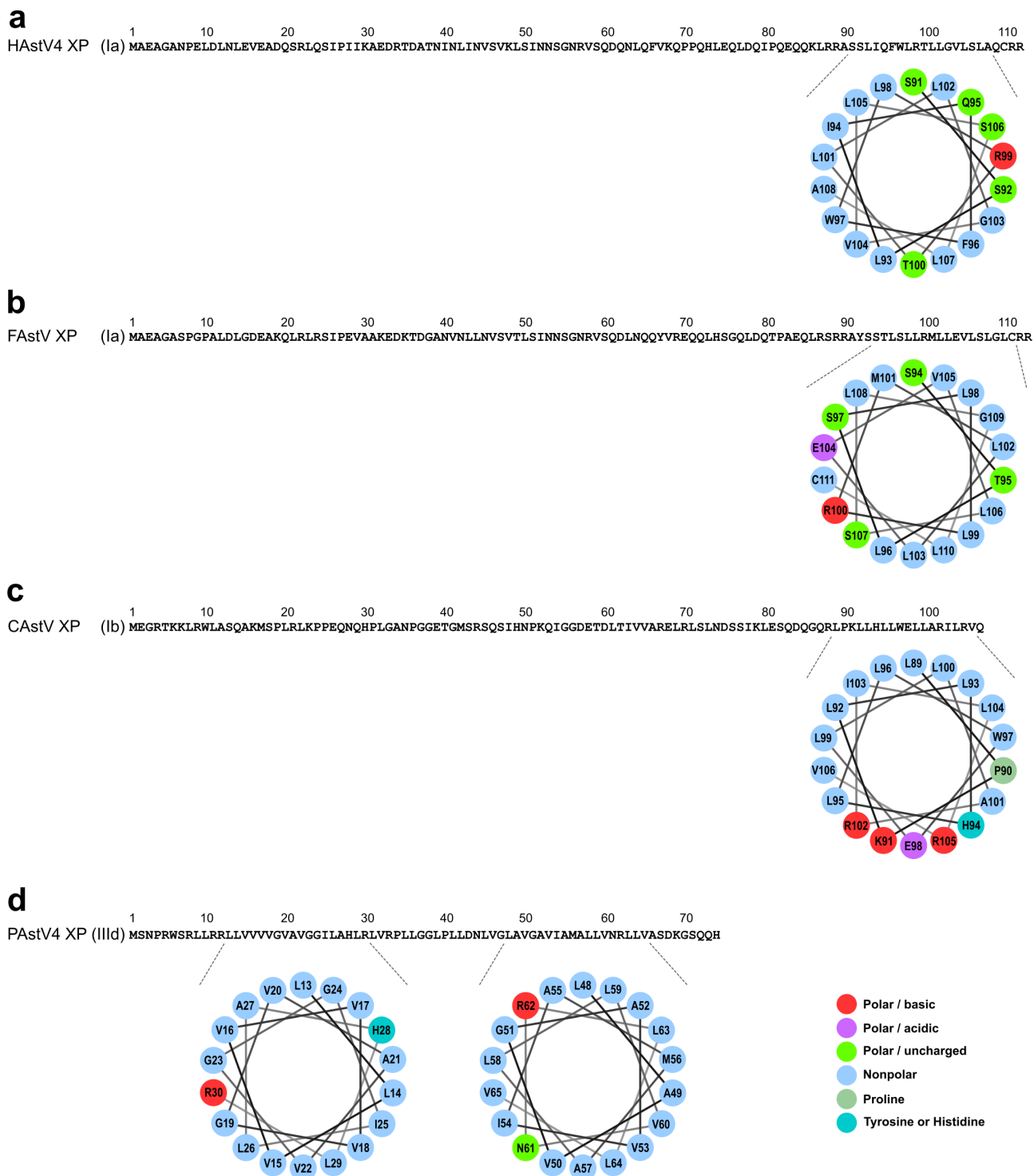
**Supplementary Figure 15 | Detection of XP-mCherry and mCherry-XP with anti-mCherry antibody.** HeLa cells were electroporated with pCAG-mCherry, pCAG-XP-mCherry or pCAG-mCherry-XP. The plasma membrane (but not internal membranes) was selectively permeabilized with digitonin, cells were stained with anti-mCherry antibody followed by Alexa 488-labelled anti-rabbit IgG antibody, and visualized by confocal microscopy. The images are averaged single plane scans. Scale bars represent 10  $\mu\text{m}$ .



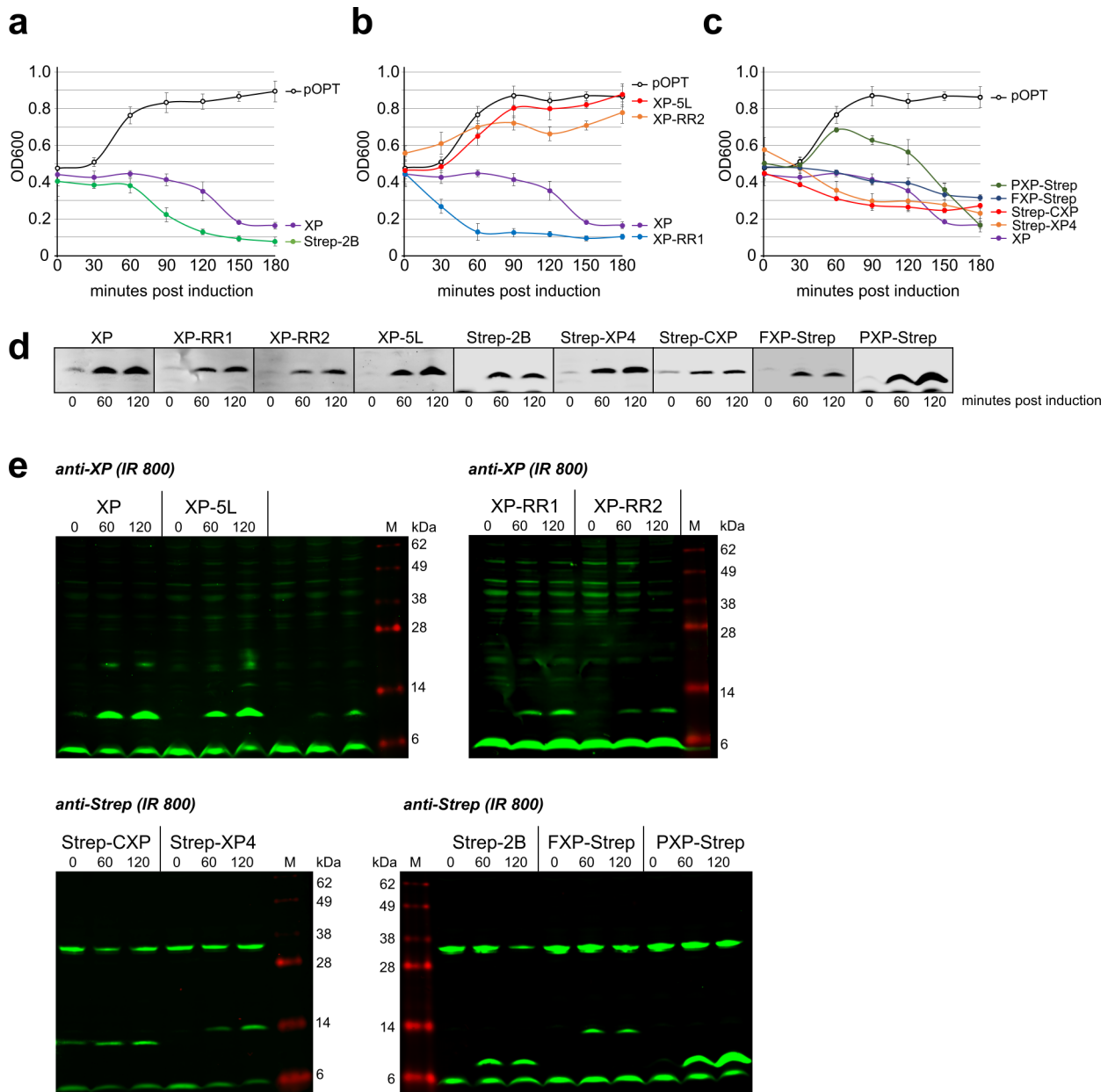
**Supplementary Figure 16 | Individual Kyte-Doolittle hydropathy plots for HAstV1-8 XPs.** Hydropathy plots for HAstV1 pAVIC1 and representative HAstV2–8 sequences (see Fig. 1f) were predicted with protscale with a 9-aa window size (<https://web.expasy.org/protscale>; May 2019).



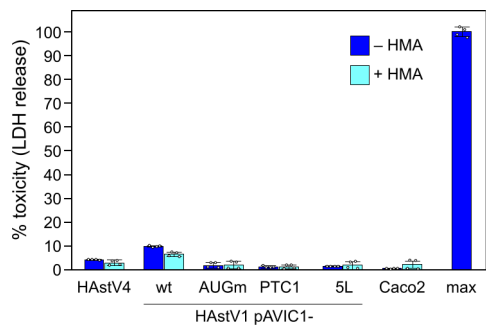
**Supplementary Figure 17 | Predicted secondary structure of HAstV XPs.** Protein secondary structures for representative sequences (see Fig. 1e) were predicted with RaptorX<sup>6</sup>.



**Supplementary Figure 18 | Helical wheel representations for TM regions of other astrovirus XPs used in Fig. 6e.** TMs were predicted with Phobius<sup>5</sup> (<http://phobius.sbc.su.se/>) and helical wheels were created and analysed using Heliquet<sup>7</sup> (<http://heliquet.ipmc.cnrs.fr>; May 2019).



**Supplementary Figure 19 | Bacterial lysis assay for viroporin-like activity. (a-c)** *E. coli* pLysS cells were transformed with pOPT constructs expressing the indicated proteins, grown to an optical density of 0.4 to 0.6, and then induced with IPTG. Optical densities were measured for induced and non-induced samples in triplicate over a time course of 180 min post induction. Graphs show means  $\pm$  s.d. from  $n = 3$  biologically independent experiments. **(d)** Non-induced, and 60 and 120 min post induction samples were also collected for protein detection by western blot. **(e)** Original western blot scans for (d). Source data are provided as a Source Data file.



**Supplementary Figure 20 | Toxicity assay for HMA-treated astrovirus-infected cells.** Caco2 cells were infected with the indicated viruses at MOI 0.2 in the presence (dark blue bars) or absence (light blue bars) of 5  $\mu$ M hexamethylene amiloride (HMA) for 48 h. Supernatant was used to measure cell viability, calculated as the ratio of released to total lactate dehydrogenase (LDH) activity (mean  $\pm$  s.d.;  $n = 4$  biologically independent experiments). Max = maximum LDH measured for fully lysed cells. Source data are provided as a Source Data file.

**Supplementary Table 1 | XP and YP statistics in different astrovirus groups**

Group	Protein	Number	Median mass (kDa)	Median pI	Median length (aa)	Predicted TM(s)
Bat astrovirus	XP	1	17.1	13.4	148	yes
Genogroup II d - bovine astrovirus etc	XP	17	10.7	12.7	91	yes
Genogroup IV - MLB astroviruses	XP	13	8.4	8.6	78	yes
Rodent astrovirus	XP	4	8.3	5.5	79	yes
Genogroup Ia - human astrovirus etc	XP	39	12.3	8.0	112	no
Genogroup Ic - California sea lion astrovirus etc	XP	5	10.8	7.9	98	no
Genogroup Ib - canine astrovirus etc	XP	10	10.1	9.7	91	no
Marmot astrovirus	XP	4	10.7	12.1	97	yes
Genogroup III d - porcine astrovirus 4 etc	XP	25	7.6	12.5	70	yes
Genogroup III a - murine astrovirus etc	XP	9	6.5	12.1	64	yes
Rodent astrovirus	XP	2	8.0	12.5	77	yes
Bovine astrovirus	XP	4	9.1	11.5	82	no
Genogroup III b - bovine astrovirus etc	XP	19	8.3	11.3	73	no
Genogroup III c - porcine astrovirus 2 etc	XP	24	8.3	11.8	75	no
Genogroup II c mamastrovirus 10 etc	YP	6	9.9	6.2	91	no

**Supplementary Table 2 | Accession numbers of the 415 astrovirus ORF2 sequences**

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AF117209 AF141381 AF248738 AF260508 AY179509 AY720891 AY720892 DQ028633  
DQ070852 DQ344027 DQ630763 EF138823 EF138824 EF138825 EF138826 EF138827 EF138828  
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FJ973620 FM213330 FM213331 FM213332 GQ267696 GQ405855 GQ405856 GQ405857  
GQ415660 GQ415661 GQ415662 GQ495608 GQ502193 GQ891990 GQ901902 GQ914773  
GU223905 GU376736 GU562296 GU732187 GU985458 HM045005 HM237363 HM447045  
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**Supplementary Table 3 | List of primers used**

Primer	Purpose	Sequence
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pAVIC-PTC1_F		CTAGGGGCGGAGATAAATCAGTGAAGATTACAGTCAATTCAAGAAACAGAG
pAVIC-PTC1_R		CTCTGTTTCTTGAATTGACTGTAATCTTCACTGATTTATCTCGGCCCTAG
pAVIC-PTC2_F		CTAGGGGCGGAGATAAATCAGTGAAGATAACAGTCAATTCAAGAAACAGAG CC
pAVIC-PTC2_R		GGCTCTGTTTCTTGAATTGACTGTTATCTTCACTGATTTATCTCGGCCCT AG
pAVIC-PTC3_F		CAGAGAGCAACAGCAACCCCTAGGGACGGTCGGGTCAAAC
pAVIC-PTC3_R		GTTTGACCCGACCGTCCCTAGGGTTGCTGTTGCTCTCTG
X-GS-mCh-F		GTTTGCCCTGTGCAGGCGCGGATCCGTGAGCAAGGGCGAGGAGG
X-SG-mCh-R:		GAGGAGTGAATTCGAAGATCTTAAGTTACTTGTACAGCTCGTCCATGCC
mCh-GS-X-F		GGCAAAGAATTAATTAAGCCACCATGGTGAGCAAGGGCGAGGAG
mCh-GS-X-R		CTTGATTTACTCCTGTTGCGGCGGATCCCTTGTACAGCTCGTCCATGCC
pAVIC-5L_F		AGTCCTCGTCAAGGACGCTACAGGAAGCACACAGTTTGGCCAGTGCAGGC GCTAGGTGCAC
pAVIC-5L_R		TGGGCCAAACTGTGTGCTTCTGTAGCGTCTTGTACGAGGACTGGGTTGAG GAGAATACACGCC
XP-5L-1R		GCTTCGGATAGCGTCTAGTTCGAGGAAGTGGTTGAGGAGAATACACGCCCTC
XP-RR-1R		CAGTAGCGTCTTAACGAGGACAGGGTTGAGGAGAATACAGCAGCCAATCT CAGTGGTGCCACTGG
XP-2R-R-BamHI		CGCGGATCCTTATGCTGCGCACAGGGCCAAACTGAGTGCTTCCAGTAGCGT CCTTAACGAGGAC
pAVIC-GNN-F		GTTTATGGAAATAACAGGCTTCTACAACACCTTCCG
pAVIC-GNN-R		GTAGAAAGCCTGTTATTTCCATAAACTACAGTGT CATAGGTC
4714-O2-2A-R		Cloning of replicon
2A-RLuc-F	CAGACTTTGAATTTTGACCTTCTCAAGCTGGCGGGAGACGTCGAGTCCAAC CCCCGGCCCCGCTTCCAAGGTGTACGACCC	
RLuc-TAG-6148-R	GCTGTGTTAGTTTTAAGTTCCACGTTACTACTGCTCGTTCCTCAGCACGC	
4714-X-2A-R	GAGAAGGTCAA AATTCAAAGTCTGTTTACC GG TGCCACAATTTCTGTTT GTGTCTGGCTTCGCGCCTGCACAGGGCC	
SINV_F_Aat2	SINV-repC cloning	CCGACAGATTGTTGACGTC AAG
SINV_R_XhoI		TTCCCCTCGAGGAATTCCTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTT TTTTTGAATG
CP_R_NdeBam		ATTAGGATCCATATATATACATATGTGCGGACCACTCTTCTGTCC
CP_F_NdeBam		tccgcaCATATGTATATATATGGATCCTAATCCGCTACGCCCAATGATCC G
mCh-Nde-F		GGGAATTCATATGGTGAGCAAGGGCGAGG
mCh-Bam-R		CGCGGATCCTTACTTGTACAGCTCGTCCATGCC
Strep-2B-NdeI-F	Cloning of 2B, XPs into pOPT, SINV-repC	GGGAATTCATATGTGGTCACATCCTCAGTTCGAAAAGTCCGCCGGGATCA AGGACTATGTGCAAC
2B-BamHI-R		CGCGGATCCTTATTGGCGTTCAGCCATGGGTATTC
Strep-XP4-NdeI-F		GGGAATTCATATGTGGTCACATCCTCAGTTCGAAAAGTCCGCCGCCGAAG CAGGAGCAAATCC
XP4-BamHI-R		CGCGGATCCTTAGCGCCTGCACTGGGCCAAAC
pCAG-StrXP-F-Pac	Cloning into pCAG vector	ATACCTTAATTAAGCCACCATGTCCGCTGGTCACATCCTCAGTTCGAAAA GGCCGCAACAGGAGTAAATCAAG
pCAG-XP-R-Afl		ATATACTTAAGTTAGCGCCTGCACAGGGCC
pCAG-XP-f-Pac		ATACCTTAATTAAGCCACCATGGCCGCAACAGGAGTAAATCAAG
pCAG-XP-HA-R-Afl		ATATACTTAAGTTAAGCGTAATCTGGAACATCGTATGGGTAGGCGGAGCGC CTGCACAGGGCC

pCAG-XPStr-R-Afl		ATATACTTAAGTTACTTTTTCGAACTGAGGATGTGACCAGGCCGGAGCGCCTG CACAGGGCC
pCAG-HAXP-F-Pac		ATACCTTAATTAAGCCACCATGTCCGCCTACCCATACGATGTTCCAGATTA CGCTGCCGCAACAGGAGTAAATCAAG
SINV-RT	Reverse transcription	GTTGAAGAATCCGCATTGCATGG
HAsV1-RT		TACTGCTGTAGCAATAAGGCCACG
SINV-qF	qPCR	GAAACAATAGGAGTGATAGGCA
SINV-qR		TGCATACCCCTCAGTCTTAGC
HAsV1-qF		TGCTATTGGTACTGTCATGGG
HAsV1-qR		GGTGTGAAATGGAATTGTGGG
hamGAPDH-qF		GGCAAGTTCAAAGGCACAGTC
hamGAPDH-qR		CACCAGCATCACCCCATTT

**Supplementary Table 4 | Host and virus read counts for different Ribo-Seq samples**

drug	repeat	time point	total reads	host rRNA	host mRNA	vRNA
No treatment	NT #1	12 hpi	34,261,071	23,068,841	3,960,680	778,926
	NT #2	12 hpi	41,436,501	27,367,513	5,669,906	1,035,209
Lactimidomycin	LTM #1	12 hpi	11,621,251	6,024,566	625,793	177,015
	LTM #2	12 hpi	12,903,118	9,177,384	828,982	240,408

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