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11 12	
13	Supplementary information to:
14	
15	Development of light-responsive protein binding in the monobody non-immunoglobulin
16	scaffold
17	César Carrasco-López <sup>1,#</sup> , Evan M. Zhao <sup>1,#</sup> , Agnieszka A. Gil <sup>2,#</sup> , Nathan Alam <sup>1</sup> , Jared E.
18	Toettcher <sup>2,*</sup> and José. L. Avalos <sup>1,3,*</sup>
19	
20	<sup>1</sup> Department of Chemical and Biological Engineering
21	Princeton University, Princeton NJ 08544
22	
23	<sup>2</sup> Department of Molecular Biology
24	Princeton University, Princeton NJ 08544
25	
26	<sup>3</sup> Andlinger Center for Energy and the Environment
27	Princeton University, Princeton NJ 08544
28	
29	<sup>#</sup> These authors contributed to this work equally
30	
31	* Co-corresponding Authors
32	toettcher@princeton.edu; javalos@princeton.edu
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35	Supplementary Information
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37	Supplementary Note 1
38	The AsLOV2 sequence is highlighted in blue.
39 40	HA4-AsLOV2 (SS58 insertion): αSH2-OptoMB GSSVSSVPTKI EVVAATPTSI LISWDAPMSSSSVYYYRITYGETGGNSPVOFETVPYS <mark>GL</mark>
41	ERIEKNEVITDPRLPDNPIIFASDSFLOLTEYSREEILGRNCRFLOGPETDRATVRKIRDAID
42	NOTEVTVOLINYTKSGKKFWNLFHLOPMRDOKGDVOYFIGVOLDGTEHVRDAAEREG
43	VMLIKKTAENIDEAAGSSTATISGLSPGVDYTITVYAWGEDSAGYMFMYSPISINYRTC*
44	
45	HA4-AsLOV2 (MS29 insertion with residues removed from Loop AB):
46	GSSVSSVPTKLEVVAATPTSLLISWDAPMGLERIEKNFVITDPRLPDNPIIFASDSFLOLTE
47	YSREEILGRNCRFLQGPETDRATVRKIRDAIDNQTEVTVQLINYTKSGKKFWNLFHLQP
48	MRDQKGDVQYFIGVQLDGTEHVRDAAEREGVMLIKKTAENIDEAAGSVYYYRITYGET
49	GGNSPVQEFTVPYSSSTATISGLSPGVDYTITVYAWGEDSAGYMFMYSPISINYRTC*
50	
51	HA4-AsLOV2 (MS29 insertion):
52	GSSVSSVPTKLEVVAATPTSLLISWDAPM <mark>GLERIEKNFVITDPRLPDNPIIFASDSFLQLTE</mark>
53	YSREEILGRNCRFLQGPETDRATVRKIRDAIDNQTEVTVQLINYTKSGKKFWNLFHLQP
54	MRDQKGDVQYFIGVQLDGTEHVRDAAEREGVMLIKKTAENIDEAAG <mark>SSSSVYYYR</mark> ITY
55	GETGGNSPVQEFTVPYSSSTATISGLSPGVDYTITVYAWGEDSAGYMFMYSPISINYRTC
56	*
57	
58	HA4-AsLOV2 (GN46 insertion with residues removed from Loop CD):
59	GSSVSSVPTKLEVVAATPTSLLISWDAPMSSSSVYYYRITYGETG <mark>GLERIEKNFVITDPRL</mark>
60	PDNPIIFASDSFLQLTEYSREEILGRNCRFLQGPETDRATVRKIRDAIDNQTEVTVQLINYT
61	KSGKKFWNLFHLQPMRDQKGDVQYFIGVQLDGTEHVRDAAEREGVMLIKKTAENIDE
62	AAGNSPVQEFTVPYSSSTATISGLSPGVDYTITVYAWGEDSAGYMFMYSPISINYRTC*
63	
64	HA4-ASLOV2 (1N45 insertion with residues removed from Loop CD):
65	GSSVSSVP1KLEVVAA1P1SLLISWDAPMSSSSVYYYKI1YGE1GLEKIEKNFVI1DPKLP
66 67	DNPIIFASDSFLQLTEY SKEEILGKNCKFLQGPETDKATVKKIKDAIDNQTEV TVQLINYTK
0/ 60	SOKKFWNLFHLQPWKDQKODVQIFIOVQLDOTEHVKDAAEKEOVWLIKKTAENIDEA
00 60	AOINSPVQEFIVFISSSTATISOLSPOVDITIIVIAWOEDSAGIMFMISFISINIKIC
70	$HAA = A \le OV2$ (SP68 insertion with residues removed from L oon EE):
70	GSSVSSVPTKI EVVA A TPTSI I ISWDA PMSSSSVVVVRITVGETGGNSPVOFETVPVSSST
72	ATISGS GLERIEKNEVITOPRI PONPIJEASOSELOLTEVSREEJI GRNCRELOGPETORATV
73	RKIRDAIDNOTEVTVOLINYTKSGKKFWNLEHLOPMRDOKGDVOVFIGVOLDGTEHVR
74	DAAEREGVMLIKKTAENIDEAAGPVDYTITVYAWGEDSAGYMFMYSPISINYRTC*
75	
76	HA4-AsLOV2 (PT18 insertion with residues removed from Loop AB):
77	GSSVSSVPTKLEVVAAPGLERIEKNFVITDPRLPDNPIIFASDSFLOLTEYSREEILGRNCRF
78	LQGPETDRATVRKIRDAIDNOTEVTVQLINYTKSGKKFWNLFHLQPMRDQKGDVQYFIG

79	VQLDGTEHVRDAAEREGVMLIKKTAENIDEAA <mark></mark> GTSLLISWDAPMSSSSVYYYRITYGET
80	GGNSPVQEFTVPYSSSTATISGLSPGVDYTITVYAWGEDSAGYMFMYSPISINYRTC*
81	
82	
83	AsLOV2 domain:
84	GLERIEKNFVITDPRLPDNPIIFASDSFLOLTEYSREEILGRNCRFLOGPETDRATVRKIRD
85	AIDNOTEVTVOLINYTKSGKKFWNLFHLOPMRDOKGDVOYFIGVOLDGTEHVRDAAER
86	EGVMLIKKTAENIDEAAG
87	
88	Monobody HA4:
89	GSSVSSVPTKLEVVAATPTSLLISWDAPMSSSSVYYYRITYGETGGNSPVOEFTVPYSSST
90	ATISGLSPGVDYTITVYAWGEDSAGYMFMYSPISINYRTC*
91	
92	His <sub>6</sub> -YFP-SH2:
93	HHHHHHSSGENLYFOGHMASKVSKGEELFTGVVPILVELDGDVNGHKFSVSGEGEGDA
94	TYGKLTLKFICTTGKLPVPWPTLVTTFGYGLQCFARYPDHMKQHDFFKSAMPEGYVQE
95	RTIFFKDDGNYKTRAEVKFEGDTLVNRIELKGIDFKEDGNILGHKLEYNYNSHNVYIMA
96	DKQKNGIKVNFKIRHNIEDGSVQLADHYQQNTPIGDGPVLLPDNHYLSTQSALSKDPNE
97	KRDHMVLLEFVTAAGITLGMDELYKSLEKHSWYHGPVSRNAAEYLLSSGINGSFLVRES
98	ESSPGQRSISLRYEGRVYHYRINTASDGKLYVSSESRFNTLAELVHHHSTVADGLITTLH
99	YPAPKRNKPTVYGVSPNY*
100	
101	HA4-AsLOV2 (SS30 insertion):
102	GSSVSSVPTKLEVVAATPTSLLISWDAPMS <mark>GLERIEKNFVITDPRLPDNPIIFASDSFLQLT</mark>
103	<b>EYSREEILGRNCRFLQGPETDRATVRKIRD</b> AIDNQTEVTVQLINYTKSGKKFWNLFHLQP
104	MRDQKGDVQYFIGVQLDGTEHVRDAAEREGVMLIKKTAENIDEAAG <mark>SSSVYYYR</mark> ITYG
105	ETGGNSPVQEFTVPYSSSTATISGLSPGVDYTITVYAWGEDSAGYMFMYSPISINYRTC*
106	
107	HA4-AsLOV2 (NS47 insertion):
108	GSSVSSVPTKLEVVAATPTSLLISWDAPMSSSSVYYYRITYGETGGN <mark>GLERIEKNFVITDP</mark>
109	${\it RLPDNPIIFASDSFLQLTEYSREEILGRNCRFLQGPETDRATVRKIRDAIDNQTEVTVQLIN}$
110	YTKSGKKFWNLFHLQPMRDQKGDVQYFIGVQLDGTEHVRDAAEREGVMLIKKTAENI
111	DEAAG SPVQEFTVPYSSSTATISGLSPGVDYTITVYAWGEDSAGYMFMYSPISINYRTC*
112	
113	HA4-AsLOV2 (SA84 insertion):
114	GSSVSSVPTKLEVVAATPTSLLISWDAPMSSSSVYYYRITYGETGGNSPVQEFTVPYSSST
115	ATISGLSPGVDYTITVYAWGEDS <mark>GLERIEKNFVITDPRLPDNPIIFASDSFLQLTEYSREEIL</mark>
116	GRNCRFLQGPETDRATVRKIRDAIDNQTEVTVQLINYTKSGKKFWNLFHLQPMRDQKG
117	DVQYFIGVQLDGTEHVRDAAEREGVMLIKKTAENIDEAAGAGYMFMYSPISINYRTC*
118	
119	YFP-SH2:
120	MASKVSKGEELFTGVVPILVELDGDVNGHKFSVSGEGEGDATYGKLTLKFICTTGKLPV
121	PWPTLVTTFGYGLQCFARYPDHMKQHDFFKSAMPEGYVQERTIFFKDDGNYKTRAEVK
122	FEGDTLVNRIELKGIDFKEDGNILGHKLEYNYNSHNVYIMADKQKNGIKVNFKIRHNIED
123	GSVQLADHYQQNTPIGDGPVLLPDNHYLSTQSALSKDPNEKRDHMVLLEFVTAAGITLG

124 MDELYKSLEKHSWYHGPVSRNAAEYLLSSGINGSFLVRESESSPGQRSISLRYEGRVYH

125 126 127	$\label{eq:constraint} YRINTASDGKLYVSSESRFNTLAELVHHHSTVADGLITTLHYPAPKRNKPTVYGVSPNY*$
127	$Hig_{\ell-}H\Lambda\Lambda$
120	HIMHINGSCENI VECCHASCSSVSSVDTVI EVVAATDTSI I ISWDADMSSSSVVVVDITV
129	CETCCNSDUCEETUDVSSSTATISCI SDCUDVTITUVAWCEDSACVMEMVSDISINVDTC
130	*
131	
132	
133	HA4-ASLOV2 (1644 insertion):
134	GSSVSSVPIKLEVVAAIPISLLISWDAPMSSSSVYYYKIIIGEIGLEKIEKNFVIIDPKLP
135	DNPIIFASDSFLQLTEY SKEEILGKNCKFLQGPETDKATVKKIKDAIDNQTEVTVQLINY IK
136	SGKKFWNLFHLQPMKDQKGDVQYFIGVQLDGTEHVKDAAEKEGVMLIKKTAENIDEA
13/	AGGGNSPVQEFIVPYSSSIAIISGLSPGVDYIIIVYAWGEDSAGYMFMYSPISINYRIC*
138	
139	HA4-AsLOV2 (SG65 insertion):
140	GSSVSSVPIKLEVVAAIPISLLISWDAPMSSSSVYYYRIIYGEIGGNSPVQEFIVPYSSSI
141	ATISGLERIEKNFVITDPRLPDNPIIFASDSFLQLTEYSREEILGRNCRFLQGPETDRATVRK
142	IRDAIDNQTEVTVQLINYTKSGKKFWNLFHLQPMRDQKGDVQYFIGVQLDGTEHVRDA
143	<mark>AEREGVMLIKKTAENIDEAAG</mark> GLSPGVDYTITVYAWGEDSAGYMFMYSPISINYRTC*
144	
145	HA4-AsLOV2 (DA26 insertion):
146	GSSVSSVPTKLEVVAATPTSLLISWD <mark>GLERIEKNFVITDPRLPDNPIIFASDSFLQLTEYSRE</mark>
147	EILGRNCRFLQGPETDRATVRKIRDAIDNQTEVTVQLINYTKSGKKFWNLFHLQPMRDQ
148	KGDVQYFIGVQLDGTEHVRDAAEREGVMLIKKTAENIDEAAGAPMSSSSVYYYRITYGE
149	TGGNSPVQEFTVPYSSSTATISGLSPGVDYTITVYAWGEDSAGYMFMYSPISINYRTC*
150	
151	HA4-AsLOV2 (ED82 insertion):
152	GSSVSSVPTKLEVVAATPTSLLISWDAPMSSSSVYYYRITYGETGGNSPVQEFTVPYSSST
153	ATISGLSPGVDYTITVYAWGEGLERIEKNFVITDPRLPDNPIIFASDSFLQLTEYSREEILGR
154	NCRFLQGPETDRATVRKIRDAIDNQTEVTVQLINYTKSGKKFWNLFHLQPMRDQKGDV
155	QYFIGVQLDGTEHVRDAAEREGVMLIKKTAENIDEAAGDSAGYMFMYSPISINYRTC*
156	
157	HA4-AsLOV2 (DS83 insertion):
158	GSSVSSVPTKLEVVAATPTSLLISWDAPMSSSSVYYYRITYGETGGNSPVQEFTVPYSSST
159	ATISGLSPGVDYTITVYAWGEDGLERIEKNFVITDPRLPDNPIIFASDSFLQLTEYSREEILG
160	RNCRFLQGPETDRATVRKIRDAIDNQTEVTVQLINYTKSGKKFWNLFHLQPMRDQKGD
161	VQYFIGVQLDGTEHVRDAAEREGVMLIKKTAENIDEAAG <mark>SAGYMFMYSPISINYRTC*</mark>
162	
163	HA4-AsLOV2 (GG45 insertion):
164	GSSVSSVPTKLEVVAATPTSLLISWDAPMSSSSVYYYRITYGET <mark>GGLERIEKNFVITDPRL</mark>
165	PDNPIIFASDSFLQLTEYSREEILGRNCRFLQGPETDRATVRKIRDAIDNQTEVTVQLINYT
166	KSGKKFWNLFHLQPMRDQKGDVQYFIGVQLDGTEHVRDAAEREGVMLIKKTAENIDE
167	AAGGNSPVQEFTVPYSSSTATISGLSPGVDYTITVYAWGEDSAGYMFMYSPISINYRTC*
168	
169	HA4-AsLOV2 (PT18 insertion):
170	GSSVSSVPTKLEVVAATP <mark>GLERIEKNFVITDPRLPDNPIIFASDSFLQLTEYSREEILGRNCR</mark>

1	FLQGPETDRATVRKIRDAIDNQTEVTVQLINYTKSGKKFWNLFHLQPMRDQKGDVQYFI
2	GVQLDGTEHVRDAAEREGVMLIKKTAENIDEAAGTSLLISWDAPMSSSSVYYYRITYGE
3	TGGNSPVQEFTVPYSSSTATISGLSPGVDYTITVYAWGEDSAGYMFMYSPISINYRTC*
4	
5	HA4-AsLOV2 (SP68 insertion):
5	GSSVSSVPTKLEVVAATPTŚLLISWDAPMSSSSVYYYRITYGETGGNSPVQEFTVPYSSST
	ATISGLSGLERIEKNFVITDPRLPDNPIIFASDSFLQLTEYSREEILGRNCRFLQGPETDRAT
	VRKIRDAIDNOTEVTVOLINYTKSGKKFWNLFHLOPMRDOKGDVOYFIGVOLDGTEHV
	RDAAEREGVMLIKKTAENIDEAAGPGVDYTITVYAWGEDSAGYMFMYSPISINYRTC*
	AsLOV2-HA4 (N-terminus):
	GLERIEKNFVITDPRLPDNPIIFASDSFLQLTEYSREEILGRNCRFLQGPETDRATVRKIRD
	AIDNQTEVTVQLINYTKSGKKFWNLFHLQPMRDQKGDVQYFIGVQLDGTEHVRDAAER
	EGVMLIKKTAENIDEAAGGSSVSSVPTKLEVVAATPTSLLISWDAPMSSSSVYYYRITYG
	ETGGNSPVQEFTVPYSSSTATISGLSPGVDYTITVYAWGEDSAGYMFMYSPISINYRTC*
	AsLOV2-HA4 (C-terminus):
	GSSVSSVPTKLEVVAATPTSLLISWDAPMSSSSVYYYRITYGETGGNSPVOEFTVPYSSST
	ATISGLSPGVDYTITVYAWGEDSAGYMFMYSPISINYRTCGLERIEKNFVITDPRLPDNPII
	FASDSFLOLTEYSREEILGRNCRFLOGPETDRATVRKIRDAIDNOTEVTVOLINYTKSGKK
	FWNLFHLOPMRDOKGDVOYFIGVOLDGTEHVRDAAEREGVMLIKKTAENIDEAAG*
	HA4-AsLOV2 (SS59 insertion):
	GSSVSSVPTKLEVVAATPTSLLISWDAPMSSSSVYYYRITYGETGGNSPVOEFTVPYSS
	LERIEKNFVITDPRLPDNPIIFASDSFLOLTEYSREEILGRNCRFLOGPETDRATVRKIRDAI
	DNOTEVTVOLINYTKSGKKFWNLFHLOPMRDOKGDVOYFIGVOLDGTEHVRDAAERE
	GVMLIKKTAENIDEAAGSTATISGLSPGVDYTITVYAWGEDSAGYMFMYSPISINYRTC*
	HA4-AsLOV2 (MY90 insertion).
	GSSVSSVPTKLEVVAATPTSLLISWDAPMSSSSVYYYRITYGETGGNSPVOEFTVPYSSST
	ATISGLSPGVDYTITVYAWGEDSAGYMFMGLERIEKNFVITDPRLPDNPIIFASDSFLOLT
	EYSREEILGRNCRFLOGPETDRATVRKIRDAIDNOTEVTVOLINYTKSGKKFWNLFHLOP
	MRDOKGDVOYFIGVOLDGTEHVRDAAEREGVMLIKKTAENIDEAAG <mark>YSPISINYRTC*</mark>
	Hise-HA4-AsLOV2 (SS58 insertion)
	HHHHHHSSGENLYFOGHASGSSVSSVPTKLEVVAATPTSLLISWDAPMSSSSVYYYRITY
	GETGGNSPVOEFTVPYS <mark>GLERIEKNEVITDPRLPDNPIIFASDSFLOLTEYSREEILGRNCRE</mark>
	LOGPETDRATVRKIRDAIDNOTEVTVOLINYTKSGKKEWNLEHLOPMRDOKGDVOVEIG
	VOLDGTEHVRDAAEREGVMLIKKTAENIDEAAGSSTATISGI SPGVDVTITVVAWGEDS
	AGYMEMYSPISINYRTC*
	HA4-AsIOV2 (VS57 insertion):
	GSSVSSVPTKI EVVA A TPTSI I ISWDA PMSSSSVVVVPITVCETCCNSDVOEETVDVCI E
	DISTORY I TALLY VAATI ISLLIS WDATWISSSSY I I TATI I UETUUNSFY VEFTY I UL PIEKNEVITOPPI DONDHEASOSELOL TEVSDEEH GONGDELOGDETOD ATVOKIDDADON
	NIENNTYH DI NEI DINHII ASDSTEQUIE I SNEDILONNUKTEQUIE I DKATYKNIKDAIDN OTEVTVOLINIVTZSCZZZWNI EULODMDDOZCDVOVELOVOLDCTEUVDDA AEDECV
	VILVIVUUVUTIKSUKKEWNLITILVPWKUVUUVVITIUVUUULEHVKUAAEKEGV

216 MLIKKTAENIDEAAGSSSTATISGLSPGVDYTITVYAWGEDSAGYMFMYSPISINYRTC\*

- 218 His<sub>6</sub>-HA4-AsLOV2 (SS58 insertion, C450V mutant): HHHHHHSSGENLYFQGHASGSSVSSVPTKLEVVAATPTSLLISWDAPMSSSSVYYYRITY 219 220 GETGGNSPVQEFTVPYSGLERIEKNFVITDPRLPDNPIIFASDSFLQLTEYSREEILGRNVR FLOGPETDRATVRKIRDAIDNOTEVTVOLINYTKSGKKFWNLFHLOPMRDOKGDVOYFI 221 GVQLDGTEHVRDAAEREGVMLIKKTAENIDEAAGSSTATISGLSPGVDYTITVYAWGED 222 223 SAGYMFMYSPISINYRTC\* 224 225 SH2-PDC1: 226 MGSLEKHSWYHGPVSRNAAEYLLSSGINGSFLVRESESSPGQRSISLRYEGRVYHYRINT 227 ASDGKLYVSSESRFNTLAELVHHHSTVADGLITTLHYPAPKRNKPTVYGVSPNYASSEIT 228 LGKYLFERLKQVNVNTVFGLPGDFNLSLLDKIYEVEGMRWAGNANELNAAYAADGYA 229 RIKGMSCIITTFGVGELSALNGIAGSYAEHVGVLHVVGVPSISAOAKOLLLHHTLGNGDF 230 TVFHRMSANISETTAMITDIATAPAEIDRCIRTTYVTQRPVYLGLPANLVDLNVPAKLLQ 231 TPIDMSLKPNDAESEKEVIDTILALVKDAKNPVILADACCSRHDVKAETKKLIDLTQFPA 232 FVTPMGKGSIDEQHPRYGGVYVGTLSKPEVKEAVESADLILSVGALLSDFNTGSFSYSY 233 KTKNIVEFHSDHMKIRNATFPGVQMKFVLQKLLTTIADAAKGYKPVAVPARTPANAAV 234 PASTPLKOEWMWNOLGNFLOEGDVVIAETGTSAFGINOTTFPNNTYGISOVLWGSIGFT 235 TGATLGAAFAAEEIDPKKRVILFIGDGSLQLTVQEISTMIRWGLKPYLFVLNNDGYTIEKL 236 IHGPKAQYNEIQGWDHLSLLPTFGAKDYETHRVATTGEWDKLTQDKSFNDNSKIRMIEI 237 MLPVFDAPQNLVEQAKLTAATNAKQ\* 238 239 His<sub>6</sub>-SUMO-HA4-AsLOV2 (SS58 insertion, V416I, G528A, N538E mutant): HHHHHHGSGSGSDQEAKPSTEDLGDKKEGEYIKLKVIGQDSSEIHFKVKMTTHLKKLKE 240 241 SYCQRQGVPMNSLRFLFEGQRIADNHTPKELGMEEEDVIEVYQEQTGGHMASKGSSVSS 242 VPTKLEVVAATPTSLLISWDAPMSSSSVYYYRITYGETGGNSPVOEFTVPYS<mark>GLERIEKN</mark> FIITDPRLPDNPIIFASDSFLOLTEYSREEILGRNCRFLOGPETDRATVRKIRDAIDNOTEVT 243 244 VOLINYTKSGKKFWNLFHLOPMRDOKGDVOYFIGVOLDGTEHVRDAAEREAVMLIKK 245 TAEEIDEAAGSSTATISGLSPGVDYTITVYAWGEDSAGYMFMYSPISINYRTC\* 246 247 His<sub>6</sub>-HA4-AsLOV2 (SS58 insertion, V416I mutant): HHHHHHSSGENLYFQGHASGSSVSSVPTKLEVVAATPTSLLISWDAPMSSSSVYYYRITY 248 GETGGNSPVQEFTVPYSGLERIEKNFIITDPRLPDNPIIFASDSFLQLTEYSREEILGRNCRF 249 LQGPETDRATVRKIRDAIDNQTEVTVQLINYTKSGKKFWNLFHLQPMRDQKGDVQYFIG 250 251 VQLDGTEHVRDAAEREGVMLIKKTAENIDEAAGSSTATISGLSPGVDYTITVYAWGEDS 252 AGYMFMYSPISINYRTC\* 253 254 HA4-irFP 255 MGSSVSSVPTKLEVVAATPTSLLISWDAPMSSSSVYYYRITYGETGGNSPVQEFTVPYSS 256 STATISGLSPGVDYTITVYAWGEDSAGYMFMYSPISINYRTCGGGAEGSVARQPDLLTC 257 DDEPIHIPGAIOPHGLLLALAADMTIVAGSDNLPELTGLAIGALIGRSAADVFDSETHNRL 258 TIALAEPGAAVGAPITVGFTMRKDAGFIGSWHRHDQLIFLELEPPQRDVAEPQAFFRRTN 259 SAIRRLQAAETLESACAAAAQEVRKITGFDRVMIYRFASDFSGEVIAEDRCAEVESKLGL 260 HYPASTVPAQARRLYTINPVRIIPDINYRPVPVTPDLNPVTGRPIDLSFAILRSVSPVHLEF 261 MRNIGMHGTMSISILRGERLWGLIVCHHRTPYYVDLDGRQACELVAQVLAWQIGVMEE
- 262 AAATPTCNMRD\*

263 264 265 266 HA4-AsLOV2-irFP (SS58 insertion) MGSSVSSVPTKLEVVAATPTSLLISWDAPMSSSSVYYYRITYGETGGNSPVOEFTVPYS<mark>G</mark> 267 268 LERIEKNFVITDPRLPDNPIIFASDSFLQLTEYSREEILGRNCRFLQGPETDRATVRKIRDAI 269 DNQTEVTVQLINYTKSGKKFWNLFHLQPMRDQKGDVQYFIGVQLDGTEHVRDAAERE <u>GVMLIKKTAENIDEAA</u>GSSTATISGLSPGVDYTITVYAWGEDSAGYMFMYSPISINYRTC 270 271 GGGAEGSVARQPDLLTCDDEPIHIPGAIQPHGLLLALAADMTIVAGSDNLPELTGLAIGA 272 LIGRSAADVFDSETHNRLTIALAEPGAAVGAPITVGFTMRKDAGFIGSWHRHDQLIFLEL 273 **EPPQRDVAEPQAFFRRTNSAIRRLQAAETLESACAAAAQEVRKITGFDRVMIYRFASDFS** 274 GEVIAEDRCAEVESKLGLHYPASTVPAQARRLYTINPVRIIPDINYRPVPVTPDLNPVTGR 275 PIDLSFAILRSVSPVHLEFMRNIGMHGTMSISILRGERLWGLIVCHHRTPYYVDLDGROA 276 CELVAQVLAWQIGVMEEAAATPTCNMRD\* 277 278 SH2-mCherry-CAAX 279 MSLEKHSWYHGPVSRNAAEYLLSSGINGSFLVRESESSPGQRSISLRYEGRVYHYRINTA 280 SDGKLYVSSESRFNTLAELVHHHSTVADGLITTLHYPAPKRNKPTVYGVSPNYVSKGEE 281 DNMAIIKEFMRFKVHMEGSVNGHEFEIEGEGEGRPYEGTQTAKLKVTKGGPLPFAWDIL 282 SPQFMYGSKAYVKHPADIPDYLKLSFPEGFKWERVMNFEDGGVVTVTQDSSLQDGEFI

- 283 YKVKLRGTNFPSDGPVMQKKTMGWEASSERMYPEDGALKGEIKQRLKLKDGGHYDAE
  284 VKTTYKAKKPVQLPGAYNVNIKLDITSHNEDYTIVEQYERAEGRHSTGGMDELYKGSG
  285 SGSKKKKKKSKTKCVIM
  286
- 287

### 288 Supplementary Note 2

289 We performed a search for monobodies in the Protein Data Bank (PDB), using "monobody" as

290 key word. Our search resulted in a total of 51 different structures of monobodies, each one in

291 complex with their cognate target. From this total, only 32 were monobodies with unique protein

- sequences, which are included in Supplementary Table 3. A close inspection of the mode of
- binding of each unique monobody in complex with its target revealed that 23 (72%) of them bind
- their cognate targets in a side-binding mode, including monobody HA4 (see Supplementary
- Figure 6). The side-binding mode involves the Loop FG, one or more of the  $\beta$ -strands from  $\beta$ S2
- and, occasionally, the CD loop.
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- 298
- 299
- 300
- 301

## **302** Supplementary Tables

303

### 304 Supplementary Table 1. List of HA4-AsLOV2 chimeras showing insertion positions.

Chimera Name	Positions in HA4 where AsLOV2 was inserted
SS30	Inserted between S30 and S31
NS47	Inserted between N47 and S48
SA84	Inserted between S84 and A85
TG44	Inserted between T44 and G45
SS58	Inserted between S58 and S59
SG65	Inserted between S65 and G66
DA26	Inserted between D26 and A27
MS29	Inserted between M29 and S30
MS29-3	Inserted between M29 and S33 residues 30-32 were removed
ED82	Inserted between E82 and D83
DS83	Inserted between D83 and S84
GG45	Inserted between G45 and G46
GN45	Inserted between G45 and N48 residues 46-47 were removed
TN44	Inserted between T44 and N48 residues 45-47 were removed
PT18	Inserted between P18 and T19
SP68	Inserted between S68 and P69
SP68-1	Inserted between S68 and P69 residue 67 was removed
N-terminus	Inserted at N-terminus
C-terminus	Inserted at C-terminus
SS59	Inserted between S59 and S60
PT18	Inserted between P18 and T19 residue 17 was removed
MY90	Inserted between M90 and Y91
YS57	Inserted between Y57 and S58

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### 307 Supplementary Table 2. Rate and dissociation constants from all individual BLI

308 experiments.

Variant	State measured	Illumination	kon (μM <sup>-1</sup> s <sup>-1</sup> )	$k_{\rm off}(\rm s^{-1})$	$K_d(\mu M)$
Monobody HA4	-	ambient	0.0631	0.0145	0.23
OptoMB 1	Lit conformation	ambient*	< 0.001	0.34	>100
OptoMB 2	Lit conformation	450 nm light	< 0.001	0.21	>100
OptoMB <sub>C450V</sub> 1	Dark conformation	ambient**	0.11	0.006	0.06
OptoMB <sub>C450V</sub> 2	Dark conformation	ambient**	0.056	0.013	0.23
OptoMB <sub>C450V</sub> 3	Dark conformation	450 nm light	0.048	0.013	0.27
OptoMB <sub>V416L</sub> 1	Lit conformation	ambient*	0.0029	0.25	87
OptoMB <sub>V416L</sub> 2	Lit conformation	450 nm light	0.0033	0.20	60
OptoMB <sub>V416L</sub> 3	Lit conformation	450 nm light	0.0058	0.24	42
<b>#</b> OptoMBv416I_G528A_N538E 1	Lit conformation	ambient*	< 0.001	0.10	>100
#OptoMBv416I_G528A_N538E 2	Lit conformation	ambient*	< 0.001	0.13	>100

309 \*Due to the sensitivity of these constructs, ambient light in the laboratory as well as internal light from the digital

310 panels of the Octet (BLI instrument) was sufficient to trigger the lit conformation.\*\*This is a light-insensitive mutant

and neither ambient nor LED light affected its basal dark conformation.

312 #SUMO tagged

# 313 Supplementary Table 3. Structurally characterized monobodies and their target binding

# 314 mode (source: PDB).

PDB	Monobody	Target	Binding	Library*	Ref.
code			Mode		
20CF	E2#23	Human estrogen receptor	Side Binding	FG-Loop (FG7)	Koide <i>et al.</i> , 2002 <sup>1</sup>
3K2M	HA4	Abl1 SH2 domain	Side Binding	Loops-only	Wojcik <i>et al.</i> , 2010 <sup>2</sup>
3QHT	ySMB-1	Yeast SUMO	Side Binding	Loops-only	Gilbreth <i>et al.</i> , 2011 <sup>3</sup>
3RZW	ySMB-9	Human SUMO	Loops Binding	Loops-only	Koide <i>et al.</i> , 2012 <sup>4</sup>
3T04	7c12	Abl1 SH2 domain	Side Binding	Loops-only	Grebien <i>et al.</i> , 2011 <sup>5</sup>
3UYO	SH13	Abl1 SH2 domain	Side Binding	Side and Loop	Koide <i>et al.</i> , 2012 <sup>4</sup>
4HUK	MbMATE1	MATE multidrug transporter	Side Binding	Loops-only	Lu <i>et al.</i> , 2013 <sup>6</sup>
4JE4	NSa1	NHP2 N-SH2 domain	Side Binding	Side and Loop	Sha <i>et al.</i> , 2013 <sup>7</sup>
4JEG	CS1	NHP2 C-SH2 domain	Side Binding	Side and Loop	Sha <i>et al.</i> , 2013 <sup>7</sup>
5A40	Bpe-S7	Fluoride Ion channel	Side Binding	Side and Loop	Stockbridge <i>et al.</i> , $2014^8$ and $2015^9$
5A43	Bpe-L2	Fluoride Ion channel	Loops Binding	Loops-only	Stockbridge <i>et al.</i> , $2014^8$ and $2015^9$
5DC0	GG3	Abl1 SH2 domain	Side Binding	Side and Loop	Wojcik <i>et al.</i> , 2016 <sup>10</sup>
5DC4	AS25	Abl1 SH2 domain	Side Binding	Side and Loop	Wojcik <i>et al.</i> , 2016 <sup>10</sup>
5E95	NS1	H-RAS	Side Binding	Side and Loop	Spencer-Smith et al., 2017 <sup>11</sup>
5ECJ	S4	Prdm14	Side Binding	Not specified	Nady <i>et al.</i> , 2015 <sup>12</sup>
5G15	Mb1	Aurora A	Loops Binding	Not specified	Zorba <i>et al.</i> , 2019 <sup>13</sup>
5KBN	S9	Fluoride channel F80I Mutant	Loops Binding	Side and Loop	Last <i>et al.</i> , 2016 <sup>14</sup>
5KVM	α5	GPR56 ERC	Side Binding	Side and Loop	Salzman <i>et al.</i> , 2016 <sup>15</sup>
5MTJ	Yes_1	Yes1-SH2	Side Binding	Side and Loop	Kükenshöner et al., 2017 <sup>16</sup>
5MTM	Lck_3	Lck-SH2	Side Binding	Side and Loop	Kükenshöner et al., 2017 <sup>16</sup>
5MTN	Lck_3	Lck-SH2	Side Binding	Side and Loop	Kükenshöner et al., 2017 <sup>16</sup>
5N7E	Bcr-DH_4	Dbl homology domain of Bcr-Abl	Side Binding	Side and Loop	Reckel <i>et al.</i> , 2017 <sup>17</sup>
5NKQ	Ec2-S9	Fluoride Ion channel	Side Binding	Side and Loop	Stockbridge et al., 20159
50C7	Bcr-PH_4	Pleckstrin-homology domain of Bcr-Abl	Side Binding	Not specified	Reckel <i>et al.</i> , 2017 <sup>17</sup>

5V7P	MbRas1	Ras Methyltransferase	Loops	Not	Diver <i>et al.</i> , 2018 <sup>18</sup>
		ICMT	Binding	specified	
6APX	YSX1	Human phosphatase 1	Loops	Loops-only	Gumpena <i>et al.</i> , 2017 <sup>19</sup>
		catalytic domain	Binding		
6BQO	Bpe-S8	Fluoride Ion channel	Side	Side and	Stockbridge et al., 20148 and
			Binding	Loop	MclIwain <i>et al.</i> , 2017 <sup>20</sup>
6BX4	S9m	Fluoride Ion channel	Loops	Side and	Turman <i>et al.</i> , 2018 <sup>21</sup>
			Binding	Loop	
6BX5	S12	Fluoride Ion channel	Loops	Side and	Turman <i>et al.</i> , 2018 <sup>21</sup>
			Binding	Loop	
6BYN	S4	WDR5	Side	Side and	Gupta et al., 2018 <sup>22</sup>
			Binding	Loop	-
6C83	Mb2	Aurora A	Loops	Not	Zorba <i>et al.</i> , 2019 <sup>13</sup>
			Binding	specified	
6D0J	X1	CLC-Fluoride proton	Side	Not	Last <i>et al.</i> , 2018 <sup>23</sup>
		antiporter	Binding	specified	

315 \*Loops-only library diversified Loops BC, DE and FG; the Side and Loop libraries have at least two versions reported,

one includes diversification of FG loop and several positions in  $\beta$ S2; another library diversifies the Loop CD in addition to FG and the positions in  $\beta$ S2; the FG-Loop library diversifies only the Loop FG.

Plasmid	Tag	Protein	Marker	Vector type
EZ-L663	None	HA4	Kanamycin	pCri-7b
EZ-L664	His <sub>6</sub>	YFP-SH2	Kanamycin	pCri-8b
EZ-L703	None	YFP-SH2	Kanamycin	pCri-7b
EZ-L704	His <sub>6</sub>	HA4	Kanamycin	pCri-8b
EZ-L706	None	HA4-AsLOV2 (SS58 insertion)	Kanamycin	pCri-7b
EZ-L736	None	HA4-AsLOV2 (MS29 insertion, residues 30-32 were removed)	Kanamycin	pCri-7b
EZ-L747	None	HA4-AsLOV2 (SS59 insertion)	Kanamycin	pCri-7b
EZ-L765	His <sub>6</sub>	HA4-AsLOV2 (SS58 insertion)	Kanamycin	pCri-8b
EZ-L830	His6-SUMO	HA4-AsLOV2 (SS58 insertion, V416L mutant)	Kanamycin	pCri-11b
EZ-L884	His <sub>6</sub>	HA4-AsLOV2 (SS58 insertion, C450V mutant)	Kanamycin	pCri-8b
EZ-L886	None	SH2-PDC1	Kanamycin	pCri-7b
EZ-L889	His6-SUMO	HA4-AsLOV2 (SS58 insertion, V416I, G528A, N538E mutant)	Kanamycin	pCri-11b
EZ-L892	His <sub>6</sub>	HA4-AsLOV2 (SS58 insertion, V416I mutant)	Kanamycin	pCri-8b
AG-pHR1	None	HA4-iRFP	Ampicillin	pHR
AG-pHR2	None	HA4-AsLOV2-iRFP (SS58 insertion)	Ampicillin	pHR
AG-pHR3	CAAX	SH2-mCherry-CAAX	Ampicillin	pHR

# 348 Supplementary Table 4. Constructs used in this study.

### 358 Supplementary Figures



360 Supplementary Fig. 1: Representative SDS-PAGE gels of pull-down screens of HA4-

361 AsLOV2 chimeras. a, Complete SDS-PAGE gel of the results shown in Fig. 1d, including

chimeras with the AsLOV2 domain inserted in positions SS58, SS59 and MS29 of HA4 (with

- residues S30 to S32 of Loop BC deleted). **b-d**, A representative sample of SDS-PAGE gels of
- 364 other chimeras with the AsLOV2 domain inserted in different positions that either do not bind or
- 365 bind poorly to SH2 (TG44, NS47, SA84, YS57 and PT18) including at two positions in Loop EF
- 366 (SG65, SP68); or bind well to SH2, but show no difference in binding between light conditions
- 367 (SS30). All pull-downs were repeated at least twice observing similar results.

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370	Supplementary Fig. 2: In vitro characterization of OptoMB binding. a, Time course of
371	fluorescence microscopy images of YFP-SH2 binding to agarose beads conjugated with
372	OptoMB <sub>V416L</sub> (left panel) or HA4 as control (right panel). Starting with beads incubated in the
373	dark, time course begins upon blue light stimulation ( $t = 0$ ), followed by a sequence of images
374	taken every 2 min for a total of 8 min. Scale bar (white) represents 100 µm. b, BLI
375	measurements of binding (left) and unbinding (right) of YFP-SH2 to immobilized monobody
376	HA4, and several replicates for OptoMB, OptoMB <sub>C450V</sub> , OptoMB <sub>V416L</sub> and
377	OptoMB <sub>V416I_G528A_N538E</sub> in different light conditions (see Supplementary Table 2). $c$ ,
378	Comparison of binding $(k_{on})$ , unbinding $(k_{off})$ , and dissociation $(K_d)$ constants calculated from
379	BLI measurements for different OptoMB variants, corresponding to dark (gray or black) or lit
380	(blue) states, including replicates. The red asterisks highlight measurements of $k_{on}$ below the
381	limit of detection of 0.001 $\mu$ M <sup>-1</sup> s <sup>-1</sup> . To calculate $K_d$ values from measurements below this limit,
382	we used $k_{on} = 0.001 \ \mu M^{-1} s^{-1}$ . OptoMB <sub>Triple</sub> refers to the OptoMB <sub>V416I_G528A_N538E</sub> variant. <b>d</b> ,
383	Comparison of the binding and unbinding rate constants, $k_{on}$ and $k_{off}$ , for replicate measurements
384	of the different OptoMB variants in their respective dark (black points) or lit (blue points) states.
385	Measurements of $k_{on}$ below the limit of detection (red dashed line) where assigned the limit $k_{on} =$
386	0.001 $\mu$ M <sup>-1</sup> s <sup>-1</sup> (red points). Source data are provided as a Source Data file.
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401 Supplementary Fig. 3: In vitro characterization of OptoMB binding in solution by size 402 exclusion chromatography. a, Experimental setup for dark or lit size exclusion chromatography 403 experiments using a Superdex 200 16/300 column (GE®). For experiments in the dark (left), the 404 column was wrapped with thick aluminum foil, and the chromatography refrigerator covered 405 with a black blanket to avoid light contamination (not shown). For experiments in the light 406 (right), the column was wrapped with blue LEDs. b, Size exclusion chromatography control 407 experiments for SUMO-tagged OptoMB<sub>V416I</sub> G528A N538E (OptoMB<sub>Triple</sub>) in light (blue) and dark 408 (red). c, Size exclusion chromatography control experiments to show identical elution profiles 409 for the HA4-YFP-SH2 complex in dark (black) or light (blue) conditions. Purified YFP-SH2 410 (yellow) and HA4 monobody (red) are also shown as controls. Source data are provided as a 411 Source Data file.



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413 Supplementary Fig. 4: Light-Controlled Affinity Chromatography (LCAC) to purify SH2tagged proteins using OptoMB immobilized on Co<sup>2+</sup> agarose beads. a, Schematic diagram of 414 LCAC procedure using a column packed with OptoMB-coated agarose beads. After flowing 415 416 through crude extract and washing in the dark, elution is carried out by applying blue light to the 417 surface of the column. b, SDS-PAGE gel of YFP-SH2 purified using a column packed with 418 OptoMB agarose beads. c, SDS-PAGE gel of YFP-SH2 purified using a column packed with 419 SUMO-tagged OptoMB<sub>V416I</sub> G528A N538E (OptoMB<sub>Triple</sub>) agarose beads. d, SDS-PAGE gel of 420 SH2-Pdc1p purified using a column packed with SUMO-tagged OptoMB<sub>V416I</sub> G528A N538E agarose 421 beads. The "negative control" c,d, are final elutions from columns that were not exposed to 422 crude extracts, but were otherwise treated the same. Purifications were repeated at least three 423 times observing similar results.



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425 Supplementary Fig. 5: Light-controlled affinity chromatography to purify SH2-tagged

426 proteins using cyanogen bromide (CNBr)-conjugated OptoMB. a, Schematic diagram of

427 OptoMBv416I\_G528A\_N538E with SUMO tag (S) covalently linked to CNBr beads through surface-

428 exposed primary amines, and their use in LCAC as described before. **b**, **c**, SDS-PAGE gel of

429 YFP-SH2 (b) or SH2-PDC1 (c) purified using SUMO-tagged OptoMB<sub>V416I\_G528A\_N538E</sub>

430 conjugated to CNBr beads, in batch. Molecular weight markers (M), lysate (L), unbound flow

431 through (FT), washing step in the dark (Dark Wash), two consecutives light elution aliquots

432 (Light Elution 1 and 2) and heat-treated beads resolved in SDS-PAGE gel (12%

433 polyacrylamide). Purifications were repeated at least three times observing similar results.

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Supplementary Fig. 6: Structures of the other twenty-two unique monobodies displaying
side-binding modes. Each monobody is shown (cartoons) in complex with their cognate targets
(white surfaces) and labeled with their corresponding PDB code. The regions involved in targetbinding interactions (orange) and the DE loop (red loop) where AsLOV2 is inserted in our
OptoMB are highlighted.

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