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Supplemental Information

Kinase Interaction Network Expands

Functional and Disease Roles of Human Kinases

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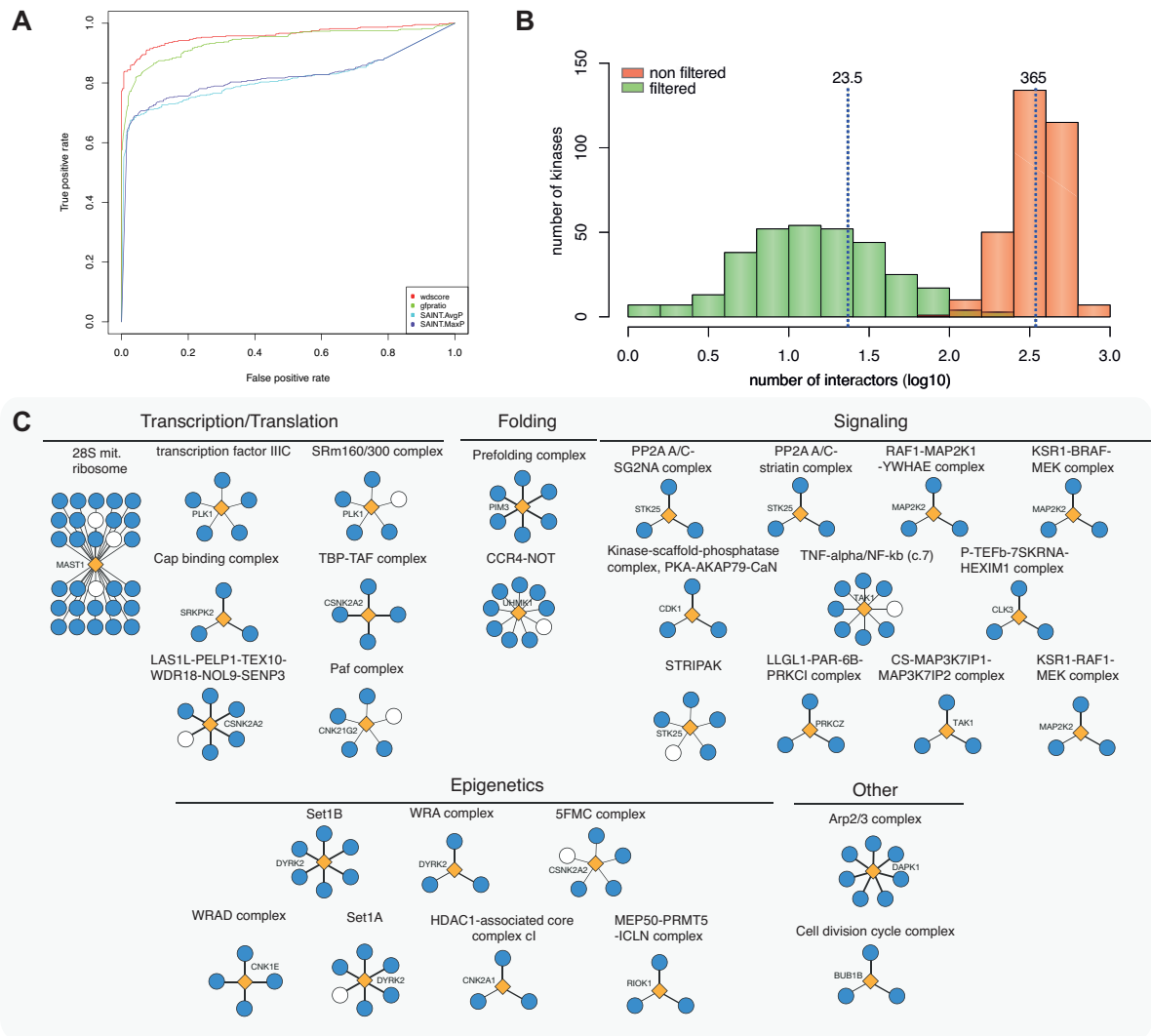


Figure S1. Data filtering and detection of known stable interactions, Related to Figure 1. **A.** Receiver operator characteristic curves for different filtering approaches that were applied on the initial interaction dataset. **B.** Histograms depicting the number of interactors before and after filtering show a reduction of nearly 95% of the interactors due to filtering. **C.** Proteins copurified with the same kinase bait can form stable complexes. CORUM protein complexes (>2 subunits) with at least 80% coverage that was achieved with a single pulldown. Some partially redundant complexes are not shown.

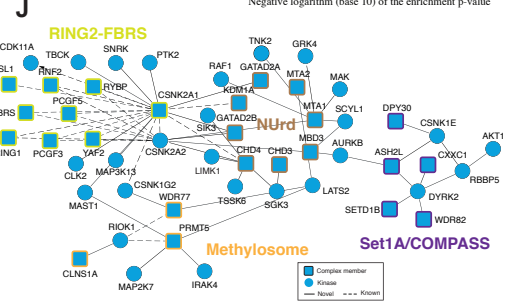
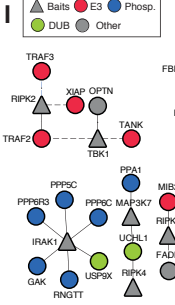
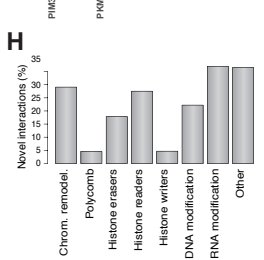
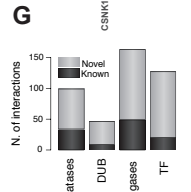
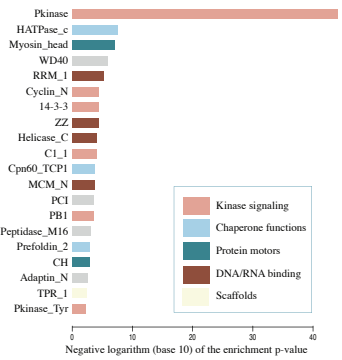
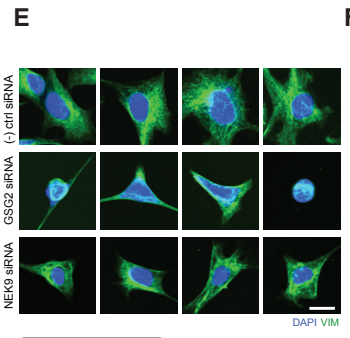
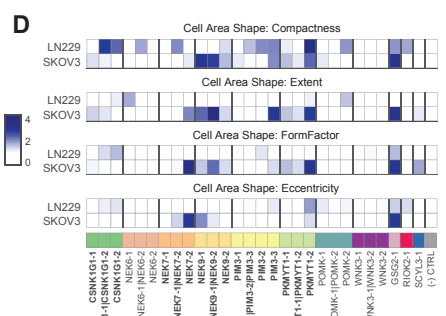
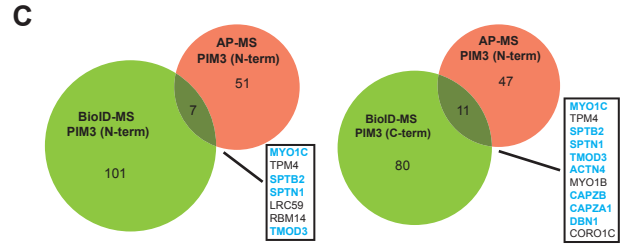
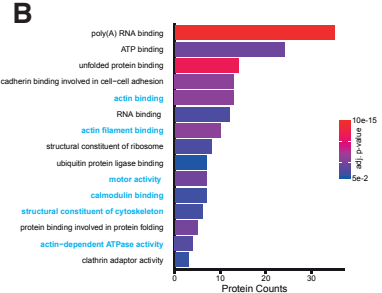
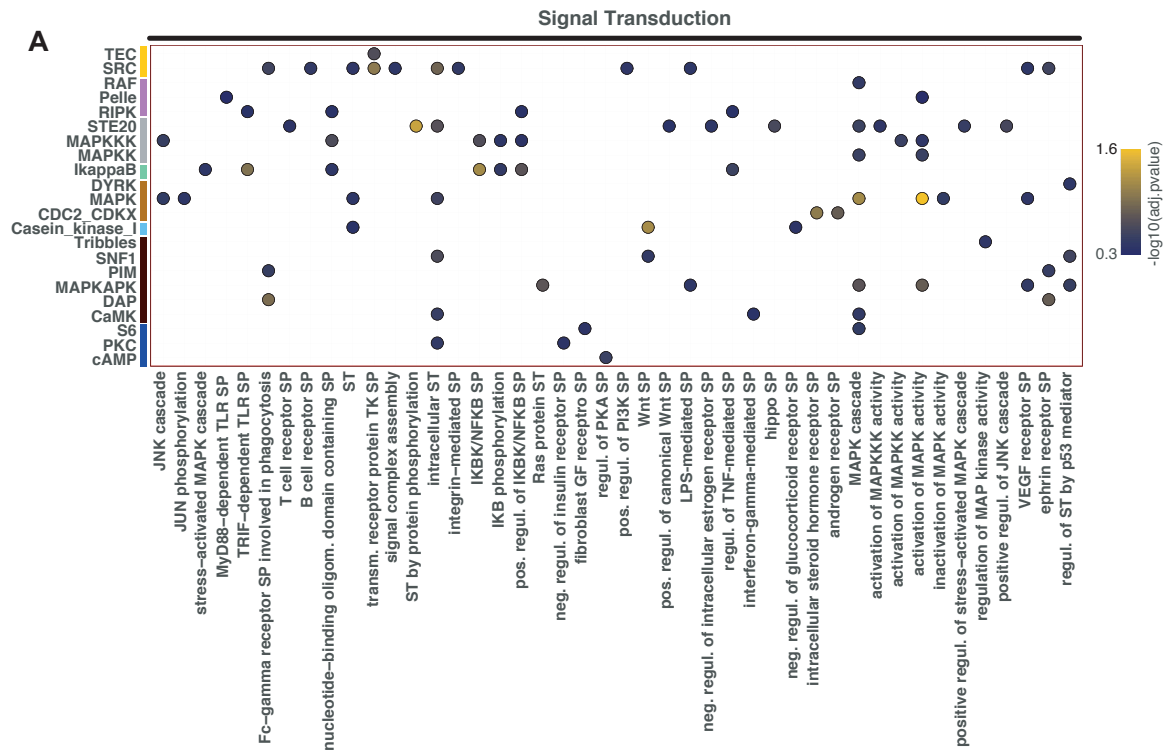
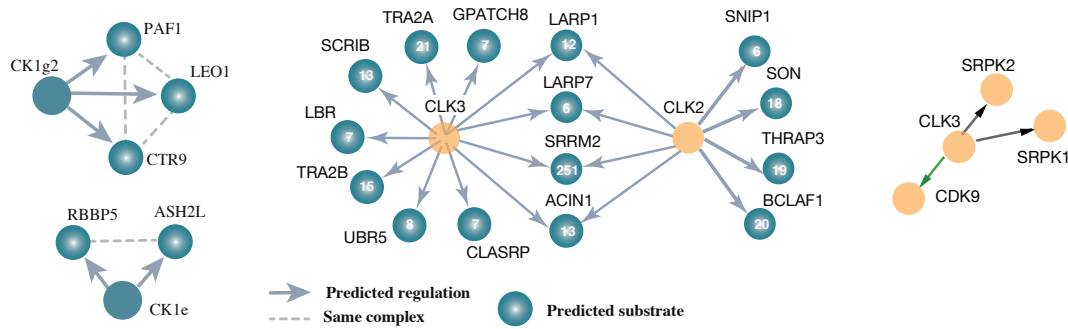


Figure S2. Functional landscape of proteins included in the Kinase interaction network, Related to Figure 3. **A.** Dotplot diagram of all terms which were collapsed to the GO term “signal transduction” in Figure 3A. SP=Signaling/Signaling Pathway; ST=Signal Transduction. **B.** Functional GO-term analysis (“Molecular Function_direct”) of the combined (N- and C-terminal) BioID-MS interaction network of PIM3. GO-terms that involve actin related molecular functions are shown in blue. **C.** Venn diagram is illustrating the overlap of the filtered AP-MS interaction network of PIM3 and the identified BioID-MS network for N-terminal (left panel) or C-terminal (right panel) FLAG-BirA*-tagged PIM3. Proteins highlighted in blue are assigned with the GO-term “actin binding” (GO: 0003779). **D.** Heat maps depict significant changes in cell phenotypes after gene silencing of target genes relative to the negative siRNA control. Example data for four different area and shape features, which were defined and measured by CellProfiler are shown. Rows represent measurements on two independent cell lines LN299 and SKOV3 and columns correspond to individual siRNA gene targets where different siRNAs for the same gene are indicated with -1 or -2 and combinations of siRNAs are denoted by the symbol ‘|’. Color-coded bars represent $-\log_{10}$ of the calculated p-values. Data is derived from n=3 replicate wells. siRNA targets that elicit significant phenotypes following siRNA depletion are indicated in bold at the bottom. **E.** Representative images of cells treated with negative control siRNAs and with siRNA constructs for knockdown of the GSG2 and NEK9 genes. **F.** Barplot of most significantly enriched domains among kinase prey proteins **G.** Stacked barchart with the fraction of novel vs already characterized interactions for the indicated protein classes. **H.** Fraction of novel interactions in the defined subclasses of epigenetic factors (see (Medvedeva et al., 2015) for classification) **I.** NfKB activation-related modules with previously reported (dashed line) and novel (solid line) interactions. **J.** Novel vs known interactions involving epigenetic complexes.

A Regulation of protein complex members and substrates with multiple motifs



B Kinase-kinase regulatory network

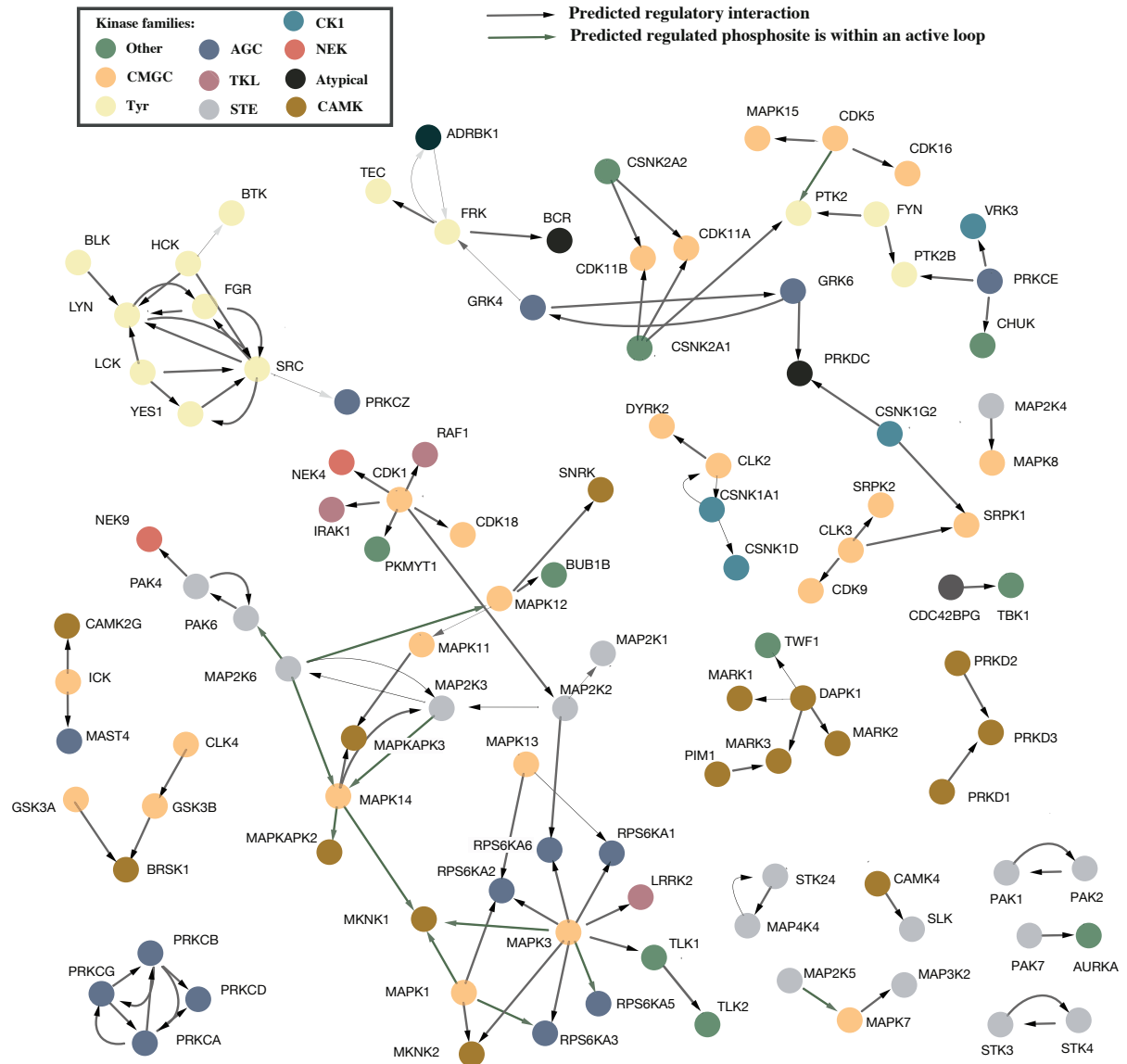


Figure S3. Regulatory interactions in the kinase network, Related to Figure 4. A. Highlighted regulatory examples: Similar to CK2 substrates, several predicted substrates of casein kinases CK1e and CK1g2 also had a role in transcription regulation. Three proteins from the Paf complex, which is involved in transcription elongation, were predicted to be regulated by the CK1g2 kinase. Furthermore, another example for kinases that had substrates with multiple corresponding phosphomotifs were CDC-like kinases CLK2 and CLK3. There, 12 out of 15 predicted substrates with multiple CLK2/3 phosphomotifs were associated with the GO term “poly(A) RNA binding”. **B.** Predicted kinase-kinase regulatory network. Dark arrows represent directed phosphorylation events, green arrows instances where a phosphosite maps to an activation loop. Kinase families are colored according to the scheme in **Figure 1**. Only regulatory modules with at least three kinases are included.

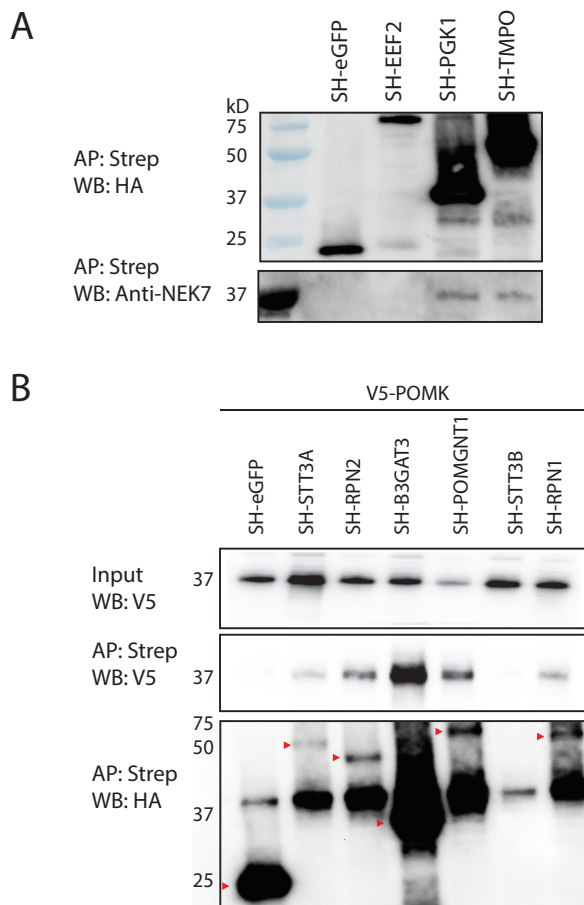


Figure S4. Validation of novel interactions by copurification and Western blotting, Related to Figure 6. A. Reciprocal validation of newly identified interactions with endogenous NEK7 in A549 cells. Proteins indicated at the top were transiently expressed as SH-tagged proteins for subsequent Streptactin affinity purification. Purified proteins were detected with anti HA antibodies (upper panel, bait expression control) or with anti NEK7 antibodies (lower panel). Lower panel validates the novel interaction of PGK1 and TMPO **B.** Validation of POMK interactors by reciprocal copurification and Western blotting in A549 cells. V5-tagged POMK kinase were transiently expressed with indicated SH-tagged proteins for subsequent copurification and Western blotting. Expression of V5-POMK5 was confirmed by anti V5 Western blotting of the transfected lysates (upper panel). Proteins purified by Streptactin affinity purification were detected by Western blotting with anti V5 antibodies for the detection of copurified V5-POMK (middle panel) and anti HA for the detection of purified SH tagged bait proteins (lower panel). Red arrow indicates expected size of SH tagged proteins. The detected V5-POMK signal confirms the known interaction with the POMGNT1 protein from the O-glycosylation pathway, as well as the novel interactions with STT3A, RPN2, RPN1 from the N-glycosylation pathway and with the B3GAT3 protein that is linked to multiple diseases. Of note, these are all membrane proteins (residing either in the ER or Golgi membranes). POMK (protein O-mannose kinase) itself is localized in the ER membrane, where it is involved in building protein-linked glycosyl chains.

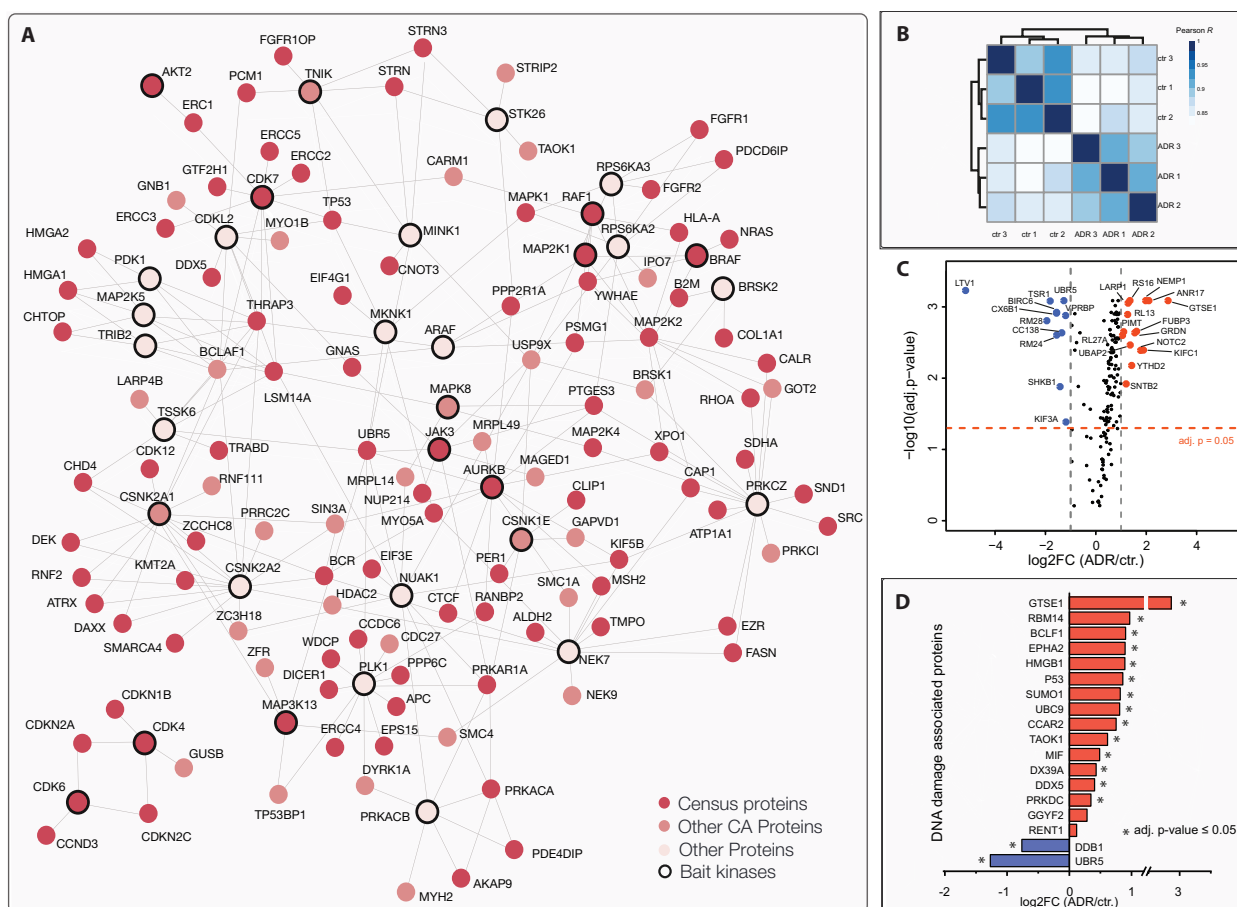


Figure S5. Cancer-associated kinase network, Related to Figure 7. **A.** All significant modules detected at a p -value < 0.05 are shown. This includes the modules depicted in **Figure 7** as well as additional examples. Bold circles indicate kinases with a significant enrichment of CA proteins. Coloring corresponds to the scheme introduced in **Figure 7**. This network highlights several kinases that are not in the Cancer Gene Census, but whose interaction neighborhoods are enriched in CA proteins. **B.** Correlation of the bait-normalized precursor peptide MS1 intensity of the different conditions and three replicates of Dyrk2 BioID-MS experiments shown in **Figure 7** indicated a high reproducibility of the measurements. **C.** Volcano plot is illustrating interaction changes in the Dyrk2 network upon induction of genotoxic stress with Adriamycin (ADR). Upregulated interactions are presented as red dots and downregulated ones as blue dots ($\log_2FC(ADR/ctr) \geq 1$ and $adj. p\text{-value} \leq 0.05$). **D.** Barplot is showing all significant interaction changes to proteins associated with DNA damage related cellular processes (GO: 0006974; “cellular response to DNA damage stimulus”) observed with BioID upon ADR treatment.