

Supplementary Information for

Cxcl9l and Cxcr3.2 regulate recruitment of osteoclast progenitors to bone matrix in a medaka osteoporosis model

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Other supplementary materials for this manuscript include the following:

Movies S1 to S13

A

	baseMean		Fold change RNAseq		Fold change qPCR	
	<i>col10a1</i> 10 dpf	<i>ctsk</i> 12 dpf	<i>col10a1</i> 10 dpf	<i>ctsk</i> 12 dpf	<i>col10a1</i> 10 dpf	<i>ctsk</i> 12 dpf
<i>cxc19l</i>	1575	182	12.85 up	1.4 down	9.1 up *	n. s.
<i>cxcr3.2</i>	30	1695	1.3 down	1.4 up	1.5 up *	6.9 up **

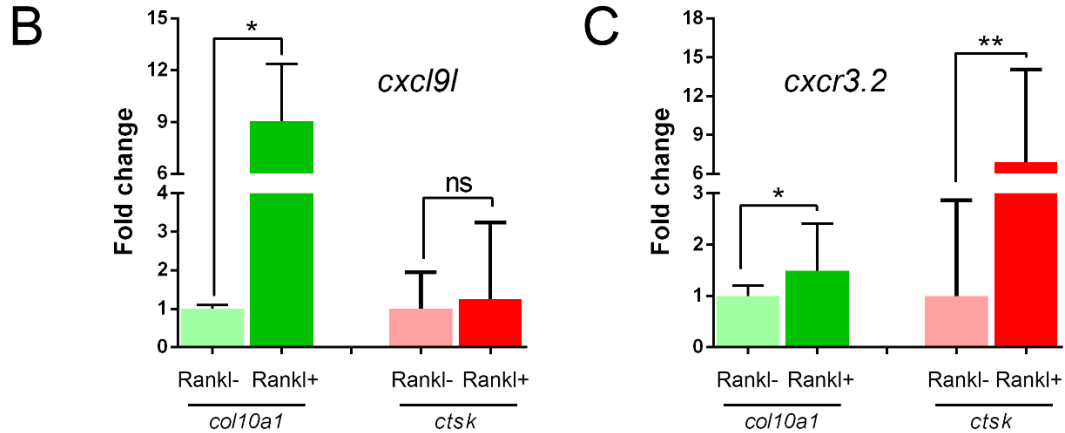


Fig. S1. qPCR validation of *cxc19l* and *cxcr3.2* regulation in osteoblast progenitors (*col10a1*) and osteoclasts (*ctsk*) after Rankl induction. (A) Table summarizing baseMean and fold change values obtained by RNAseq and qPCR analysis, respectively. Samples used for RNAseq and qPCR were from independent experiments. The 'baseMean' value indicates average of transcript numbers in controls and Rankl-induced cell types for both cell types, respectively. (B) qPCR analysis of *cxc19l* transcription in FAC-sorted *col10a1* (at 10 dpf) and *ctsk* cells (at 12 dpf) with and without Rankl induction. (C) qPCR analysis of *cxcr3.2* transcription in FAC-sorted *col10a1* (at 10 dpf) and *ctsk* cells (at 12 dpf) with and without Rankl induction. Three biological and three technical replicates each were analyzed. Error bars show mean value \pm SD; * $p < 0.05$, ** $p < 0.01$; n.s. = no significant difference, Student's t-test.

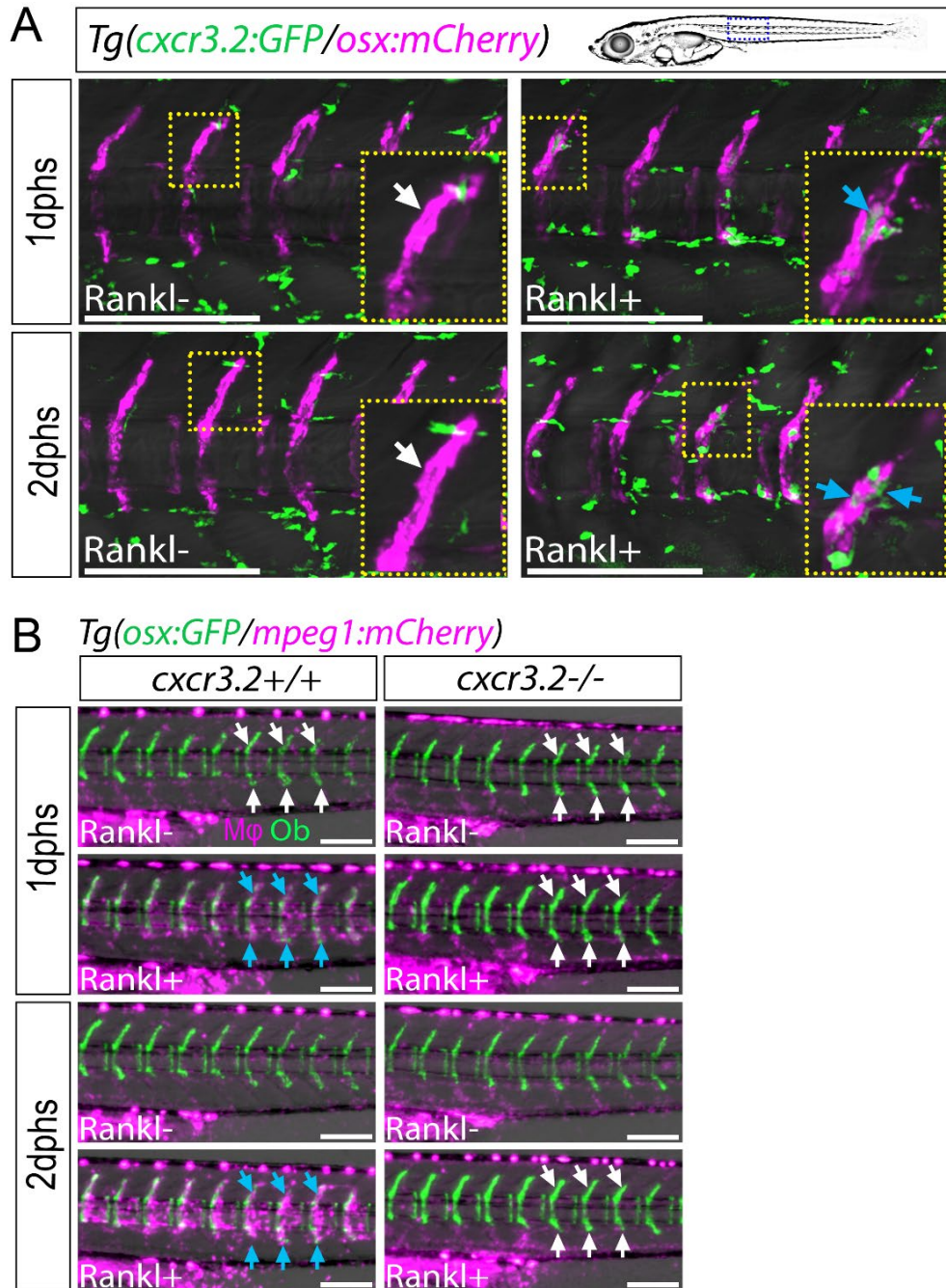


Fig. S2. *Cxcr3.2* expression and function in osteoblasts. (A) *cxcr3.2:GFP* does not overlap with *osx:mCherry* expression in osteoblasts without (left) or with *Rankl* induction (right). Upon *Rankl* induction, *cxcr3.2:GFP* cell numbers are increased in centra and neural arches at 1 and 2 dphs. Note tight interaction of *cxcr3.2:GFP* cells with *osx:mCherry* osteoblasts in arches (blue arrows). Inset shows higher magnification view of boxed area. (B) In the absence of ectopic *Rankl*, *osx:GFP* osteoblasts are similarly distributed in vertebrae of *cxcr3.2*^{+/+} and *cxcr3.2*^{-/-} siblings at 1 and 2 dphs (white arrows). Upon *Rankl* induction, *osx:GFP* cells are redistributed in *cxcr3.2*^{+/+} siblings (blue arrows), as a consequence of ectopic osteoclast differentiation. In contrast, osteoblast distribution in *cxcr3.2*^{-/-} mutants is not affected (right column, white arrows). Scale bars: 200 μ m.

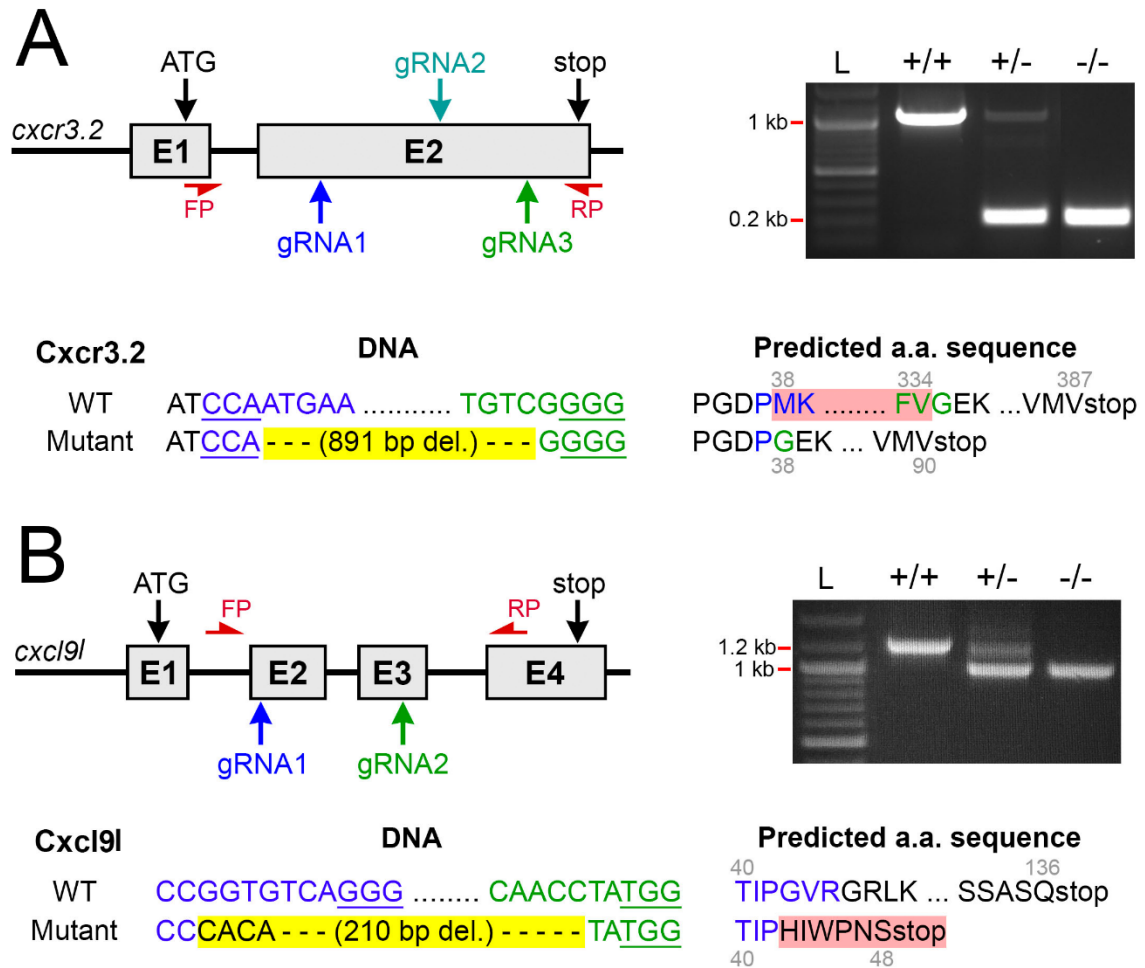


Fig. S3. Generation of medaka *cxcl9l* and *cxcr3.2* mutants. (A) Generation of *cxcr3.2* CRISPR/Cas9 mutants using three guide RNAs (gRNA1-3) targeting exon 2 (E2). Red arrows indicate position of forward (FP) and reverse (RP) primers for PCR genotyping (WT (+/+): approx. 1.1 kb; homozygous mutants (-/-): approx. 0.2 kb; heterozygous carriers (+/-): 1.1 and 0.2 kb). Mutants have a 891 bp deletion (yellow) between gRNA1 and gRNA3 target sites, leading to a predicted a.a. sequence missing aa 38 to 334. (B) Generation of *cxcl9l* mutants. Two guide RNAs (blue, green) targeting *cxcl9l* exon 2 (E2) and exon 3 (E3) were injected. PCR genotyping identified WT (+/+; approx. 1.15 kb), homozygous mutants (-/-; approx. 0.95 kb) and heterozygous siblings (+/-; 1.15 and 0.95 kb). Mutants have a 214 bp indel, leading to a predicted prematurely truncated aa sequence (red). PAM sequences are underlined. Sequences of guide RNAs and primers are listed in Table S5.

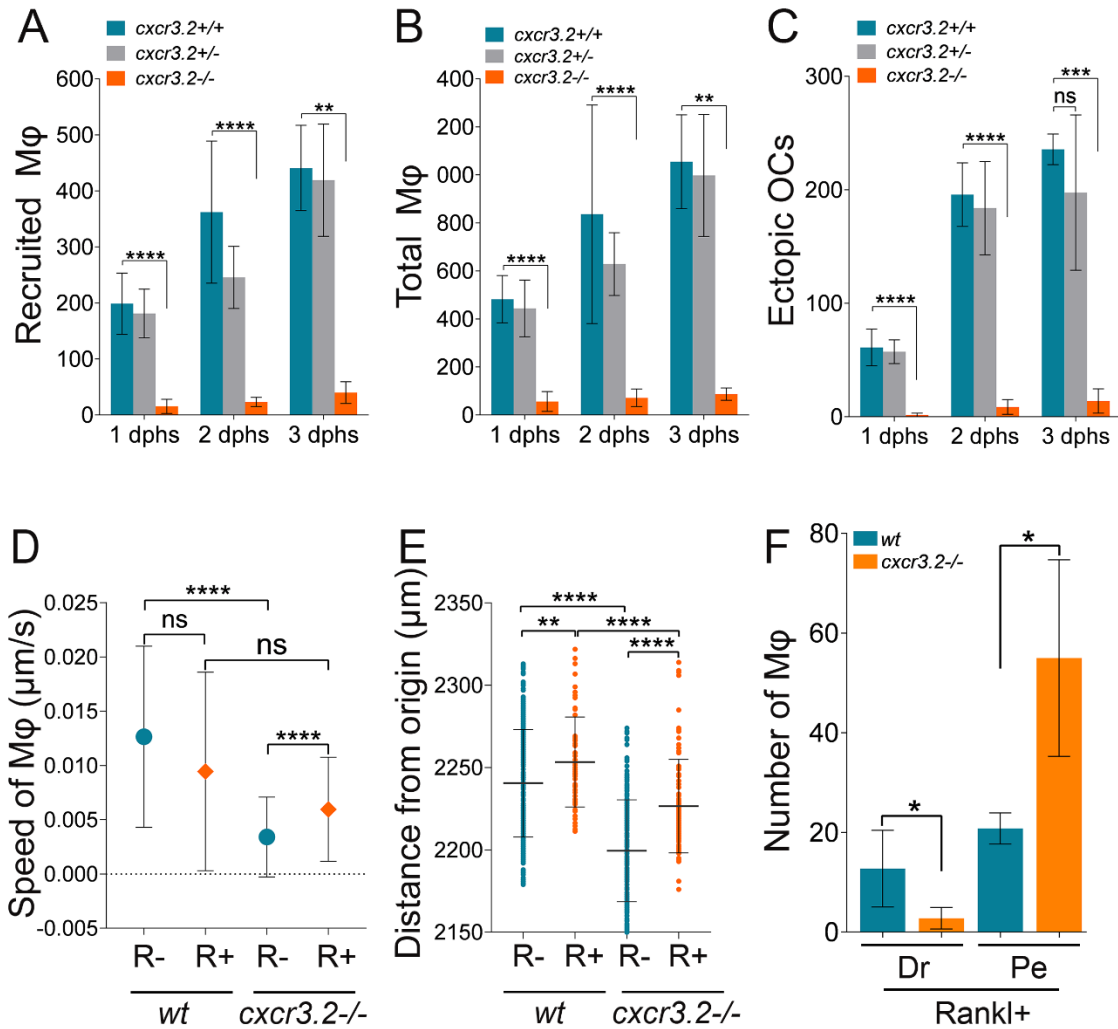


Fig. S4. Quantification of macrophage dynamics and differentiation in *cxcr3.2* mutants. (A-C) Sibling embryos obtained from *cxcr3.2* heterozygous incrosses were induced for Rankl, imaged for cell counting and genotyped. Numbers of recruited macrophages (A), total macrophages (B) and ectopic osteoclasts (C) are shown. Error bars show mean number of cells \pm SD, Mann-Whitney test, ** $p < 0.01$, *** $p < 0.001$, **** $p < 0.0001$, ns: non-significant, $7 \leq N_{cxcr3.2+/+larvae} \leq 13$, $5 \leq N_{cxcr3.2-/-larvae} \leq 20$, from three independent experiments. (D-E) Speed and distance from origin of macrophages were extracted from time-lapse movies at six hours after the start of the movies (i.e. 10 hphs). Although speed and traveling distance of *cxcr3.2-/-* macrophages increased upon Rankl induction, both parameters were lower compared to that of wide type siblings. (F) Number of macrophages moving from AGM towards vertebral column (10-14 hphs), and cells retaining in the AGM (at 14 hphs), measured in three vertebral bodies), and cells retaining in the AGM (at 14 hphs). More macrophages migrated directly from the AGM to the vertebral column in wild type siblings, while most *cxcr3.2-/-* mutant macrophages remained in the AGM. Error bars show mean values \pm SD, Mann-Whitney test, * $p < 0.05$, ** $p < 0.01$, **** $p < 0.0001$, ns: non-significant; $2 \leq N_{movies} \leq 5$. R-: Rankl negative; R+: Rankl positive; Dr: Directed recruitment towards vertebral column; Pe: Persistence in the AGM; Mφ: macrophage; OCs: osteoclasts.

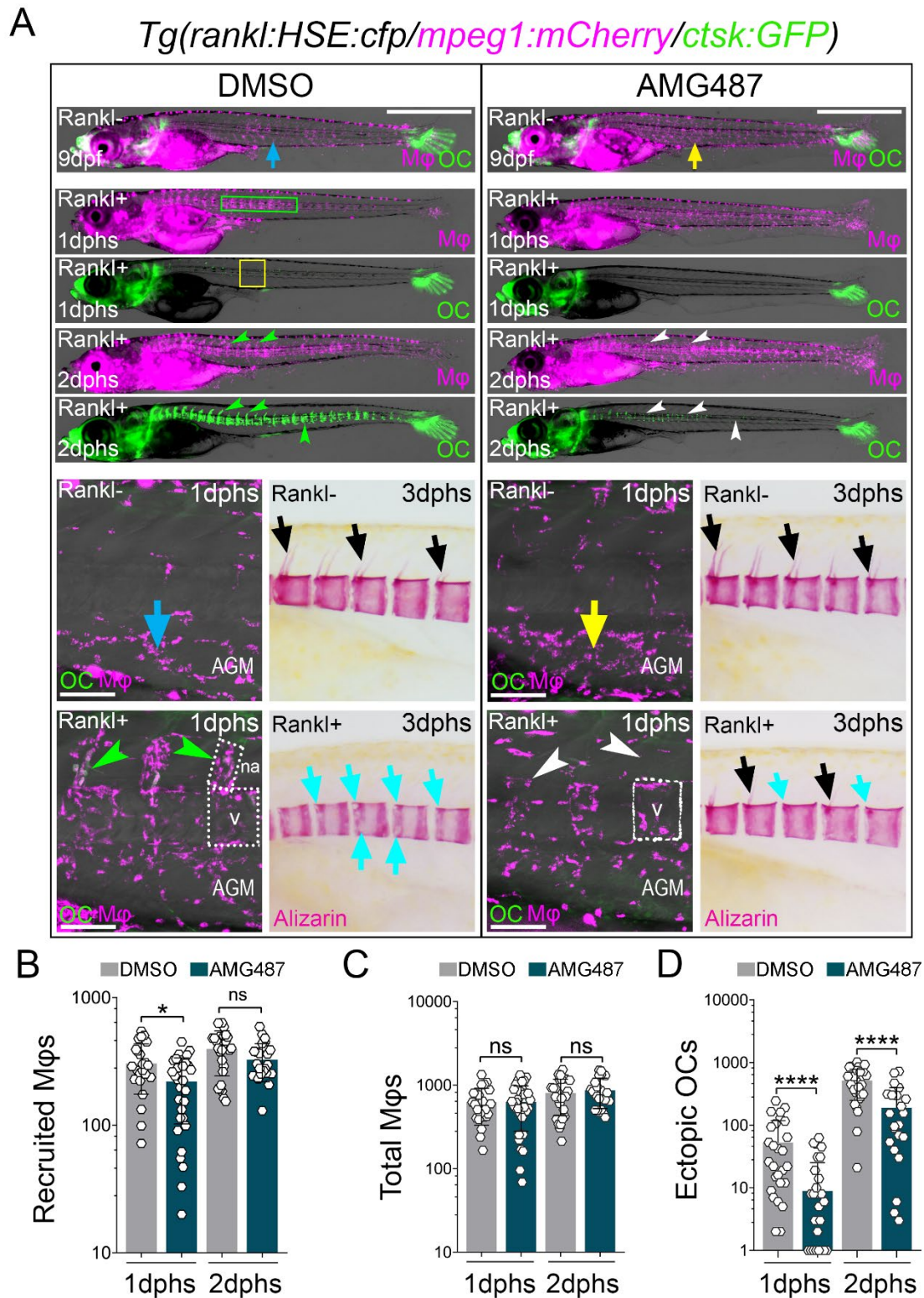


Fig. S5. AMG487 treatment protects bone upon Rankl induction. (A) 20 μ M AMG487 was injected into the yolk of *rankl:HSE:cfp/mpeg1:mCherry/ctsk:GFP* transgenic embryos two hours prior to heat-shock for Rankl induction. In Rankl- embryos, AMG487 treatment does not affect

development but mildly alters macrophage distribution in the AGM (arrows). Upon Rankl induction, recruitment of macrophages and osteoclast formation is considerably reduced in AMG487 treated embryos (white arrowheads at 2 dphs) when compared to controls (green arrowheads). Alizarin Red staining of mineralized matrix shows severe bone lesions in control embryos (cyan arrows) and less lesions after AMG487 treatment (black arrows). Green box indicates region for cell quantification; yellow box shows region imaged with higher magnification. AGM: aorta-gonad-mesonephros, M ϕ : macrophage; OC: osteoclast; na: neural arch, V: vertebra. Scale bar: 1 mm (whole embryo image), 100 μ m (zoom-in image). (B) Number of macrophages recruited to vertebral column. (C) Total number of macrophages in trunk (including AGM). (D) Number of ectopic osteoclasts in vertebral column. Y axes show cell numbers and error bars indicate mean values \pm SD, unpaired two-tailed Student's t-test (B-C), Mann-Whitney test (D), *P<0.05, ****P<0.0001, ns: non-significant, N_{DMSO Larvae} = 23, N_{AMG487 Larvae} = 32, four independent experiments.

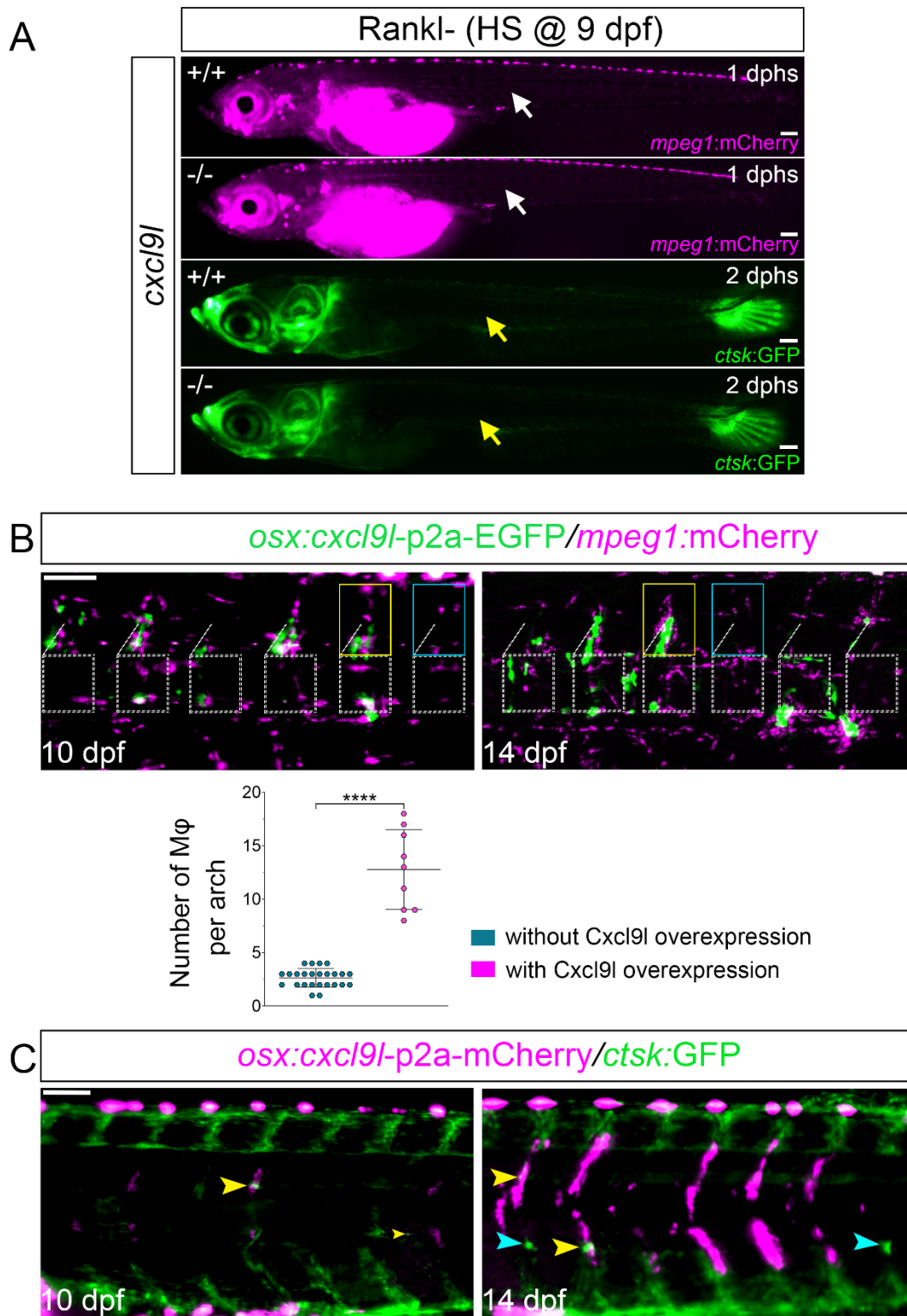
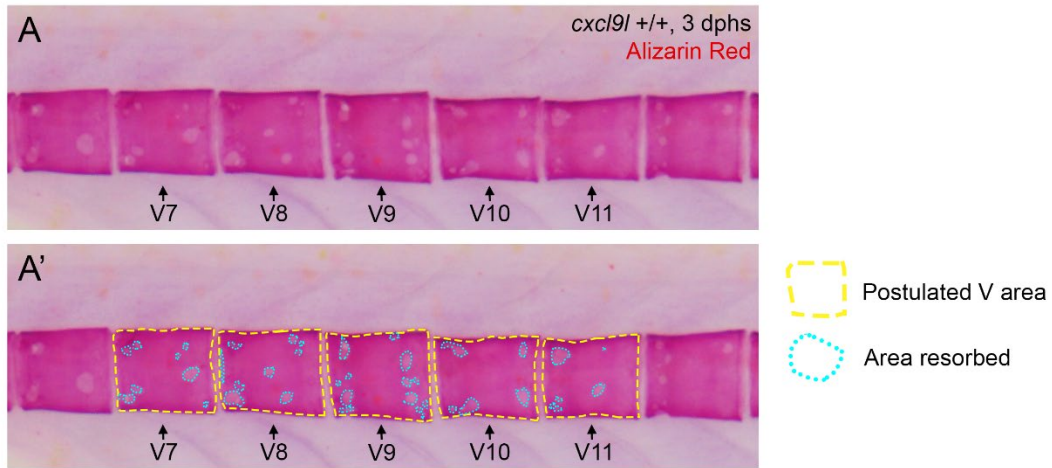


Fig. S6. Macrophage and osteoclast distribution in *cxc19l*^{-/-} mutants without Rankl induction and after ectopic Cxcl9l overexpression. (A) In the absence of Rankl induction, *cxc19l*^{-/-} mutants develop normally and show similar distribution of *mpeg1:mCherry* and *ctsk:GFP* cells as *cxc19l*^{+/+} siblings. Scale bars: 100 μ m. (B) Distribution of *cxc19l-EGFP* expressing cells (green) and

mpeg1:mCherry macrophages (magenta) in vertebral bodies of *osx:cxcl9-p2a-GFP* injected larva at 10 and 14 dpf. Yellow boxes demarcate neural arches with *Cxcl9l* overexpression, cyan boxes indicate neural arch regions without *Cxcl9l* overexpression, white boxes indicate centra of vertebral bodies. Quantification of macrophages in neural arches without and with *Cxcl9l* overexpression. (C). Distribution of *cxcl9l-mCherry* expressing cells (magenta) and *ctsk:GFP* positive osteoclasts (green) in vertebral bodies of *osx:cxcl9-p2a-mCherry* injected larva at 10 and 14 dpf. Yellow arrowheads indicate osteoclasts in vicinity of *Cxcl9l* producing cells, cyan arrowheads label osteoclasts away from *Cxcl9l* producing cells. Scale bars: 100 μm in (B) and (C).



B

$$\% \text{ bone degeneration} = 100\% * \left(\frac{\text{Total area resorbed}}{\text{Total postulated V area}} \right)_{V7-V11}$$

Fig. S7. Quantification of bone degeneration. (A) Example of Alizarin Red stained vertebral column used for quantification of bone degeneration. V7-V11, 7th to 11th vertebral bodies. (A') To quantify bone degeneration, the postulated V area (enclosed by yellow dotted lines) and area resorbed (enclosed by cyan dotted lines) for V7-V11 were determined using ImageJ. (B) Equation for calculation of percentage (%) bone degeneration using values obtained from the analysis indicated in (A').

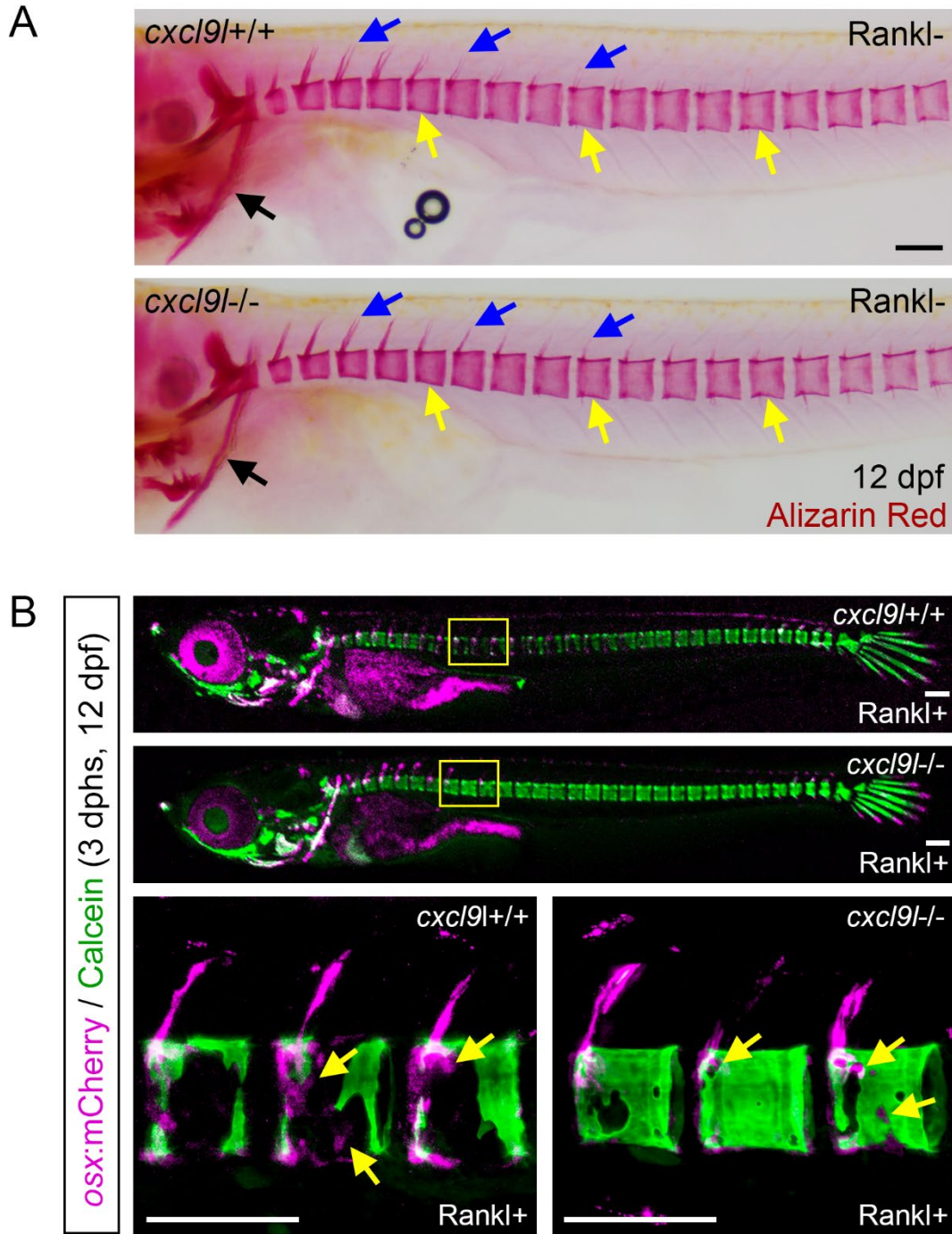


Fig. S8. Bone development in *cxc19l* mutants. (A) In the absence of Rankl induction, Alizarin Red staining revealed normal bone mineralization in the cleithrum (black arrows), neural arches (blue arrows) and vertebral bodies (yellow arrows) of *cxc19l*^{-/-} mutants as compared to *cxc19l*^{+/+} siblings at 12 dpf. (B) After Rankl induction, calcein staining revealed less severe bone lesions in *cxc19l*^{-/-} mutants due to reduced osteoclast formation. Overview and magnified confocal images showing *osx:mCherry*-expressing osteoblasts accumulating at bone lesion sites (arrows) in both *cxc19l*^{+/+} and *cxc19l*^{-/-} siblings (3 dphs, 12 dpf). Yellow boxes indicate regions shown in the magnified views. Scale bars: 100 μ m.

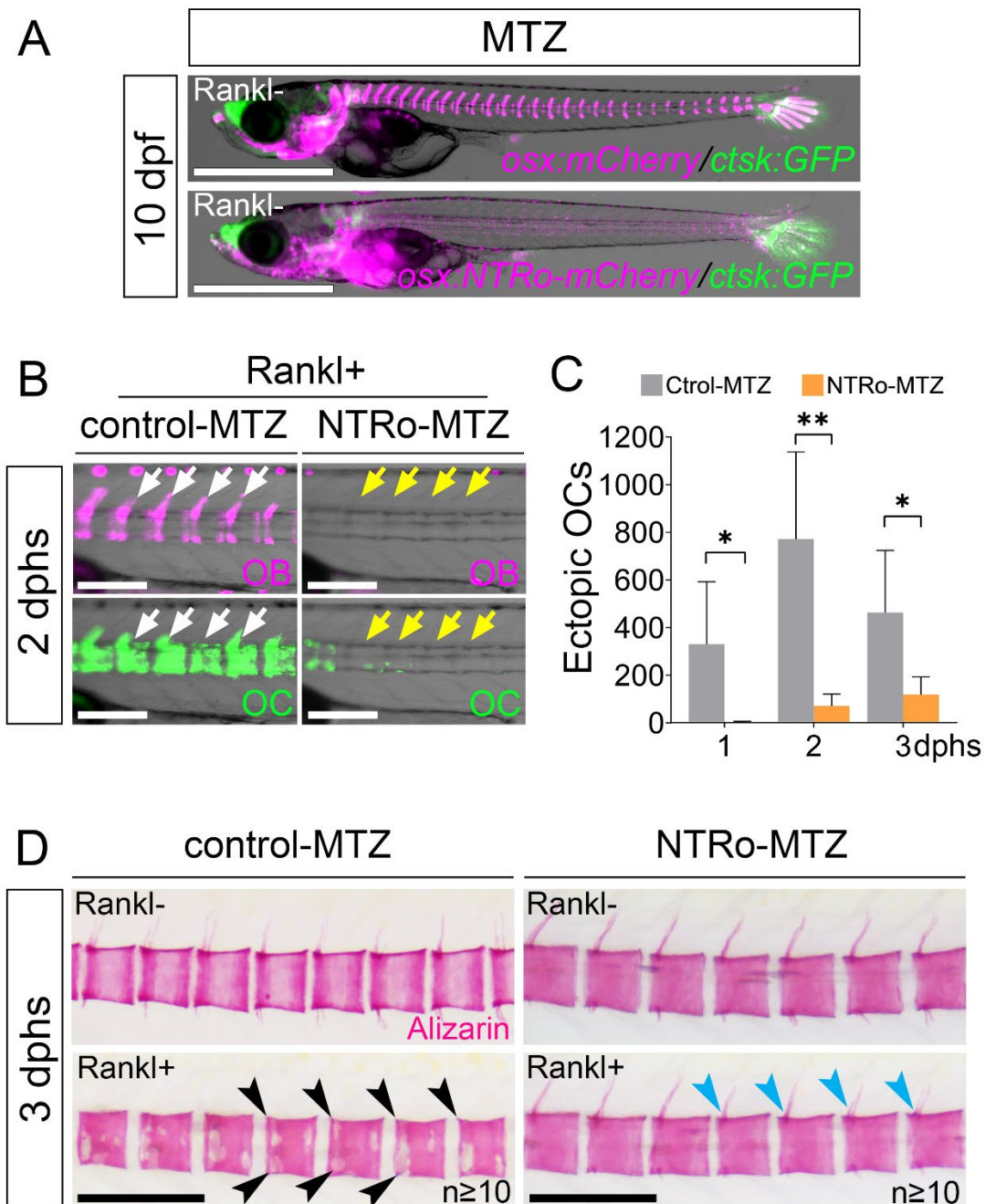


Fig. S9. Osteoclast formation depends on the presence of osteoblasts. (A) Metronidazole (MTZ) treatment results in efficient ablation of osteoblasts in *osx:mCherry-NTRo/ctsk:GFP* embryos when compared to *osx:mCherry/ctsk:GFP* control embryos. (B) After ablation of osteoblasts, Rankl fails to stimulate the formation of ectopic osteoclasts (yellow arrows) compared to massive osteoclast formation in control embryos (white arrows). (C) Quantification of osteoclast numbers showing mean number \pm SD, Brown-Forsythe ANOVA with Games Howell's test, * $p < 0.05$, ** $p < 0.01$, $9 \leq N \leq 11$, from two independent experiments. OB: osteoblast; OC: osteoclast. (D) MTZ treatment does not affect normal bone development (control-MTZ; Rankl-). Ablation of osteoblasts protects bone from lesions by Rankl induction (Rankl+; arrowheads). Scale bars: 1 mm (A), 200 μ m (B-D).

Table S1. Genes exclusively up-regulated in *col10a1* cells or *osx* cells at 10 dpf, or commonly up-regulated in both cell types.

Up-regulated exclusively in "col10 10dpf" (n=32)		Up-regulated in "col10 10dpf" and "osx 10dpf" (n=13)		Up-regulated exclusively in "osx 10dpf" (n=15)	
symbol	description	symbol	description	symbol	description
cd74	HLA class II histocompatibility antigen gamma chain	itgb8	integrin beta-8	LOC101160009	calcium/calmodulin-dependent protein kinase type II subunit beta isoform X1
psmb9-like	proteasome subunit%2C beta type 9-like	c6	complement component C6 isoform X1	LOC101160009	calcium/calmodulin-dependent protein kinase type II subunit beta isoform X2
orla-uaa	major histocompatibility complex class I-related gene protein-like precursor	c6	complement component C6 isoform X2	LOC101160009	calcium/calmodulin-dependent protein kinase type II subunit beta isoform X3
orla-uba	major histocompatibility complex class I-related gene protein-like isoform X1	LOC101156669	collagenase 3	LOC101160009	calcium/calmodulin-dependent protein kinase type II subunit beta isoform X4
orla-uba	major histocompatibility complex class I-related gene protein-like precursor	LOC101168980	C-X-C motif chemokine 2	LOC101160009	calcium/calmodulin-dependent protein kinase type II subunit beta isoform X5
LOC101172593	tumor necrosis factor-like	LOC110016535	no description	LOC101156421	neurofilament light polypeptide
str-3	stromelysin-3 precursor	LOC101159531	tumor necrosis factor-like	relt	tumor necrosis factor receptor superfamily member 19L
LOC101169056	C5a anaphylatoxin chemotactic receptor 1-like	LOC101157923	insulin-like growth factor-binding protein 5	cutc	copper homeostasis protein cutC homolog
LOC101163430	interferon regulatory factor 1 isoform X1	LOC101162522	bactericidal permeability-increasing protein	LOC101166002	uncharacterized protein
LOC101163430	interferon regulatory factor 1 isoform X2	cdk5rap1	CDK5 regulatory subunit-associated protein 1 isoform X1	LOC111949221	no description
LOC101171022	transcription regulator protein BACH1	cdk5rap1	CDK5 regulatory subunit-associated protein 1 isoform X2	otog	otogelin isoform X1
LOC101163059	heme-binding protein 2	mmp9	matrix metalloproteinase-9 precursor	otog	otogelin isoform X2
LOC101166736	disintegrin and metalloproteinase domain-containing protein 8	nfe2	transcription factor NF-E2 45 kDa subunit	LOC101162414	adenylate cyclase type 8-like
steap4	metalloreductase STEAP4			LOC101160286	heme-binding protein 1
mmp14	matrix metalloproteinase-14 precursor			LOC111947687	no description
LOC101163459	mamu class II histocompatibility antigen DR alpha chain				

adamts5	A disintegrin and metalloproteinase with thrombospondin motifs 5
map3k8	mitogen-activated protein kinase kinase kinase 8
tank	TRAF family member-associated NF-kappa-B activator
orla-b2m	b2-microglobulin isoform X1
orla-b2m	b2-microglobulin precursor
LOC101156150	HLA class II histocompatibility antigen%2C DP alpha 1 chain
prdm1b	uncharacterized protein LOC101173518
prdm1b	uncharacterized protein LOC101173518 isoform X1
tnfrsf21	tumor necrosis factor receptor superfamily member 21
LOC110017367	no description
cd276	CD276 antigen
LOC101166708	podocin
alpl	alkaline phosphatase%2C tissue-nonspecific isozyme
LOC101170656	olfactomedin-4
il34	interleukin-34
mafb	transcription factor MafB

Table S2. Top up-regulated genes in *col10a1* cells at 10 dpf (log2 FC >1; baseMean >= 10; adj p-value < 0.05).

geneID	baseMean	log2 FC	p adj	symbol	description
gene11555	2593.35	4.11	0.00	cd74	HLA class II histocompatibility antigen gamma chain
gene12485	1019.15	3.53	0.00	itgb8	integrin beta-8
gene12651	16.64	2.97	0.03	psmb9-like	proteasome subunit%2C beta type 9-like
gene12654	399.86	1.63	0.04	orla-uaa	major histocompatibility complex class I-related gene protein-like precursor
gene12655	453.16	1.85	0.00	orla-uba	major histocompatibility complex class I-related gene protein-like isoform X1
gene12655	453.16	1.85	0.00	orla-uba	major histocompatibility complex class I-related gene protein-like precursor
gene12867	44.57	4.71	0.00	LOC101172593	tumor necrosis factor-like
gene13945	179.45	1.57	0.00	str-3	stromelysin-3 precursor
gene14071	2666.33	4.77	0.00	c6	complement component C6 isoform X1
gene14071	2666.33	4.77	0.00	c6	complement component C6 isoform X2
gene15152	131.20	3.28	0.01	LOC101169056	C5a anaphylatoxin chemotactic receptor 1-like
gene15223	1052.85	5.91	0.00	LOC101156669	collagenase 3
gene15559	306.69	1.32	0.00	LOC101163430	interferon regulatory factor 1 isoform X1
gene15559	306.69	1.32	0.00	LOC101163430	interferon regulatory factor 1 isoform X2
gene15803	205.87	2.06	0.00	LOC101171022	transcription regulator protein BACH1
gene16569	516.26	2.47	0.03	LOC101163059	heme-binding protein 2
gene17282	1574.72	3.68	0.00	LOC101168980	C-X-C motif chemokine 2
gene17283	822.76	2.15	0.02	LOC110016535	no description
gene17293	197.66	3.02	0.03	LOC101166736	disintegrin and metalloproteinase domain-containing protein 8
gene18053	3924.33	1.45	0.00	steap4	metalloreductase STEAP4
gene18583	884.44	4.54	0.00	LOC101159531	tumor necrosis factor-like
gene19596	4639.77	1.81	0.02	mmp14	matrix metalloproteinase-14 precursor
gene2037	1005.57	4.64	0.00	LOC101157923	insulin-like growth factor-binding protein 5
gene21042	162.58	3.41	0.03	LOC101163459	mamu class II histocompatibility antigen%2C DR alpha chain
gene2216	903.66	1.65	0.00	adamts5	A disintegrin and metalloproteinase with thrombospondin motifs 5
gene22512	498.72	2.23	0.00	map3k8	mitogen-activated protein kinase kinase kinase 8
gene23741	289.11	1.72	0.01	tank	TRAF family member-associated NF-kappa-B activator
gene25463	821.34	1.85	0.00	orla-b2m	b2-microglobulin isoform X1
gene25463	821.34	1.85	0.00	orla-b2m	b2-microglobulin precursor
gene2562	431.02	3.75	0.00	LOC101156150	HLA class II histocompatibility antigen%2C DP alpha 1 chain
gene26177	35.32	2.79	0.04	prdm1b	uncharacterized protein LOC101173518
gene26177	35.32	2.79	0.04	prdm1b	uncharacterized protein LOC101173518 isoform X1
gene26348	832.01	2.12	0.01	tnfrsf21	tumor necrosis factor receptor superfamily member 21

gene2926	19.73	3.19	0.01	LOC110017367	no description
gene3342	2116.96	1.27	0.00	cd276	CD276 antigen
gene4117	75.73	2.86	0.00	LOC101166708	podocin
gene5496	4243.16	4.38	0.00	LOC101162522	bactericidal permeability-increasing protein
gene5497	737.83	3.33	0.00	cdk5rap1	CDK5 regulatory subunit-associated protein 1 isoform X1
gene5497	737.83	3.33	0.00	cdk5rap1	CDK5 regulatory subunit-associated protein 1 isoform X2
gene5832	1363.09	1.75	0.04	alpl	alkaline phosphatase%2C tissue-nonspecific isozyme
gene6029	110.24	4.09	0.01	LOC101170656	olfactomedin-4
gene6853	246.58	3.14	0.00	il34	interleukin-34
gene7554	14765.11	5.17	0.00	mmp9	matrix metalloproteinase-9 precursor
gene7663	2198.70	2.65	0.00	nfe2	transcription factor NF-E2 45 kDa subunit
gene7750	1791.19	1.48	0.01	mafb	transcription factor MafB

Table S3: Top up-regulated genes in osx cells at 10 dpf (log2 FC >1; baseMean >= 10; adj p-value < 0.05).

geneID	baseMean	logFC	p adj	symbol	description
gene12485	523.76	3.00	0.00	itgb8	integrin beta-8
gene13252	123.30	1.78	0.01	LOC101160009	calcium/calmodulin-dependent protein kinase type II subunit beta isoform X1
gene13252	123.30	1.78	0.01	LOC101160009	calcium/calmodulin-dependent protein kinase type II subunit beta isoform X2
gene13252	123.30	1.78	0.01	LOC101160009	calcium/calmodulin-dependent protein kinase type II subunit beta isoform X3
gene13252	123.30	1.78	0.01	LOC101160009	calcium/calmodulin-dependent protein kinase type II subunit beta isoform X4
gene13252	123.30	1.78	0.01	LOC101160009	calcium/calmodulin-dependent protein kinase type II subunit beta isoform X5
gene13829	10.25	4.03	0.00	LOC101156421	neurofilament light polypeptide
gene14071	1334.32	2.02	0.00	c6	complement component C6 isoform X1
gene14071	1334.32	2.02	0.00	c6	complement component C6 isoform X2
gene15005	14.41	3.68	0.01	re1t	tumor necrosis factor receptor superfamily member 19L
gene15223	772.16	2.84	0.04	LOC101156669	collagenase 3
gene17248	61.84	3.05	0.03	cutc	copper homeostasis protein cutC homolog
gene17282	1396.00	3.85	0.00	LOC101168980	C-X-C motif chemokine 2
gene17283	763.32	1.97	0.01	LOC110016535	no description
gene18056	299.45	1.80	0.00	LOC101166002	uncharacterized protein LOC101166002
gene18583	1181.39	3.73	0.00	LOC101159531	tumor necrosis factor-like
gene2037	451.82	4.46	0.00	LOC101157923	insulin-like growth factor-binding protein 5
gene20621	29.80	3.87	0.01	LOC111949221	no description
gene2435	68.68	3.63	0.00	otog	otogelin isoform X1
gene2435	68.68	3.63	0.00	otog	otogelin isoform X2
gene26204	21.14	2.94	0.04	LOC101162414	adenylate cyclase type 8-like
gene5496	3946.75	1.95	0.02	LOC101162522	bactericidal permeability-increasing protein
gene5497	718.80	1.68	0.00	cdk5rap1	CDK5 regulatory subunit-associated protein 1 isoform X1
gene5497	718.80	1.68	0.00	cdk5rap1	CDK5 regulatory subunit-associated protein 1 isoform X2
gene7554	9157.18	3.00	0.01	mmp9	matrix metalloproteinase-9 precursor
gene7663	1796.27	1.68	0.00	nfe2	transcription factor NF-E2 45 kDa subunit
gene8	148.46	1.44	0.02	LOC101160286	heme-binding protein 1
gene8360	13.82	4.49	0.00	LOC111947687	no description

Table S4: Top regulated genes in *ctsk* cells at 12 dpf (log2 FC <1/>1; baseMean >= 10; adj p-value < 0.05).

Down-regulated						Up-regulated					
geneID	baseMean	logFC	p adj	symbol	description	geneID	baseMean	logFC	p adj	symbol	description
gene10078	297.84	-1.21	0.04	LOC101161893	no description	gene12655	154.29	1.56	0.00	orla-uba	major histocompatibility complex class I-related gene protein-like precursor
gene11327	241.08	-1.38	0.00	gdpd2	glycerophosphoinositol inositolphosphodiesterase GDPD2	gene151	111.54	1.06	0.04	LOC111947943	histone H1-like
gene11841	79.20	-1.41	0.04	LOC101159027	N-acetyllactosaminide beta-acetylglucosaminyltransferase 2	gene2304	98.30	2.42	0.01	cfap47	cilia- and flagella-associated protein 47 isoform X1
gene1186	180844.67	-1.08	0.01	LOC101159648	keratin%2C type I cytoskeletal 13	gene4068	1080.94	1.05	0.03	comp	cartilage oligomeric matrix protein isoform X1
gene13325	552.45	-1.29	0.04	LOC101174360	proteinase-activated receptor 2	gene4068	1080.94	1.05	0.03	comp	cartilage oligomeric matrix protein isoform X2
gene14417	378.81	-1.02	0.01	LOC101167722	suppressor of tumorigenicity 14 protein homolog	gene6944	137.50	1.45	0.00	b3gnt9	UDP-GlcNAc:betaGal beta-acetylglucosaminyltransferase 9
gene14893	30.76	-2.71	0.00	LOC101175669	thymus-specific serine protease-like	gene7162	2397.26	1.23	0.01	ddost	dolichyldiphosphooligosaccharide—protein glycosyltransferase 48 kDa subunit
gene15169	33.74	-2.14	0.03	LOC101171510	CMP-N-acetylneuraminat-beta-galactosamide-alpha-sialyltransferase 4-like						
gene15879	75.86	-1.94	0.04	zpld1	zona pellucida-like domain-containing protein 1 isoform X1						
gene15879	75.86	-1.94	0.04	zpld1	zona pellucida-like domain-containing protein 1 isoform X2						
gene15949	8465.15	-1.25	0.00	LOC101165663	lipocalin						
gene16700	139.57	-1.11	0.02	LOC101156521	hydroxycarboxylic acid receptor 2 isoform X1						
gene16700	139.57	-1.11	0.02	LOC101156521	hydroxycarboxylic acid receptor 3 isoform X2						
gene16751	63.96	-1.52	0.01	LOC105358576	B-cadherin-like						
gene18443	78.71	-1.52	0.01	LOC101168736	cannabinoid receptor 2						

gene18722	231.12	-1.85	0.03	LOC10535 7484	apoptosis-associated speck- like protein containing a CARD
gene18735	526.23	-1.12	0.04	LOC10116 2953	uncharacterized protein LOC101162953 isoform X1
gene18735	526.23	-1.12	0.04	LOC10116 2953	uncharacterized protein LOC101162953 isoform X2
gene18735	526.23	-1.12	0.04	LOC10116 2953	uncharacterized protein LOC101162953 isoform X3
gene18735	526.23	-1.12	0.04	LOC10116 2953	uncharacterized protein LOC101162953 isoform X4
gene18735	526.23	-1.12	0.04	LOC10116 2953	uncharacterized protein LOC101162953 isoform X5
gene20384	27.73	-2.04	0.03	LOC10115 6610	uncharacterized protein LOC101156610 isoform X1
gene20384	27.73	-2.04	0.03	LOC10115 6610	uncharacterized protein LOC101156610 isoform X2
gene20677	45.34	-1.41	0.04	LOC10116 1585	ephexin-1
gene21132	337.74	-1.20	0.02	LOC10117 4954	nectin-4 isoform X1
gene21132	337.74	-1.20	0.02	LOC10117 4954	nectin-4 isoform X2
gene21239	85.81	-2.26	0.04	LOC10535 8224	no description
gene21278	195.27	-1.07	0.02	LOC11194 6396	no description
gene21279	105.51	-1.29	0.01	LOC11194 6395	no description
gene22836	26.24	-3.16	0.00	LOC10116 5601	uncharacterized protein LOC101165601
gene24481	205.32	-1.31	0.01	LOC10535 6917	interleukin-31 receptor subunit alpha-like
gene26296	43.82	-2.02	0.03	LOC10117 0320	sodium-dependent glucose transporter 1-like
gene2654	123.55	-1.10	0.03	ano1	anoctamin-1 isoform X1
gene2654	123.55	-1.10	0.03	ano1	anoctamin-1 isoform X2
gene2654	123.55	-1.10	0.03	ano1	anoctamin-1 isoform X3
gene26752	300.20	-1.30	0.02	rhov	rho-related GTP-binding protein RhoV

gene3536	35.81	-1.67	0.03	LOC10116 3707	aerolysin-like protein
gene3539	168.60	-1.25	0.01	LOC11001 3331	aerolysin-like protein
gene3541	37.85	-1.67	0.04	LOC10116 0881	aerolysin-like protein
gene4324	1311.78	-1.34	0.00	LOC10116 5224	epiplakin isoform X1
gene4324	1311.78	-1.34	0.00	LOC10116 5224	epiplakin isoform X2
gene4467	4415.93	-1.41	0.00	LOC10117 3929	dimethylaniline monooxygenase [N-oxide- forming] 5
gene6482	613.99	-1.16	0.00	LOC10116 5389	plakophilin-3 isoform X1
gene6482	613.99	-1.16	0.00	LOC10116 5389	plakophilin-3 isoform X2
gene6958	285.25	-1.46	0.03	LOC10117 5077	carbohydrate sulfotransferase 6-like
gene7240	175297.4 0	-1.06	0.00	LOC10116 7707	intermediate filament protein ON3
gene7832	130.39	-1.44	0.02	il17rc	interleukin-17 receptor C
gene8486	375.87	-1.42	0.04	LOC10115 6896	tryptase

Table S5: Used oligos.

Name of oligo	Sequence 5' to 3'	Application
mpeg1.2F	ATAGCGAGCGGATCCAGAACAGTTTCTGGCAGAAGTTAGTC	Promoter cloning
mpeg1.2R	AATATGCGGCCGCTGAATTCATCCATGTTCCGCAGGTTGTC	Promoter cloning
cxcl9IUP	TGATCCGCTGCTGGGATAGA	qPCR
cxcl9IDOWN	TTAGTCCCAAGGGCTTGTC	qPCR
cxcr3.2UP	CTAATATGGGTTACACACATCCACCAC	qPCR
cxcr3.2Down	CTCAGAACAAGGGGCTCCAATC	qPCR
β -actinUP	GCCAACAGGGGAGAAGATGAC	qPCR
β -actinDOWN	CATCACCAGAGTCCATGACG	qPCR
cxcl9IF	GTTTAGCTTGTCTCCCGGGCAGGCCACC	Full length cDNA
cxcl9IR	TTGTGAGGCTGAGGAGGA	Full length cDNA
cxcl9I crRNA1	TCAGACAATACCCGGTGTCAAGG	CRISPR mutant
cxcl9I crRNA2	TTTGAATCCAGAGCAACCTATGG	CRISPR mutant
cxcr3.2 crRNA1	CCAATGAAGATTGGAGCCCCCTTG	CRISPR mutant
cxcr3.2 crRNA2	CCTCGGTTTAAGCGGAGTGGACA	CRISPR mutant
cxcr3.2 crRNA3	TCTGCTGTACGGATTTGTCGGGG	CRISPR mutant
cxcl9IgenoF	TCTGTATAGTGAGGGAAACGGACGG	genotyping
cxcl9IgenoR	CCTGGGCTTTCTTTTGAACAATGC	genotyping
cxcr3.2genoF	TGTTTGTGCAGAAAATAGGCATTCC	genotyping
cxcr3.2genoR	TCGGAGCCTCTTTCCTTTCC	genotyping

Movie S1. Dynamics of *cxcr3.2:GFP* positive cells in medaka embryo without Rankl induction. A *cxcr3.2:GFP/mpeg1:mCherry* transgenic embryo was heat-shocked at 9 dpf (2 hours, 39°C). Time-lapse started at 2 hphs. The majority of macrophages remained in the AGM while single cells were positioned in neural arches and centra. Most *mpeg1:mCherry* macrophages express *cxcr3.2* but at different levels. Box: vertebral body; VC: vertebral column; AGM: aorta-gonad-mesonephros; time: hh-mm-ss. Scale bar: 50 µm.

Movie S2. Dynamics of *cxcr3.2:GFP* positive cells in medaka embryo after Rankl induction. A *rankl:HSE:cfp/cxcr3.2:GFP/mpeg1:mCherry* transgenic embryo was heat-shocked at 9 dpf (2 hours, 39°C). Time-lapse started at 2 hphs. Upon Rankl induction, *cxcr3.2:GFP* cells migrate towards the mineralized matrix of the vertebral column, where they differentiate into osteoclasts (see Movie S3). Scale bar: 50 µm.

Movie S3. Differentiation of *cxcr3.2:GFP* cells into *ctsk:mCherry* osteoclasts. A *rankl:HSE:cfp/cxcr3.2:GFP/ctsk:mCherry* transgenic embryo was heat-shocked at 9 dpf (2 hours, 39°C). Time-lapse started at 24 hphs. The recruited *cxcr3.2* positive cells gradually express *ctsk:mCherry* during osteoclast maturation (yellow arrowheads). Time: hh-mm-ss. Scale bar: 50 µm.

Movie S4. Downregulation of *cxcr3.2:GFP* in maturing osteoclasts. A *rankl:HSE:cfp/cxcr3.2:GFP/ctsk:mCherry* transgenic embryo was heat-shocked at 9 dpf (2 hours, 39°C). Time-lapse started at 1 dphs. *cxcr3.2:GFP* expression in *ctsk:mCherry* positive cells becomes weaker with time (white arrow). Note cellular protrusions characteristic for maturing osteoclast. Time: hh-mm-ss. Scale bar: 50 µm.

Movie S5. Macrophage (*cxcr3.2+/+*) dynamics in Rankl- embryo. A *mpeg1:mCherry* transgenic embryo was heat-shocked at 9 dpf (2 hours, 39°C). Time-lapse was started at 5 hphs. Most macrophages reside in the AGM, while only few cells are located in neural arches and centra. Scale bar: 50 µm.

Movie S6. Macrophage (*cxcr3.2+/+*) dynamics in Rankl+ embryo. A *rankl:HSE:cfp/mpeg1:mCherry* transgenic embryo was heat-shocked at 9 dpf (2 hours, 39°C). Time-lapse started at 5 hphs. Upon Rankl induction, increasing numbers of macrophages are present in vertebral bodies. Scale bar: 50 µm.

Movie S7. Mutant macrophage (*cxcr3.2-/-*) dynamics in Rankl- embryo. A *cxcr3.2-/-* mutant embryo with *mpeg1:mCherry* transgenic background was heat-shocked at 9 dpf (2 hours, 39°C). Time-lapse started at 5 hphs. Most macrophages reside in the AGM with only few cells located in neural arches and centra. Compared to wild-type macrophages, *cxcr3.2-/-* macrophages appear less dynamic, especially in the AGM. Scale bar: 50 µm.

Movie S8. Mutant macrophage (*cxcr3.2-/-*) migration in Rankl+ embryo. A *cxcr3.2-/-* mutant embryo with a *rankl:HSE:cfp/mpeg1:mCherry* transgenic background was heat-shocked at 9 dpf (2 hours, 39°C). Time-lapse was started at 5 hphs. Upon Rankl induction, *cxcr3.2-/-* macrophages become more motile but do not move towards vertebral column. Most cells remain in the AGM. Scale bar: 50 µm.

Movie S9. Macrophages (*cxcr3.2+/+*) clear *E. coli* cells. A *cxcr3.2+/+* wild-type sibling with *mpeg1:mCherry* transgenic background was injected into muscle with non-pathogenic *E.coli* K12

expressing GFP. Time-lapse was started at 20 minutes post infection (mpi). White arrows indicate macrophages engulfing and clearing bacteria. Time: hh-mm-ss.

Movie S10. Macrophages (*cxcr3.2+/-*) clear *E. coli* cells. Same set-up as in Movie S9 but with a *cxcr3.2+/-* embryo. Time: hh-mm-ss.

Movie S11. Mutant macrophages (*cxcr3.2-/-*) clear *E. coli* cells. Same set-up as in Movies S9 and S10 but with a *cxcr3.2-/-* embryo. Time: hh-mm-ss.

Movie S12. Macrophages (*cxcl9+/-*) clear *E. coli* cells. Same set-up as in Movies S9-S11 but with a heterozygous *cxcl9+/-* sibling with *mpeg1:mCherry* transgenic background. Time: hh-mm-ss.

Movie S13. Mutant macrophages (*cxcl9-/-*) clear *E. coli* cells. Same set-up as in Movies S9-S12 but with a *cxcl9-/-* sibling with *mpeg1:mCherry* transgenic background. Time: hh-mm-ss.