

## SUPPLEMENTAL TABLES

Supplemental Table 1. Extracellular forms of PD-L1 and outcomes in multiple malignancies.

	High sPD-L1 prognosis	PD-L1 protein-to-mRNA prognosis <sup>1</sup>	First Line IO	Non-First Line IO
<b>Adrenocortical</b>		↑		
<b>Bladder</b>		↑	Cisplatin-ineligible CPS10: P <sup>2</sup> A <sup>3</sup>	P <sup>2</sup> N <sup>4</sup>
<b>Breast</b>	Poor <sup>5</sup>	↑	TNBC unresectable PD-L1 > 1%: A <sup>3</sup>	
<b>Biliary</b>	Poor <sup>6</sup>			
<b>Cervical</b>		↗		P <sup>2</sup>
<b>Colon</b>			MSI-high: P <sup>2</sup>	
<b>Gastric</b>	Mainly poor <sup>7-11</sup> Poor in IO <sup>12</sup> Poor with EVs <sup>13</sup>	↓		P <sup>2</sup>
<b>Glioma (low-grade)</b>		↑		
<b>Head/Neck SCC</b>		↗	P <sup>2</sup>	N <sup>4</sup>
<b>Hepatocellular</b>	Poor DFS and OS <sup>14-18</sup> or equivocal <sup>19</sup>	↑		P <sup>2</sup> N <sup>4</sup>
<b>Lung</b>	Poor <sup>20-23</sup> Poor response in IO <sup>12,24,25</sup>		Metastatic NSCLC no muts: P <sup>2</sup> A <sup>3</sup> SCLC: A <sup>3</sup>	P <sup>2</sup> N <sup>4</sup>
<b>Lymphoma</b>	BCL poor OS <sup>26</sup> poor response <sup>27</sup> decrease on tx positive <sup>28</sup> HL poor PFS <sup>29</sup> PTCL poor OS/PFS/response <sup>30,31</sup> NK/T poor OS/PFS <sup>32,33</sup>	↗		HL: P <sup>2</sup> N <sup>4</sup>
<b>Melanoma</b>	Poor <sup>34</sup> Poor in IO <sup>35</sup>	↗	Unresectable/metastatic: P <sup>2</sup> N <sup>4</sup> Adjuvant if LNs after resection: P <sup>2</sup>	
<b>Mesothelioma</b>		↑		
<b>Myeloma</b>	Poor PFS <sup>36,37</sup>			
<b>Ovarian (epithelial)</b>	Poor OS and PFS in cisplatin-resp <sup>38</sup>			
<b>Pancreatic</b>	Decrease on tx positive <sup>39</sup>			
<b>Prostate</b>		↗		
<b>Rectal</b>	Poor DFS <sup>40</sup>			
<b>Renal</b>		↗	Advanced with axitinib: P <sup>2</sup>	N <sup>4</sup>
<b>Sarcoma</b>		↑		

Supplemental Table 1. Extracellular PD-L1 and survival. Cancers for which serum sPD-L1 and/or PD-L1 protein-to-mRNA ratios affect(s) prognosis in a previous study of Cancer Genome

Atlas (TCGA) data (increased improves survival)<sup>1</sup> and/or in which first-line or common second-line PD-(L)1 inhibitor therapy is FDA approved as a partial list. OS: overall survival. PFS: progression-free survival. IO: immunotherapy. A: atezolizumab. N: nivolumab. P: pembrolizumab. References:

1. Orme, J. et al. ADAM10 and ADAM17 cleave PD-L1 to mediate PD-(L)1 inhibitor resistance. *Oncoimmunology* **In press**, DOI 10.1080/2162402X.2020.1744980 (2020).
2. *Keytruda: Highlights of Prescribing Information*. (2017).
3. Genentech. *Atezolizumab: Highlights of Prescribing Information*. (2018).
4. *Nivolumab: Highlights of Prescribing Information*. (2018).
5. Li, Y. et al. Serum sPD-1 and sPD-L1 as Biomarkers for Evaluating the Efficacy of Neoadjuvant Chemotherapy in Triple-Negative Breast Cancer Patients. *Clin. Breast Cancer* **19**, 326-332.e1 (2019).
6. Ha, H. et al. Soluble programmed death-ligand 1 (sPDL1) and neutrophil-to-lymphocyte ratio (NLR) predicts survival in advanced biliary tract cancer patients treated with palliative chemotherapy. *Oncotarget* **7**, 76604–76612 (2016).
7. Zheng, Z. et al. Level of circulating PD-L1 expression in patients with advanced gastric cancer and its clinical implications. *Chin. J. Cancer Res.* **26**, 104–11 (2014).
8. Ito, M. et al. Is high serum programmed death ligand 1 level a risk factor for poor survival in patients with gastric cancer? *Ann. Gastroenterol. Surg.* (2018). doi:10.1002/ags3.12175
9. Bang, Y.-J. et al. Relationship between PD-L1 expression and clinical outcomes in patients with advanced gastric cancer treated with the anti-PD-1 monoclonal antibody pembrolizumab (MK-3475) in KEYNOTE-012. *J. Clin. Oncol.* **33**, 4001 (2015).
10. Shigemori, T. et al. Soluble PD-L1 Expression in Circulation as a Predictive Marker for Recurrence and Prognosis in Gastric Cancer: Direct Comparison of the Clinical Burden Between Tissue and Serum PD-L1 Expression. *Ann. Surg. Oncol.* **26**, 876–883 (2019).
11. Choi, Y. Y. et al. Microsatellite Instability and Programmed Cell Death-Ligand 1 Expression in Stage II/III Gastric Cancer: Post Hoc Analysis of the CLASSIC Randomized Controlled study. *Ann. Surg.* **270**, 309–316 (2019).
12. Ando, K. et al. Plasma Levels of Soluble PD-L1 Correlate With Tumor Regression in Patients With Lung and Gastric Cancer Treated With Immune Checkpoint Inhibitors. *Anticancer Res.* **39**, 5195–5201 (2019).
13. Fan, Y. et al. Exosomal PD-L1 Retains Immunosuppressive Activity and is Associated with Gastric Cancer Prognosis. *Ann. Surg. Oncol.* **26**, 3745–3755 (2019).
14. Finkelmeier, F. et al. High levels of the soluble programmed death-ligand (sPD-L1) identify hepatocellular carcinoma patients with a poor prognosis. *Z. Gastroenterol.* **53**, A4\_19 (2015).
15. El-Gebaly, F. et al. Study of Serum Soluble Programmed Death Ligand 1 as a prognostic Factor in Hepatocellular Carcinoma in Egyptian Patients. *Curr. Cancer Drug Targets* (2019). doi:10.2174/1568009619666190718141647
16. Chang, B. et al. The correlation and prognostic value of serum levels of soluble programmed death protein 1 (sPD-1) and soluble programmed death-ligand 1 (sPD-L1) in patients with hepatocellular carcinoma. *Cancer Immunol. Immunother.* **68**, 353–363 (2019).
17. Han, X. et al. Pre-treatment serum levels of soluble programmed cell death-ligand 1 predict prognosis in patients with hepatitis B-related hepatocellular carcinoma. *J. Cancer Res. Clin. Oncol.* **145**, 303–312 (2019).

18. Kim, H. J., Park, S., Kim, K.-J. & Seong, J. Clinical significance of soluble programmed cell death ligand-1 (sPD-L1) in hepatocellular carcinoma patients treated with radiotherapy. *Radiother. Oncol.* **129**, 130–135 (2018).
19. Elmezayen, H. A. *et al.* Clinical role of serum programmed death ligand 1 in patients with hepatocellular carcinoma: Where does it come from? *Surg. Today* (2019). doi:10.1007/s00595-019-01920-8
20. Okuma, Y. *et al.* High plasma levels of soluble programmed cell death ligand 1 are prognostic for reduced survival in advanced lung cancer. *Lung Cancer* **104**, 1–6 (2017).
21. Zhang, J. *et al.* Circulating PD-L1 in NSCLC patients and the correlation between the level of PD-L1 expression and the clinical characteristics. *Thorac. Cancer* **6**, 534–538 (2015).
22. Costantini, A. *et al.* Predictive role of plasmatic biomarkers in advanced non-small cell lung cancer treated by nivolumab. *Oncoimmunology* e1452581 (2018). doi:10.1080/2162402X.2018.1452581
23. Zhao, J. *et al.* Plasma levels of soluble programmed death ligand-1 may be associated with overall survival in nonsmall cell lung cancer patients receiving thoracic radiotherapy. *Medicine (Baltimore)*. **96**, e6102 (2017).
24. Costantini, A. *et al.* Predictive role of plasmatic biomarkers in advanced non-small cell lung cancer treated by nivolumab. *Oncoimmunology* **7**, e1452581 (2018).
25. Okuma, Y. *et al.* Soluble Programmed Cell Death Ligand 1 as a Novel Biomarker for Nivolumab Therapy for Non-Small-cell Lung Cancer. *Clin. Lung Cancer* **19**, 410-417.e1 (2018).
26. Rossille, D. *et al.* High level of soluble programmed cell death ligand 1 in blood impacts overall survival in aggressive diffuse large B-Cell lymphoma: results from a French multicenter clinical trial. *Leukemia* **28**, 2367–75 (2014).
27. Rossille, D. *et al.* High level of soluble programmed cell death ligand 1 in blood impacts overall survival in aggressive diffuse large B-Cell lymphoma: results from a French multicenter clinical trial. *Leukemia* **28**, 2367–2375 (2014).
28. El-Ghammaz, A. M. S., Gadallah, H. A., Kamal, G., Maher, M. M. & Mohamad, M. A. Impact of serum soluble programmed death ligand 1 on end of treatment metabolic response of diffuse large B cell lymphoma patients. *Clin. Exp. Med.* **18**, 505–512 (2018).
29. Guo, X. *et al.* High Serum Level of Soluble Programmed Death Ligand 1 is Associated With a Poor Prognosis in Hodgkin Lymphoma. *Transl. Oncol.* **11**, 779–785 (2018).
30. Zhang, X. *et al.* Plasma soluble programmed death ligand 1 levels predict clinical response in peripheral T-cell lymphomas. *Hematol. Oncol.* **37**, 270–276 (2019).
31. Shen, H. *et al.* Soluble programmed death-ligand 1 are highly expressed in peripheral T-cell lymphoma: a biomarker for prognosis. *Hematology* **24**, 392–398 (2019).
32. Bi, X.-W. *et al.* PD-L1 is upregulated by EBV-driven LMP1 through NF- $\kappa$ B pathway and correlates with poor prognosis in natural killer/T-cell lymphoma. *J. Hematol. Oncol.* **9**, 109 (2016).
33. Wang, H. *et al.* High post-treatment serum levels of soluble programmed cell death ligand 1 predict early relapse and poor prognosis in extranodal NK/T cell lymphoma patients. *Oncotarget* **7**, 33035–45 (2016).
34. Dronca, R.; Leontovich, A.; Harrington, S.; Jegapragasan, M.; Kottschade, L.; Nevala, W.; Enninga, E.; Markovic, S.; Dong, H. Soluble PD-L1 (sPD-L1) is associated with decreased survival in metastatic melanoma. *Pigment Cell Melanoma Res.* **28**, 768–769

- (2015).
35. Zhou, J. *et al.* Soluble PD-L1 as a Biomarker in Malignant Melanoma Treated with Checkpoint Blockade. *Cancer Immunol. Res.* **5**, 480–492 (2017).
  36. Wang, L. *et al.* Serum levels of soluble programmed death ligand 1 predict treatment response and progression free survival in multiple myeloma. *Oncotarget* **6**, 41228–36 (2015).
  37. Huang, S.-Y. *et al.* Soluble PD-L1: A biomarker to predict progression of autologous transplantation in patients with multiple myeloma. *Oncotarget* **7**, 62490–62502 (2016).
  38. Buderath, P. *et al.* Soluble Programmed Death Receptor Ligands sPD-L1 and sPD-L2 as Liquid Biopsy Markers for Prognosis and Platinum Response in Epithelial Ovarian Cancer. *Front. Oncol.* **9**, 1015 (2019).
  39. Park, H. *et al.* Prognostic implications of soluble programmed death-ligand 1 and its dynamics during chemotherapy in unresectable pancreatic cancer. *Sci. Rep.* **9**, 11131 (2019).
  40. Tominaga, T. *et al.* Clinical significance of soluble programmed cell death-1 and soluble programmed cell death-ligand 1 in patients with locally advanced rectal cancer treated with neoadjuvant chemoradiotherapy. *PLoS One* **14**, e0212978 (2019).

**Supplemental Table 2. Baseline patient characteristics in the melanoma cohort.**

	High sPD-L1 (n=216)	Low sPD-L1 (n=61)	Statistic
Age, years	60 (50, 69)	58 (50, 70)	F=0.10, P = 0.756 <sup>1</sup>
Sex: M	137/216 (63)	31/60 (52)	X <sup>2</sup> = 2.73, P=0.10 <sup>2</sup>
Stage			X <sup>2</sup> = 3.95, P=0.139 <sup>2</sup>
M1a	31/214 (14.5)	14/59 (23.7)	
M1b	54/214 (25.2)	17/59 (28.8)	
M1c	129/214 (60.3)	28/59 (47.5)	
sPD-L1, ng/uL	0.589 (0.370, 3.153)	0.216 (0.173, 0.249)	F=272.68 P < 0.001 <sup>1</sup>
LDH, U/L	209 (168, 395)	187 (163, 298)	F=2.05 P = 0.154 <sup>1</sup>

**Supplemental Table 2. Baseline patient characteristics in the melanoma cohort.** Patients with melanoma from three studies with retrospective data are compared by starting sPD-L1 level above or below the discovered survival cutoff (0.277ng/mL). Characteristics collected at entry into the study included age in years, sex, stage, sPD-L1, and LDH (lactate dehydrogenase).

<sup>1</sup>Kruskal-Wallis. <sup>2</sup>Pearson.

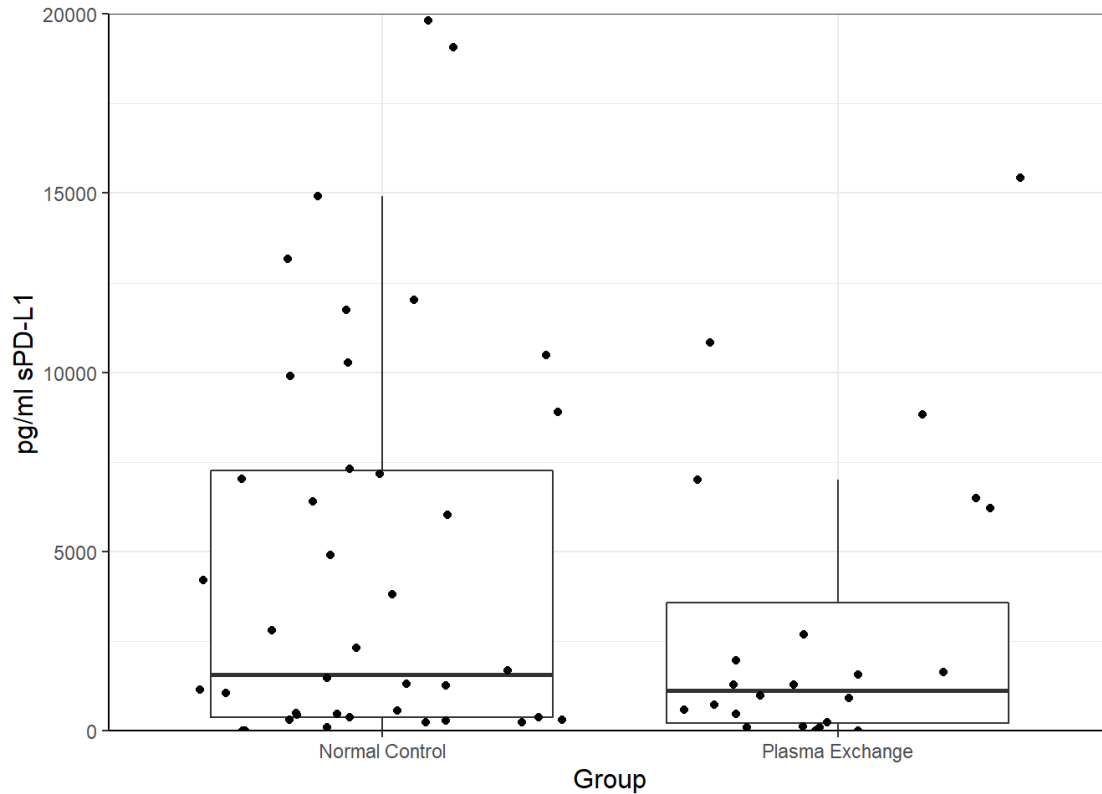
<b>Supplemental Table 3. Multivariate analysis in melanoma survival at diagnosis.</b>			
	<b>Hazard Ratio</b>	<b>P value</b>	
<b>Age &gt; 60 years</b>	1.00 (0.76-1.32)	0.93	
<b>Sex: M</b>	1.19 (0.90-1.58)	0.25	
<b>Stage</b>			
<b>M1a</b>	1		
<b>M1b</b>	1.42 (0.91-2.23)	0.11	
<b>M1c</b>	1.94 (1.28-2.96)	0.002 **	
<b>sPD-L1 &gt; 277ng/mL</b>	1.47 (1.05-2.07)	0.025 *	
<b>LDH &gt; 180U/L</b>	1.53 (1.53, 2.11)	0.010 **	

**Supplemental Table 3.** Multivariate analysis in melanoma survival at diagnosis. Cox proportional Hazards multivariate analysis was performed.

<b>Supplemental Table 4. Reduction in EVs by subtype per exchange.</b>	
<b>% Reduction in total EVs per exchange</b>	<b>(n=55)</b>
Mean (SD)	33.5 (89.4)
Median [Min, Max]	60.9 [-468, 99.1]
<b>% Reduction PD-L1-positive EVs per exchange</b>	<b>(n=13)</b>
Mean (SD)	73.1 (14.6)
Median [Min, Max]	72.3 [50.0, 98.5]
<b>% Reduction in ADAM10-positive EVs per exchange</b>	<b>(n=55)</b>
Mean (SD)	5.91 (126)
Median [Min, Max]	42.4 [-709, 99.5]

**Supplemental Table 4. Reduction in EVs by subtype per exchange.** Percent reduction in EVs for TPE sessions in which over one million pre-TPE EVs were present (*i.e.* above background noise levels) and no FFP was given.

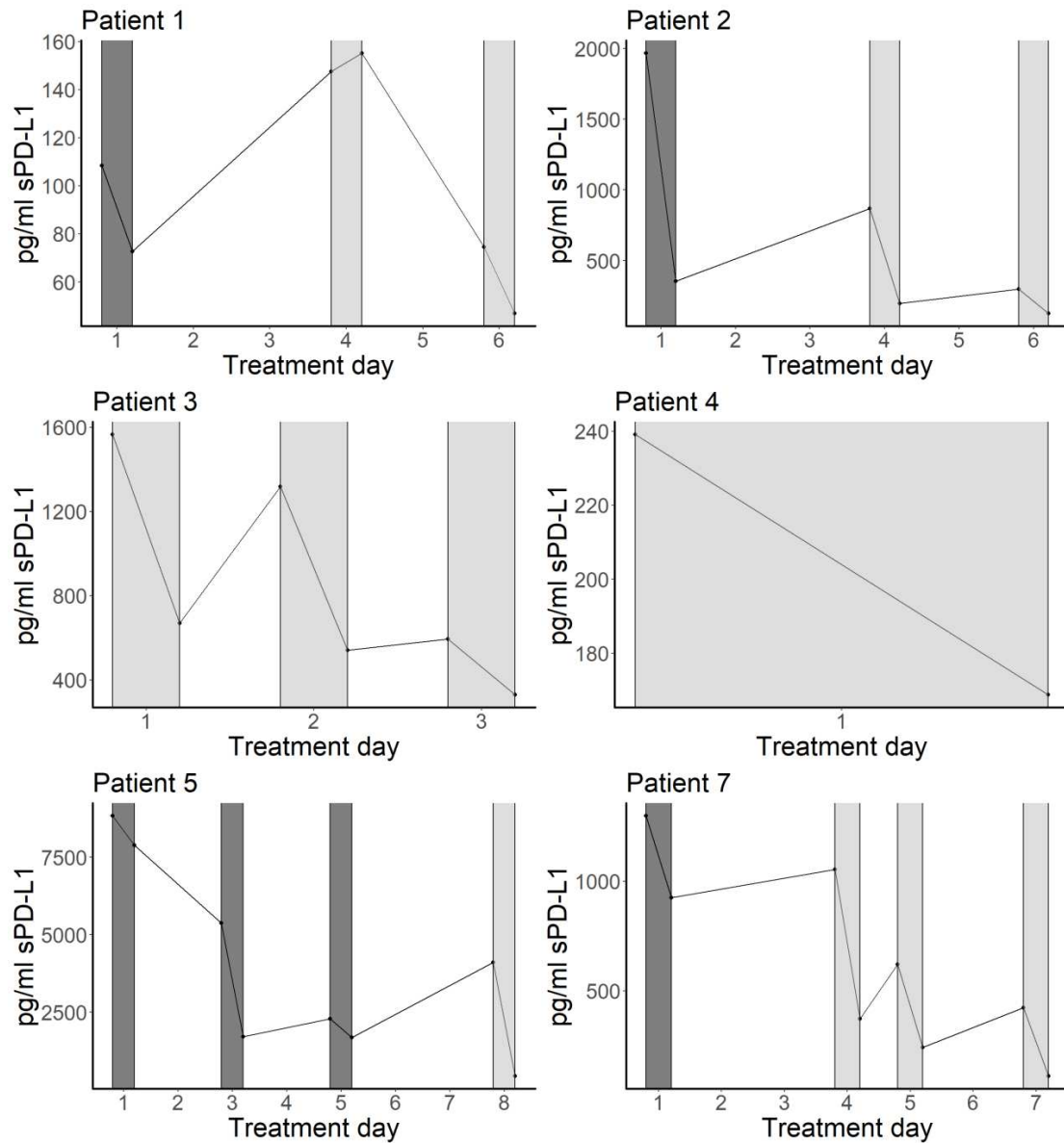
## SUPPLEMENTAL FIGURES

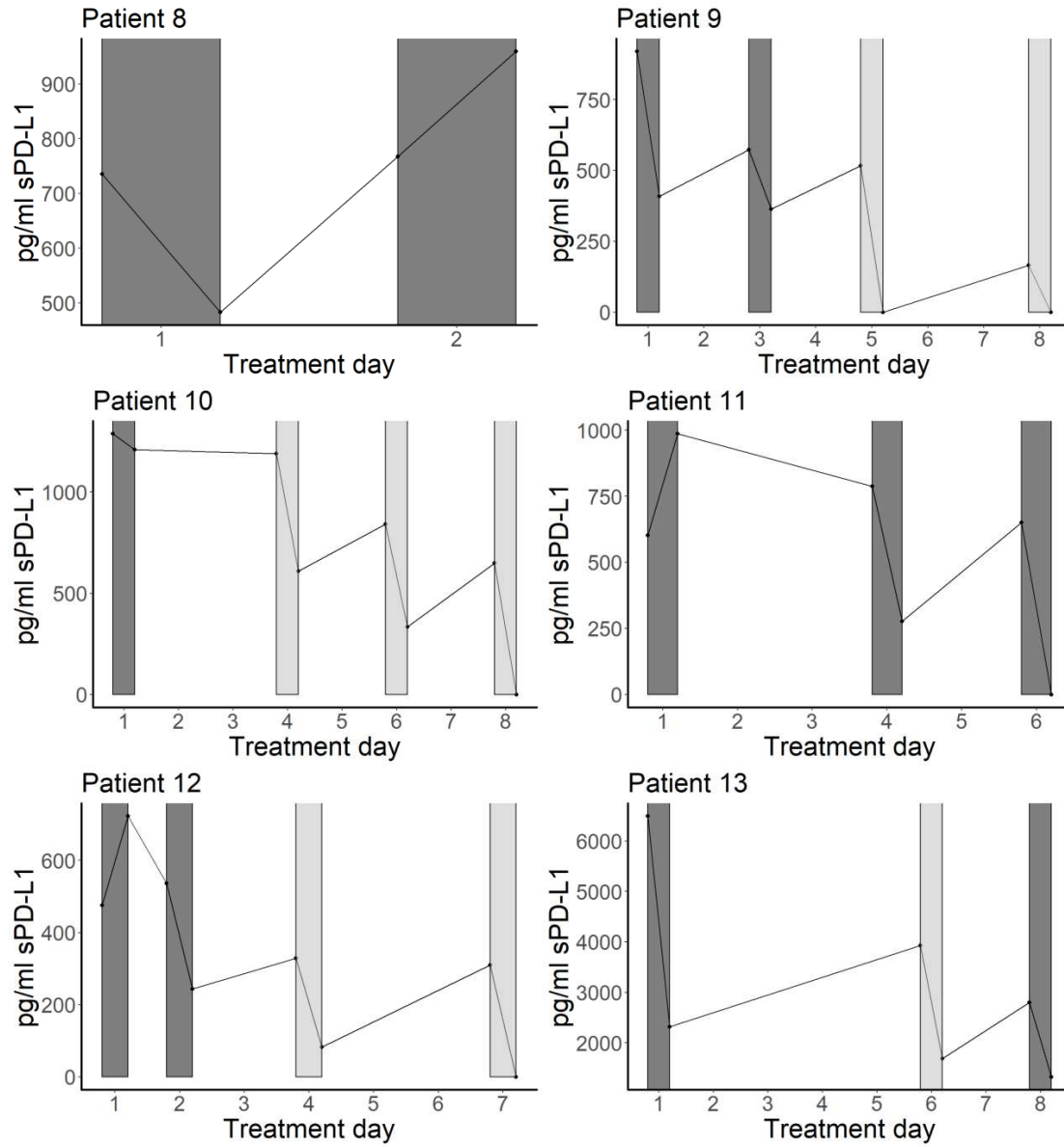
**Supplemental Figure 1. Baseline plasma sPD-L1 in normal controls versus patients undergoing TPE.**

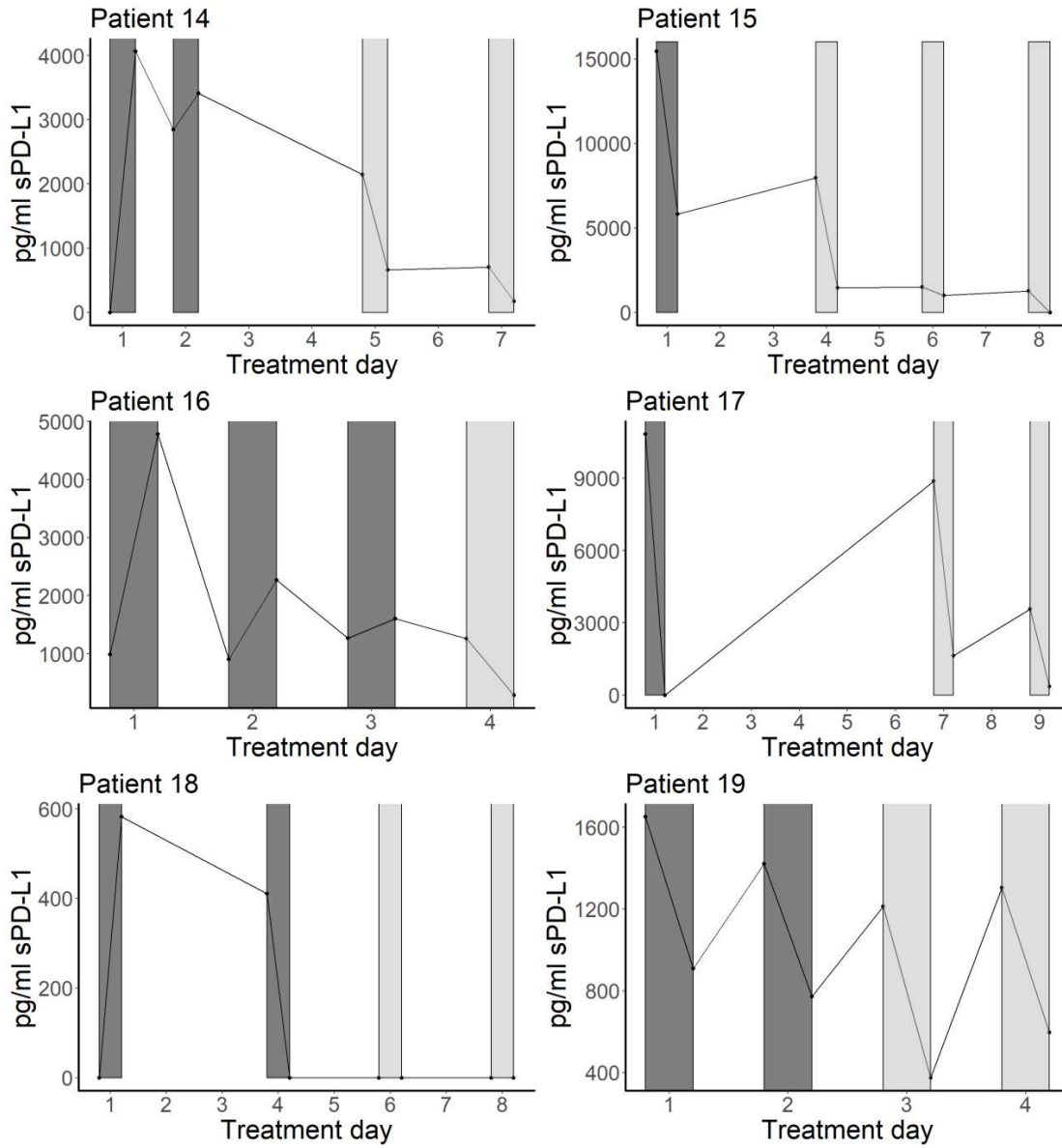
Baseline plasma sPD-L1 in normal controls versus patients undergoing TPE.					
	P value	mean	median	lower95CI	upper95CI
Normal Control	1	8188.0	2002.0	3167.9	13208.1
Plasma Exchange	0.1044	2898.8	1135.5	1554.4	4243.2

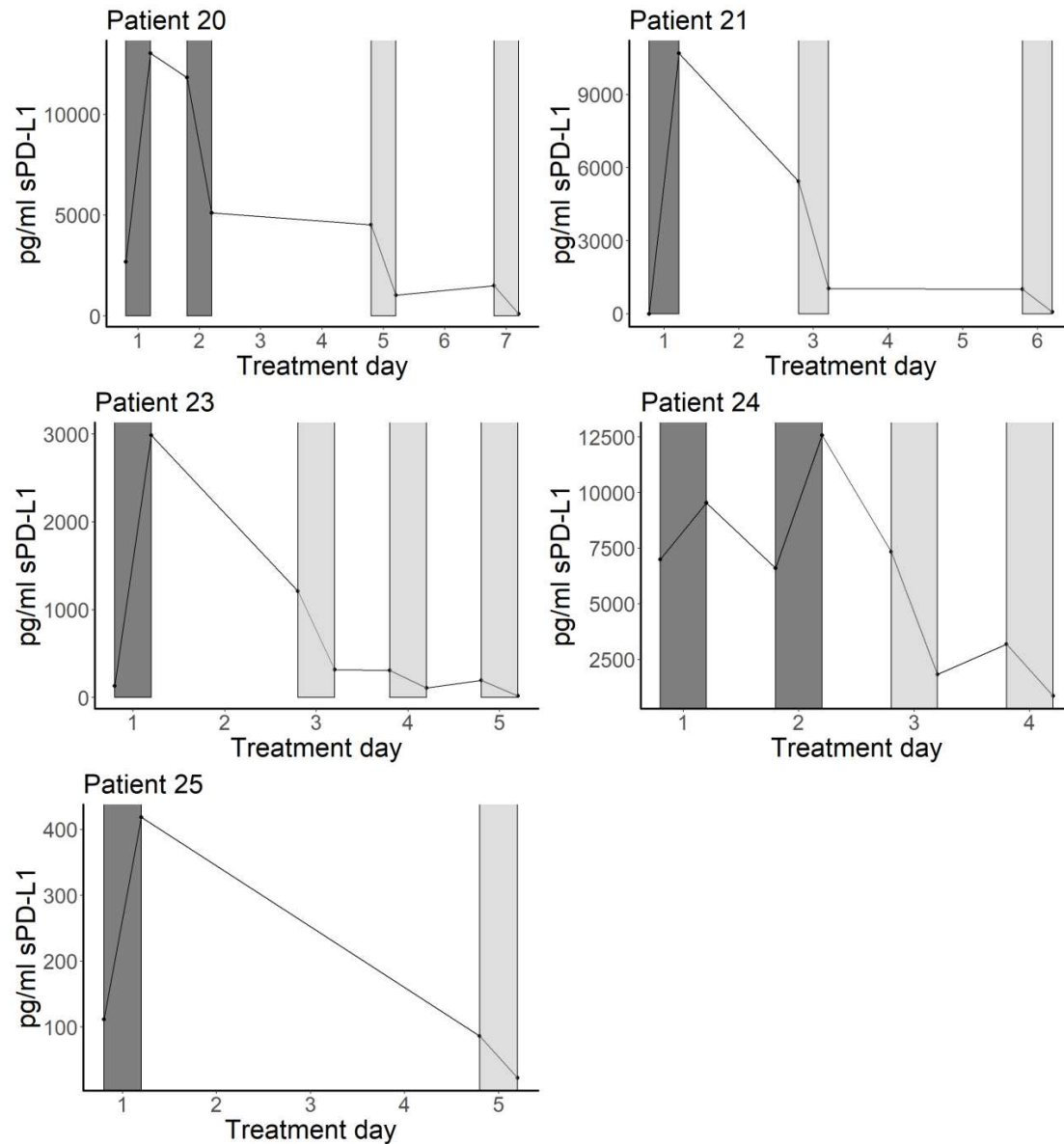
**Supplemental Figure 1. Baseline plasma sPD-L1 in normal controls versus patients undergoing TPE.** Levels of PD-L1 in patients undergoing TPE were not significantly lower than those of matched normal controls. Statistical table including p value (two-sided Student's *t* test), mean, and 95% confidence intervals are shown.



**Supplemental Figure 2. Plasma sPD-L1 in all TPE treatment courses (including sessions involving FFP).**

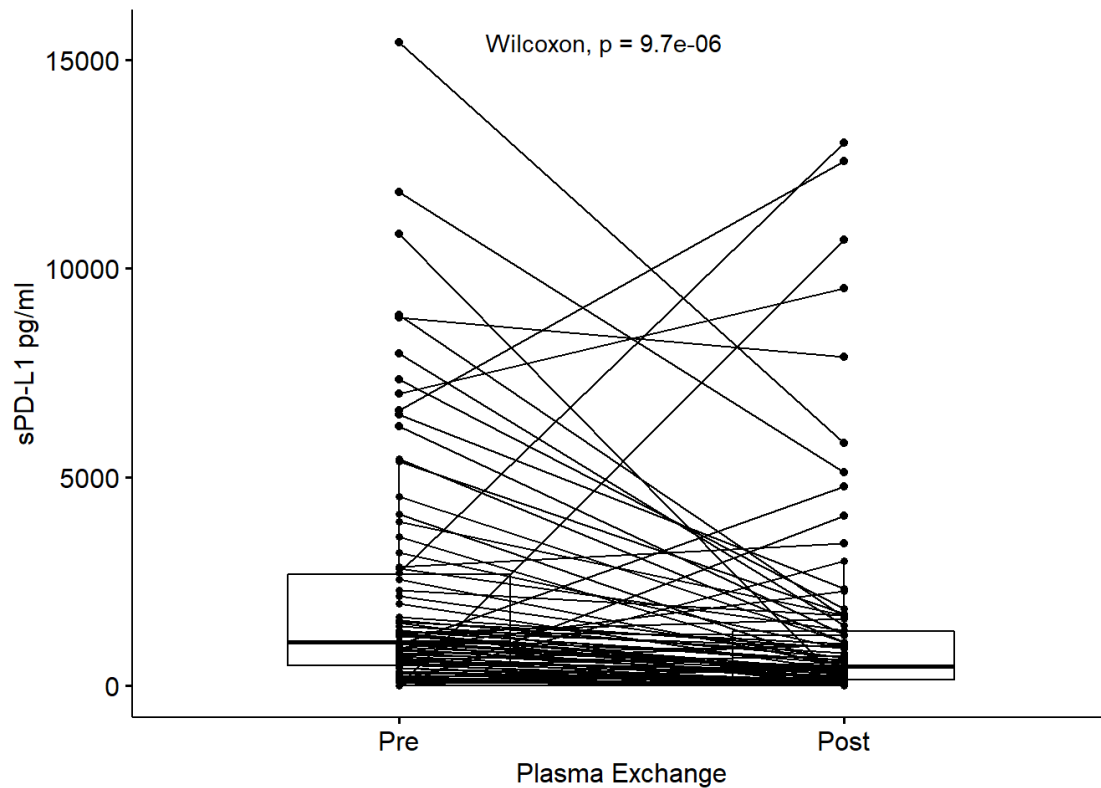




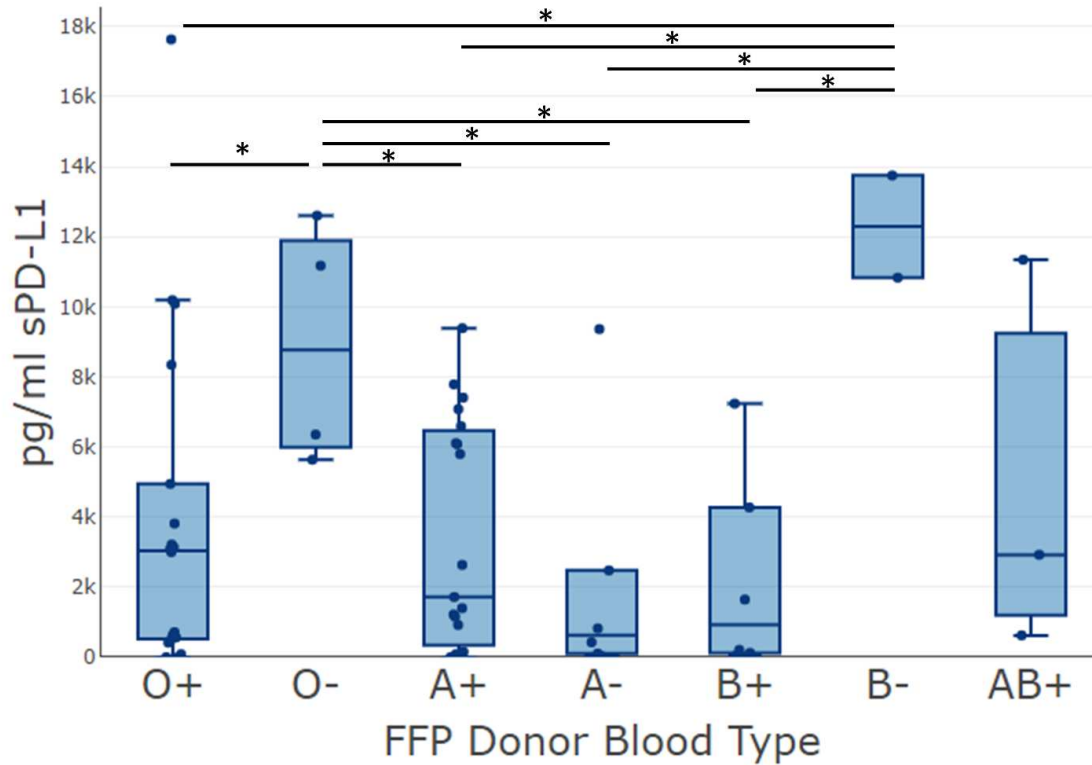


**Supplemental Figure 2. Plasma sPD-L1 in all TPE treatment courses (including sessions involving FFP).** Treatment courses of each patient are shown. Dark gray bars represent TPE sessions in which FFP was given. Light gray bars represent TPE sessions in which no FFP (*i.e.* only albumin) replacement was given. Patient 22 treatment course is shown in Fig 2D. Patient 6 was excluded for biotin use.

**Supplemental Figure 3. Plasma sPD-L1 in all TPE treatment courses (including sessions requiring FFP).**



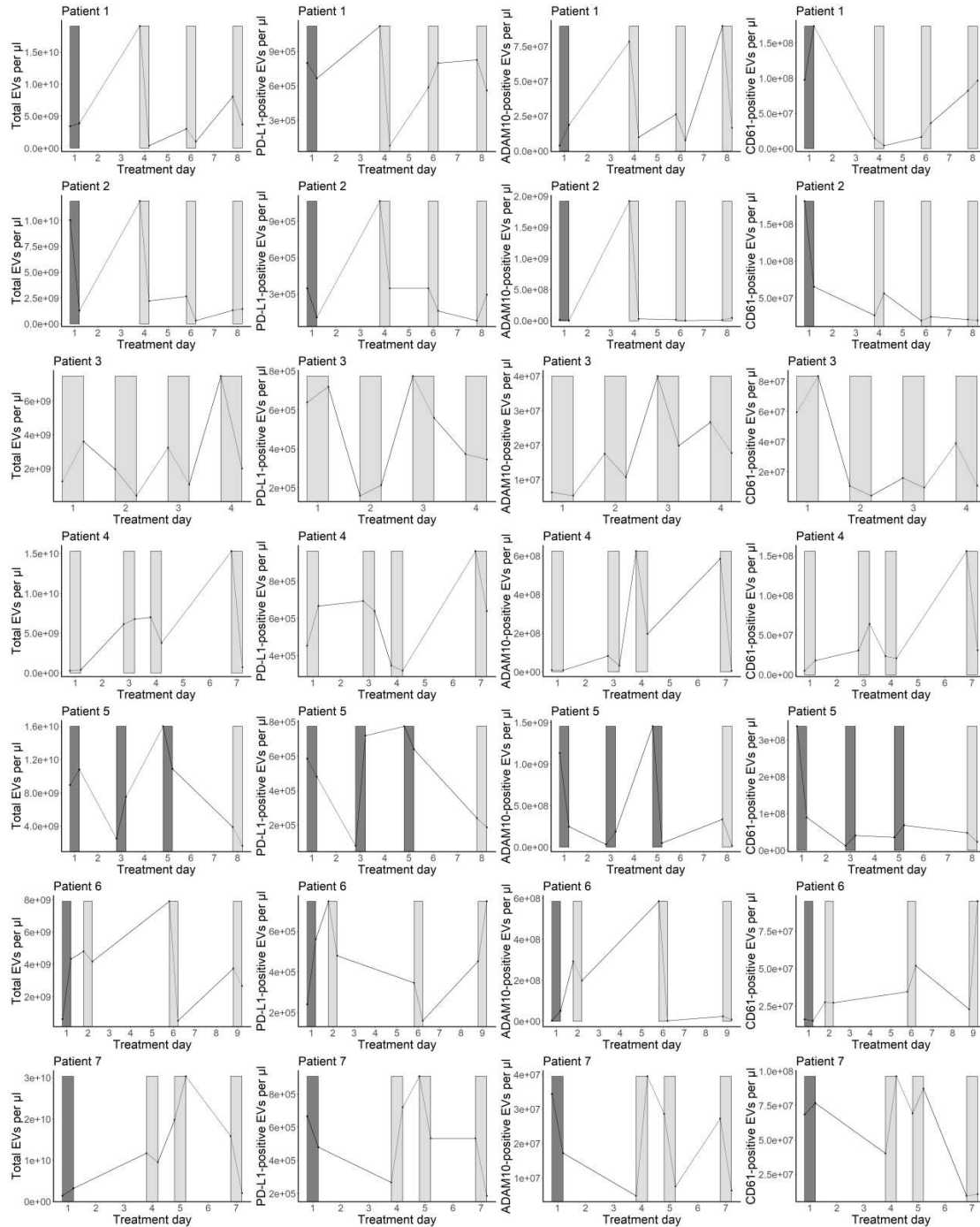
**Supplemental Figure 3. Plasma sPD-L1 in all TPE treatment courses (including sessions requiring FFP).** TPE significantly reduced sPD-L1 levels in all sessions, including those in which patients received donor fresh frozen plasma (FFP).

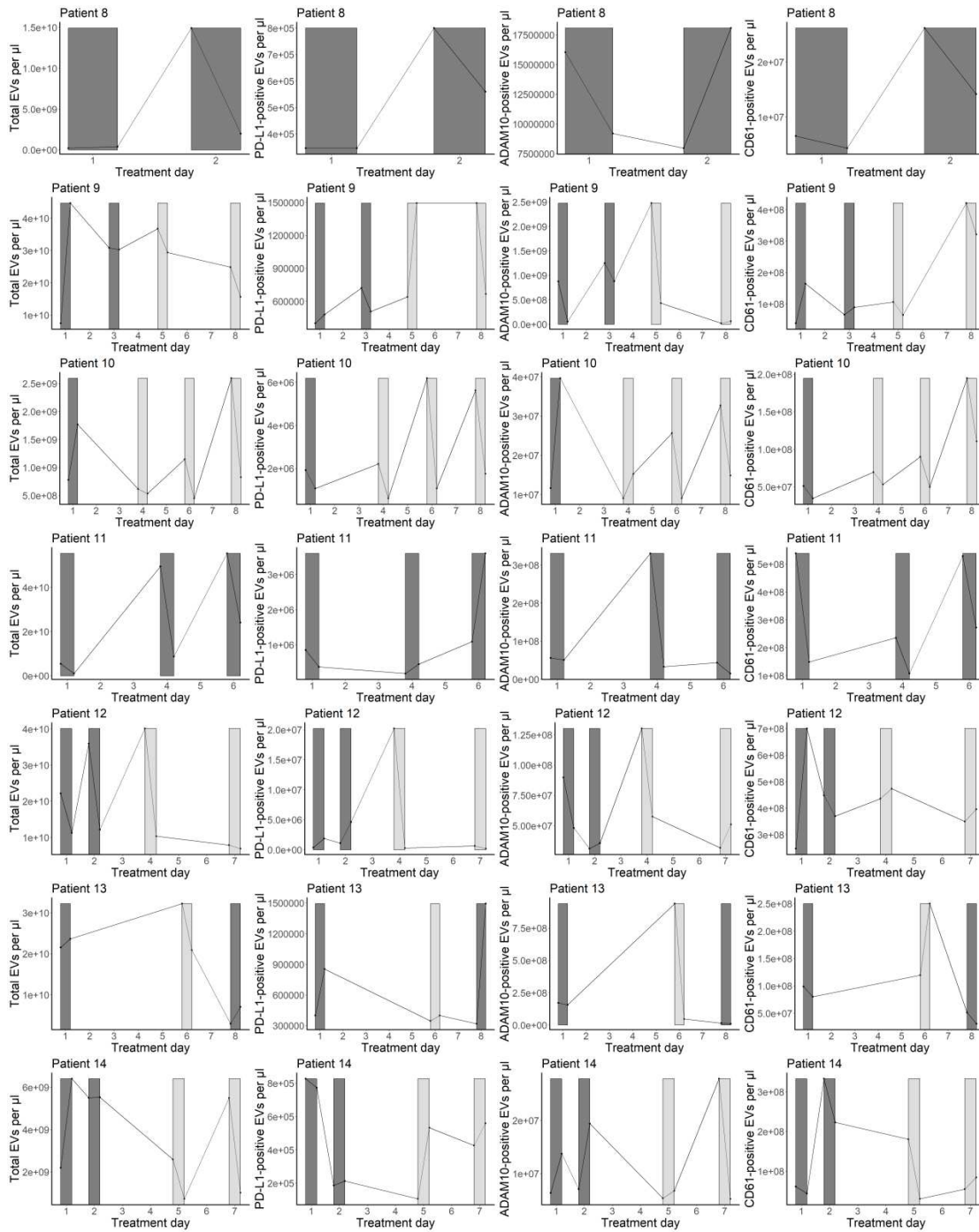
**Supplemental Figure 4. FFP donor sPD-L1 correlates with blood type.**

	P	mean	lower95CI	upper95CI		p	mean	lower95CI	upper95CI
O+	0.05	3909	2051	5767	O+	0.03	3909	2051	5767
O-	1	8942	6093	11791	O-	0.22	8942	6093	11791
A+	0.04	3448	2207	4689	A+	0.05	3448	2207	4689
A-	0.02	2205	0	4636	A-	0.01	2205	0	4636
B+	0.02	2258	293	4223	B+	0.02	2258	293	4223
B-	0.22	12289	9890	14688	B-	1	12289	9890	14688
AB+	0.35	4963	0	10323	AB+	0.14	4963	0	10323

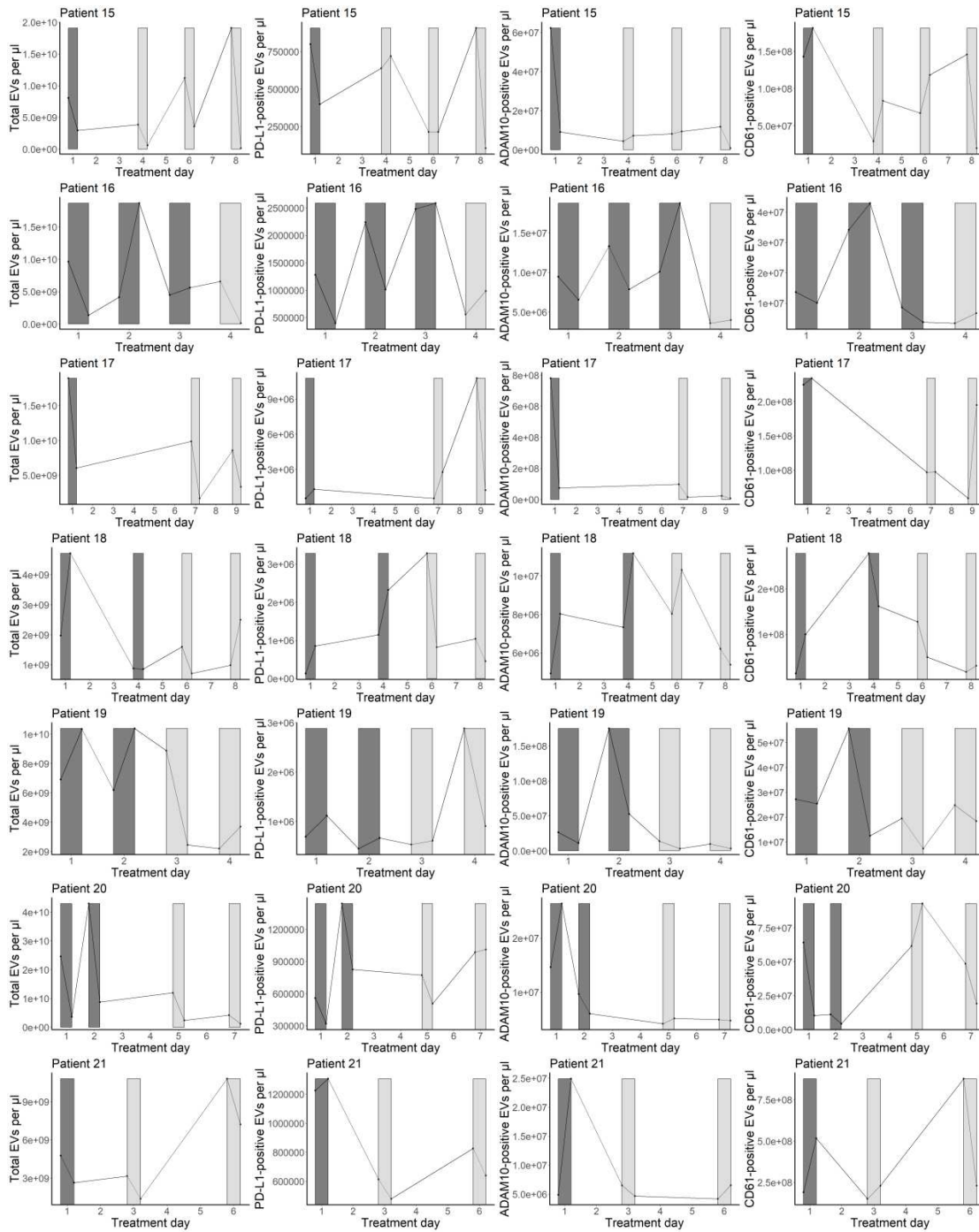
**Supplemental Figure 4. FFP donor sPD-L1 correlates with blood type.** Plasma from donors of fresh frozen plasma (FFP) showed variable levels of sPD-L1 by blood type.

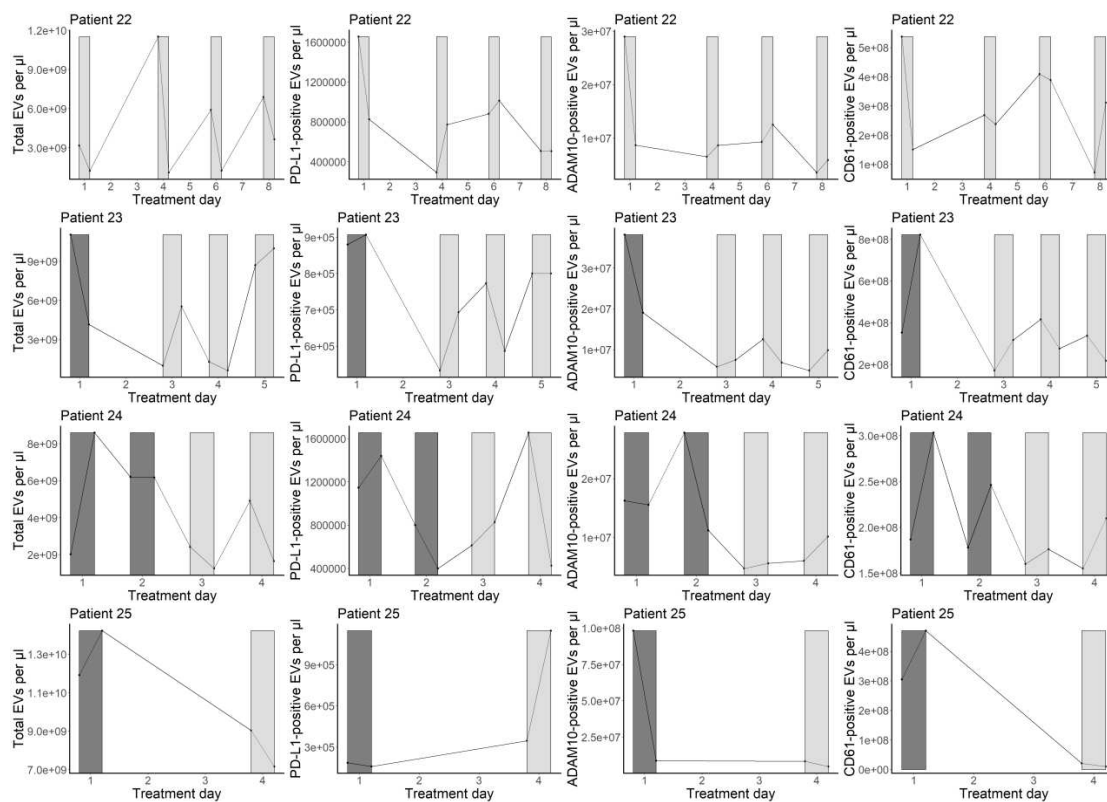
**Supplemental Figure 5. Plasma EVs in all TPE treatment courses (including sessions requiring FFP).**



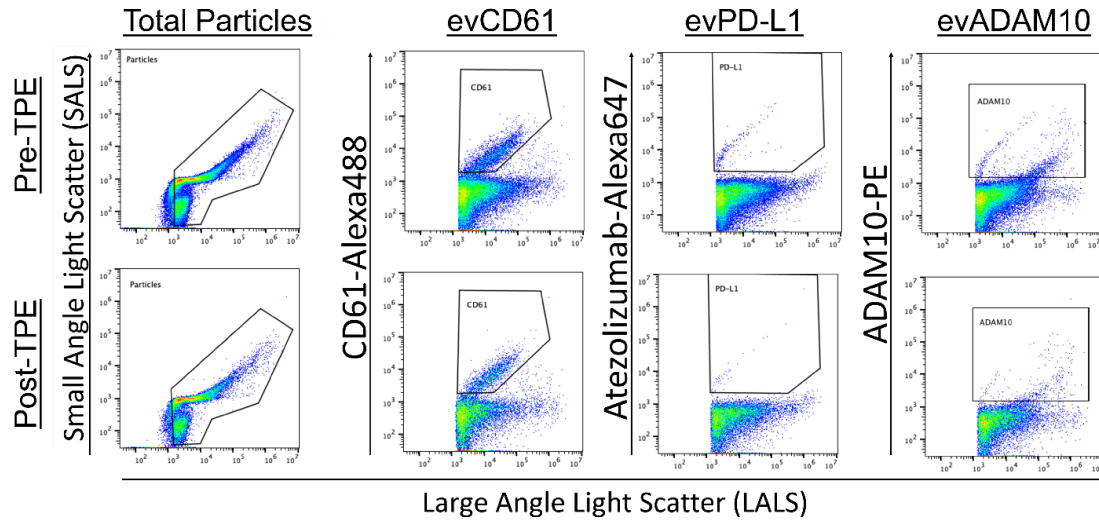




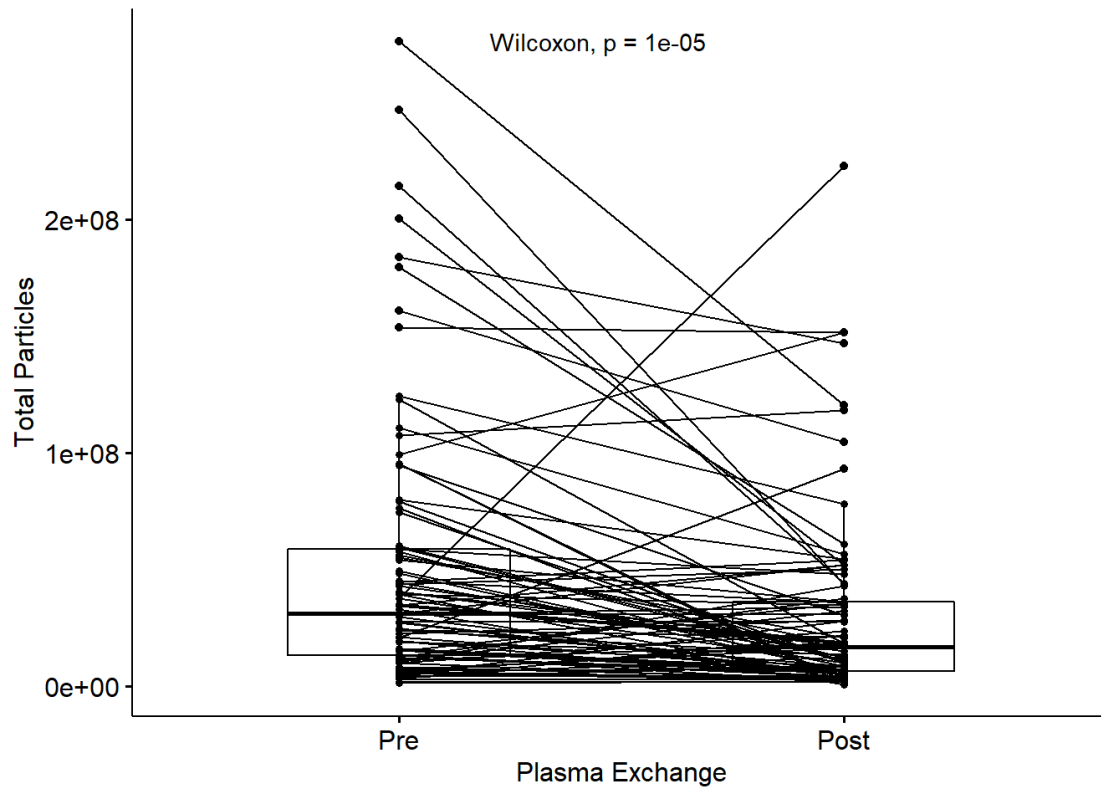




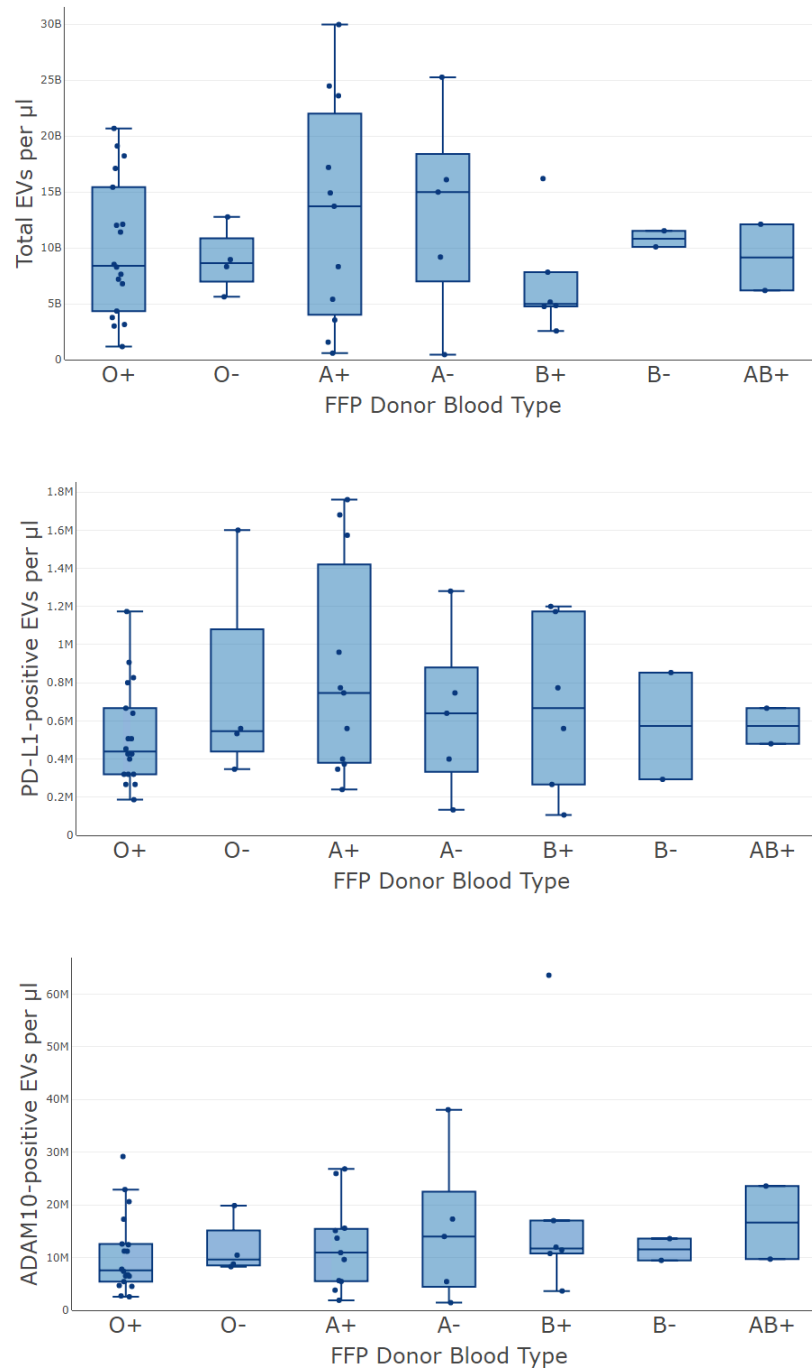
**Supplemental Figure 5. Plasma EVs in all TPE treatment courses (including sessions requiring FFP).** Plasma EV (total, PD-L1-positive, ADAM10-positive, and CD61-positive) levels over the course of TPE treatment are shown. Bars indicate TPE sessions, usually lasting 2 hours. Light-colored bars indicate that no FFP was received during the TPE session; conversely, dark bars indicate that FFP was received during the TPE session.

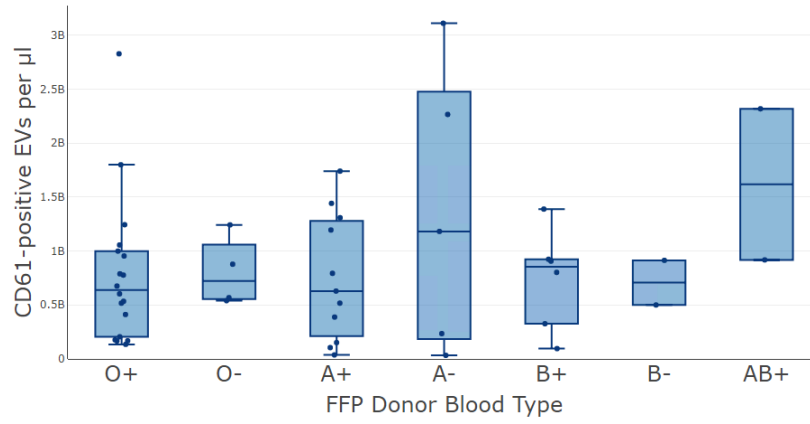
**Supplemental Figure 6. Example plasma EV nanoflow plots.****Supplemental Figure 6. Example plasma EV nanoflow plots.** Patient 10 pre- (top row) and post- (bottom row) TPE nanoflow cytometry detecting total EVs, evCD61, evPD-L1, and evADAM10.

**Supplemental Figure 7. Plasma EVs in all TPE treatment courses (including sessions requiring FFP).**



**Supplemental Figure 7. Plasma EVs in all TPE treatment courses (including sessions requiring FFP).** Plasma total EV per microliter levels over the course of TPE treatment are shown including those sessions in which FFP was received.

**Supplemental Figure 8. FFP donor EV concentrations do not correlate with blood type**



**Supplemental Figure 8. FFP donor EV concentrations do not correlate with blood type.**