

SUPPLEMENTAL INFORMATION

Phospholipase A₂ Inhibitor-Loaded
Phospholipid Micelles Abolish
Neuropathic Pain

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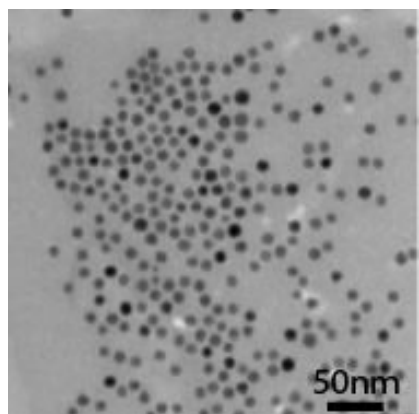


Figure S1. Transmission electron microscopy (TEM) image of individual hydrophobic SPION.

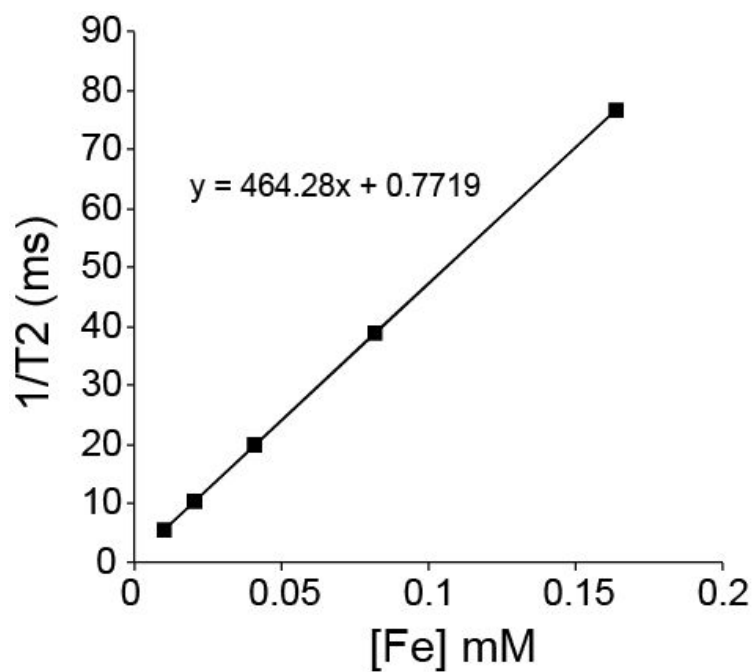


Figure S2. Relaxivity determination for sPLA₂ inhibitor-loaded micelles.

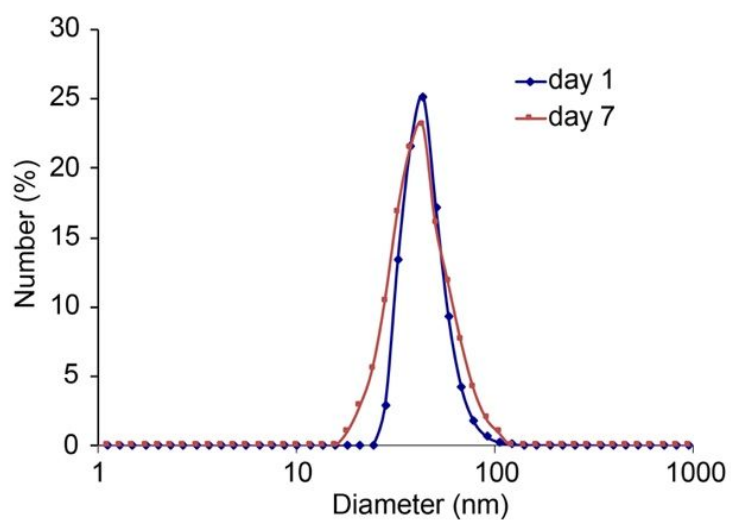


Figure S3. Incubation of sPLA₂ inhibitor-loaded micelles in buffer (0.1 M PBS, pH 7.4) does not change micelle stability for at least a week with no observable change in the hydrodynamic diameter of the micelle during incubation.

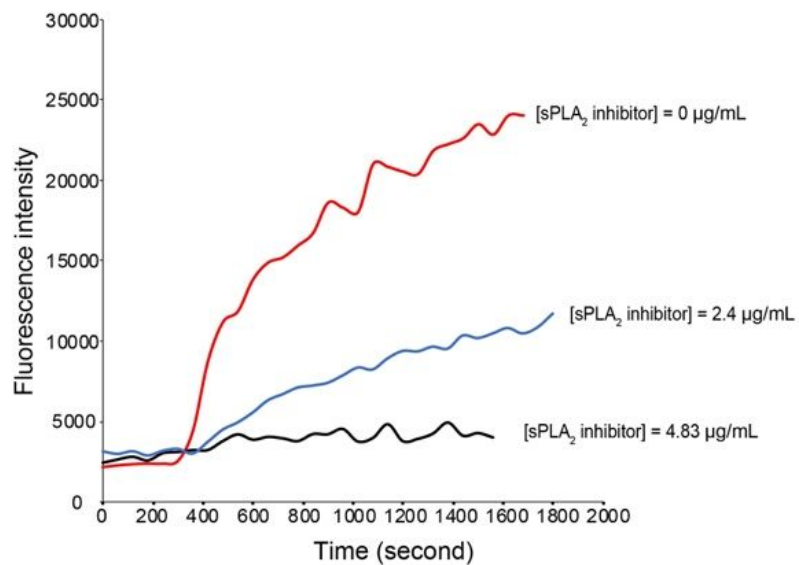


Figure S4. sPLA₂ inhibitor-loaded micelles inhibited sPLA₂ activity in a concentration dependent manner as observed by a reduction in fluorescence activation.

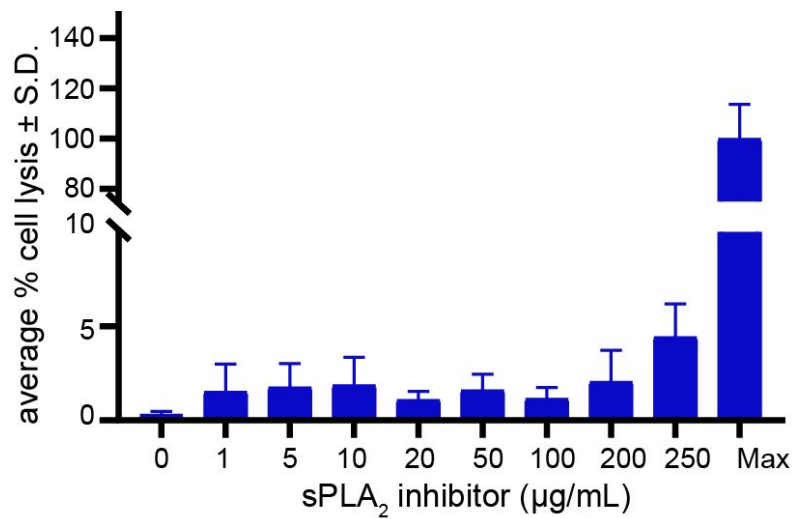


Figure S5. Incubation with sPLA₂ inhibitor-loaded micelles does not significantly increase the average percent of cell lysis observed in primary DRG cultures, with no significant differences ($p > 0.38$) for any micelle concentrations compared to control.

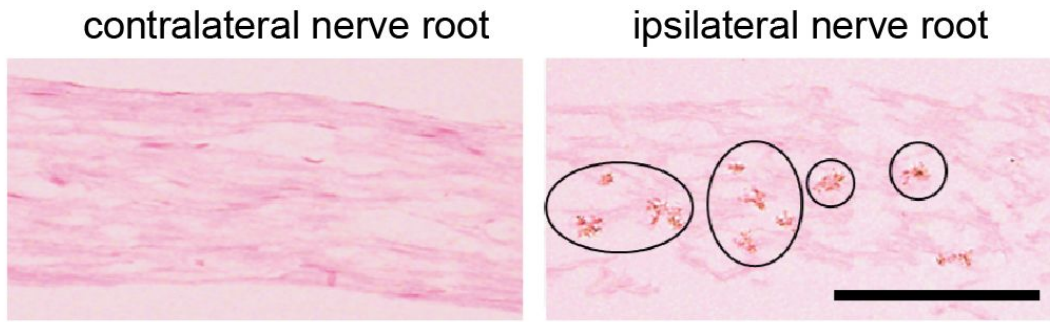


Figure S6. Unloaded micelles (*i.e.* without sPLA₂ inhibitor) are found to localize at day 7 to the injured nerve root after local administration at the time of injury (day 0). Iron is detected (circles) only in the ipsilateral C7 dorsal nerve root after a nerve root compression that is treated with unloaded micelles. There is no evidence of iron in the contralateral C7 nerve root after either type of treatment. The scale bar is 100 μm and applies to all panels.

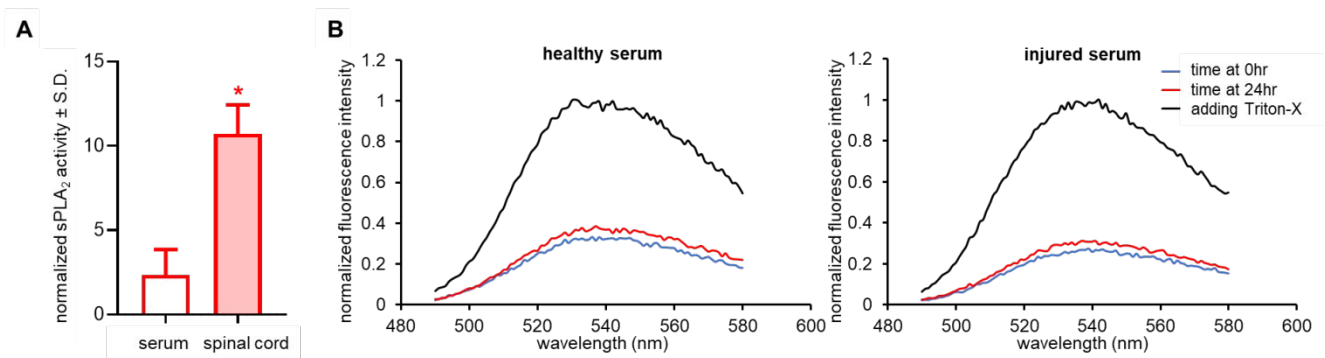


Figure S7. (A) sPLA₂ activity assay of serum and spinal cord tissue collected one day after a painful injury shows significantly increased ($p = 0.0004$) activity in the spinal cord compared to serum. (B) Incubation sPLA₂-responsive phospholipid liposomes with serum from injured rats does not affect the fluorescence release from sPLA₂ responsive phospholipid liposomes at 0 or 24 hours compared to incubation with healthy serum and is lower than Triton-X positive controls.

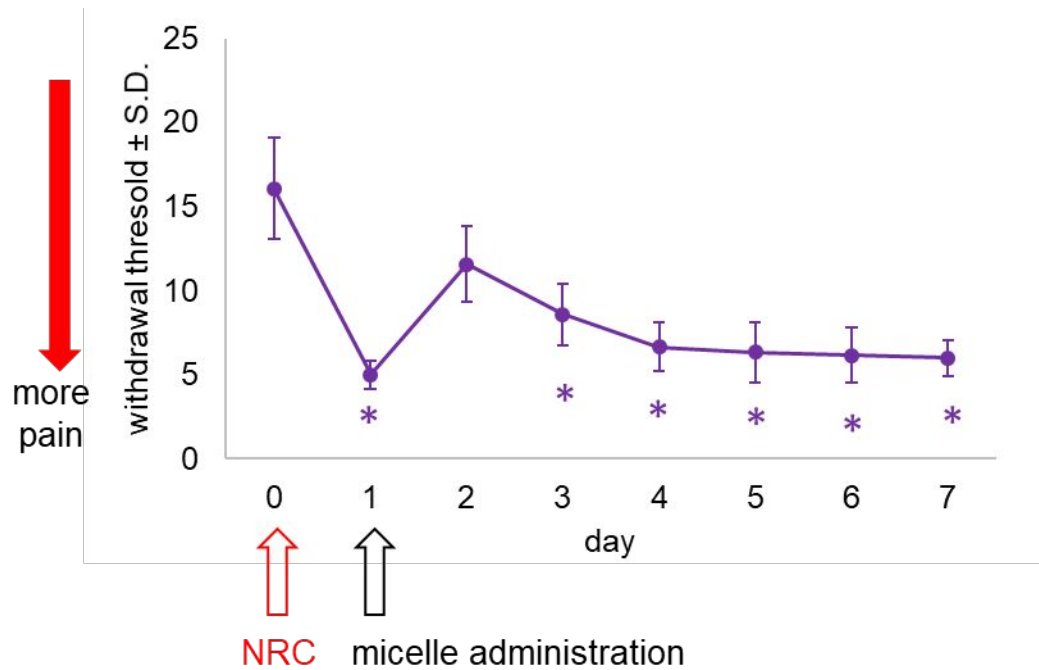


Figure S8. Following painful nerve root compression on day 0, withdrawal thresholds on day 1 are significantly lower ($p < 0.0002$) than pre-injury (day 0) levels. Administration of a single dose of sPLA₂ inhibitor-loaded micelles on day 1 transiently increases thresholds on day 2, but thresholds are significantly lower ($p < 0.0007$) on days 3 through 7 indicating the presence of pain on those day.

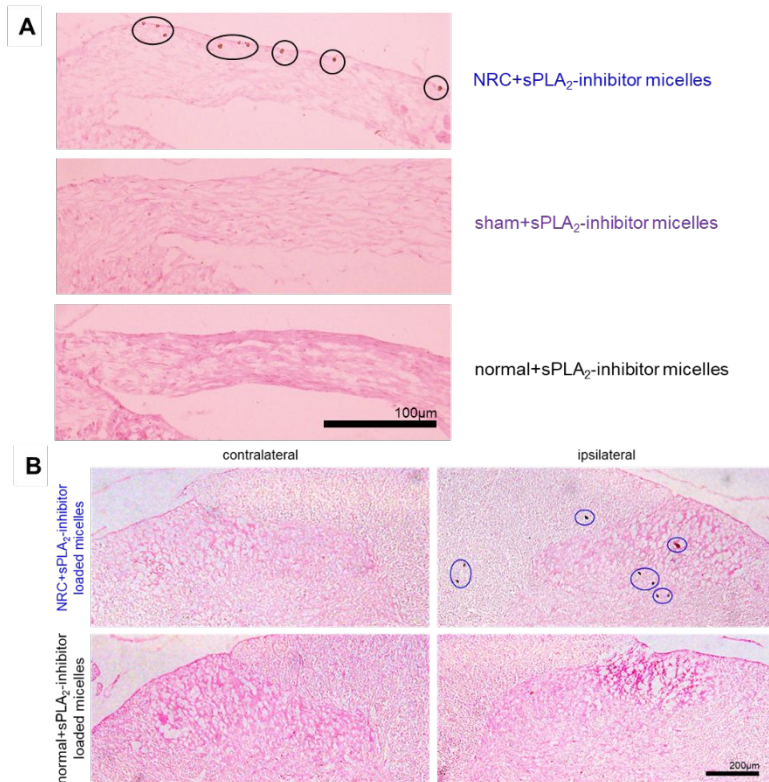


Figure S9. (A) At day 7 after intravenous administration of sPLA₂ inhibitor-loaded micelles, Prussian Blue- labeled SPIOs only localize to the injured nerve root and not to uncompressed sham and normal nerve roots. (B) Positive SPIO labeling is also observed in the white and gray matter only in the ipsilateral spinal cord following intravenous administration of sPLA₂ inhibitor-loaded micelles. There was no SPIO labeling evident in the ipsilateral or contralateral spinal sections of naïve rats treated with sPLA₂ inhibitor-loaded micelles.

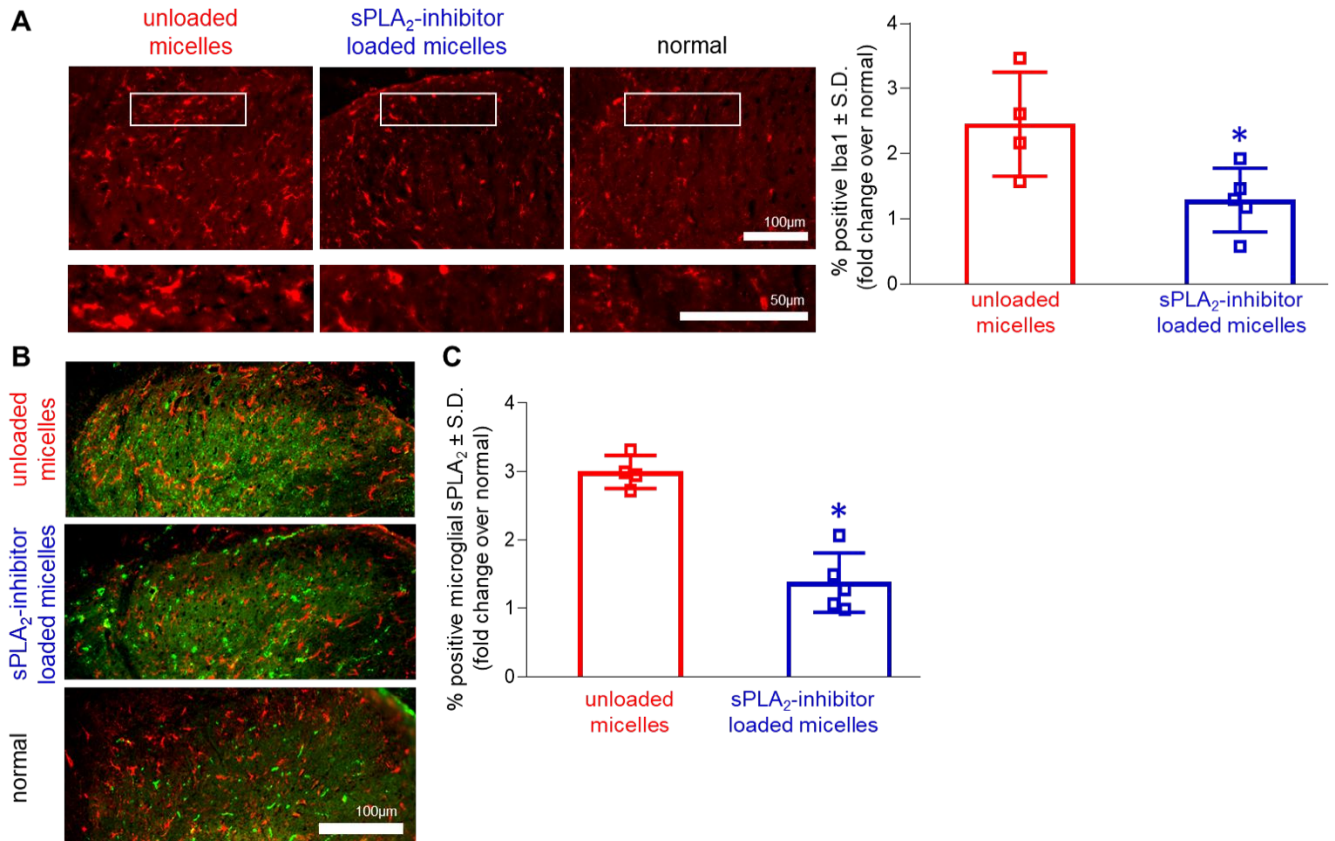


Figure S10. (A) Spinal Iba1 expression at day 7 is significantly elevated ($*p < 0.0002$) following systemic treatment with unloaded micelles compared to systemic treatment with sPLA₂ inhibitor-loaded micelles. The images on the bottom row are higher magnification of the enclosed boxes to show individual cells. (B) Representative images of the spinal dorsal horn show a reduction in microglial sPLA₂ (yellow) following treatment with sPLA₂ inhibitor-loaded micelles. (C) Spinal microglial sPLA₂ expression is significantly reduced ($*p = 0.0002$), to normal levels, with sPLA₂ inhibitor-loaded micelles compared to treatment with unloaded micelles.

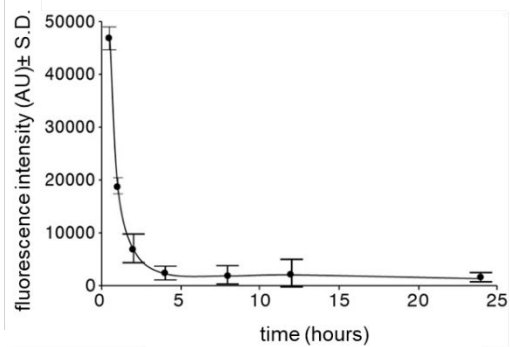
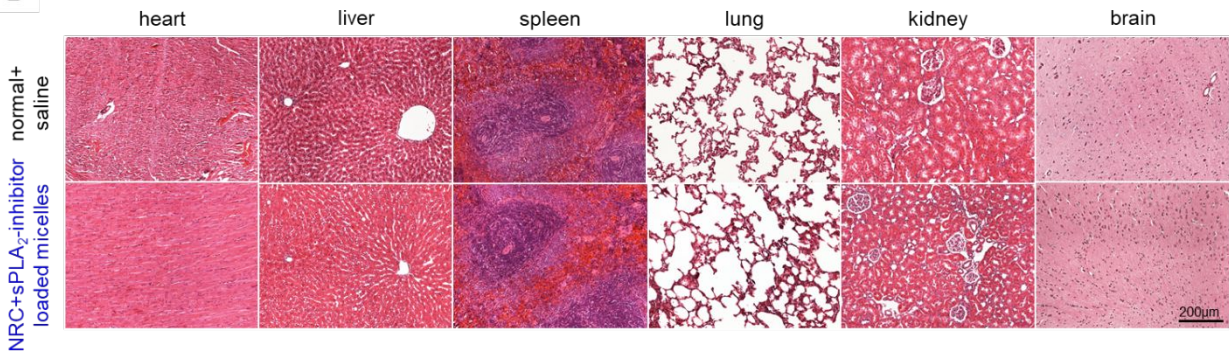
A**B**

Figure S11. (A) The pharmacokinetic profile of rhodamine-labeled sPLA₂ inhibitor-loaded micelles in whole blood following intravenous administration indicates that micelles have an *in vivo* half-life of around 45 minutes. (B) *H&E* staining of rat heart, liver, spleen, lung, kidney and brain at 7 days after intravenous injection of sPLA₂ inhibitor-loaded micelles or saline show no abnormalities.