

Supplemental Figure S1, related to Figure 1.

(A) Venn diagrams showing overlap between called peaks of two independent ChIP-seq replicates (Rep1 and Rep2) for each Nup, as labeled, in S2 cells (left) and respective principal component analysis (PCA) for ChIP-seq Nup peak datasets (right). (B) Western Blot of S2 cells treated with dsWhite and dsNup107 RNAi for 3 rounds (total of 6 days), and blotted against Nup107 antibody (top). Graph depicts guantification of Nup107 western signal normalized to tubulin signal (bottom). Molecular weight markers are 250, 150, 100, 75, 55 and 37 kDa, topbottom. Bars represent mean +/- SD from 3 biological replicates. (C) Representative IF images of cells treated with RNAi against control (dsW), or Nup107 (dsNup107). Cells are stained for 414 (green), Nup107 (red) and DNA (DAPI, blue) (D) ChIP-gPCR of S2 cells treated with RNAi against control (dsWhite) or Nup107 transcripts (dsNup107) using generated Nup107 antibody or rabbit IgG, at genes identified as targets of Nup107 (left). RT-qPCR of Nup107 transcript levels in RNAi treatments is shown at right. Bars represent mean ChIP signal normalized to input (left) and mean expression levels normalized to dsWhite (right) + SEM (3 biological replicates). For (B)-(D) RNAi treatments were performed for 3 rounds, 144 hours total. (E) Venn diagram comparing overlap between Elys/Nup93 ChIP-seq peaks with Elys/Nup107 ChIP-seq peaks. (F) Distribution of Nup ChIP-seq peaks relative to genomic elements as labeled (left), and relative to the 4 chromosomes and heterochromatic/repetitive elements of the fly genome (right).

Chromatin Color P - Values								
	Yellow	Red	Green	Blue	Black			
Elys	P < 0.01	P < 0.01	P = 1.0	P = 1.0	P = 1.0			
Nup107	P < 0.01	P < 0.01	P = 1.0	P = 1.0	P = 1.0			
Elys/Nup107	P < 0.01	P < 0.01	P = 0.54	P = 1.0	P = 1.0			
Nup93	P = 1.0	P < 0.01	P = 1.0	P < 0.01	P = 1.0			
Elys/Nup93	P = 1.0	P < 0.01	P = 1.0	P < 0.01	P = 1.0			

Chromatin State P - Values									
	State 1	State 2	State 3	State 4	State 5	State 6	State 7	State 8	State 9
Elys	P < 0.01	P = 1.0	P < 0.01	P = 1.0	P = 1.0	P = 1.0	P = 1.0	P = 1.0	P = 1.0
Nup107	P < 0.01	P = 1.0	P = 1.0	P = 1.0	P = 1.0	P = 1.0	P = 1.0	P = 1.0	P = 1.0
Elys/Nup107	P < 0.01	P = 1.0	P = 0.54	P = 1.0	P = 1.0	P = 1.0	P = 1.0	P = 1.0	P = 1.0
Nup93	P < 0.01	P = 1.0	P < 0.01	P = 1.0	P = 1.0	P < 0.01	P = 1.0	P = 1.0	P = 1.0
Elys/Nup93	P = 1.0	P = 1.0	P < 0.01	P = 1.0	P = 1.0	P < 0.01	P = 1.0	P = 1.0	P = 1.0



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Top 20 most enriched GO terms for Nup93-Pc targets			
GO:0010092~specification of organ identity			
GO:0048098~antennal joint development			
GO:0061332~Malpighian tubule bud morphogenesis			
GO:0007383~specification of segmental identity, antennal segment			
GO:0042693~muscle cell fate commitment			
GO:1902339~positive regulation of apoptotic process involved in morphogenesis			
GO:0035223~leg disc pattern formation			
GO:0042684~cardioblast cell fate commitment			
GO:0042682~regulation of compound eye cone cell fate specification			
GO:0048735~haltere morphogenesis			
GO:0042659~regulation of cell fate specification			
GO:0035288~anterior head segmentation			
GO:0035284~brain segmentation			
GO:0007487~analia development			
GO:0007483~genital disc morphogenesis			
GO:0030539~male genitalia development			
GO:0009997~negative regulation of cardioblast cell fate specification			
GO:0007380~specification of segmental identity, head			
GO:0035051~cardiocyte differentiation			
GO:0007384~specification of segmental identity, thorax			



Supplemental Figure S2, related to Figure 2.

(A) P-values of enrichment of chromatin states (according to 5-state model (top) and 9-state model (bottom)) for each Nup peaks set, as labeled, calculated by Permutation test, which involves comparison to 100 random shuffles of the same number of peaks on the genome. (B) Box plots of ChIP signal of selected data sets as labeled, at Nup93 peaks and its random shuffled control (left 3 plots) and at Nup107 peaks and its random shuffled control (right 3 plots). P-values were calculated using two-tailed nonparametric t test (Mann Whitney test). The null hypothesis assumes that the Polycomb change may occur in both the same and opposite directions as the Nup change. (C) Top 20 GO Analysis terms of genes co-bound by Nup93 and Pc (GO terms sorted by fold enrichment). (D) Genome browser tracks showing previously defined Polycomb Response Elements (PREs) (work of Kassis lab, as referenced in text), relative to Nup93, Pc, H3K27Me3 ChIP-seq and Blue chromatin state.



Supplemental Figure S3, related to Figure 3.

(A-B) Distribution of chromatin colors (5 color model (A) or 9 color model (B)) among Nup93 or Nup107 ChIP-seq peaks within 5 Kb boundaries of previously defined LADs. (C) Enrichment heatmaps of Lamin DamID signal at ChIP-seq identified peaks of Nup93. Peaks are sorted by Lamin signal (left) and Nup93 peak size/width (right). (D) Enrichment heatmaps showing comparison of previously defined LADs, derived from Lamin DamID in Kc cells (described in text), to ChIP-seq of Lamin in Kc and S2 cells, using the L7 anti-Lamin antibody (cited in text). For both (C) and (D), y-axis units correspond to ChIP-seg/DamID signal intensity (rpm/bp) and the x-axis corresponds to distance from the peak (Kb (C), Mb (D)). (E) Heatmap table plotting Pearson correlations between S2 and Kc cell Lamin ChIP-seq replicates (3 replicates per cell type). (F) Representative GB snapshots of LADs defined by Lamin DamID along with Kc and S2 cell Lamin ChIP-seq replicates, and other tracks as indicated, at the Soxn locus (330 Kb). (G) Representative IF images of WT S2 cells stained against DNA (DAPI, blue), Nup93 (left column)/Nup107 (right column) and mAb414 (green). (H) Representative line graphs of florescence intensity of 414 and Nup93 or Nup107 across the diameter of an S2 cell nucleus, represented as fraction of total intensity signal. (I) Plot of Pearson correlation coefficients (PCCs) between 414 signal and Nup107 (left) or Nup93 (middle) (PCCs for Nup107/414 mean=0.74, Nup107/DAPI mean=0.04, and randomized pixels mean=0, n=52, 2 biological replicates; PCCs for Nup93/414 mean=0.71, Nup93/DAPI mean=-0.11, and randomized pixels mean=0, n=47, 2 biological replicates). (J) Plot of Manders overlap coefficients (MOCs) for fraction of Nup107 or Nup93 that overlaps with 414 signal (Nup107 MOC mean=0.94, n=52, 2 biological replicates; Nup93 MOC mean=0.98, n=45, 2 biological replicates). Middle line demarcates mean ± SD. P values are from a two tailed unpaired non-parametric t test. (K) Quantification of internal florescence signal of Nup93 or Nup107 as a fraction of total florescence signal calculated from line graphs in H using 414 signal to define periphery (for Nup93 mean=6.0%, n=26; for Nup107 mean=2.7%, n=32).

Pc ChIP-seq Peaks that Overlap with Nup93 or LAD



Supplemental Figure S4, related to Figure 4.

(A) Distribution of Pc ChIP-seq peaks (described in text) containing Nup93 ChIP-seq peaks (Nup93), LADs with Nup93 ChIP-seq signal within or at 5 Kb boundaries (LAD+Nup93), LADs without Nup93 ChIP-seq signal within or at 5 Kb boundaries (LAD-Nup93), or none of these (pie chart at left), and averaged Pc ChIP-seq signal intensities among the four groups (box and whisker plot at right). Nup93 ChIP-seq signal at peaks of three well-characterized PcG targets *Abd-B, Antp* and *ss* are depicted by the dots imbedded in the Nup93 box and whisker plot. (B) Predominant chromatin colors of LADs with and without Nup93 within or at 5 Kb boundaries. (C) Quantification of Nup93 (left) and Nup107 (right) IP efficiency, from Co-IPs depicted in figure 4E. Bars represent mean percentages of IP signal relative to 100% input (calculated from 5% input signal), +/- SEM (3 biological replicates).











Fraction of cells with locus > 1 µm from nuclear periphery



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Peripheral distance (µm)







Peripheral distance (µm)

Figure S5, related to Figure 5.

(A) Representative images of Kc cells treated with RNAi against control (dsW), Pc/Ph (dsPcPh) or Nup93 (dsNup93) as described in Figure 5. Cells are co-labeled with Oligopaints FISH probes (red or cyan, probes as labeled), and IF for Lamin (green) and DNA (DAPI, blue). (B) Schematic of quantification scheme depicting defined distances between probe signal and the nuclear periphery. Average nuclear size is shown, with lines delimiting 1 µm intervals. Mean diameter for Kc cell nuclei demarcated by Lamin = 6.02µm, calculated from >400 cells per replicate, 4 biological replicates. (C) Normalized RT-qPCR analysis of Nup93-1, Nup93-2, Ph and Pc expression in Kc cells treated for 4 days with RNAi against indicated genes, relative to control cells treated with RNAi against dsWhite (dsW). Bars represent mean + SEM. (D) Fraction of cell population with a probe to periphery distance larger than 1 µm, for cells treated with RNAi described in 5F and 5G, for the 4 utilized probes. (E) Distributions of measured probe to periphery distances, for the 2 indicated control probes, in cells treated with control (dsW) and Nup93 RNAi (dsNup93), using DAPI to define the nuclear periphery, plotted as Fraction of cell population with the distance within an assigned bin (labeled on the bottom axis). Bars represent mean + SEM (3 biological replicates). P values are from a two-tailed unpaired non-parametric t test. Null hypothesis is "not being further away from periphery".



Supplemental Figure S6, related to Figure 6.

(A) Hi-C interaction map of *D. melanogaster* genome (see text for reference), with locations of Pc regions bound by Nup93 (red, Pc+Nup93), not bound by Nup93 (blue, Pc-Nup93), and of non-Pc regions (grey). (B) Averaged amounts of inter-chromosomal Hi-C interactions between Pc domains that contain Nup93 ChIP-seq peaks (Nup93+), Pc domains that do not (Nup93-) and regions that do not contain Pc. Average inter-chromosomal Hi-C interactions between Pc domains that do not have Nup93 ChIP-seq peaks (Nup93-) and the whole genome were also analyzed (plotted below). P-values were calculated using two-tailed nonparametric t test (Mann Whitney test). (C) GB snapshot of the Bithorax/Fab-7 and ss loci, with arrows representing previously identified log-range interactions (see text). ChIP-seg of Nup93, Pc, and H3K27Me3, and Blue chromatin color domains are shown above. (D) Distributions of measured probe to periphery distances, for Fab-7 and ss loci, in cells treated with control (dsW) and lamin B/C RNAi (dslamin) using DAPI to define the periphery. Distributions are plotted as fractions of the cell population within assigned bins. Bars represent mean + SEM (> 300 cells per condition, 2 biological replicates). P values are from two-tailed unpaired non-parametric t test. Null hypothesis is "not being further away from nuclear periphery". RNAi treatments were performed for 2 rounds, 96 hours total (E) Distributions of measured probe to periphery distances, for Fab-7 and ss loci, in Kc cells treated with control (dsW) and Nup107 RNAi (dsNup107) using lamin IF to define the nuclear periphery. RNAi treatments were performed for 3 rounds, 144 hours total. Distributions are plotted as fractions of the cell population within assigned bins (left and middle). Violin plot of measured distances between centers of Fab-7 and ss Oligopaint probes in KC cells, treated with RNAi against control and Nup107 RNAi (right). Bars represent Mean + SEM (left and middle) and, lines depict median and quartiles (right). P values are from a twotailed unpaired non-parametric t test. Measurements are from >300 cells per condition; 4 biological replicates. In case of periphery distance measurements, null hypothesis is "not being further away from nuclear periphery".



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Supplemental Figure S7, related to Figure 7.

(A) Normalized RT-qPCR analysis of Nup93 and Ph expression in S2 cells treated with RNAi against control (dsWhite), Nup93 (dsNup93) or Ph (dsPh), for 2 rounds/96 hours total. Bars depict mean + SEM (5 biological replicates). (B) Average RNA FISH signal for *Abd-B* probe in S2 cells, calculated as an average number of probe-labeled dots per cell in indicated RNAi treatments (plot at left), and fraction of cells with at least one RNA FISH dot of *Abd-B* present in the 3 RNAi treatments (plot at right). 50 cells analyzed per condition. (C) Representative images of S2 cells treated with RNAi against control (dsW), Pc/Ph (dsPcPh) or Nup93 (dsNup93). Cells are labeled with Abd-B RNA FISH probes (red) and stained DNA (DAPI, blue). (D) IF of S2 cells treated with RNAi against control (dsW) or Nup93 (dsNup93), as described for Figure 7, using antibodies to nuclear pore marker mAb414 (red) and Pc or Ph (green). (E) Normalized RT-qPCR analysis of Nup93 and Nup107 expression from wing discs of wandering 3rd instar larvae with Nubbin-Gal4 driven RNAi against Nup93 or Nup107, relative to control Nubbin-Gal4 lines. Bars represent mean + SEM (3 biological replicates, 25+ animals each).