Figure S1. Monitoring of the retention of transplanted EVs by the live animal imaging system. (A) EVs was labeled with Dil. (B) At the indicated time points, the mice were sacrificed immediately, and the colon tissues were examined using the live animal imaging system. EVs, extracellular vesicles.

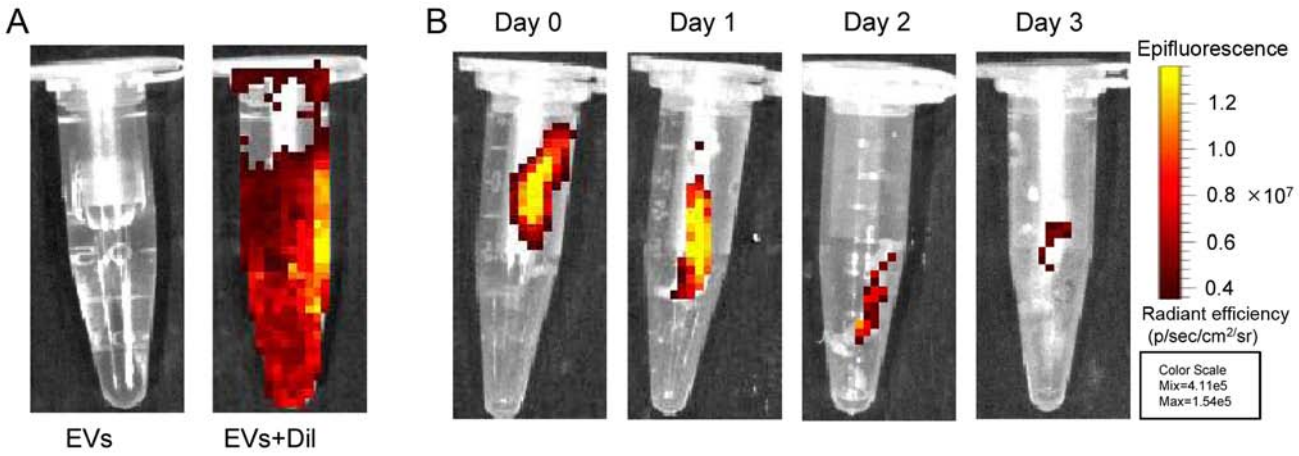

Figure S2. Densitometric analyses of the protein bands in Fig. 3B. Data are expressed as the means ± SD of 3 independent experiments. $^{*}P<0.05$, $^{**}P<0.01$ and $^{***}P<0.001$ vs. sham group. EVs, extracellular vesicles.

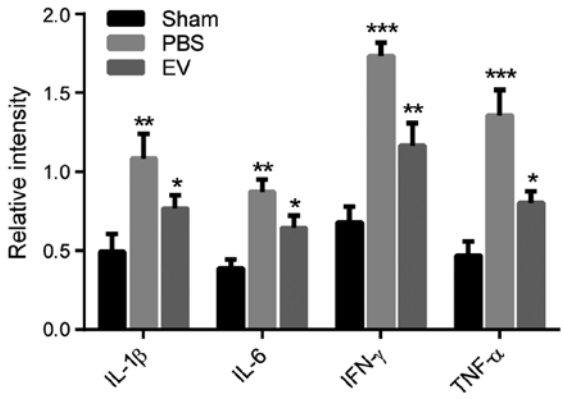

Figure S3. The mean of integrate optical density (IOD) of MPO was measured with image analysis and then statistically analyzed. Data are expressed as the means ± SD; (n=8). **P<0.01 vs. sham group; $^{\#}$P<0.05 vs. PBS group. EVs, extracellular vesicles.

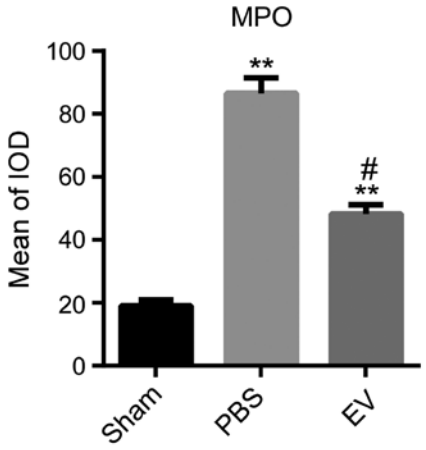

Figure S4. The mean of IOD of caspase-3 was measured with image analysis and then statistically analyzed. Data are expressed as the means ± SD; (n=8). *P<0.05, **P<0.01 vs. sham group; $^{\#\#}$P<0.01 vs. PBS group. EVs, extracellular vesicles.

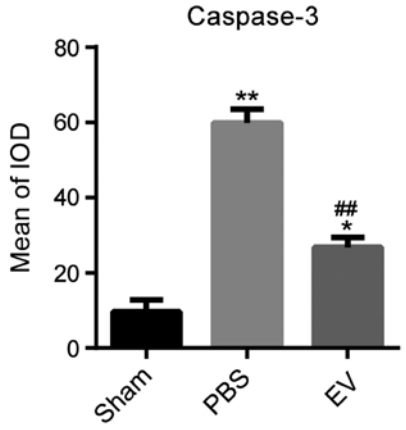

Figure S5. Detection of the expression of EpCAM in the colon tissues. (A) Representative images of immunohistochemical staining of EpCAM in the colon tissues of mice in the sham, PBS and EV groups; (n=8). Scale bar, 100 μm. The specific positive coloration of EpCAM is brown. (B) Representative images of immunofluorescence staining of EpCAM in the colon tissues of mice in sham, PBS and EV groups; (n=8) Scale bar, 100 μm. EVs, extracellular vesicles.

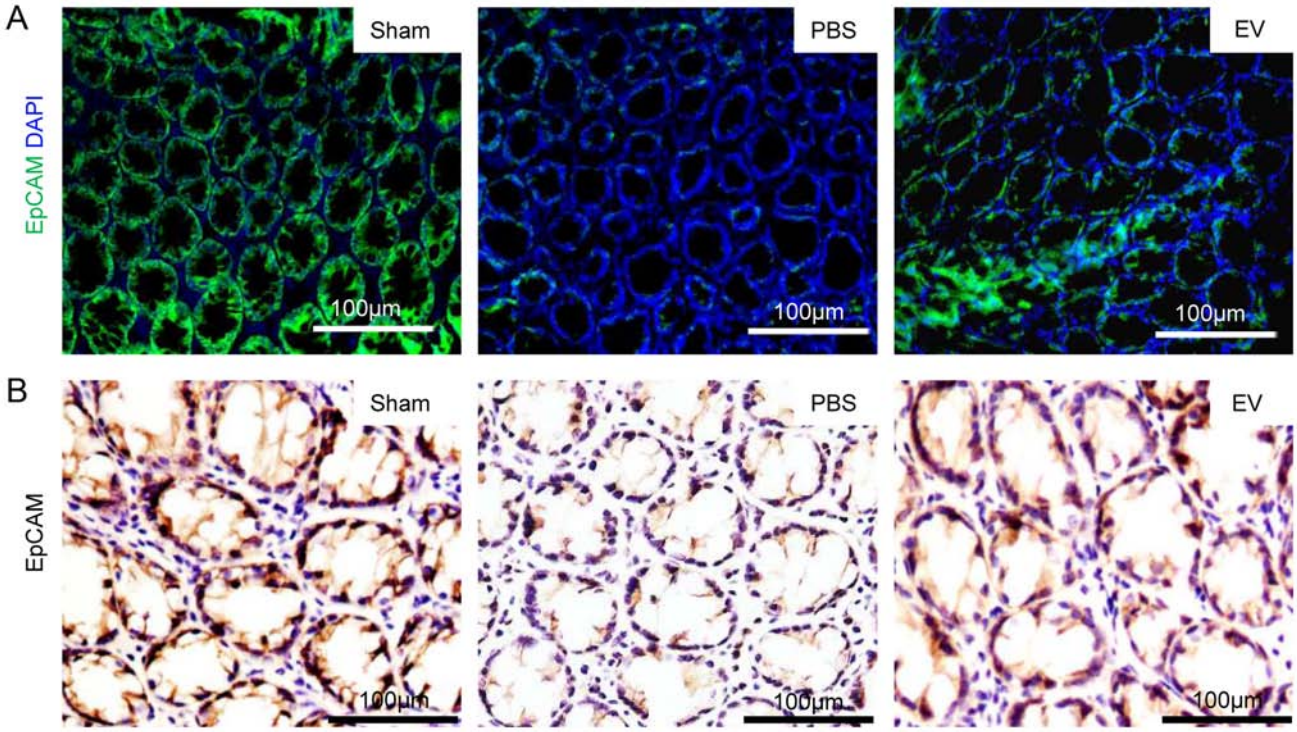

Figure S6. The mean of IOD of Ki67 were measured with image analysis and then statistically analyzed. Data are expressed as the means ± SD; (n=8). *P<0.05 vs. sham group; #P<0.05 vs. PBS group. EVs, extracellular vesicles.

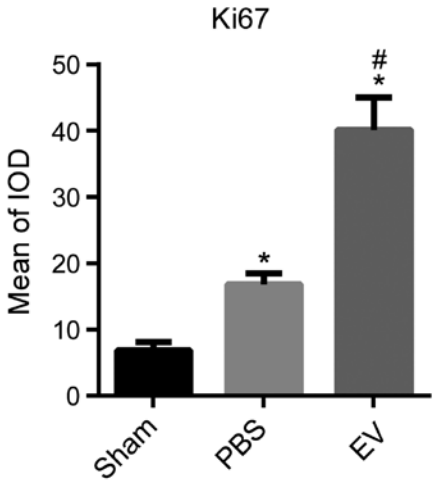

Table SI. Primers used for RT-qPCR.

Gene	Primer sequences
mIL-10	Forward: CTTACTGACTGGCATGAGGATCA Reverse: GCAGCTCTAGGAGCATGTGG
mTGF-β	Forward: CAACGCAATCTATGACAAAACC Reverse: TGAGCACTGAAGCGAAAGC
mIL-1β	Forward: GAAATGCCACCTTTTGACAGTG Reverse: TGGATGCTCTCATCAGGACAG
mIL-6	Forward: CTGCAAGAGACTTCCATCCAG Reverse: AGTGGTATAGACAGGTCTGTTGG
mIFN-γ	Forward: ACAGCAAGGCGAAAAAGGATG Reverse: TGGTGGACCACTCGGATGA
mTNF-α	Forward: CTGAACTTCGGGGTGATCGG Reverse: GGCTTGTCACTCGAATTTTGAGA
mOccludin	Forward: TGAAAGTCCACCTCCTTACAGA Reverse: CCGGATAAAAAGAGTACGCTGG
mClaudin	Forward: TGCCCCAGTGGAAGATTTACT Reverse: CTTTGCGAAACGCAGGACAT
mZO-1	Forward: GCTTTAGCGAACAGAAGGAGC Reverse: TTCATTTTTCCGAGACTTCACCA
mMMP-2	Forward: GGAAGCATCAAATCGGACTG Reverse: CACCCTCTTAAATCTGAAATCACC
mMMP-9	Forward: CCTGCGTATTTCCATTCATCTT Reverse: ATCCTGGTCATAGTTGGCGGT
Caspase-3	Forward: GGCACAAAGCGACTGGATG Reverse: CTGCCGTGGTACAGAACTGG
Caspase-8	Forward: ATCTGGCTCGGGGTTACTG Reverse: CTGCGTGGTGGTCATTCTC
Caspase-9	Forward: GGCTGTCTACGGCACAGATGGA Reverse: CTGGCTCGGGGTTACTGCCAG
β-actin	Forward: GTCAGGTCATCACTATCGGCAAT Reverse: AGAGGTCTTTACGGATGTCAACGT