

Electronic Supplementary Information

**Design and Synthesis of a Bivalent Probe Targeting the
Putative Mu Opioid Receptor and Chemokine Receptor
CXCR4 Heterodimer**

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Experimental

Chemical Syntheses

All reagents were purchased from Sigma-Aldrich or as otherwise stated. Melting points were obtained with a Fisher Scientific micro melting point apparatus without further correction. All IR spectra were recorded on a Nicolet iS10 FT-IR Instrument. Proton (400 MHz) and carbon-13 (100 MHz) nuclear magnetic resonance (NMR) spectra were acquired at ambient temperature with tetramethylsilane as the internal standard on a Bruker Ultrashield 400 Plus spectrometer. HRMS (ESI) analysis was performed on Perkin Elmer AxION 2 TOF mass spectrometer. HPLC analysis of the final compounds was achieved on Varian ProStar 210 system on Microsorb-MV 100-5 C18 column (250 mm × 4.6 mm) at 254 nm eluting with acetonitrile (0.01% TFA):water, 40:60 – 0:100 (1 & 3) or 45:55 – 0:100 (2), at 0.65 mL/min over 30 min. TLC analyses were carried out on Analtech Uniplate F254 plates. Chromatographic purification was accomplished on silica gel columns (230~400 mesh, Merck). Yields were not maximized.

General procedure for amide coupling – On an ice-water bath, a solution of acid in DMF, was added EDCI (2 eq.), HOBt (2 eq.), 4Å molecular sieves, and TEA (4.0 eq.) under N₂ protection. After 30 minutes, a solution of amine (1.0 eq.) in DMF was added dropwise. The resultant mixture was allowed to warm up to room temperature. After reaction completion, the mixture was filtered through celite. Solvent was removed in vacuum and the product was purified using column chromatography or recrystallization.

6β-naltrexamine hydrochloride salt The title compound was prepared following the reported procedure⁶³ in 60% yield in two steps. ¹H NMR (400 MHz, DMSO-*d*₆): δ 9.63 (s, 1H, exchangeable), 8.93 (brs, 1H, exchangeable), 8.56 (brs, 3H, exchangeable), 6.83 (d, *J* = 8.0 Hz,

1H), 6.66 (d, $J = 8.0$ Hz, 1H), 6.47 (brs, 1H), 4.71 (d, $J = 7.2$ Hz, 1H), 3.94 (d, $J = 5.6$ Hz, 1H), 3.35-3.27 (m, 2H), 3.06-3.00 (m, 2H), 2.92-2.87 (m, 1H), 2.78-2.74 (m, 1H), 2.46-2.44 (m, 2H), 2.01 (q, $J = 12.8$, 1H), 1.85 (d, $J = 14.0$ Hz, 1H), 1.79-1.75 (m, 1H), 1.44 (d, $J = 8.8$ Hz, 1H), 1.29 (td, $J_1 = 13.6$ Hz, $J_2 = 2.4$ Hz, 1H), 1.12-1.05 (m, 1H), 0.70-0.63 (m, 1H), 0.62-0.56 (m, 1H), 0.55-0.49 (m, 1H), 0.42-0.36 (m, 1H). ^{13}C NMR (100 MHz, DMSO- d_6): δ 141.62, 141.54, 129.06, 120.70, 119.83, 118.26, 88.08, 69.40, 61.28, 56.71, 52.35, 46.30, 45.62, 28.74, 27.14, 22.96, 21.39, 5.71, 5.08, 2.65.

3-(chloromethyl)-5,6-dihydro-6,6-dimethylimidazo[2,1-b]thiazole hydrochloride (5) A mixture of 4,4-dimethyl-imidazolidine-2-thione (500 mg, 3.84 mmol), 1,3-dichloroacetone (500 mg, 3.84 mmol), and acetonitrile (10 mL) was refluxed for 2 hr. The white precipitate was filtered off, dried, suspended in 1-methoxy-2-(2-methoxy-ethoxy)-ethane (5 mL), and subsequently heated at 145 °C for 2 hr. The precipitate was filtered off and washed with ether to give the hydrochloride product (571 mg, 56.3 %). ^1H NMR (400 MHz, DMSO- d_6 /D $_2$ O = 10:1): δ 7.07 (s, 1H), 4.85 (s, 2H), 4.22 (s, 2H), 1.50 (s, 6H). ^{13}C NMR (100 MHz, DMSO- d_6 /D $_2$ O = 10:1): δ 168.38, 133.30, 111.16, 69.47, 57.92, 35.96, 27.47. IR ν (Diamond, cm^{-1}) 3094, 2986, 2851, 2788, 1551, 1303, 1130. HRMS calc. m/z for $[\text{C}_8\text{H}_{11}\text{ClN}_2\text{S}]$: 202.0331, found: 203.0379 (M+H) $^+$.

trans-4-[(tert-butoxycarbonylamino)methyl]cyclohexanecarboxylic acid (6) To a solution of *trans*-4-(aminomethyl)cyclohexane-1-carboxylic acid (500 mg, 3.18 mmol) and NaHCO $_3$ (600 mg, 7.14 mmol) in 10 mL water, added a solution of Boc $_2$ O (834 mg, 3.82 mmol) in 1,4-dioxane (10 mL) at -5 °C. The pH was adjusted to 7-8 with additional NaHCO $_3$ (400 mg, 4.79 mmol). After stirred for 10 min, the solution was returned to room temperature and stirred for 24 hr. After 24 hr, 1,4-dioxane was removed under vacuum to give a white residue. The residue was extracted with

1:1 EtOAc/hexane system. The aqueous layer was cooled on ice bath and acidified to pH = 2 with HCl to give a white solid precipitate. The precipitate was filtered and washed with EtOAc to give the product (398 mg, 57%) ¹H NMR (400 MHz, DMSO-*d*₆): δ 12.01 (s, 1H, exchangeable), 6.78 (t, *J* = 5.5 Hz, 1H, exchangeable), 2.76 (t, *J* = 6.4 Hz, 2H), 2.09 (tt, *J*₁ = 3.6 Hz, *J*₂ = 12.2 Hz, 1H), 1.88-1.86 (m, 2H), 1.70-1.68 (m, 2H), 1.37 (s, 9H), 1.32-1.18 (m, 3H), 0.91-0.81 (m, 2H). ¹³C NMR (100 MHz, DMSO-*d*₆): δ 176.66, 155.65, 77.20, 45.95, 42.60, 37.33, 29.32, 28.21. IR ν (Diamond, cm⁻¹) 3366, 2924, 1686, 1522, 1251, 1166. HRMS calc. *m/z* for [C₁₃H₂₃NO₄]: 257.1627, found: 256.1551(M-H)⁻.

(*tert*-butoxy-*N*-({*trans*-4-[(phenylmethoxy)carbonyl-amino]cyclohexyl}methyl) carboxamide

(7) To a flask, **6** (250 mg, 0.971 mmol) was suspended in toluene (7 mL) and chilled in an ice-water bath under N₂. DPPA (267 mg, 0.971 mmol) and TEA (147 mg, 1.46 mmol) was added. The mixture was then warmed to room temperature and slowly heated to 80 °C. After 24 hr, the mixture was cooled to 35 °C and benzyl alcohol (340 mg, 3.15 mmol) was added via syringe. The mixture was heated to 80 °C and stirred for 24 hr. The reaction was concentrated in vacuum and the residue was treated with water (30 mL) and EtOAc (30 mL). The aqueous layer was extracted with EtOAc (30 mL). The combined organic layers were washed with concentrated HCl (25 mL), aqueous sodium bicarbonate (25 mL), and brine (25 mL), then dried over Na₂SO₄ and concentrated in vacuum to afford the white solid product (120 mg, 23.9%). ¹H NMR (400 MHz, DMSO-*d*₆): δ 7.38-7.28 (m, 5H), 7.10 (d, *J* = 8.0 Hz, 1H, exchangeable), 6.75 (t, *J* = 5.6 Hz, 1H, exchangeable), 4.99 (s, 2H), 3.25-3.16 (m, *J* = 4.0 Hz, 1H), 2.75 (t, *J* = 6.4 Hz, 2H), 1.81-1.78 (m, 2H), 1.67-1.64 (m, 2H), 1.37 (s, 9H), 1.31-1.22 (m, 1H), 1.16-1.06 (m, 2H), 0.93-0.84 (m, 2H). ¹³C NMR (100 MHz, DMSO-*d*₆): δ 155.62, 155.18, 137.22, 128.21, 127.62, 127.59, 77.19, 64.92, 49.78, 45.74, 37.04, 32.03, 28.98, 28.19. IR ν (Diamond, cm⁻¹) 3346, 2930, 1626, 1439, 1521, 1245, 1171.

HRMS calc. m/z for $[C_{20}H_{30}N_2O_4]$: 362.2206, found 385.2054 ($M+Na$)⁺.

***tert*-butyl ((*trans*-4-aminocyclohexyl)methyl)carbamate (8)** A solution of **7** (100 mg, 0.276 mmol) in methanol (15 mL) was treated with Pd/C (10 mg, 10% wt). The resultant solution was reacted at 10 psi H₂ for 3 hr at room temperature. The reaction mixture was filtered through celite pad and washed with methanol to yield a white solid (84 mg, 100%). ¹H NMR (400 MHz, CDCl₃): δ 4.59 (s, 1H), 2.97-2.94 (m, 4H), 2.68-2.61 (m, 1H), 1.90-1.88 (m, 2H), 1.77-1.74 (m, 2H), 1.43 (s, 9H), 1.39-1.35 (m, 1H), 1.17-1.07 (m, 2H), 1.02-0.91 (m, 2H). ¹³C NMR (100 MHz, CDCl₃): δ 156.21, 79.22, 50.77, 46.61, 37.83, 35.56, 29.51, 28.58. IR ν (Diamond, cm⁻¹) 3344, 2929, 2848, 1683, 1530, 1246, 1168. HRMS calc. m/z for $[C_{12}H_{24}N_2O_2]$: 228.1838, found: 229.1845 ($M+H$)⁺.

Isothiocyanatocyclohexane (9) To a solution of cyclohexylamine (1.00g, 10.08 mmol), and triethylamine (3.36 g, 33.26 mmol) in 10 mL THF, carbon disulfide (767 mg, 10.08 mmol) was added in 40 min at 0 °C. The resulted solution was stirred at room temperature for 2 hr. The mixture was cooled to 0 °C and 4-toluene sulfonyl chloride (2.11g, 11.09 mmol) was added. The mixture was allowed to warm to room temperature and stirred for 2 hr. 1M Hydrochloric acid (20 mL) was then added. The mixture was extracted with hexane and Et₂O (1:1, 20 mL). The combined organic layers were dried over Na₂SO₄ and concentrated in vacuum. The resulting residue was purified by column chromatography to yield a clear oil (893 mg, 63%). ¹H NMR (400 MHz, DMSO-*d*₆): δ 3.90 (m, $J = 4.0$ Hz, 1H), 1.89-1.83 (m, 2H), 1.60-1.57 (m, 4H), 1.44-1.34 (m, 4H). ¹³C NMR (100 MHz, DMSO-*d*₆): δ 128.39, 54.91, 32.42, 24.43, 22.59. IR ν (Diamond, cm⁻¹) 2933, 2856, 2174, 2092, 2056, 1448, 1360, 1310, 1134.

***tert*-Butyl (((1*r*,4*r*)-4-(3-cyclohexylthioureido)cyclohexyl)methyl)carbamate (10)** To a solution of **9** (100 mg, 0.71 mmol) in 10 mL CH₂Cl₂ was added **8** (162 mg, 0.71 mmol). After

stirring at room temperature for 24 hr, the solvent was removed in vacuum. The white residue was recrystallized using Et₂O to yield a white powder (189 mg, 72%). ¹H NMR (400 MHz, DMSO-*d*₆): δ 7.13 (t, *J* = 7.6 Hz, 2H, exchangeable), 6.80 (t, *J* = 5.65, 1H), 3.87 (bs, 2H), 2.76 (t, *J* = 6.4 Hz, 2H), 1.92-1.83 (m, 4H), 1.67-1.57 (m, 5H), 1.37 (m, 9H), 1.32-1.22 (s, 3H), 1.31-1.02 (m, 5H), 0.94-0.85 (m, 2H). ¹³C NMR (100 MHz, DMSO-*d*₆): δ 179.98, 155.61, 77.18, 52.02, 51.39, 45.69, 37.09, 32.24, 31.78, 28.96, 28.19, 25.12, 24.38. IR ν (Diamond, cm⁻¹) 3374, 2936, 1685, 1556, 1406, 1297, 1152. HRMS calc. *m/z* for [C₁₉H₃₅N₃O₂S]: 369.2450, found: 392.2322 (M+Na)⁺.

tert-Butyl (((1*r*,4*r*)-4-(((*Z*)-(cyclohexylamino))((6,6-dimethyl-5,6-dihydroimidazo[2,1-*b*]thiazol-3-yl)methyl)thio)methylene)amino)cyclohexyl)methyl)carbamate (11) A solution of **10** (50 mg, 0.135 mmol) and **5** (32 mg, 0.135 mmol) in 3 mL acetonitrile was refluxed for 24 hr. The precipitate was filtered off and recrystallized from methanol/ether to give the product (43 mg, 51%). ¹H NMR (400 MHz, DMSO-*d*₆/D₂O = 10:1): δ 6.88 (s, 1H), 4.88 (s, 2H), 4.27 (s, 2H), 3.88 (s, 1H), 3.67 (s, 1H), 2.78 (m, 2H), 1.71-1.54 (m, 13H), 1.50 (s, 6H), 1.37 (s, 9H), 1.30-0.95 (m, 6H). ¹³C NMR (100 MHz, DMSO-*d*₆/D₂O = 10:1): δ 168.34, 160.32, 155.72, 131.36, 110.22, 77.40, 69.29, 58.15, 56.30, 53.34, 45.61, 45.48, 40.08, 36.74, 32.07, 31.41, 30.96, 30.33, 28.51, 28.23, 28.04, 27.36, 24.51, 24.32. IR ν (Diamond, cm⁻¹) 3333, 2929, 1698, 1610, 1499, 1250, 1172, 1027. HRMS calc. *m/z* for [C₂₇H₄₅N₅O₂S₂]: 535.3015, found: 536.2936 (M+H)⁺.

aminomethyl-substituted IT1t (4) To a solution of **11** (500 mg, 0.821 mmol) in 10 mL CH₂Cl₂ was added TFA (1 mL). After being stirred for 0.5 hr, Et₂O (20 mL) was added. The precipitate

was filtered off and washed again with Et₂O to give a white solid as the trifluoroacetic acid salt (511 mg, 70.4 %). ¹H NMR (400 MHz, DMSO-*d*₆): δ 10.56 (s, 1H, exchangeable), 9.52 (s, 1H, exchangeable), 9.37 (s, 1H, exchangeable), 7.91 (s, 3H, exchangeable), 6.85 (s, 1H), 4.67 (s, 2H), 4.22 (s, 2H), 3.77-3.69 (m, 2H), 2.66 (m, 2H), 1.84-1.71 (m, 8H), 1.62-1.56 (m, 2H), 1.50 (s, 6H), 1.46-1.25 (m, 5H), 1.08-0.97 (m, 3H). ¹³C NMR (100 MHz, DMSO-*d*₆): δ 168.40, 160.50, 158.35, 131.27, 116.95, 110.24, 69.33, 58.03, 56.38, 55.76, 53.23, 52.86, 43.97, 43.85, 34.34, 32.07, 30.99, 30.01, 28.09, 27.86, 27.37, 24.55, 24.28. IR ν (Diamond, cm⁻¹) 2936, 1669, 1607, 1556, 1451, 1310, 1126. HRMS calc. m/z for [C₂₂H₃₇N₅S₂]: 435.2490, found: 436.2321 (M+H)⁺.

benzyl 6-aminohexylcarbamate (12) To a solution of 1,6-diaminohexane (2.32 g, 20 mmol) in CH₂Cl₂-MeOH (100 mL: 100 mL) was added a solution of CbzCl (3.08 g, 18 mmol) in CH₂Cl₂ (200 mL) dropwise over 8 hr while keeping the temperature below 0 °C. The mixture was stirred at the same temperature for another 16 hr before the solvent was removed under vacuum. The white residue was then dissolved in 100 mL water and acidified to pH = 2 using concentrated HCl. The resulting residue was then extracted using CH₂Cl₂ and stirred on ice. The organic solution was then basified using NaOH solution until pH = 12. The residue was then extracted again using CH₂Cl₂. The organic layer was dried with Na₂SO₄ and concentrated in vacuum to yield a white solid (2.03 g, 40.6 %) ¹H NMR (400 MHz, CDCl₃): δ 7.36-7.34 (m, 4H), 7.33-7.29 (m, 1H), 5.10 (s, 2H), 4.72 (s, 1H, exchangeable), 3.19 (q, *J* = 6.8 Hz, 2H), 2.68 (t, *J* = 6.4 Hz, 2H), 1.55-1.48 (m, 2H), 1.45-1.41 (m, 2H), 1.35-1.32 (m, 4H), 1.26 (s, 2H). ¹³C NMR (100 MHz, CDCl₃): δ 156.55, 136.87, 128.66, 128.22, 66.76, 42.21, 41.20, 33.76, 30.13, 26.71, 26.65. IR ν (Diamond, cm⁻¹) 3332, 2940, 2855, 1682, 1524, 1453, 1302, 1282, 1255, 1228, 1133, 1096, 1000. HRMS calc. m/z for [C₁₄H₂₂N₂O₂]: 250.1681, found: 251.1760 (M+H)⁺.

3,12-dioxo-1-phenyl-2,14-dioxo-4,11-diazahexadecan-16-oic acid (13) To a stirring solution of **12** (6.09 g, 24.3 mmol) in 50 mL THF was added diglycolic anhydride (2.96 g, 25.5 mmol) in three portions. The resultant mixture was stirred at ambient temperature for 23 hr. After THF was removed under reduced pressure, the residue was crystallized by EtOAc–hexane to give a white solid (6.91g, 77%). ¹H NMR (400 MHz, DMSO-*d*₆): δ 12.78 (s, 1H, exchangeable), 7.80 (bs, 1H, exchangeable), 7.40-7.28 (m, 5H), 7.19 (bs, 1H, exchangeable), 5.00 (s, 2H), 4.10 (s, 2H), 3.94 (s, 2H), 3.08 (q, *J* = 6.8 Hz, 2H), 2.98 (q, *J* = 6.8 Hz, 2H), 1.42-1.37 (m, 4H), 1.24 (br, 4H). ¹³C NMR (100 MHz, DMSO-*d*₆): δ 171.32, 168.45, 155.99, 137.26, 128.23, 127.61, 70.16, 67.88, 64.99, 38.00, 29.25, 28.97, 25.96, 25.84. IR ν (Diamond, cm⁻¹) 3338, 3305, 2959, 2928, 1690, 1651, 1541, 1264, 1237, 1172, 1141, 1008. HRMS calc. *m/z* for [C₁₈H₂₆N₂O₆]: 366.1791, found: 389.1674 (M+Na)⁺.

Benzyl (6-(2-(2-(((4R,4aS,7R,7aR)-3-(cyclopropylmethyl)-4a,9-dihydroxy-2,3,4,4a,5,6,7,7a-octahydro-1H-4,12-methanobenzofuro[3,2-*e*]isoquinolin-7-yl)amino)-2-

oxoethoxy)acetamido)hexyl)carbamate (14) The title compound was prepared according to the general amide coupling procedure by reacting **13** with 6β-naltrexamine hydrochloride salt in DMF. The resulting residue was dissolved in MeOH and 5 eq. of K₂CO₃ was added and stirred for 24 hr. After 24 hr, the K₂CO₃ was filtered out and the solution was concentrated to dryness to afford a brown solid. The solid was purified with column chromatography (CH₂Cl₂-MeOH, 30:1) and recrystallized with MeOH/Et₂O to afford an orange solid product (1.21 g, 72%) ¹H NMR (400 MHz, CDCl₃): δ 7.60 (d, *J* = 9.2 Hz, 1H, exchangeable), 7.35-7.28 (m, 5H), 7.02 (t, *J* = 5.6 Hz, 1H, exchangeable), 6.71 (m, 1H), 6.55 (m, 1H), 5.09 (s, 2H), 4.95 (t, *J* = 5.6 Hz, 1H, exchangeable), 4.43 (d, *J* = 6.0 Hz, 1H), 4.11-3.99 (m, 4H), 3.31-3.24 (m, 2H), 3.21-3.17 (m, 2H),

3.09 (d, $J = 6.0$ Hz, 1H), 3.02 (d, $J = 18.4$ Hz, 1H), 2.64-2.58 (m, 2H), 2.40-2.30 (m, 2H), 2.23-2.14 (m, 2H), 1.84-1.74 (m, 1H), 1.66-1.60 (m, 1H), 1.53-1.45 (m, 7H), 1.35-1.34 (m, 4H), 1.14-1.11 (m, 1H), 0.85-0.75 (m, 1H), 0.55-0.50 (m, 2H), 0.14-0.10 (m, 2H). ^{13}C NMR (100 MHz, CDCl_3): δ 168.82, 168.61, 156.78, 143.32, 139.61, 136.76, 130.75, 128.65, 128.23, 128.18, 124.76, 119.32, 117.89, 92.39, 77.36, 71.09, 71.06, 70.27, 66.81, 62.56, 59.54, 49.68, 47.37, 46.17, 44.03, 40.84, 38.85, 31.94, 30.00, 29.56, 29.17, 26.32, 26.11, 23.37, 22.77, 9.57, 4.12, 3.96. IR ν (Diamond, cm^{-1}) 3297, 2929, 1651, 1538, 1505, 1454, 1323, 1239, 1126. HRMS calc. m/z for $[\text{C}_{38}\text{H}_{50}\text{N}_4\text{O}_8]$: 690.3629, found: 691.3459 ($\text{M}+\text{H}$) $^+$.

N-(6-aminohexyl)-2-(2-(((4R,4aS,7R,7aR)-3-(cyclopropylmethyl)-4a,9-dihydroxy-2,3,4,4a,5,6,7,7a-octahydro-1H-4,12-methanobenzofuro[3,2-e]isoquinolin-7-yl)amino)-2-

oxoethoxy)acetamide (15) A solution of **14** (250 mg, 0.36 mmol) in methanol (20 mL) was hydrogenated in the presence of 10% Pd/C (25 mg) under a H_2 atmosphere (10 psi) at room temperature for 3 hr. The mixture was filtered, and the filtrate was concentrated to a white foam (200 mg, 99%) ^1H NMR (400 MHz, $\text{DMSO}-d_6$): δ 8.24 (d, $J = 8.4$ Hz, 1H, exchangeable), 8.05 (t, $J = 5.2$ Hz, 1H, exchangeable), 6.57 (m, 1H), 6.52 (m, 1H), 4.89 (brs, 1H, exchangeable), 4.59 (d, $J = 7.6$ Hz, 1H), 3.94 (m, 4H), 3.56-3.48 (m, 1H), 3.14 (m, $J = 6.4$ Hz, 2H), 3.01-2.94 (m, 2H), 2.60-2.56 (m, 2H), 2.38-2.28 (m, 2H), 2.15 (dt, $J_1 = 4.4$ Hz, $J_2 = 12.0$ Hz, 1H), 1.97 (dt, $J_1 = 3.2$ Hz, $J_2 = 12.0$ Hz, 1H), 1.84-1.75 (m, 1H), 1.46-1.44 (m, 4H), 1.33-1.23 (m, 9H), 0.87-0.79 (m, 1H), 0.48-0.46 (m, 2H), 0.11 (m, 2H). ^{13}C NMR (100 MHz, $\text{DMSO}-d_6$): δ 168.36, 168.19, 142.06, 140.51, 131.20, 123.30, 118.32, 117.07, 90.43, 70.40, 70.35, 69.53, 61.72, 58.35, 50.62, 46.99, 43.64, 41.43, 38.09, 33.10, 30.25, 30.02, 29.15, 26.27, 26.05, 24.56, 22.13, 9.19, 3.62, 3.49. IR ν

(Diamond, cm^{-1}) 3269, 2924, 1651, 1548, 1454, 1373, 1322, 1258, 1127, 1034. HRMS calc. m/z for $[\text{C}_{30}\text{H}_{44}\text{N}_4\text{O}_6]$: 556.3261, found: 557.3213 ($\text{M}+\text{H}$)⁺.

18-(((4R,4aS,7R,7aR)-3-(cyclopropylmethyl)-4a,9-dihydroxy-2,3,4,4a,5,6,7,7a-octahydro-1H-4,12-methanobenzofuro[3,2-e]isoquinolin-7-yl)amino)-5,14,18-trioxo-3,16-dioxo-6,13-diazaoctadecanoic acid (16) Diglycolic anhydride (125 mg, 1.08 mmol) was added to the solution of **15** (600 mg, 1.08 mmol) in DMF (5 mL). The resultant mixture was stirred at ambient temperature for 3 hr. After removal of DMF under reduced pressure, the residue was recrystallized by MeOH/Et₂O to give a white solid (586 mg, 80%). ¹H NMR (400 MHz, DMSO-*d*₆): δ 8.70 (br, 1H, exchangeable), 8.36 (d, $J = 8.4$ Hz, 1H, exchangeable), 8.20 (br, 1H, exchangeable), 6.62 (m, 1H), 6.53 (m, 1H), 4.66 (d, $J = 7.6$ Hz, 1H), 3.95 -3.93 (m, 8H), 3.56-3.48 (m, 1H), 3.15-3.00 (m, 6H), 2.70-2.60 (m, 2H), 2.55-2.52 (m, 1H), 2.43-2.39 (m, 1H), 2.20 (dt, $J_1 = 4.0$ Hz, $J_2 = 12.0$ Hz, 1H), 2.09-2.02 (m, 1H), 1.87-1.78 (m, 1H), 1.50-1.33 (m, 6H), 1.29-1.24 (m, 6H), 0.90-0.87 (m, 1H), 0.49 (m, $J = 7.2$ Hz, 2H), 0.17-0.16 (m, 2H). ¹³C NMR (100 MHz, DMSO-*d*₆): δ 168.37, 168.16, 142.04, 140.61, 130.97, 122.84, 118.37, 117.13, 90.31, 70.35, 70.31, 69.51, 61.71, 58.10, 50.56, 46.75, 44.00, 37.96, 29.96, 29.76, 28.94, 28.88, 25.90, 25.87, 24.39, 22.25, 8.62, 3.75, 3.78. IR ν (Diamond, cm^{-1}) 2931, 1651, 1599, 1504, 1432, 1393, 1253, 1126, 1034. HRMS calc. m/z for $[\text{C}_{34}\text{H}_{48}\text{N}_4\text{O}_{10}]$: 672.3370, found: 673.3519 ($\text{M}+\text{H}$)⁺.

2-(2-(methylamino)-2-oxoethoxy)acetic acid (17) To a cooled 2M solution of methylamine in THF (10 mL), diglycolic anhydride (2.00 g, 17.2 mmol) was added. The resulted solution was stirred for 5 min on ice bath and then allowed to continue at ambient temperature for 24 hr. The resulting yellow oil was concentrated under vacuum and recrystallized using MeOH-Et₂O to yield

an off white solid (1.704g, 67%). ^1H NMR (400 MHz, DMSO- d_6): δ 8.07 (s, 1H, exchangeable), 4.00 (s, 2H), 3.92 (s, 2H), 2.62 (d, J = 4.72 Hz, 3H). ^{13}C NMR (100 MHz, DMSO- d_6): δ 171.28, 169.08, 70.13, 67.83, 25.11. IR ν (Diamond, cm^{-1}) 3387, 2919, 2524, 1707, 1627, 1258, 1203, 1130, 1084. HRMS calc. m/z for $[\text{C}_5\text{H}_9\text{NO}_4]$: 147.0532, found: 146.4232 (M-H) $^-$.

benzyl (6-(2-(2-(methylamino)-2-oxoethoxy)acetamido)hexyl)carbamate (18) The title compound was prepared according to the general amide coupling procedure by reacting acid **17** with amine **12**. The residue was then purified using column chromatography (CH_2Cl_2 -MeOH, 20:1) to afford the product (2.79 g, 50%) ^1H NMR (400 MHz, DMSO- d_6): δ 7.97 (m, 2H, exchangeable), 7.35 (m, 5H), 7.27 (m, 1H, exchangeable), 5.00 (s, 2H), 3.93 (m, 4H), 3.12 (m, 2H), 2.98 (m, 2H), 2.66 (d, J = 4.68 Hz, 3H), 2.51 (m, 2H), 1.40 (m, 4H), 1.26 (m, 4H). ^{13}C NMR (100 MHz, DMSO- d_6): δ 168.87, 168.27, 156.06, 137.32, 128.28, 128.08, 127.66, 127.62, 70.32, 65.05, 40.22, 40.01, 39.80, 39.59, 39.38, 39.17, 38.96, 38.06, 29.53, 29.31, 29.15, 26.04, 25.89, 25.06. IR ν (Diamond, cm^{-1}) 3337, 2924, 2855, 1690, 1652, 1543, 1454, 1435, 1407, 1367, 1279, 1148, 1124, 1059. HRMS calc. m/z for $[\text{C}_{19}\text{H}_{29}\text{N}_3\text{O}_5]$: 379.2107, found: 402.2018 (M+Na) $^+$.

***N*-(6-aminohexyl)-2-(2-(methylamino)-2-oxoethoxy)acetamide (19)** A solution of **18** (400 mg, 1.21 mmol) and Pd/C (40 mg, 10%) in 25 mL MeOH was shaken at 60 psi of H_2 . After 24 hr the solution was filtered through celite and concentrated under vacuum. The residue was recrystallized using MeOH:Et $_2$ OH to afford a white solid (258 mg, 100%) ^1H NMR (400 MHz, DMSO- d_6): δ 8.00 (br, 2H, exchangeable), 3.90 (m, 4H), 3.13-3.08 (m, 2H), 2.65 (d, J = 4.68 Hz, 3H), 2.54-2.52 (m, 1H), 1.44-1.39 (m, 2H), 1.35-1.31 (m, 2H), 1.26 (m, 4H) ^{13}C NMR (100 MHz, DMSO- d_6): δ 70.29, 48.55, 41.32, 40.19, 39.98, 39.77, 39.56, 39.36, 39.15, 38.94, 38.07, 32.79, 29.21, 26.27,

26.04, 25.06. IR ν (Diamond, cm^{-1}) 3318, 2924, 2855, 1652, 1548, 1432, 1338, 1124, 1004. HRMS calc. m/z for $[\text{C}_{11}\text{H}_{23}\text{N}_3\text{O}_3]$: 245.1739, found: 246.1798 ($\text{M}+\text{H}$)⁺.

3,7,16-trioxo-5,18-dioxa-2,8,15-triazaicosan-20-oic acid (20) To a 25 mL flask added **19** (188 mg, 0.770 mmol), diglycolic anhydride (95 mg, 0.880 mmol), and 3 mL CH_2Cl_2 . After 24 hr of stirring at ambient temperature, the solution was concentrated under vacuum to afford a faint yellow oil. Recrystallization using MeOH:Et₂OH afforded a faint yellow solid (157 mg, 56%) ¹H NMR (400 MHz, DMSO-*d*₆): δ 7.99 (m, 2H, exchangeable), 7.82 (m, 1H, exchangeable), 4.10 (s, 2H), 3.94 (s, 2H), 3.90(s, 4H), 3.13-3.06 (m, 4H), 2.65 (d, $J = 4.68$ Hz, 3H), 1.42 (m, 4H), 1.25 (m, 4H) ¹³C NMR (100 MHz, DMSO-*d*₆): δ 171.31, 168.81, 168.47, 168.20, 70.23, 70.15, 67.87, 40.13, 39.92, 39.71, 39.50, 39.30, 39.09, 38.88, 37.97, 37.96, 29.07, 28.94, 25.96, 25.94, 25.00. IR ν (Diamond, cm^{-1}) 3310, 2930, 1651, 1538, 1469, 1417, 1362, 1338, 1124, 1036. HRMS calc. m/z for $[\text{C}_{15}\text{H}_{27}\text{N}_3\text{O}_7]$: 361.1849, found: 384.1774 ($\text{M}+\text{Na}$)⁺.

Bivalent ligand (1) The title compound was prepared according to the general amide coupling procedure by reacting **16** with the aminomethyl-substituted Itlt **4** in DMF. The resulting crude residue was purified using column chromatography (CH_2Cl_2 -MeOH, 10:1) to afford the product (104 mg, 38%). ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.01 (m, 1H, exchangeable), 8.21 (m, 1H, exchangeable), 8.19-8.01 (m, exchangeable, 3H), 6.58 (m, 1H), 6.52 (m, 1H), 4.59 (d, $J = 7.80$ Hz, 1H), 3.94 (m, 8H), 3.29 (s, 14H), 3.16 (m, 6H), 3.01 (m, 3H), 2.94 (m, 2H) 2.84 (m, 2H), 2.69 (m, 3H) 2.57 (m, 3H) 2.33 (m, 3H), 2.18-2.11 (m, 1H), 2.00-1.95 (m, 1H), 1.69 (m, 6H), 1.46 (m, 9H) 1.27 (m, 10H), 1.20 (m, 9H), 1.00-0.90 (m, 2H), 0.82 (m, 2H), 0.47 (m, 2H) 0.14 (m, 2H). ¹³C NMR (100 MHz, DMSO-*d*₆): δ 169.31, 168.50, 168.38, 168.24, 141.00, 141.10, 119.05, 117.71, 89.72, 83.47, 70.31, 69.58, 63.36, 61.65, 55.65, 55.42, 50.26, 45.73, 43.89, 43.03, 40.09, 39.88,

39.68, 39.47, 39.26, 39.05, 38.84, 38.00, 36.64, 33.85, 33.64, 33.23, 30.17, 29.42, 29.06, 28.61, 28.37, 27.67, 27.41, 26.09, 25.96, 25.43, 24.97, 23.74, 23.61, 22.77. IR ν (Diamond, cm^{-1}) 3611, 3451, 3252, 3003, 1657, 1553, 1501, 1466, 1430, 1361, 1202, 1188, 1128, 1059. HRMS calc. m/z for $[\text{C}_{56}\text{H}_{83}\text{N}_9\text{O}_9\text{S}_2]$: 1089.5755, found: 1090.5825 ($\text{M}+\text{H}$)⁺.

MOR Monovalent Control (2) The title compound was prepared according to the general amide coupling procedure by reacting **20** with 6 β -naltrexamine hydrochloride salt. After starting material was consumed, the reaction mixture was filtered through celite and concentrated under vacuum. The resulting residue was dissolved in 5 mL MeOH and stirred at room temperature with K_2CO_3 (120 mg, 0.875 mmol). After 24 hr, K_2CO_3 was filtered out and the filtrate was concentrated to dryness to afford a brown solid. The residue was then purified using column chromatography (CH_2Cl_2 -MeOH, 5:1) to afford the product (45 mg, 19%). ^1H NMR (400 MHz, $\text{DMSO}-d_6$): δ 9.01 (brs, 1H, exchangeable), 8.21 (d, $J = 8.4$ Hz, 1H, exchangeable), 8.03-7.96 (m, 3H, exchangeable), 6.58 (m, 1H), 6.52 (m, 1H), 4.88 (brs, 1H, exchangeable), 4.59 (d, $J = 7.6$ Hz, 1H), 3.95 (s, 2H), 3.93 (s, 2H), 3.91 (s, 4H), 3.56-3.48 (m, 1H), 3.17-3.09 (m, 4H), 3.01-2.95 (m, 2H), 2.65 (d, $J = 4.4$ Hz, 3H), 2.62-2.57 (m, 2H), 2.38-2.28 (m, 2H), 2.19-2.11 (m, 1H), 2.01-1.94 (m, 1H), 1.83-1.74 (m, 1H), 1.46-1.44 (m, 6H), 1.32-1.23 (m, 6H), 0.87-0.81 (m, 1H), 0.48-0.46 (m, 2H), 0.12-0.11 (m, 2H). ^{13}C NMR (100 MHz, $\text{DMSO}-d_6$): δ 168.80, 168.33, 168.22, 168.14, 142.02, 140.35, 131.22, 123.40, 118.28, 116.98, 90.43, 70.36, 70.31, 70.24, 69.51, 61.71, 58.34, 50.60, 46.97, 43.62, 38.01, 37.97, 30.24, 29.99, 29.10, 25.98, 25.01, 24.51, 22.13, 9.16, 3.59, 3.46. IR ν (Diamond, cm^{-1}) 3288, 2929, 1651, 1548, 1504, 1454, 1323, 1239, 1185, 1124, 1035. HRMS calc. m/z for $[\text{C}_{35}\text{H}_{51}\text{N}_5\text{O}_9]$: 685.3687, found: 686.3595 ($\text{M}+\text{H}$)⁺.

CXCR4 Monovalent Control (3) The title compound was prepared according to the general amide coupling procedure by reacting **20** with the aminomethyl-substituted It1t **4**. The residue was then purified using column chromatography (CH₂Cl₂-MeOH, 30:1) to afford the product (47 mg, 11%) ¹H NMR (400 MHz, DMSO-*d*₆): δ 8.03-7.95 (m, 4H, exchangeable), 3.92-3.86 (m, 8H), 3.71 (s, 2H), 3.69-3.66 (m, 1H, exchangeable), 3.36-3.12 (m, 2H), 3.15-3.07 (m, 4H), 2.99-2.95 (m, 2H), 2.93-2.87 (m, 2H), 2.83 (m, 2H), 2.65 (d, *J* = 4.6 Hz, 3H), 1.72-1.40 (m, 4H), 1.36-1.26 (m, 3H), 1.20-1.19 (m, 5H), 1.06-1.02 (m, 8H), 1.00-0.98 (d, *J* = 5.8 Hz, 6H), 0.90-0.79 (m, 3H) ¹³C NMR (100 MHz, DMSO-*d*₆): δ 168.90, 168.58, 168.48, 168.30, 148.75, 84.48, 74.29, 70.27, 70.15, 55.51, 54.79, 40.00, 39.79, 39.58, 39.37, 39.16, 38.95, 38.74, 37.87, 36.86, 33.74, 33.32, 29.07, 28.91, 25.97, 24.93, 23.67. IR ν (Diamond, cm⁻¹) 3289, 2924, 2850, 2359, 2161, 2027, 1651, 1544, 1435, 1363, 1120, 1031. HRMS calc. *m/z* for [C₃₇H₆₂N₈O₆S₂]: 778.4234, found: 779.4079 (M+H)⁺.

Radioligand Binding Assay

The competition-binding assay was conducted to determine the affinity of the bivalent compound and its monovalent control at the mu opioid receptor (MOR) using monoclonal mouse opioid receptor expressed in Chinese hamster ovary (CHO) cell lines. MOR-CHO cells were cultured and maintained in DMEM (90%), and fetal bovine serum 10% (v/v). [³H]naloxone ([³H]NLX) was used to label the MOR and a saturation assay was conducted to determine the K_d and B_{max} values for [³H]NLX at the MOR, which were measured as 1.81 ± 0.16 nM and 2.47 ± 0.20 pmol/mg, respectively. To determine the binding affinity of test compounds, 30 μg of membrane protein was incubated with [³H]NLX in the presence of different concentrations of test compounds in TME buffer (50 mM Tris, 3 mM MgCl₂, and 0.2 mM EGTA, pH 7.7) for 1.5

h at 30 °C. The bound radioligand was separated by filtration using the Brandel harvester. Specific (i.e., opioid receptor-related) binding at the KOR was determined as the difference in binding obtained in the absence and presence of 5 μ M naltrexone. The IC₅₀ values were determined and converted to K_i values using the Cheng–Prusoff equation.

Antibody Binding Assays

CHO-CXCR4 cells were maintained in RPMI1640 medium supplemented with 10% (v/v) FBS, 100 units/mL penicillin, 100 mg/mL streptomycin and 2 mM L-glutamine, and 400 μ g/mL geneticin. Following trypsinization, CHO-CXCR4 cells were washed twice with FACS buffer (0.5% BSA, 0.05% sodium azide in PBS). 5 \times 10⁵ cells per well with primary antibody (1:2000, mouse anti-human CD184 antibody, BD Biosciences, USA) were seeded in 96-well v-bottom plates at the presence of various concentrations of compounds. After incubation for 40 min on ice, cells were washed twice with FACS buffer, and then incubated with secondary antibody (1:1000, anti-mouse IgG-FITC antibody, Sigma, USA) for 30 min on ice. Cells were washed twice again and re-suspended with FACS buffer. The fluorescence (excitation 485/emission 528) was recorded using a Synergy II plate reader. IC₅₀s were calculated by GraphPad Prism from at least three independent experiments.

Calcium Mobilization Assays

The ligands were first tested with various concentrations (0.3 nM to 3 μ M) for possible agonist activity in either MOR-CHO or CXCR4-HOS cells. The protocol was the same for the antagonism study for both MOR and CXCR4 cell types, except for the addition of an agonist (either DAMGO or SDF-1).

CXCR4-HOS cells were cultured and maintained in DMEM (90%), fetal bovine serum 10% (v/v), and supplemented with 1.0 µg/ml puromycin. MOR-CHO cells were cultured and maintained in DMEM (90%), and fetal bovine serum 10% (v/v).

Either CXCR4-HOS or MOR-CHO cells were transfected with Gqi5 pcDNA1 (Addgene, Cat# 24501) using Lipofectamine 2000 (Invitrogen) according to the manufacturer's recommended procedure. Cells were incubated for 4 hr at 37 °C and 5% CO₂ and then trypsinized and transferred to a clear bottom, black 96-well plate (Greiner Bio-one) at 3x10⁶ cells per well in their respective growth media and incubated until confluent. 48 hours after transfection the growth media was decanted and cells were then incubated with 50 µL of fluo-4 AM loading buffer [24 µL 2 mM fluo-4 AM solution (Invitrogen), 12 µL 250 mM probenecid, in 6 mL assay buffer (HBSS–HEPES–Ca–Mg–probenecid)] for 45 min. Loading buffer was then decanted and cells were incubated for an additional 15 min in 20 µL of each compound in varying concentrations and 60 µL assay buffer. Ca²⁺ concentrations were monitored by RFU for 90 seconds right after addition of 20 µL of agonist (DAMGO or SDF-1) to each well in the microplate reader (FlexStation3, Molecular Devices). Peak values were obtained using SoftMaxPro software (Molecular Devices) and non-linear regression curves were generated using Prism (GraphPad) to calculate IC₅₀ values. All doses were tested with triplicates. All experiments were repeated at least 4 times to obtain standard error values.

Apr.23,2015
PROTON DMSO /opt/topspin Kang.g 42

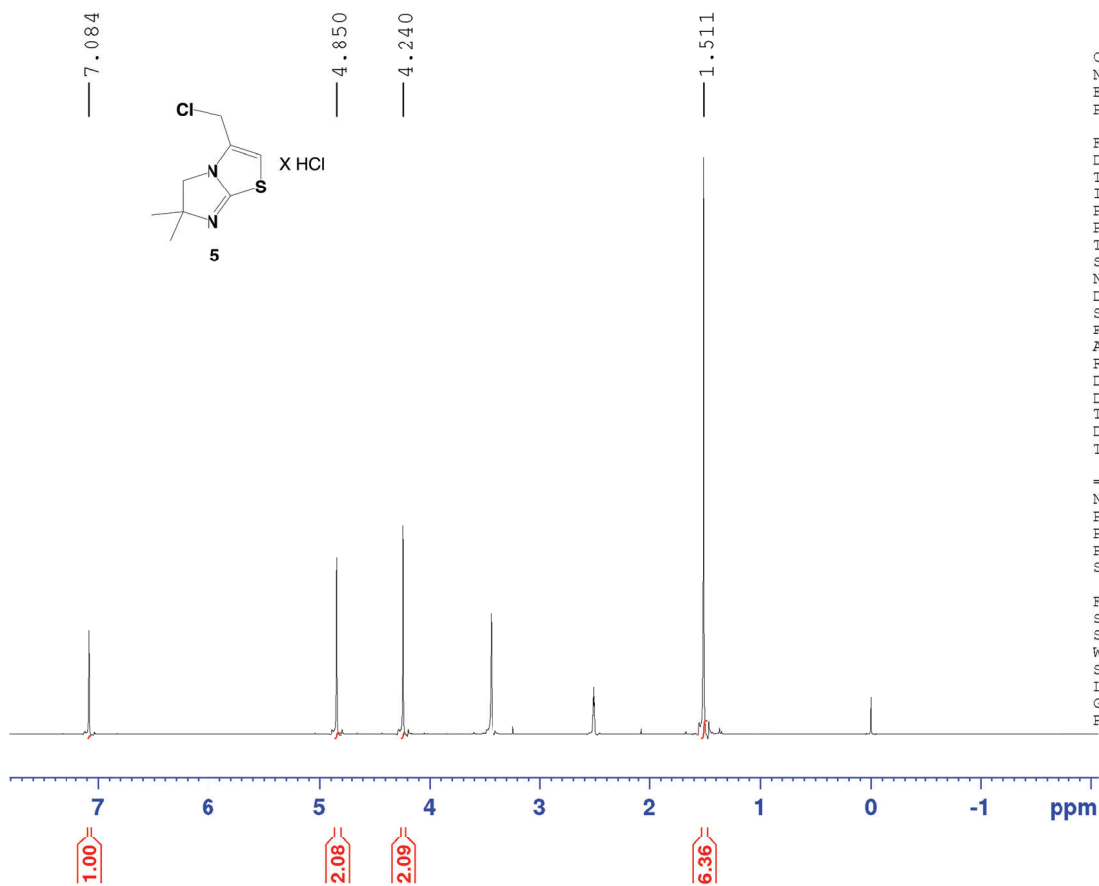


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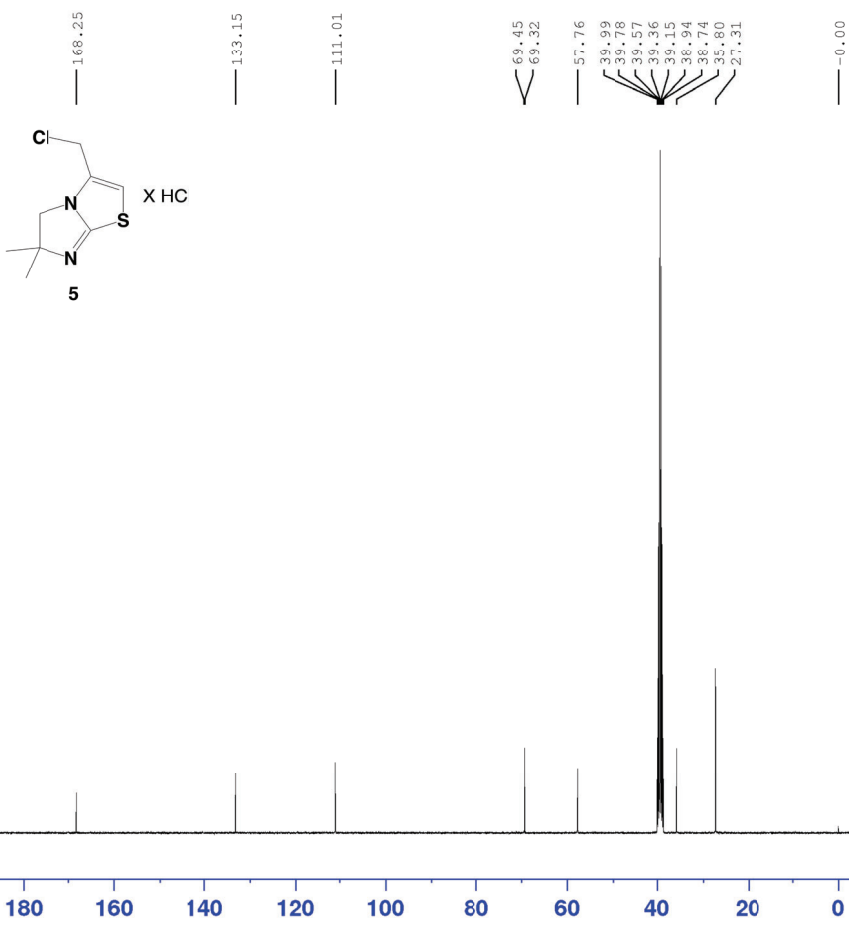
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 C13C2D DMSO /opt/topspin Kang.g 42



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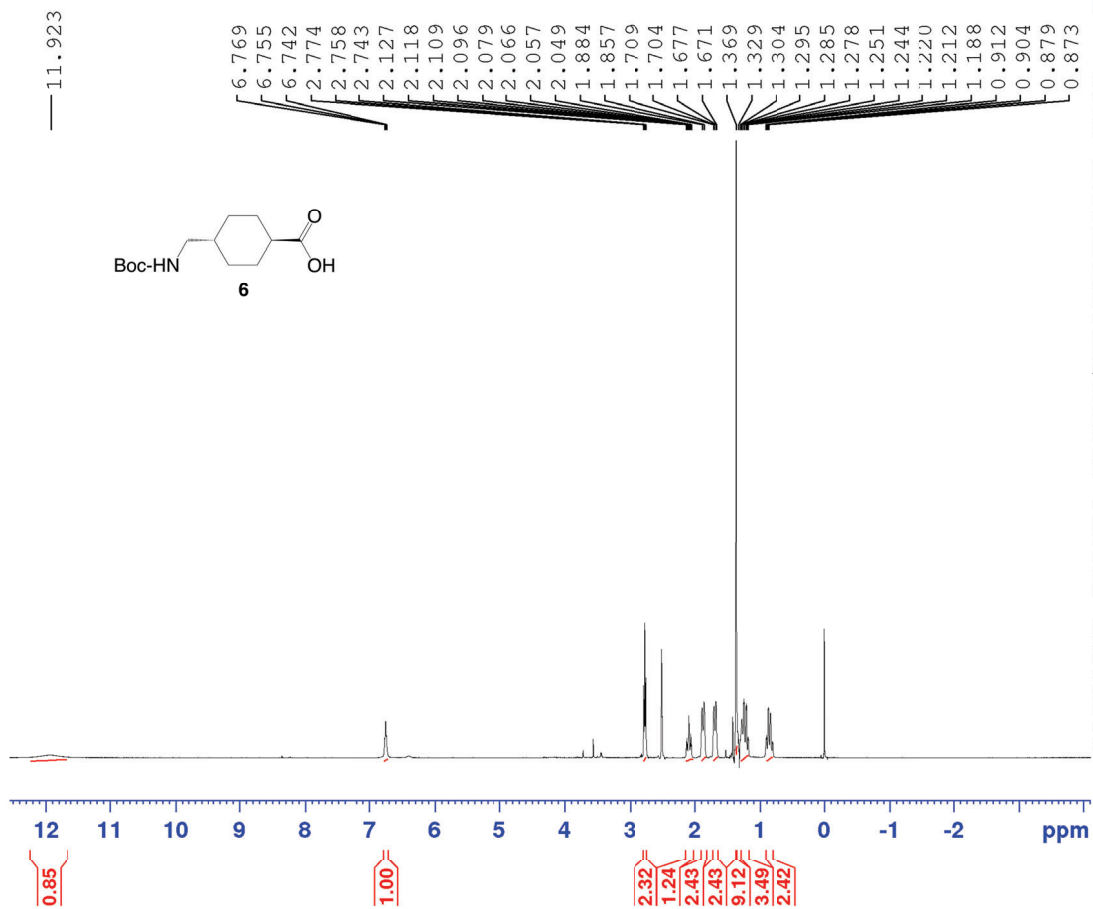
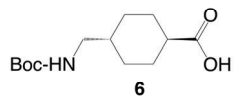
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F2 - Processing parameters
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May 27, 2015
 PROT0N DMSO /opt/topspin Kang.g 27



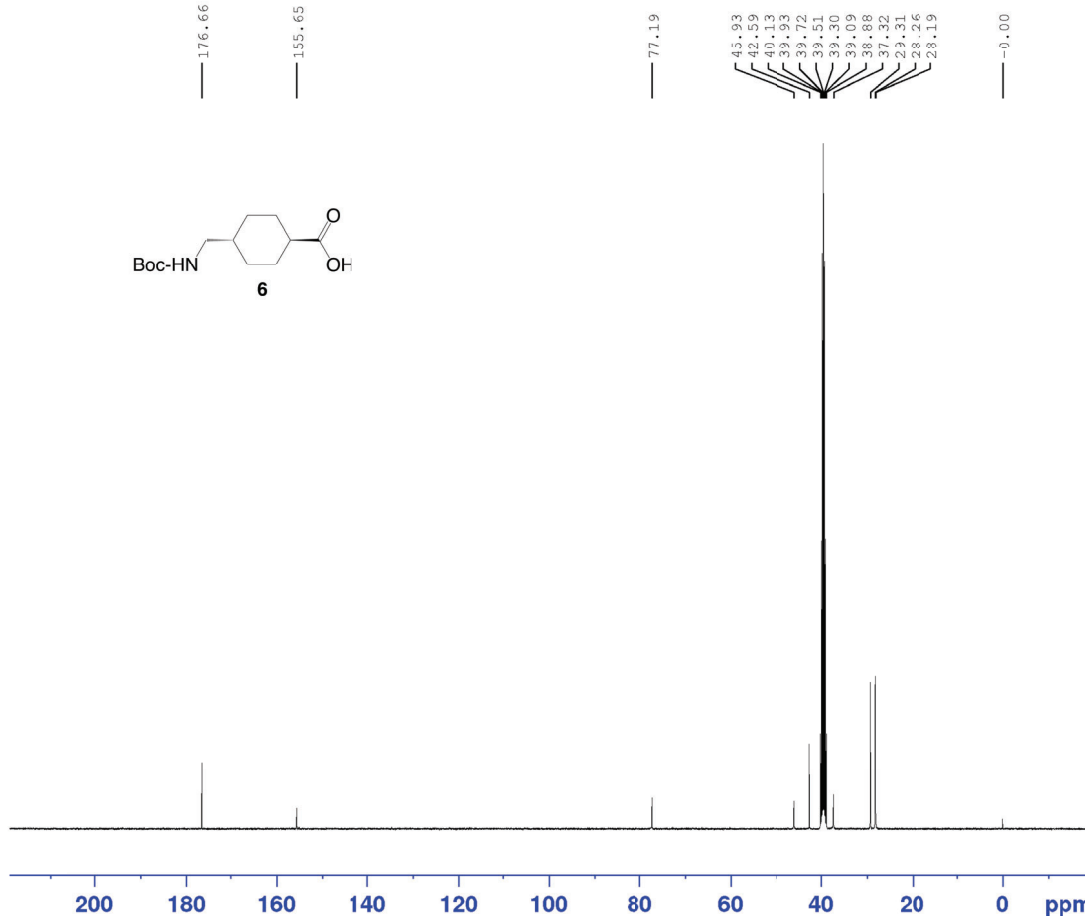
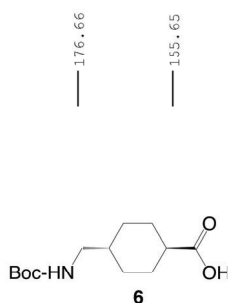
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 C13C2D DMSO /opt/topspin Kang.g 27



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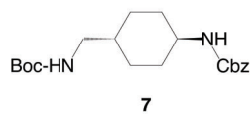
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 PROT0N DMSO /opt/topspin Kang.g 60



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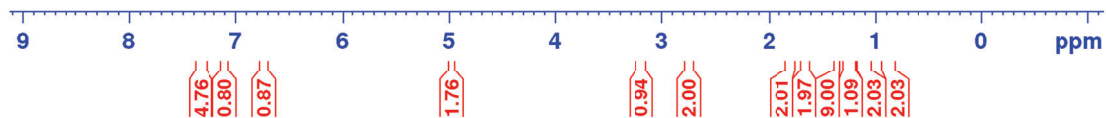


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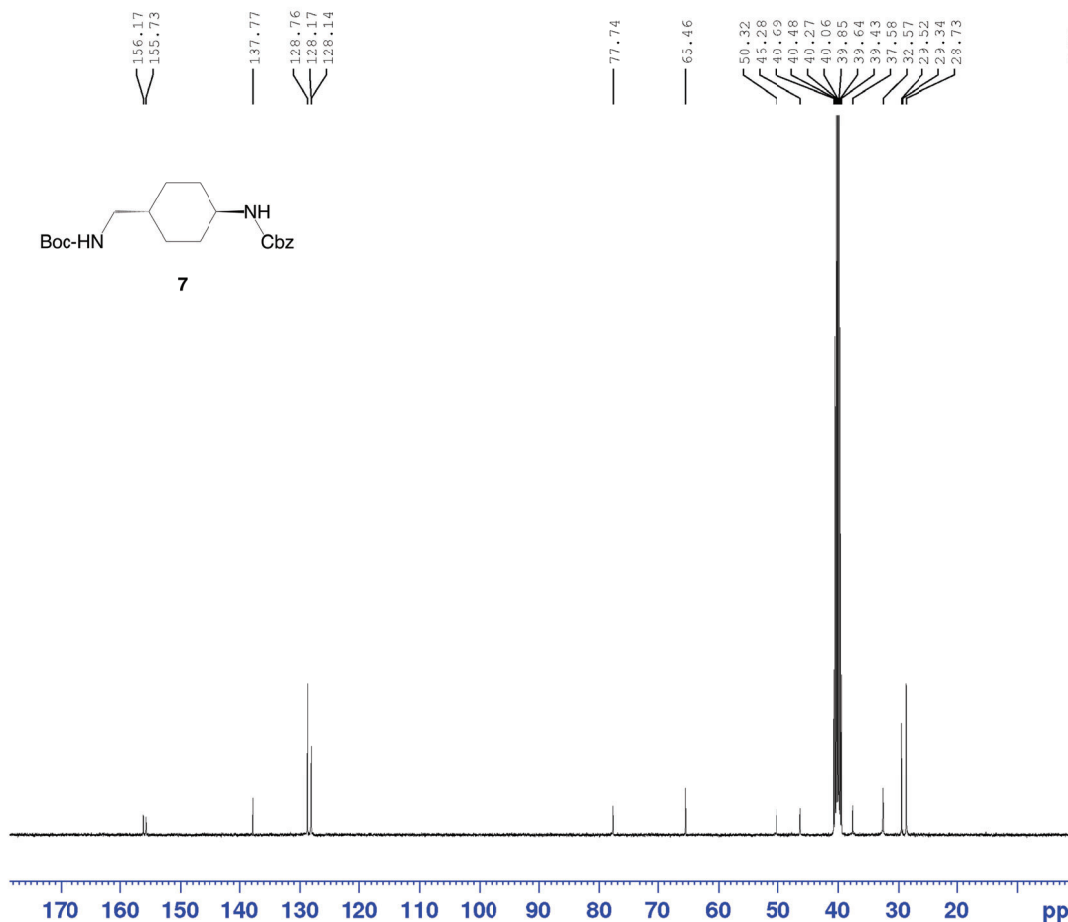
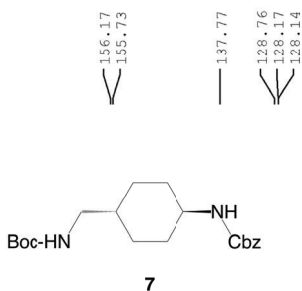
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 C13C2D DMSO /opt/topspin Kang.g 60



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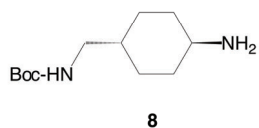
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PL12       14.50 dB
PL13       14.50 dB
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PL12W      0.33867535 W
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 DE 6.50 usec
 TE 302.7 K
 D1 1.0000000 sec
 TD0 1

----- CHANNEL f1 -----
 NUC1 1H
 P1 12.00 usec
 PL1 -2.00 dB
 PL1W 15.12807274 W
 SFO1 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300052 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



0.72
 3.80
 1.00
 2.02
 2.09
 9.27
 1.26
 2.09
 2.14

May 29, 2015
C13C2D CDC13 /opt/topspin Kang.g 26



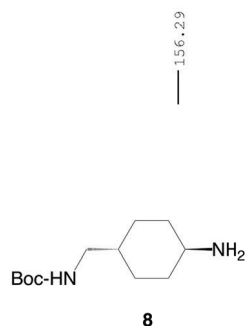
Current Data Parameters
NAME GK-II-33_CDCl3_C
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20150601
Time 5.51
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 5120
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631488 sec
RG 203
DW 20.800 usec
DE 6.50 usec
TE 303.8 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 8.50 usec
PL1 -1.70 dB
PL1W 55.73028564 W
SFO1 100.6228298 MHz

===== CHANNEL f2 =====
CPDPRG[2] waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 -2.00 dB
PL12 14.50 dB
PL13 14.50 dB
PL2W 15.12807274 W
PL12W 0.33867535 W
PL13W 0.33867535 W
SFO2 400.1316005 MHz

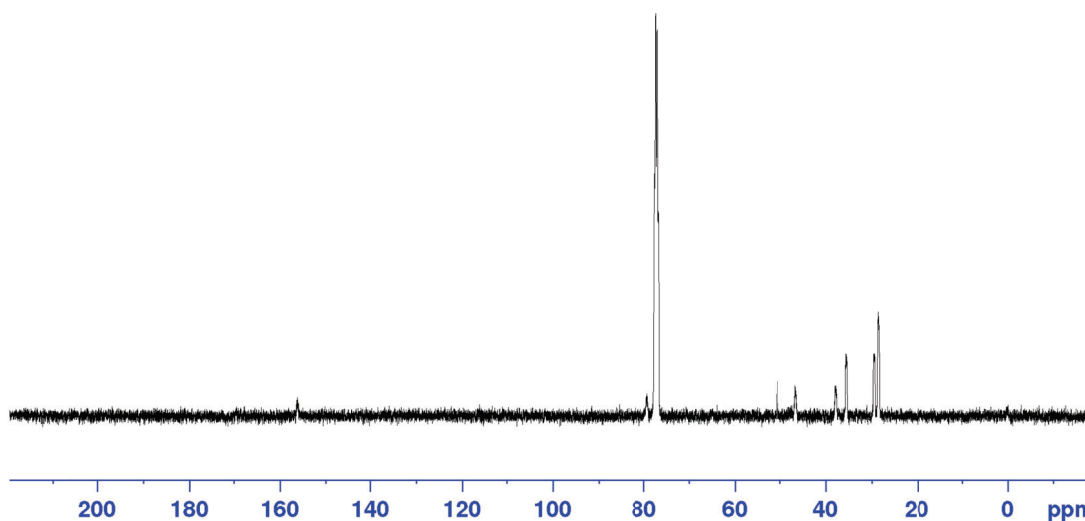
F2 - Processing parameters
SI 32768
SF 100.6127690 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



79.18
77.56
77.29
76.98

50.62
46.66

37.79
35.66
29.43
28.60

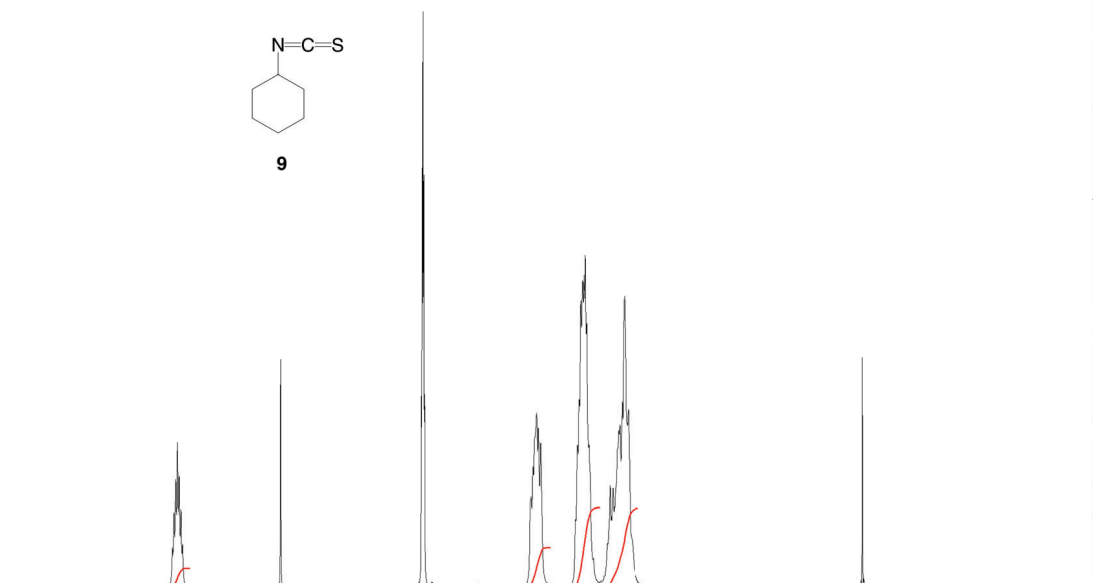
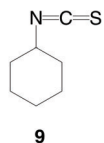


Jan.21,2015
 PROT9N DMSO /opt/topspin Kang.g 16



3.920
 3.910
 3.900
 3.891
 3.881

1.883
 1.871
 1.851
 1.841
 1.828
 1.611
 1.603
 1.592
 1.584
 1.577
 1.569
 1.412
 1.396
 1.389
 1.382
 1.368
 1.355
 1.351
 1.342
 1.334



Current Data Parameters
 NAME GK-I-71
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20150121
 Time 10.10
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 65536
 SOLVENT DMSO
 NS 32
 DS 2
 SWH 8223.685 Hz
 FIDRES 0.125483 Hz
 AQ 3.9845889 sec
 RG 203
 DW 60.800 usec
 DE 6.50 usec
 TE 298.2 K
 D1 1.0000000 sec
 TD0 1

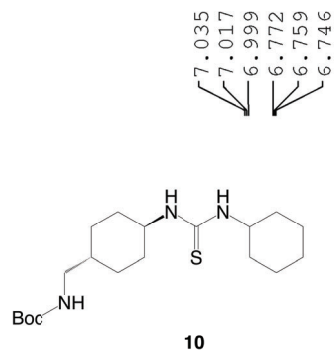
----- CHANNEL f1 -----
 NUC1 1H
 P1 12.00 usec
 PL1 -2.00 dB
 PL1W 15.12807274 W
 SFO1 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300045 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5 0.0 -0.5 ppm

1.00
 2.09
 4.24
 4.19

May 25, 2015
 PROT0N DMSO /opt/topspin Kang.g 33

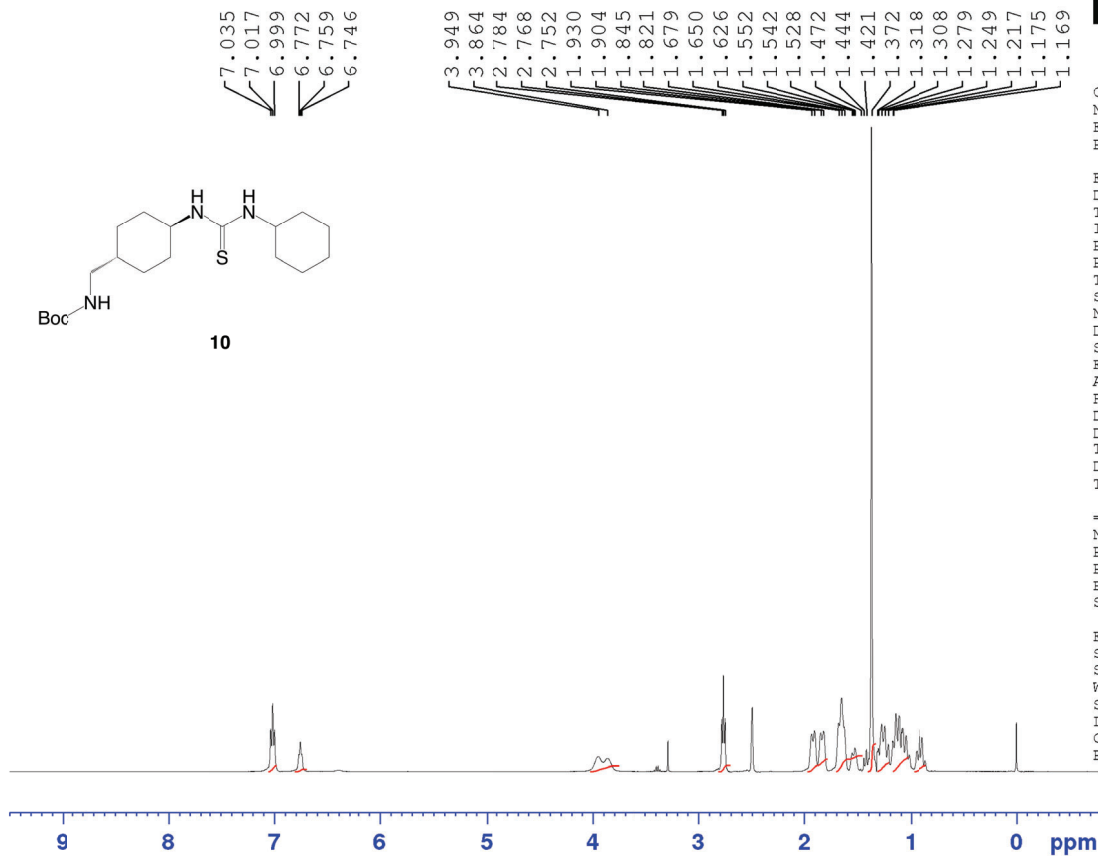


Current Data Parameters
 NAME GK-I-73b_DMSOb
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20150525
 Time 19.49
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 65536
 SOLVENT DMSO
 NS 16
 DS 2
 SWH 8223.685 Hz
 FIDRES 0.125483 Hz
 AQ 3.9845889 sec
 RG 90.5
 DW 60.800 usec
 DE 6.50 usec
 TE 305.1 K
 D1 1.00000000 sec
 TD0 1

----- CHANNEL f1 -----
 NUC1 1H
 P1 12.00 usec
 PL1 -2.00 dB
 PL1W 15.12807274 W
 SFO1 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300045 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



May 25, 2015
 C13C2D DMSO /opt/topspin Kang.g 33



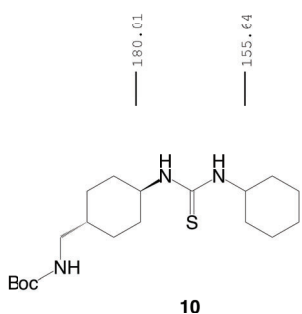
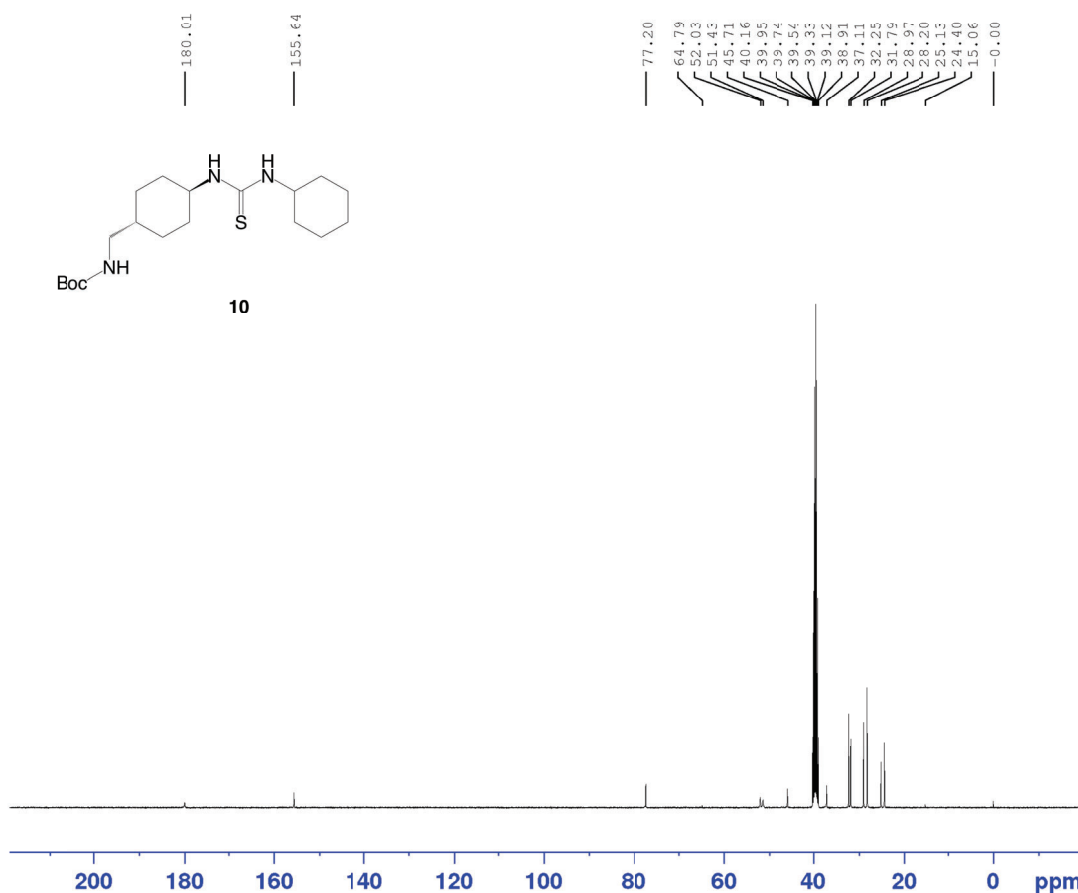
Current Data Parameters
 NAME GK-I-73b_DMSOb_C
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20150526
 Time 0.48
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT DMSO
 NS 5120
 DS 4
 SWH 24038.461 Hz
 FIDRES 0.366798 Hz
 AQ 1.3631488 sec
 RG 203
 DW 20.800 usec
 DE 6.50 usec
 TE 306.9 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 8.50 usec
 PL1 -1.70 dB
 PL1W 55.73028564 W
 SFO1 100.6228298 MHz

===== CHANNEL f2 =====
 CPDPRG[2] waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -2.00 dB
 PL12 14.50 dB
 PL13 14.50 dB
 PL2W 15.12807274 W
 PL12W 0.33867535 W
 PL13W 0.33867535 W
 SFO2 400.1316005 MHz

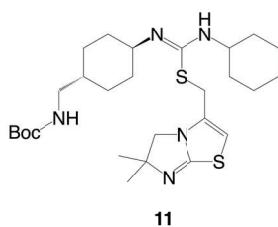
F2 - Processing parameters
 SI 32768
 SF 100.6128250 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40



Mar.17,2015
 PROT0N DMSO /opt/topspin Kang.g 29



6.884
 4.881
 4.275
 3.886
 3.675
 2.778
 2.763
 1.718
 1.607
 1.579
 1.549
 1.505
 1.371
 1.308
 1.284
 1.212
 1.092
 0.979
 0.956

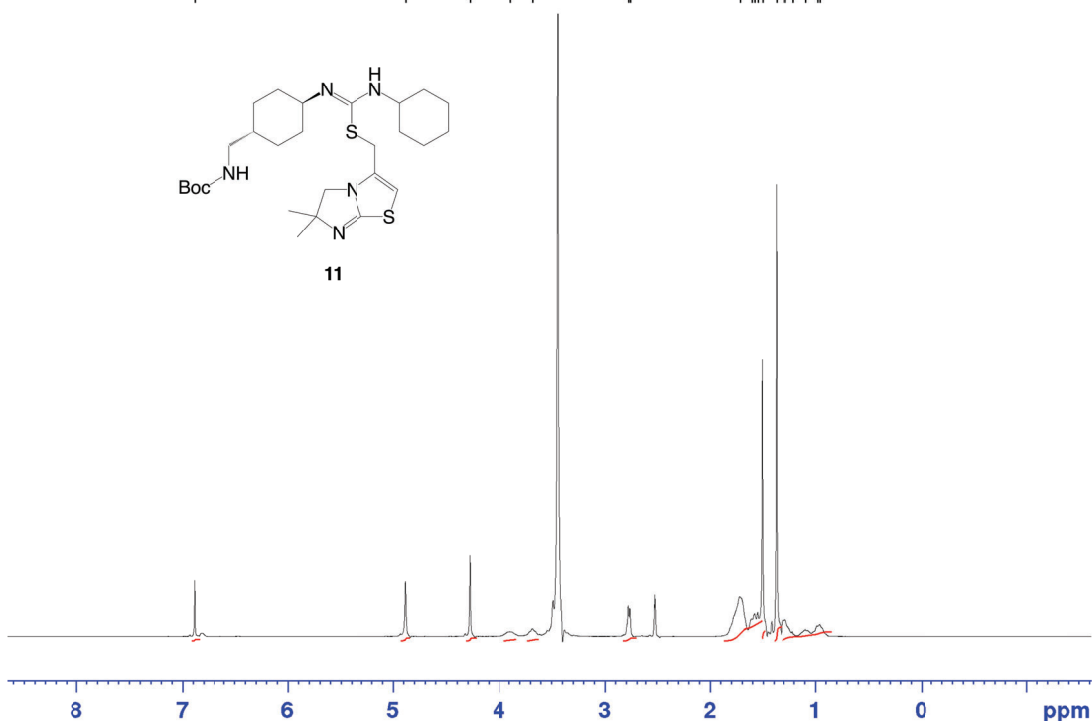


Current Data Parameters
 NAME GK-II-19b
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20150317
 Time 17.11
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 65536
 SOLVENT DMSO
 NS 8
 DS 2
 SWH 8223.685 Hz
 FIDRES 0.125483 Hz
 AQ 3.9845889 sec
 RG 80.6
 DW 60.800 usec
 DE 6.50 usec
 TE 303.2 K
 D1 1.00000000 sec
 TD0 1

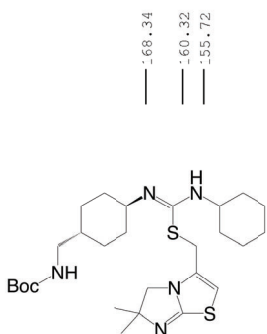
----- CHANNEL f1 -----
 NUC1 1H
 P1 12.00 usec
 PL1 -2.00 dB
 PL1W 15.12807274 W
 SFO1 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300000 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



1.00
 1.86
 1.86
 0.87
 1.02
 1.87
 13.30
 6.31
 6.90
 5.81

Mar.17,2015
C13C2D DMSO /opt/topspin Kang.g 29



— .68.34
— .60.32
— .55.72

— .31.36

— .10.22

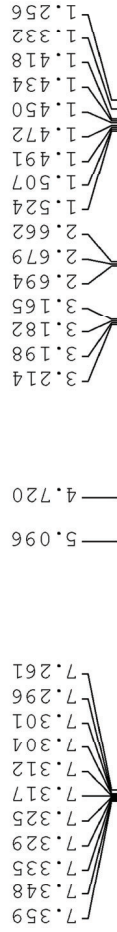
77.40
69.29
58.15
56.30
53.34
45.61
45.48
40.08
39.87
38.67
38.45
38.23
38.05
38.02
36.74
32.07
31.41
30.96
30.39
28.51
28.23
28.04
27.36
24.51
24.32



Current Data Parameters
NAME GK-II-19b_C
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20150319
Time 1.54
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT DMSO
NS
DS
SWH
FIDRES
AQ

Apr.02,2015
PROTON CDCl3 /opt/topspin Kang.g 59

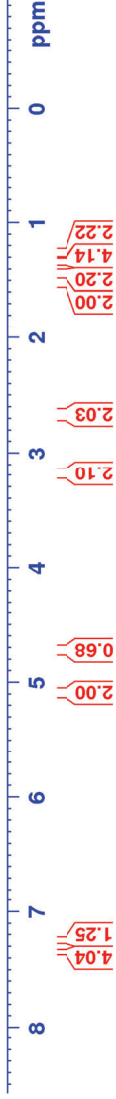


Current Data Parameters
NAME GK-II-29
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20150402
Time 16.54
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65336
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9845889 sec
RG 203
DW 60.800 usec
DE 6.50 usec
TE 306.5 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 12.00 usec
PL 2.00 dB
FLL 15.12607274 W
SFO1 400.1324710 MHz

F2 - Processing parameters
SI 32768
SF 400.1300115 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



Apr.12,2015
C13CPD CDCl3 /opt/topspin Kang.g 29



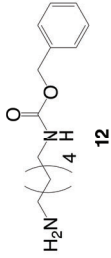
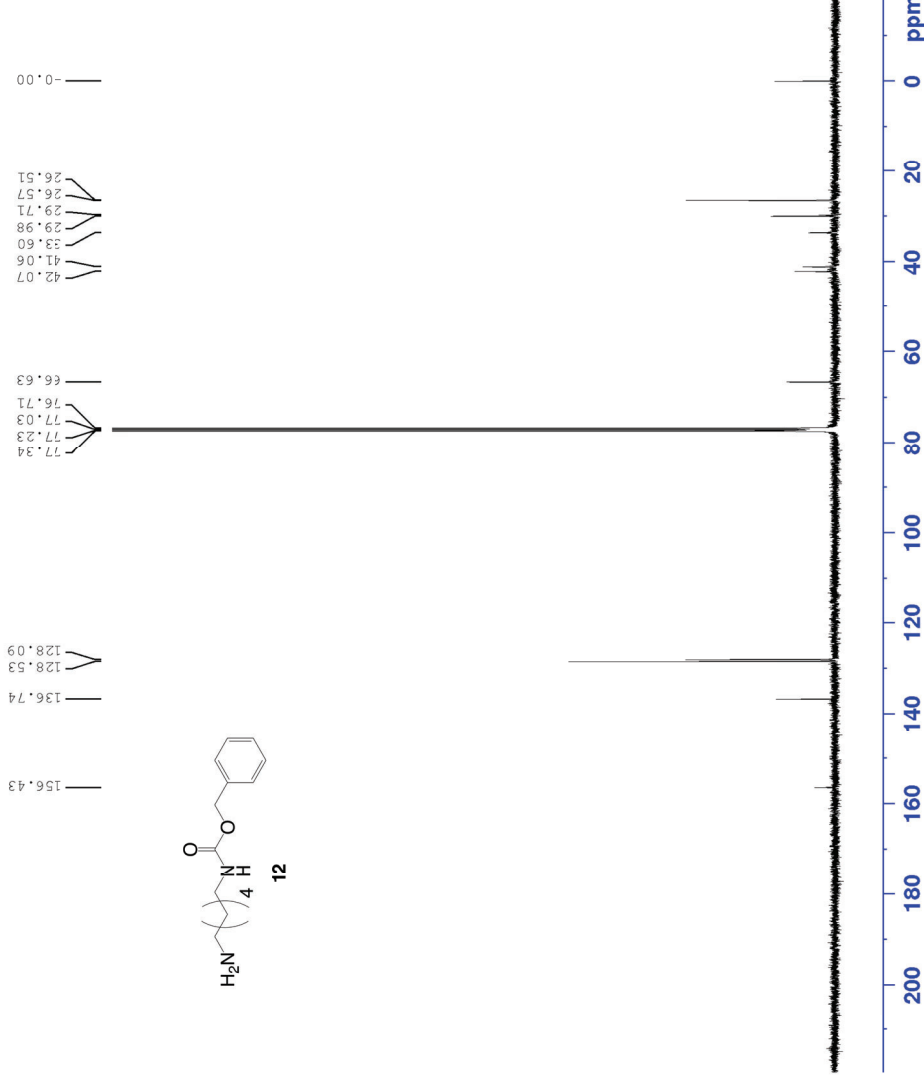
Current Data Parameters
NAME GK-II-29_C
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20150413
Time 23.03
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 5210
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631488 sec
RG 203
DW 20.800 usec
DE 6.50 usec
TE 304.4 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

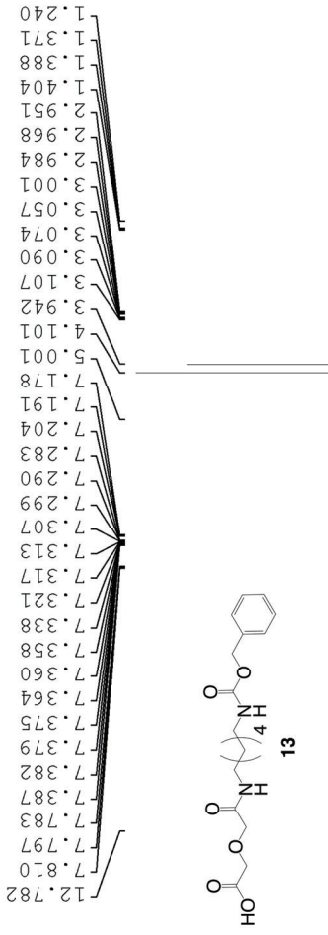
===== CHANNEL f1 =====
NUC1 13C
P1 8.50 usec
PL1 -1.70 dB
PL1W 55.73028564 W
SFO1 100.6228298 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 -2.00 dB
PL2 14.50 dB
PL3 14.50 dB
PL2W 15.12807274 W
PL12W 0.33867535 W
PL13W 0.33867535 W
SFO2 400.1316005 MHz

F2 - Processing Parameters
SI 32768
SF 100.6127655 MHz
RGW 0
SSB 0
LB 0
GB 0
PC 1.40



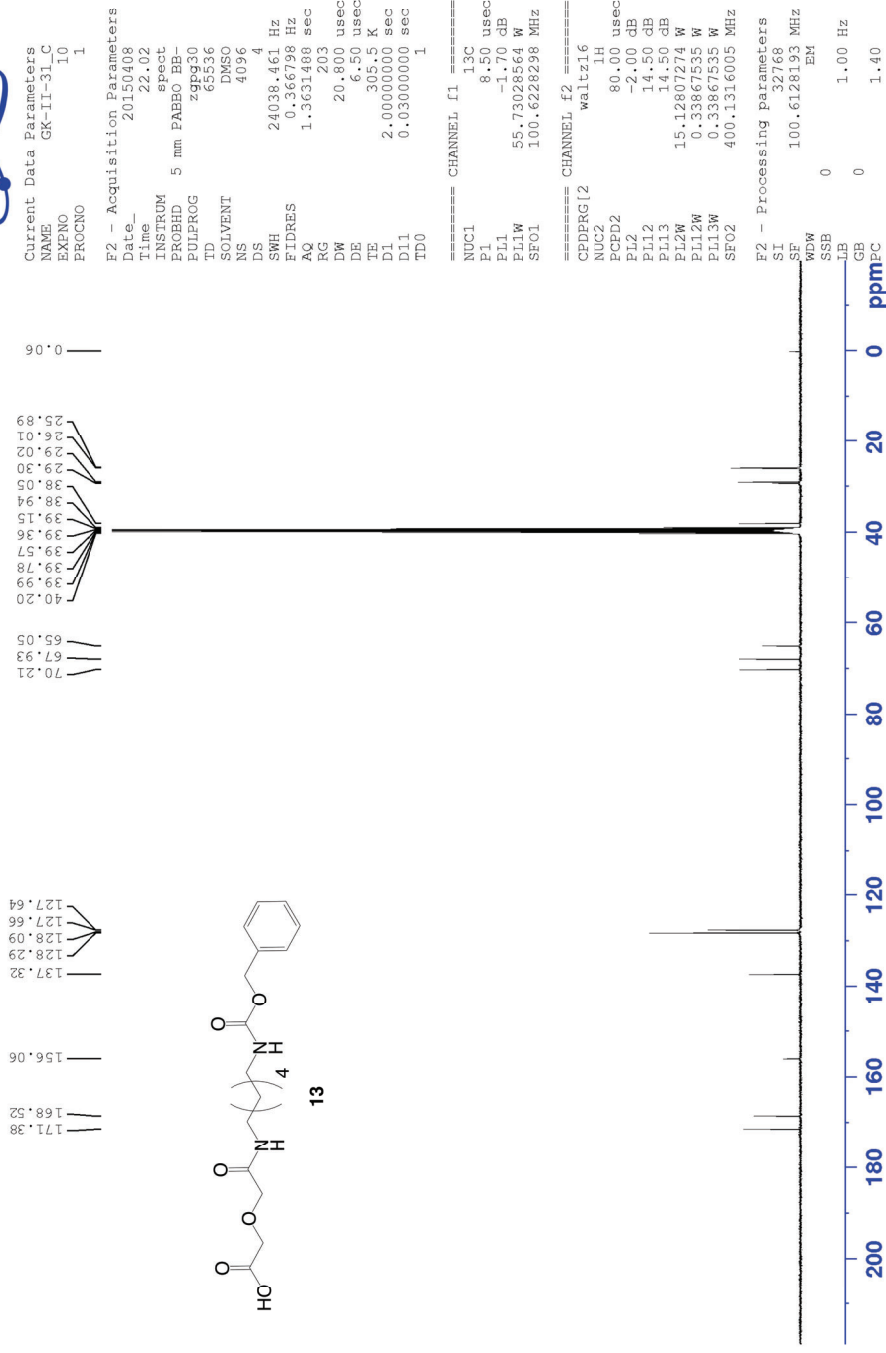
Apr.07, 2015
PROTON DMSO /opt/topspin Kang.g 33



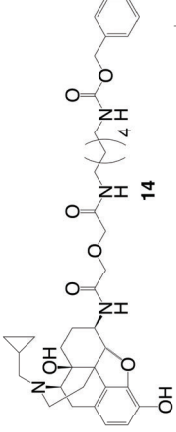
Current Data Parameters
NAME GK-II-31
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20150407
Time 17.27
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT DMSO
NS 8
DS 2
SWH 8223.685 H
FIDRES 0.125483 H

Apr.07,2015
C13CPD DMSO /opt/topspin Kang.g 33



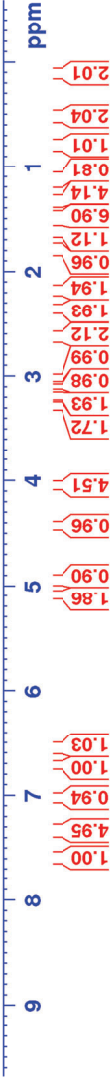
Apr.25, 2015
 PROTON CDCl3 /opt/topspin Kang.g 58



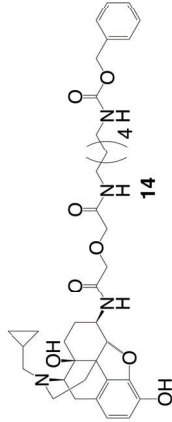
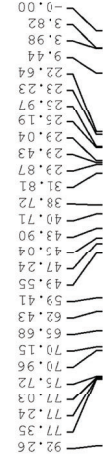
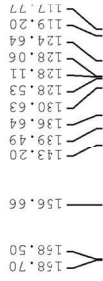
F2 - Acquisition Parameters
 Date_ 20150425
 Time_ 16.31
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 65336
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 8223.685 Hz
 FIDRES 0.125483 Hz
 AQC 3.9845889 sec
 RG 181
 DW 60.800 usec
 DE 6.50 usec
 TE 298.4 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 12.00 usec
 PL 2.00 dB
 FLL 15.12607274 W
 SFO1 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300087 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



Apr.25,2015
 C13CPD CDC13 /opt/topspin Kang.g 58



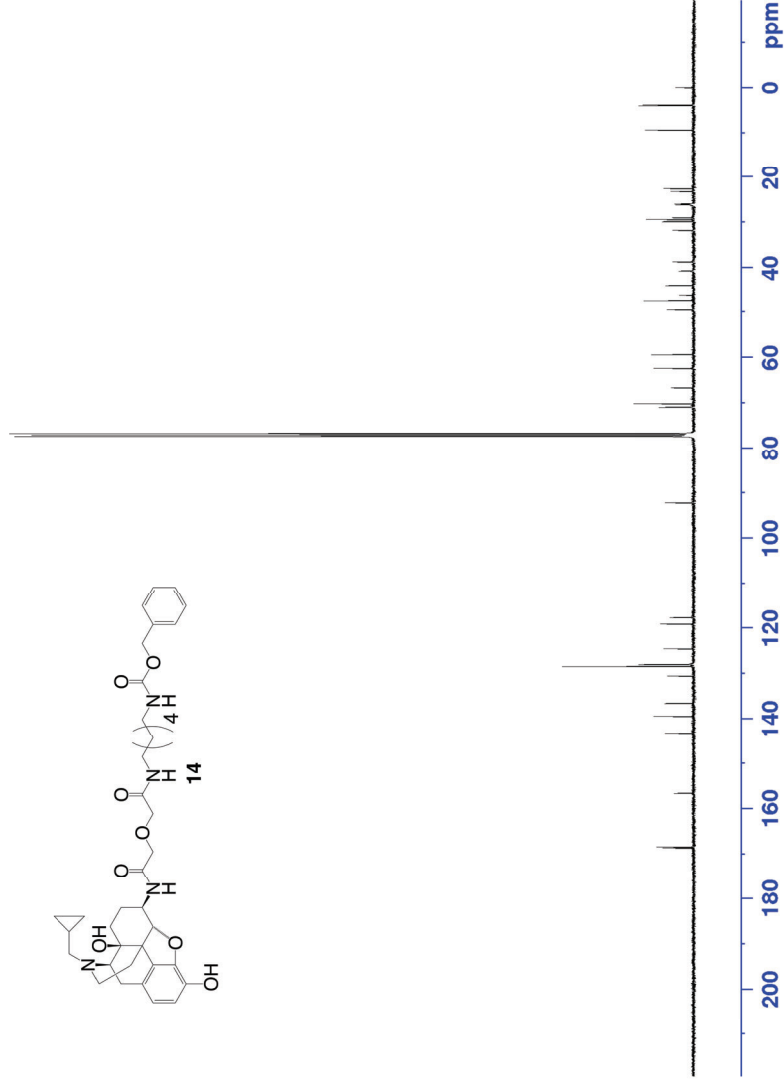
Current Data Parameters
 NAME GR-II-34s_C
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20150426
 Time 7.50
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDC13
 NS 4096
 DS 4
 SWH 24038.461 Hz
 FIDRES 0.366798 Hz
 AQ 1.3631488 sec
 RG 203
 DW 20.800 usec
 DE 6.50 usec
 TE 302.1 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TD0 1

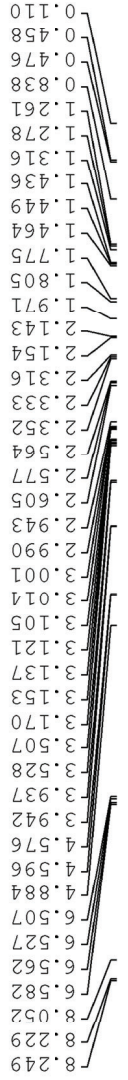
===== CHANNEL f1 =====
 NUC1 13C
 P1 8.50 usec
 PL1 -1.70 dB
 PL1W 55.73028564 W
 SFO1 100.6228298 MHz

===== CHANNEL f2 =====
 CPDPRG[2] waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -2.00 dB
 PL12 14.50 dB
 PL13 14.50 dB
 PL1W 15.12807274 W
 PL12W 0.33867535 W
 PL13W 0.33867535 W
 SFO2 400.1316005 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6127670 MHz
 NCBW
 SSB 0
 LB 0 1.00 Hz
 GB 0
 PC 1.40



Apr.28,2015
 PROTON DMSO /opt/topspin Kang.g 30

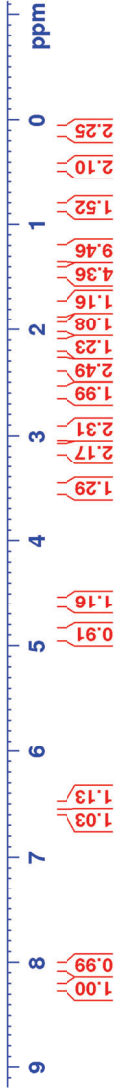
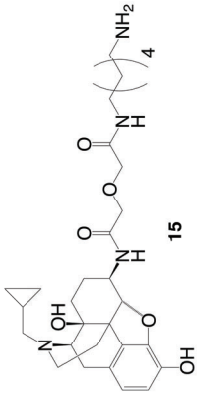


Current Data Parameters
 NAME GK-II-40a
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20150428
 Time 9.06
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 65336
 SOLVENT DMSO
 NS 16
 DS 2
 SWH 8223.685 Hz
 FIDRES 0.125483 Hz
 AQC 3.9845889 sec
 RG 203
 DW 60.800 usec
 DE 6.50 usec
 TE 299.6 K
 D1 1.00000000 sec
 TD0 1

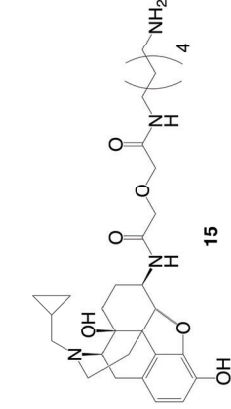
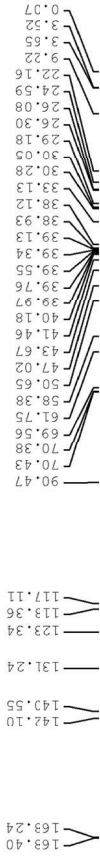
===== CHANNEL f1 =====
 NUC1 1H
 P1 12.00 usec
 PL -2.00 dB
 FLL 15.12607274 W
 SFO1 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300043 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



1.00
 0.99
 1.03
 1.13
 0.91
 1.16
 1.29
 2.17
 2.31
 1.99
 2.49
 1.23
 1.08
 1.16
 4.36
 9.46
 1.52
 2.10
 2.25

Apr.28,2015
C13CPD DMSO /opt/topspin Kang.g 30



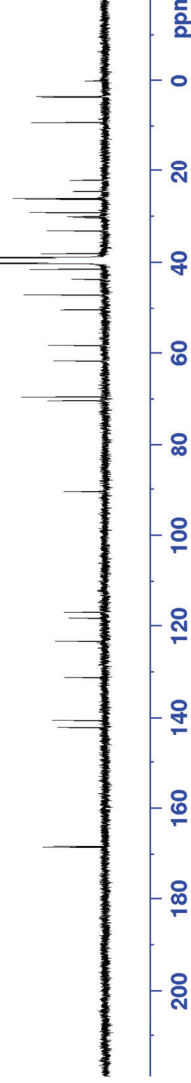
Current Data Parameters
NAME GR-II-40s_C
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20150429
Time 7.51
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT DMSO
NS 10240
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631488 sec
RG 203
DW 20.800 usec
DE 6.50 usec
TE 301.8 K
D1 2.0000000 sec
D11 0.0300000 sec
TD0 1

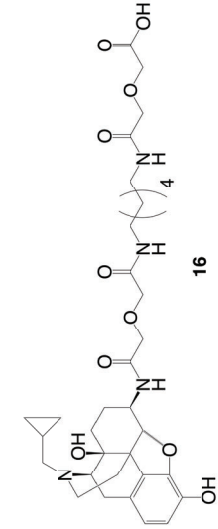
===== CHANNEL f1 =====
NUC1 13C
P1 8.50 usec
PL1 -1.70 dB
PL1W 55.73028564 W
SFO1 100.6228298 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 -2.00 dB
PL2 14.50 dB
PL13 14.50 dB
PL12W 15.12807274 W
PL12M 0.33867535 W
PL13W 0.33867535 W
SFO2 400.1316005 MHz

F2 - Processing parameters
SI 32768
SF 100.6128193 MHz
WDW EM
SSB 0
GB 0
PC 1.00 Hz
PR 1.40



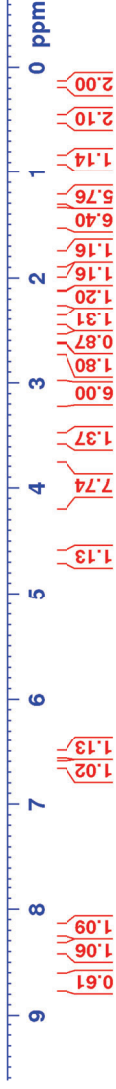
May 08, 2015
 PROTON DMSO /opt/topspin Kang.g 37



F2 - Acquisition Parameters
 Date_ 20150508
 Time_ 15.27
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 65336
 SOLVENT DMSO
 NS 16
 DS 2
 SWH 8223.685 Hz
 FIDRES 0.125483 Hz
 AQ 3.9845889 sec
 RG 203
 DW 60.800 usec
 DE 6.50 usec
 TE 301.0 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 12.00 usec
 PL -2.00 dB
 FLL 15.12807274 W
 SFO1 400.1324710 MHz

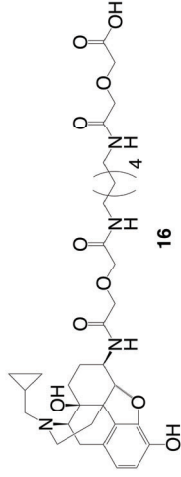
F2 - Processing parameters
 SI 32768
 SF 400.1300041 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



May 07, 2015
 C13CPD DMSO /opt/topspin Kang.g 37



169.15
 147.02
 147.59
 133.95
 122.82
 118.95
 117.11
 90.29
 70.32
 70.29
 69.49
 61.69
 58.08
 50.53
 46.73
 43.97
 38.17
 37.93
 29.93
 29.73
 28.91
 28.85
 25.87
 25.84
 24.36
 22.23
 22.23
 8.59
 3.72
 3.35



Current Data Parameters
 NAME GR-II-47B_C
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20150508
 Time 6.43
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT DMSO
 NS 8192
 DS 4
 SWH 24038.461 Hz
 FIDRES 0.366798 Hz
 AQ 1.3631488 sec
 RG 203
 DW 20.800 usec
 DE 6.50 usec
 TE 302.8 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TD0 1

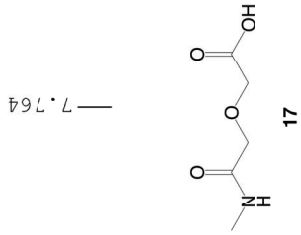
===== CHANNEL f1 =====
 NUC1 13C
 P1 8.50 usec
 PL1 -1.70 dB
 PL1W 55.73028564 W
 SFO1 100.6228298 MHz

===== CHANNEL f2 =====
 CPDPRG[2] waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -2.00 dB
 PL2 14.50 dB
 PL13 14.50 dB
 PL2W 15.12807274 W
 PL12W 0.33867535 W
 PL13W 0.33867535 W
 SFO2 400.1316005 MHz

F2 - Processing Parameters
 SI 32768
 SF 100.6128279 MHz
 SSB 0
 GB 0
 PC 1.40



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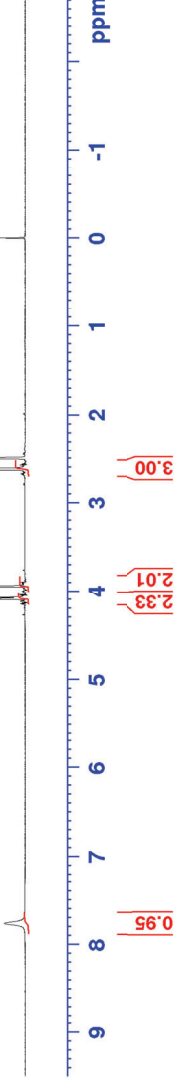
7.764
4.096
3.941
2.630
2.619

Current Data Parameters
NAME GK-II-54a
EXPNO 10
PROCNO 1

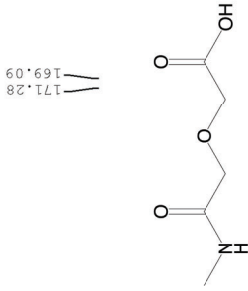
F2 - Acquisition Parameters
Date_ 20150610
Time 17.09
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 6536
SOLVENT DMSO
NS 8
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9845889 sec
RG 181
DW 60.800 usec
DE 6.50 usec
TE 302.4 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 12.00 usec
PL -2.00 dB
FL1 15.12607274 W
SFO1 400.1324710 MHz

F2 - Processing parameters
SI 32768
SF 400.1300039 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



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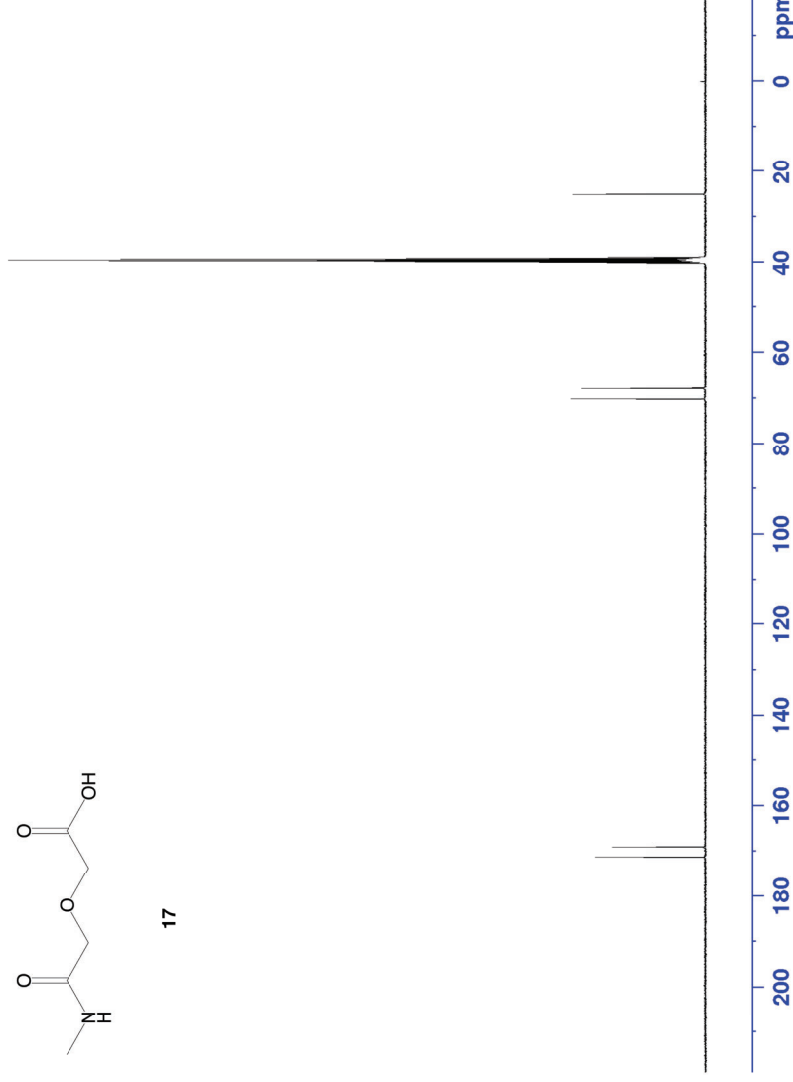
Current Data Parameters
NAME GR-II-54a_C
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20150611
Time 1.52
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT DMSO
NS 4096
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631488 sec
RG 203
DW 20.800 usec
DE 6.50 usec
TE 304.2 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

==== CHANNEL f1 =====
NUC1 13C
P1 8.50 usec
PL1 -1.70 dB
PL1W 55.73028564 W
SFO1 100.6228298 MHz

==== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 -2.00 dB
PL2W 14.50 dB
PL13 14.50 dB
PL12W 15.12807274 W
PL12W 0.33867535 W
PL13W 0.33867535 W
SFO2 400.1316005 MHz

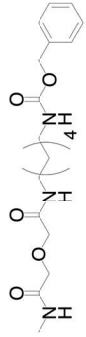
F2 - Processing parameters
SI 32768
SF 100.6128193 MHz
WDW EM
SSB 0
GB 0
PC 1.40



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7.986
 7.972
 7.957
 7.343
 7.338
 7.203
 7.190
 7.177
 5.002
 3.908
 3.126
 3.110
 3.093
 3.076
 3.004
 2.987
 2.971
 2.955
 2.657
 2.645
 2.503
 2.499
 2.494
 1.410
 1.402
 1.394
 1.376
 1.250

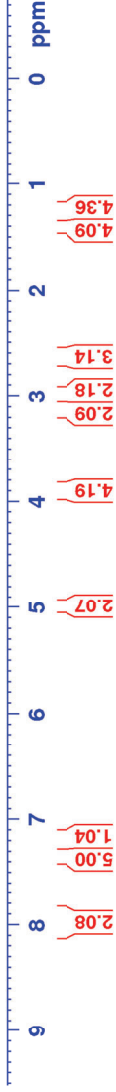


Current Data Parameters
 NAME GK-II-57b
 EXPNO 10
 PROCNO 1

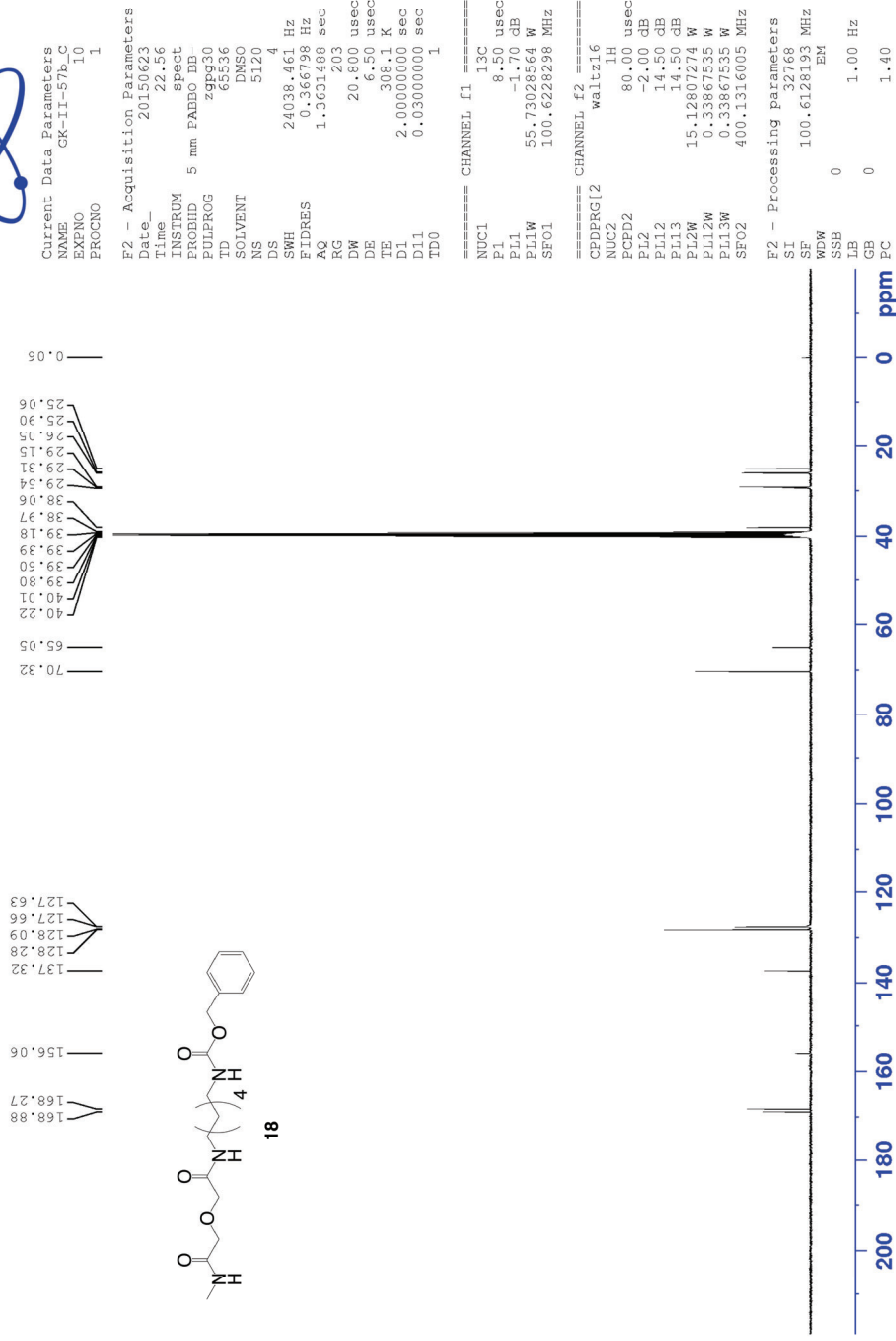
F2 - Acquisition Parameters
 Date_ 20150622
 Time_ 14.54
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 65336
 SOLVENT DMSO
 NS 8
 DS 2
 SWH 8223.685 Hz
 FIDRES 0.125483 Hz
 AQ 3.9845889 sec
 RG 161
 DW 60.800 usec
 DE 6.50 usec
 TE 304.9 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 12.00 usec
 PL -2.00 dB
 FLL 15.12607274 W
 SFO1 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300055 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



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 C13CPD DMSO /opt/topspin Kang.g 17

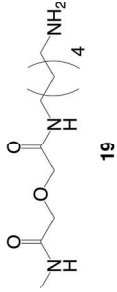


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8.000
7.987

3.908
3.906
3.132
3.115
3.098
3.081
2.658
2.646
2.541
2.526
1.465
1.447
1.430
1.413
1.395
1.358
1.343
1.327
1.310
1.266
1.257
1.200

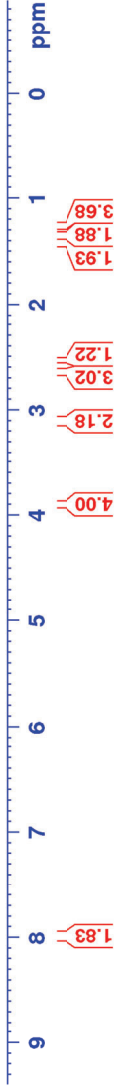


Current Data Parameters
NAME GK-II-62
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20150625
Time 17.26
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65336
SOLVENT DMSO
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9845889 sec
RG 161
DW 60.800 usec
DE 6.50 usec
TE 304.4 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 12.00 usec
PL 2.00 dB
FL1 15.12607274 W
SFO1 400.1324710 MHz

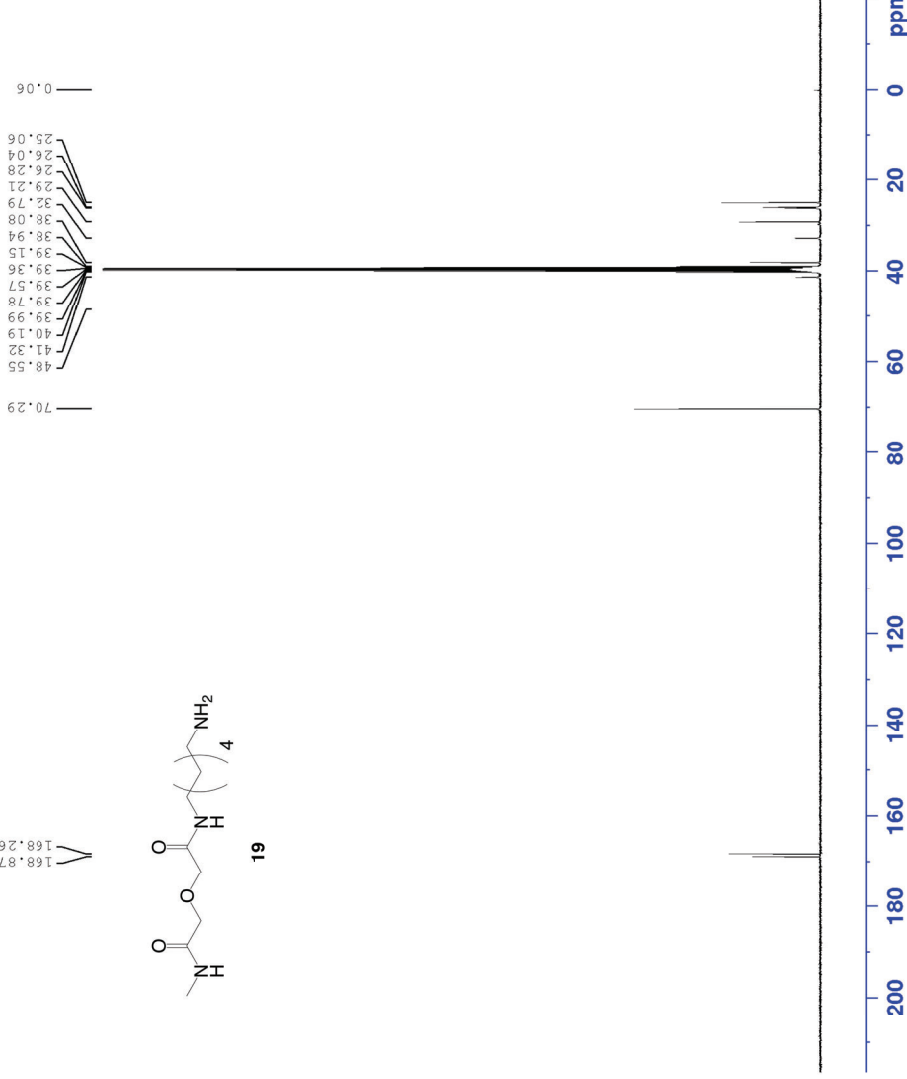
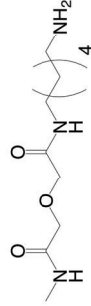
F2 - Processing parameters
SI 32768
SF 400.1300047 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



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C13CPD DMSO /opt/topspin Kang.g 22



168.26
168.87
168.87



Current Data Parameters
NAME GK-11-62
EXPNO 20
PROCNO 1

F2 - Acquisition Parameters
Date_ 20150626
Time 7.53
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT DMSO
NS 7168
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631488 sec
RG 203
DW 20.800 usec
DE 6.50 usec
TE 304.8 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 8.50 usec
PL1 -1.70 dB
PL1W 55.73028564 W
SFO1 100.6228298 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 -2.00 dB
PL2 14.50 dB
PL13 14.50 dB
PL2W 15.12807274 W
PL12W 0.33867535 W
PL13W 0.33867535 W
SFO2 400.1316005 MHz

F2 - Processing parameters
SI 32768
SF 100.6128193 MHz
RGW 0
SSB 0
LB 0
GB 0
PC 1.40

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Current Data Parameters
NAME GK-II-63
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20150701
Time 17.18
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 6536
SOLVENT DMSO
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9845889 sec
RG 161
DW 60.800 usec
DE 6.50 usec
TE 304.0 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 12.00 usec
PL 2.00 dB
FL1 15.12607274 W
SFO1 400.1324710 MHz

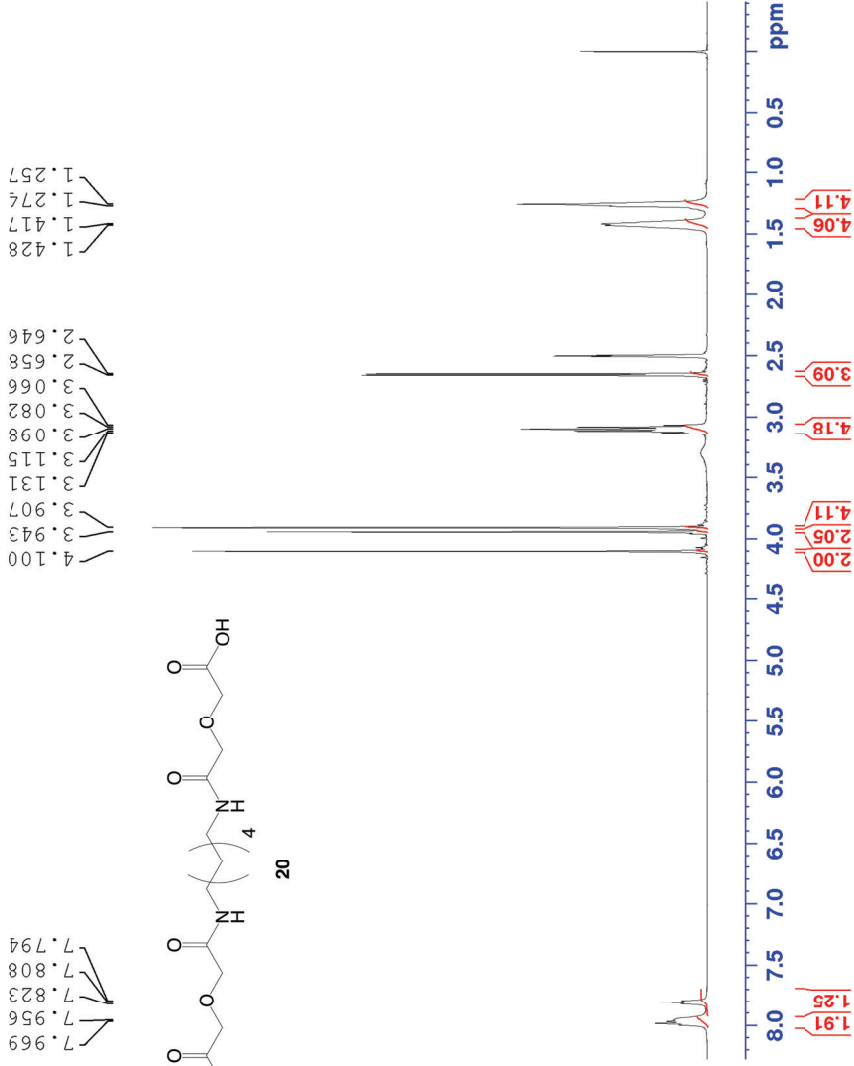
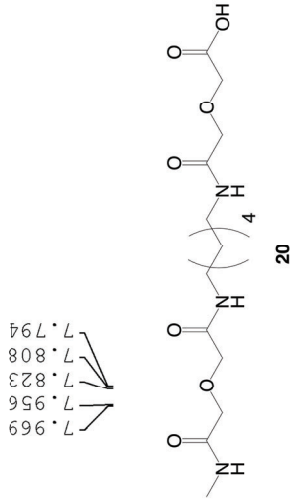
F2 - Processing parameters
SI 32768
SF 400.1300047 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

7.969
7.956
7.823
7.808
7.794

4.100
3.943
3.907
3.131
3.115
3.098
3.082
3.066
2.658
2.646

1.428
1.417
1.274
1.257

2.00
2.05
4.11
1.91
4.18
3.09
4.06
4.11



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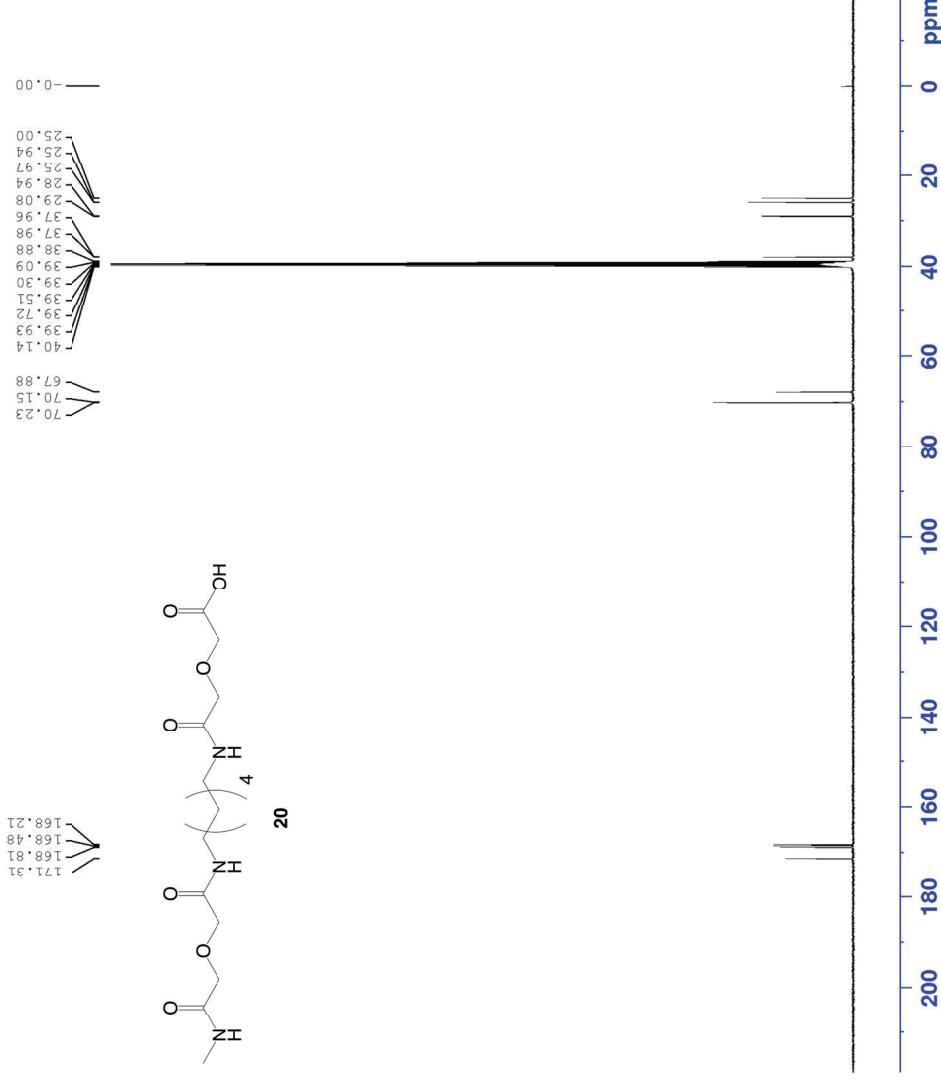
Current Data Parameters
NAME GK-II-63_C
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20150702
Time 1.51
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT DMSO
NS 8192
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631488 sec
RG 203
DW 20.800 usec
DE 6.50 usec
TE 308.5 K
D1 2.0000000 sec
D11 0.0300000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 8.50 usec
PL1 -1.70 dB
PL1W 55.73028564 W
SFO1 100.6228298 MHz

===== CHANNEL f2 =====
CPDPRG[2] waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 -2.00 dB
PL2 14.50 dB
PL13 14.50 dB
PL2W 15.12807274 W
PL12W 0.33867535 W
PL13W 0.33867535 W
SFO2 400.1316005 MHz

F2 - Processing parameters
SI 32768
SF 100.6128256 MHz
RGW 0
SSB 0
LB 0
GB 0
PC 1.40



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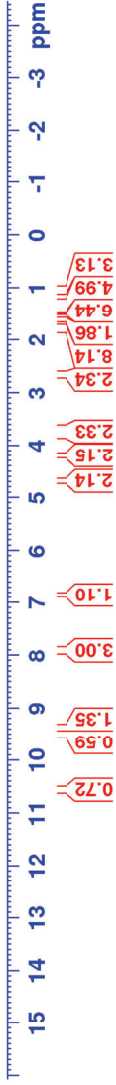
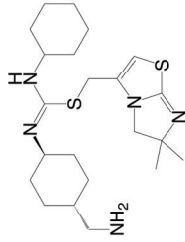
Current Data Parameters
NAME GK-II-25b
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20150322
Time 13.06
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT DMSO
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9845889 sec
RG 203
DW 60.800 usec
DE 6.50 usec
TE 303.3 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 12.00 usec
PL1 -2.00 dB
FL1 15.12807274 W
SFO1 400.1324710 MHz

F2 - Processing parameters
SI 32768
SF 400.1300000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

10.570
10.444
10.399
9.530
9.521
9.377
9.210
9.186
7.920
7.220
6.863
4.680
4.229
3.886
3.793
3.740
3.709
3.646
2.675
1.848
1.816
1.746
1.675
1.631
1.600
1.516
1.419
1.388
1.371
1.354
1.326
1.293
1.263
1.047
1.017



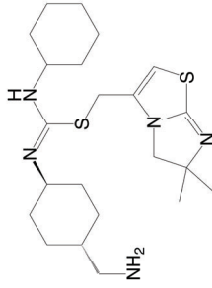
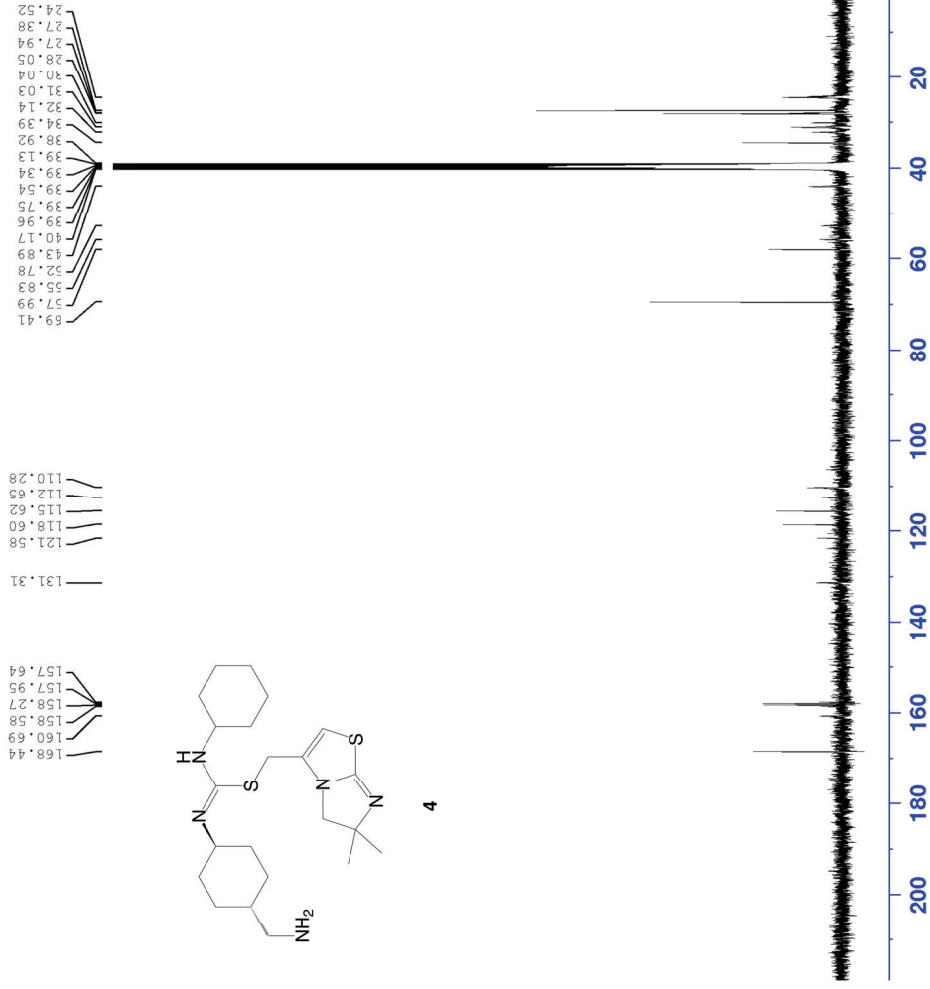
Mar.22,2015
C13CPD DMSO /opt/topspin Kang.g 45



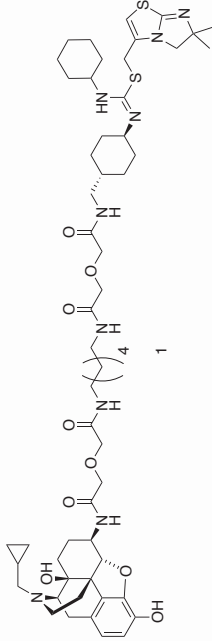
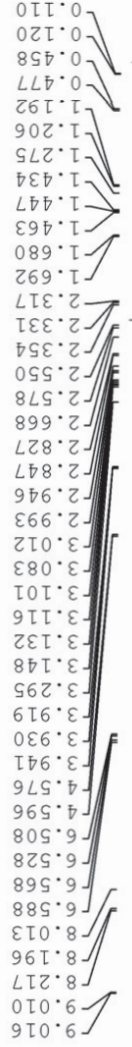
Current Data Parameters
NAME GR-II-256_C
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20150324
Time 1.52
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT DMSO
NS 8192
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631488 sec
RG 203
DW 20.800 usec
DE 6.50 usec
TE 304.2 K
D1 2.0000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 8.50 usec
PL1 -1.70 dB
PL1W 55.73028564 W
SFO1 100.6228298 MHz
===== CHANNEL f2 =====
CPDPRG[2] waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 -2.00 dB
PL2 14.50 dB
PL13 14.50 dB
PL2W 15.12807274 W
PL12W 0.33867535 W
PL13W 0.33867535 W
SFO2 400.1316005 MHz
F2 - Processing parameters
SI 32768
SF 100.6128193 MHz
WDW EM
SSB 0
GB 0
PC 1.00 Hz
LB 0
PC 1.40



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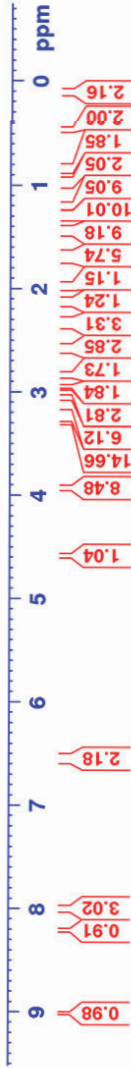


Current Data Parameters
 NAME GK-II-50e
 EXPNO 10
 PROCNO 1

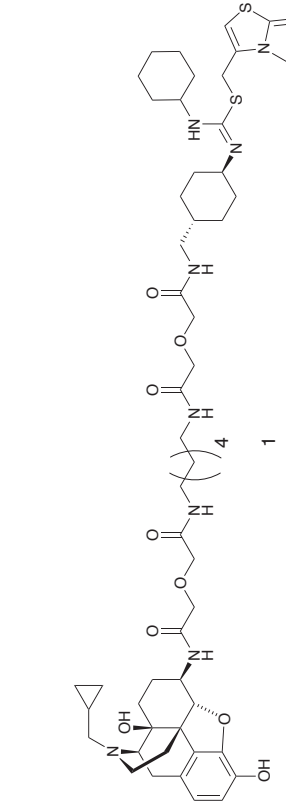
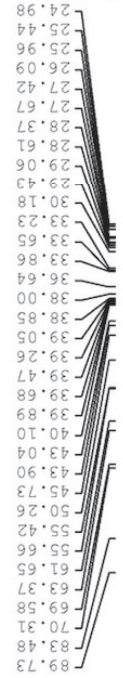
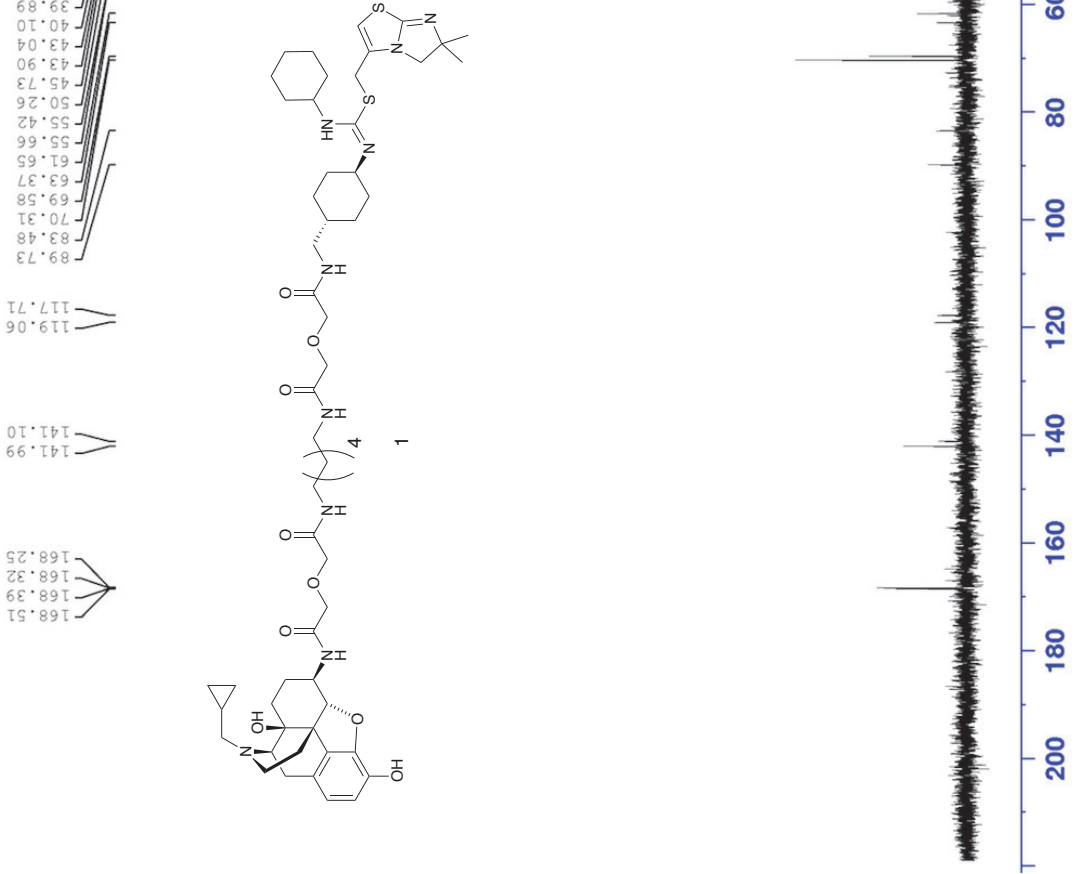
F2 - Acquisition Parameters
 Date_ 20150526
 Time_ 9.17
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 ID 65536
 SOLVENT DMSO
 NS 128
 DS 2
 SWH 8223.685 Hz
 FIDRES 0.125483 Hz
 AQ 3.9845889 sec
 RG 203
 DW 60.800 usec
 DE 6.50 usec
 TE 302.2 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 12.00 usec
 PL1 -2.00 dB
 FLLW 15.12807274 W
 SF01 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300050 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



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 C13CPD DMSO /opt/topspin Kang.g 10



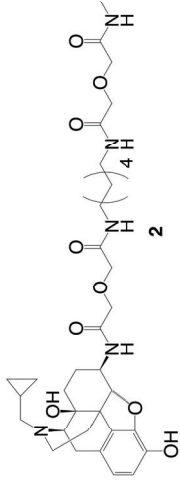
F2 - Acquisition Parameters
 Date_ 20150516
 Time_ 1.51
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT DMSO
 NS 8192
 DS 4
 SWH 24038.461 Hz
 FIDRES 0.366798 Hz
 AQ 1.3631488 sec
 RG 203
 DW 20.800 usec
 DE 6.50 usec
 TE 301.7 K
 D1 2.0000000 sec
 D11 0.0300000 sec
 TDO 1

==== CHANNEL f1 =====
 NUC1 13C
 P1 8.50 usec
 PL1 -1.70 dB
 PL1W 55.73028564 W
 SF01 100.6228298 MHz

==== CHANNEL f2 =====
 CPDPRG[2] waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -2.00 dB
 PL2 14.50 dB
 PL3 14.50 dB
 PL2W 15.12807274 W
 PL12W 0.33867535 W
 PL13W 0.33867535 W
 SFO2 400.1316005 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6128271 MHz
 EM
 SSB 0
 GB 0
 LB 1.00 Hz
 PC 1.40

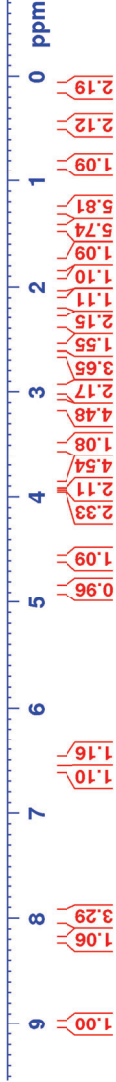
Aug.23, 2015
 PROTON DMSO /opt/topspin Kang.g 2



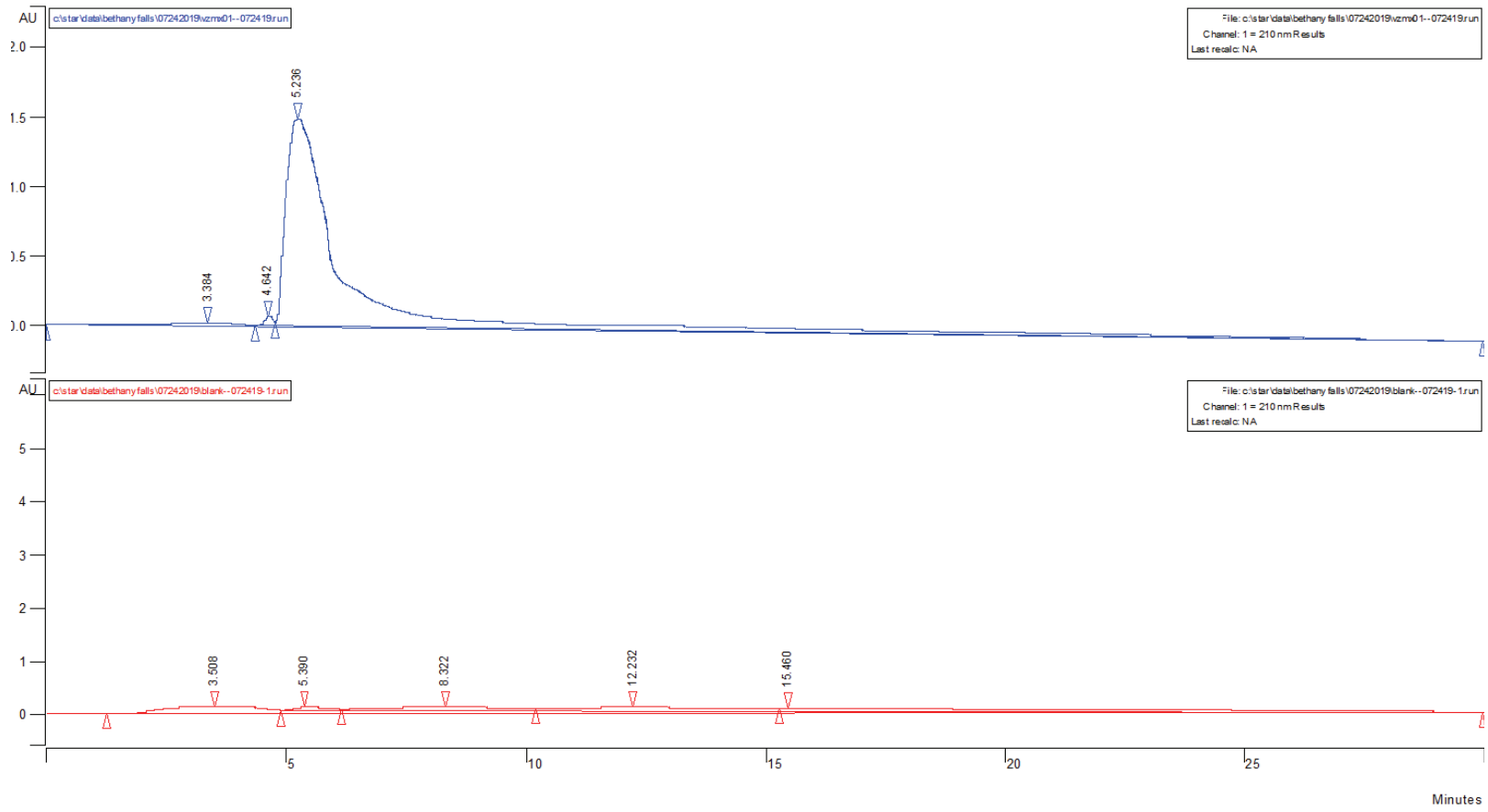
F2 - Acquisition Parameters
 Date_ 20150823
 Time_ 16.37
 INSTRUM spect
 PROBD 5 mm PABBO BB-
 PULPROG zg30
 TD 65336
 SOLVENT DMSO
 NS 16
 DS 2
 SWH 8223.685 Hz
 FIDRES 0.125483 Hz
 AQ 3.9845889 sec
 RG 161
 DW 60.800 usec
 DE 6.50 usec
 TE 303.0 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 12.00 usec
 PL -2.00 dB
 FLL 15.12607274 W
 SFO1 400.1324710 MHz

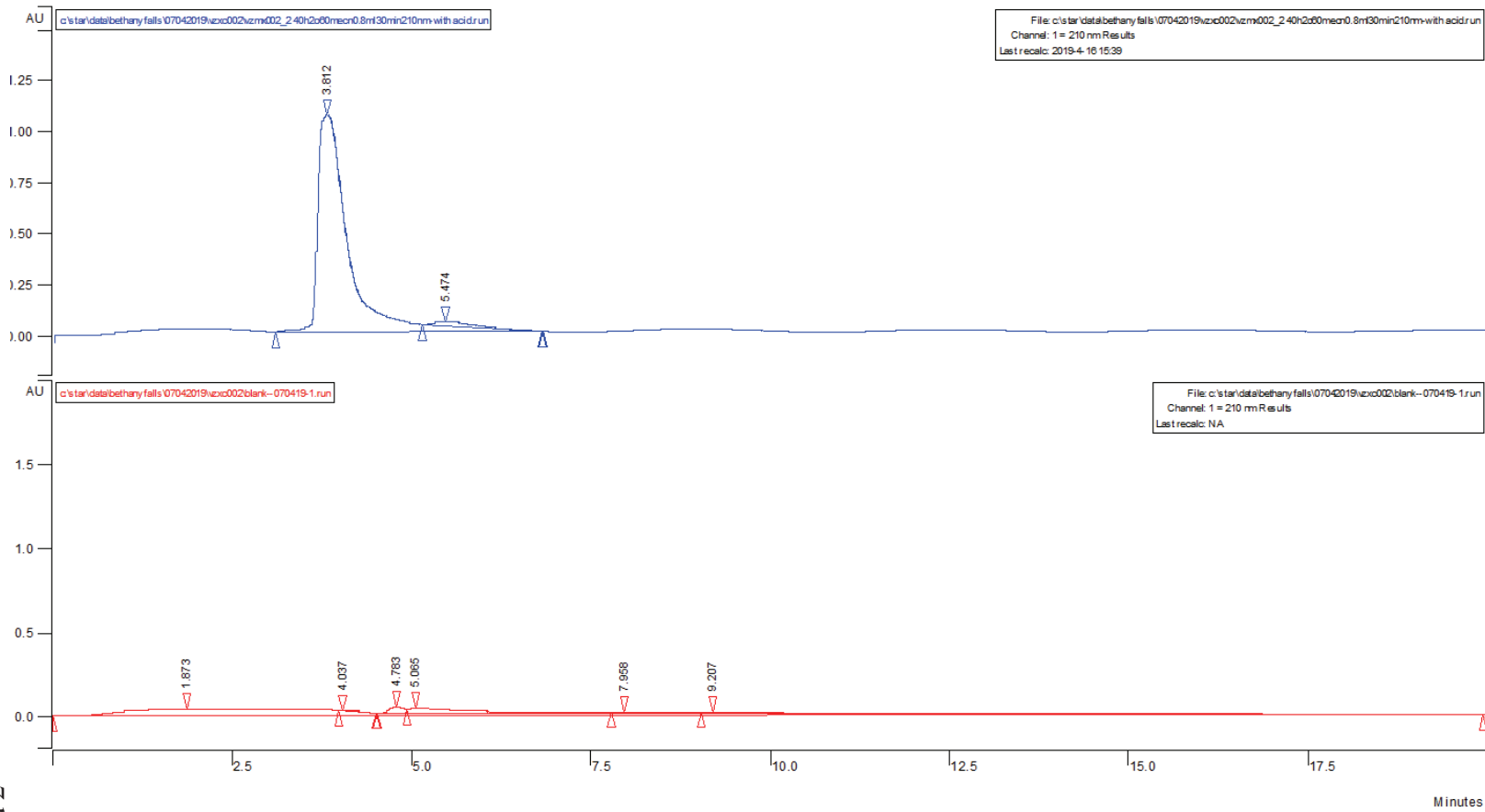
F2 - Processing parameters
 SI 32768
 SF 400.1300044 MHz
 WDW EM
 SSB 0
 LB 0
 GB 0
 PC 1.00



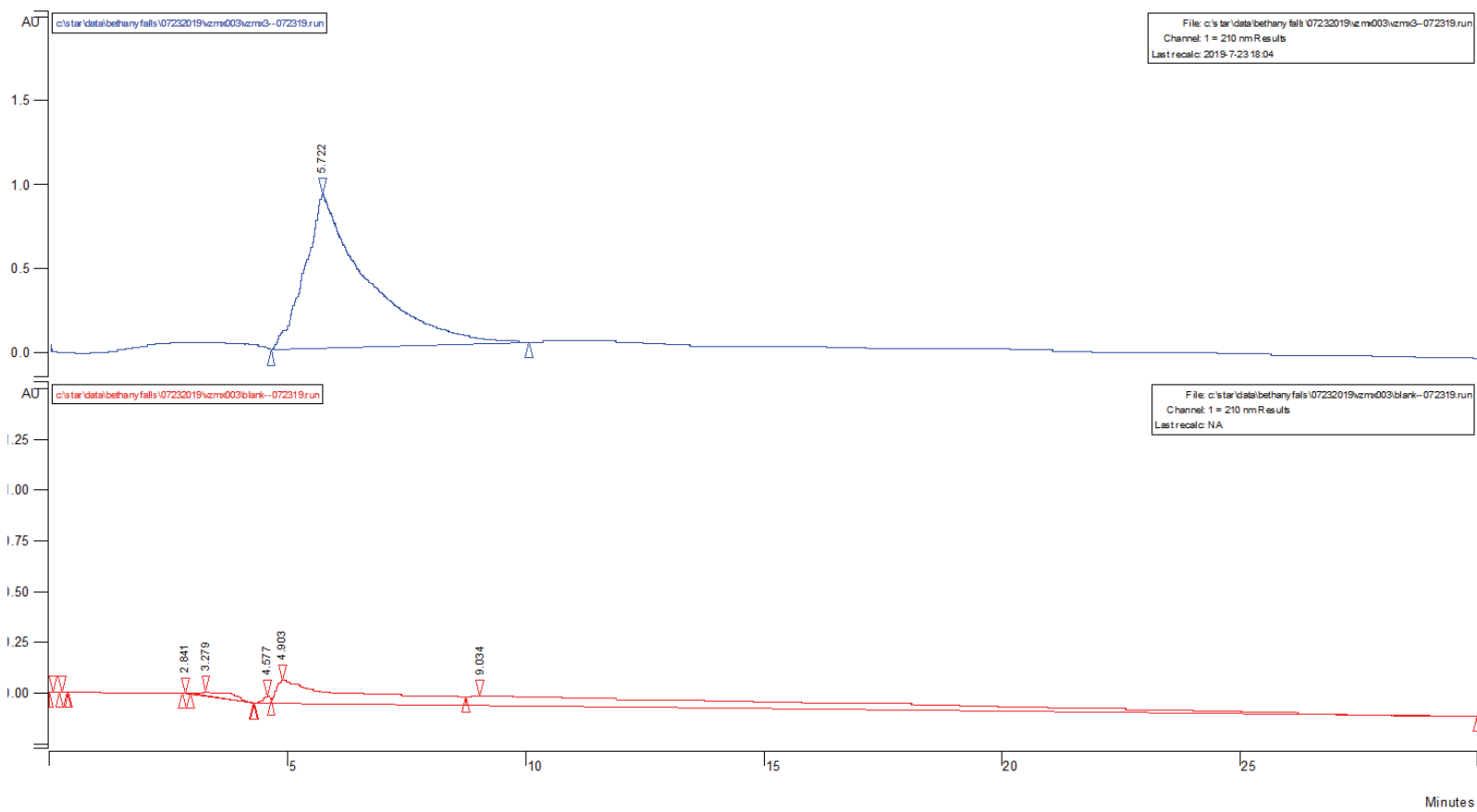
Compound 1



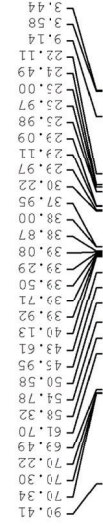
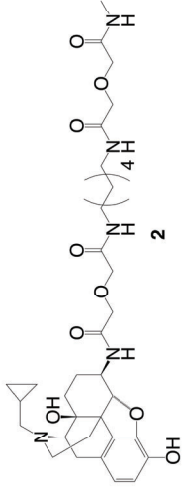
Compound 2



Compound 3



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 C13CPD DMSO /opt/topspin Kang.g 2



Current Data Parameters
 NAME GK-II-82_C
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20150823
 Time 19.11
 INSTRUM spect
 PROBHD 5 mm FAPBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT DMSO
 NS 10240
 DS 4
 SWH 24038.461 Hz
 FIDRES 0.366798 Hz
 AQ 1.3631488 sec
 RG 203
 DW 20.800 usec
 DE 6.50 usec
 TE 304.5 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TD0 1

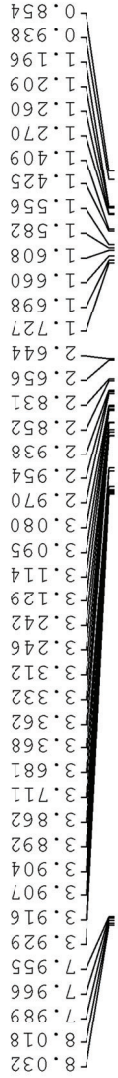
==== CHANNEL f1 =====
 NUC1 13C
 P1 8.50 usec
 PL1 -1.70 dB
 PL1W 55.73028564 W
 SF01 100.6228298 MHz

==== CHANNEL f2 =====
 CPDPRG[2] waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -2.00 dB
 PL12 14.50 dB
 PL13 14.50 dB
 PL14 14.50 dB
 PL1Z 15.12807274 W
 PL1Z 0.33867535 W
 PL13W 0.33867535 W
 SF0Z 400.1316005 MHz

F2 - Processing parameters
 SI 32766
 SF 100.6128261 MHz
 NDW 0
 SSB 0
 LB 0
 GB 0
 PC 1.40



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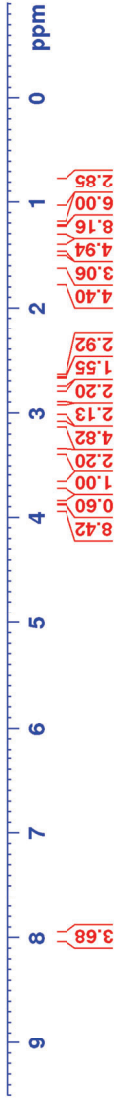
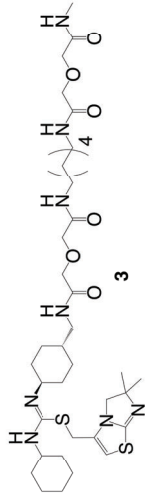


Current Data Parameters
NAME Bf-II-ititmono
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20171107
Time 10.49
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT DMSO
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9845889 sec
RG 203
DW 60.800 usec
DE 6.50 usec
TE 299.8 K
D1 1.00000000 sec
TD0 1

==== CHANNEL f1 =====
NUC1 1H
P1 12.00 usec
PL 2.00 dB
FLL 15.12807274 W
SFO1 400.1324710 MHz

F2 - Processing parameters
SI 32768
SF 400.1300022 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00





Current Data Parameters
 NAME BF-III-itlmono
 EXPNO 60
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20171114
 Time 19.02
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT DMSO
 NS 1024
 DS 4
 SWH 24038.461 Hz
 FIDRES 0.366798 Hz
 AQA 1.3631488 sec
 RG 203
 DW 20.800 usec
 DE 6.50 usec
 TE 300.2 K
 D1 2.0000000 sec
 D11 0.0300000 sec
 TD0 1

==== CHANNEL f1 =====
 NUC1 13C
 P1 8.50 usec
 PL1 -1.70 dB
 PL1W 55.73028564 W
 SFO1 100.6228298 MHz

==== CHANNEL f2 =====
 CPDPRG[2] waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -2.00 dB
 PL12 14.50 dB
 PL13 14.50 dB
 PL1W 15.12807274 W
 PL12W 0.33867535 W
 PL13W 0.33867535 W
 SFO2 400.1316005 MHz

F2 - Processing Parameters
 SI 32768
 SF 100.6128193 MHz
 GB 0
 SSB 0
 LB 0
 GB 0
 PC 1.40
 LB 1.40

