

Supporting Information for

Original article

Evodiamine-inspired dual inhibitors of histone deacetylase 1 (HDAC1) and topoisomerase 2 (TOP2) with potent antitumor activity

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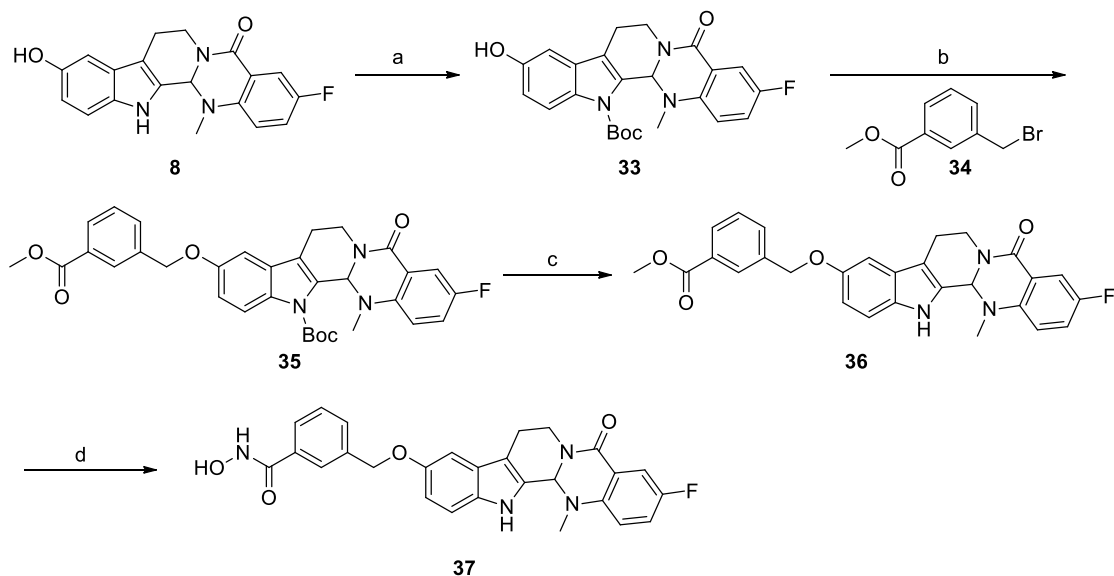
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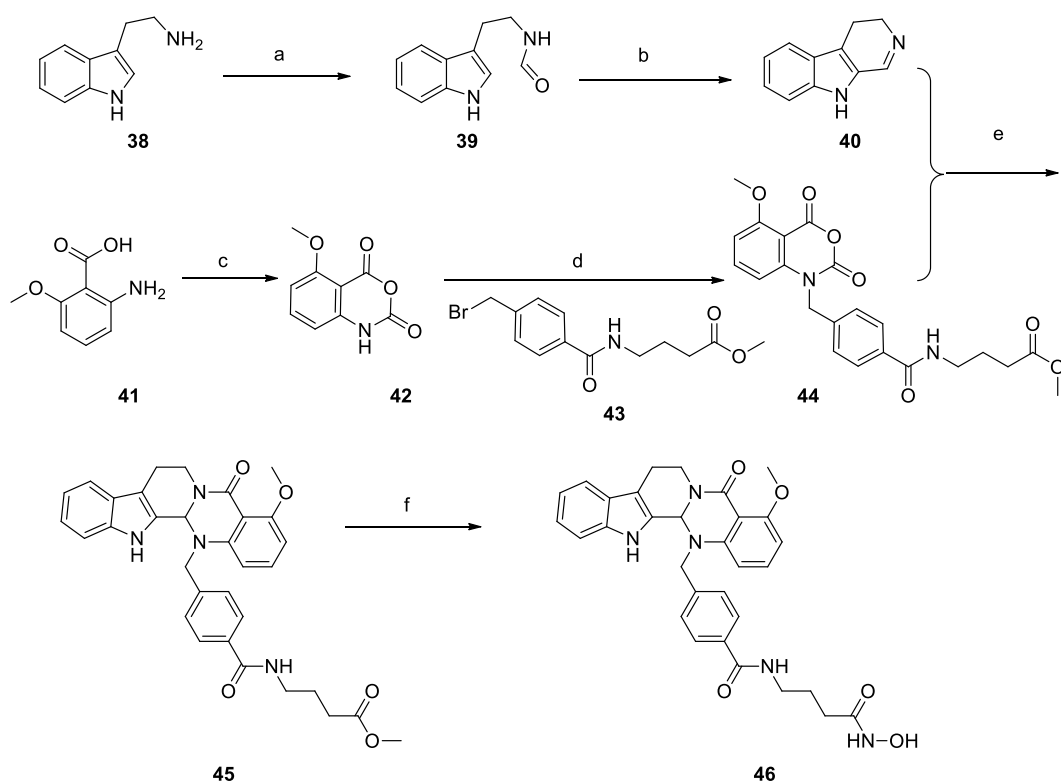
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Scheme S1 Synthesis of compound **37**. Reagents and conditions: (a) $(\text{Boc})_2\text{O}$, DMAP, NaOH, THF, K_2CO_3 , reflux, r.t., 4 h, 69%; (b) K_2CO_3 , DMF, 12 h, 66%–88%; (c) TFA, DCM, r.t., 6 h, 64%–69%; (d) NH_2OH , CH_3OH , 45 °C, 1 h, 52%–81%.



Scheme S2 Synthesis of compound **46**. Reagents and conditions: (a) ethyl formate, reflux, 12 h, 90%; (b) POCl_3 , DCM, 0 °C, 6 h, 68%; (c) triphosgene, THF, reflux, 12 h, 75%; (d) K_2CO_3 , DMF, 2.5 h, 59%–68%; (e) DCM, r.t., 12 h, 39%–56%; (f) $\text{NH}_2\text{OH}\cdot\text{HCl}$, KOH, MeOH, 1 h, 39%–48%.

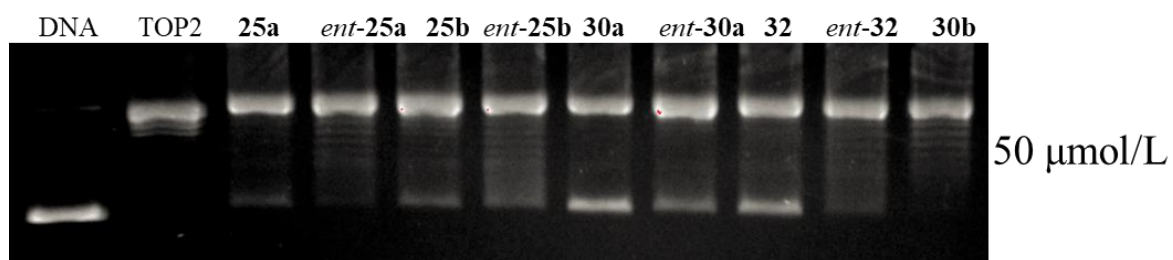


Figure S1 TOP2 inhibitory potency of the target compounds at 50 $\mu\text{mol/L}$. Lane 1, supercoiled plasmid DNA; lane 2, DNA + TOP2; lanes 3–11, DNA + TOP2 + compound.

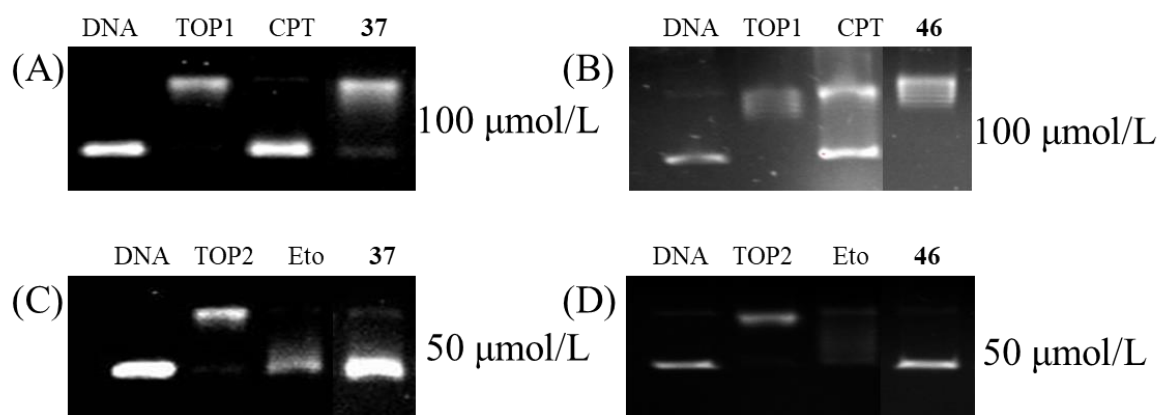


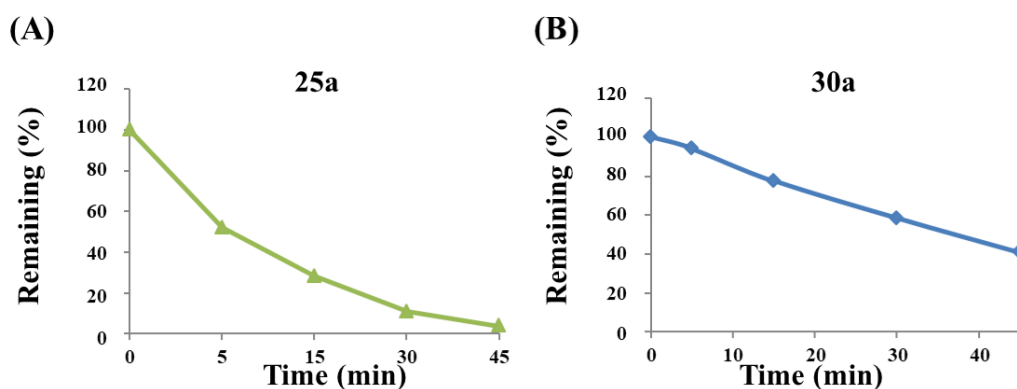
Figure S2 TOP inhibitory potency of the compounds **37** and **46**. TOP1 inhibitory activity of compounds **37** (A) and **46** (B) at 100 $\mu\text{mol/L}$; TOP2 inhibitory activity of compounds **37** (C) and **46** (D) at 50 $\mu\text{mol/L}$.

Table S1 *In vitro* antiproliferative activity of compounds **37** and **46**.

Compd.	IC ₅₀ (μmol/L)			
	HDAC1	HCT116	MCF-7	A549
37	1.9	> 20	> 20	> 20
46	> 1	> 10	> 10	> 10

Table S2 *In vitro* antiproliferative activity of compounds **25a** and **30a**.

Compd.	IC ₅₀ (μmol/L)			
	HEL	HL60	K562	HUVEC
25a	0.34 ± 0.04	0.63 ± 0.15	1.0 ± 1.2	7.4 ± 1.1
30a	0.66 ± 0.05	0.56 ± 0.06	1.9 ± 0.65	48 ± 3.2
1	0.21 ± 0.03	0.49 ± 0.04	0.58 ± 0.20	5.9 ± 2.5
6	1.61 ± 0.16	1.30 ± 0.05	1.34 ± 0.42	3.7 ± 2.2
1 + 6 (1:1)	0.19 ± 0.06	0.61 ± 0.11	0.47 ± 0.08	1.0 ± 0.1

**Figure S3** The mice microsomal stability of compounds **25a** and **30a**.

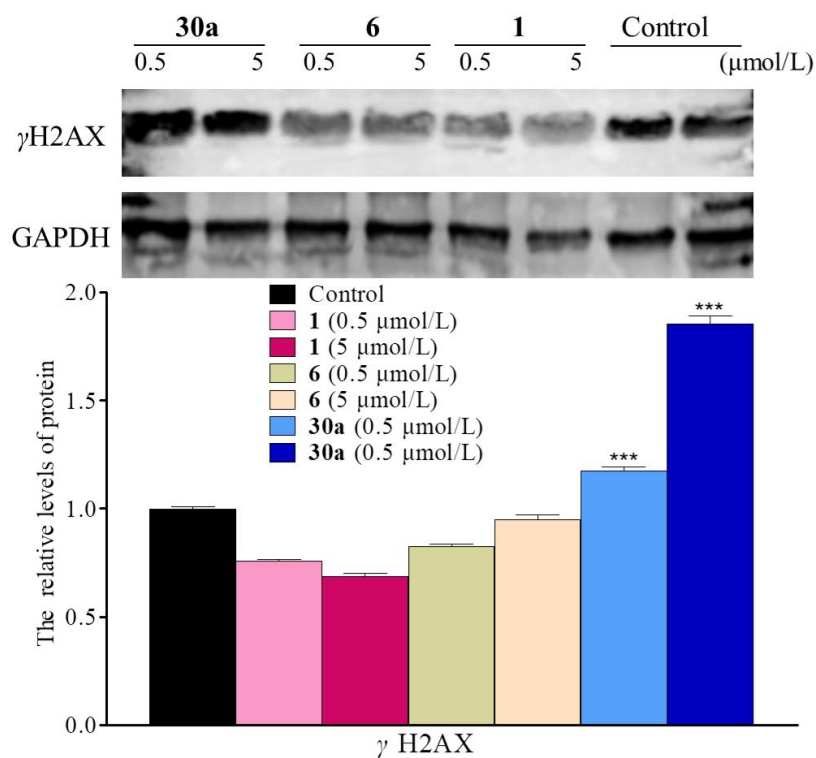


Figure S4 Western blot probing for γ H2AX in HCT116 cells after 24 h treatment with compounds **1**, **6** and **30a**. Data were expressed as mean \pm SD ($n = 3$). * $P < 0.05$, ** $P < 0.01$ and *** $P < 0.001$ vs. the control group, determined with student's t test.

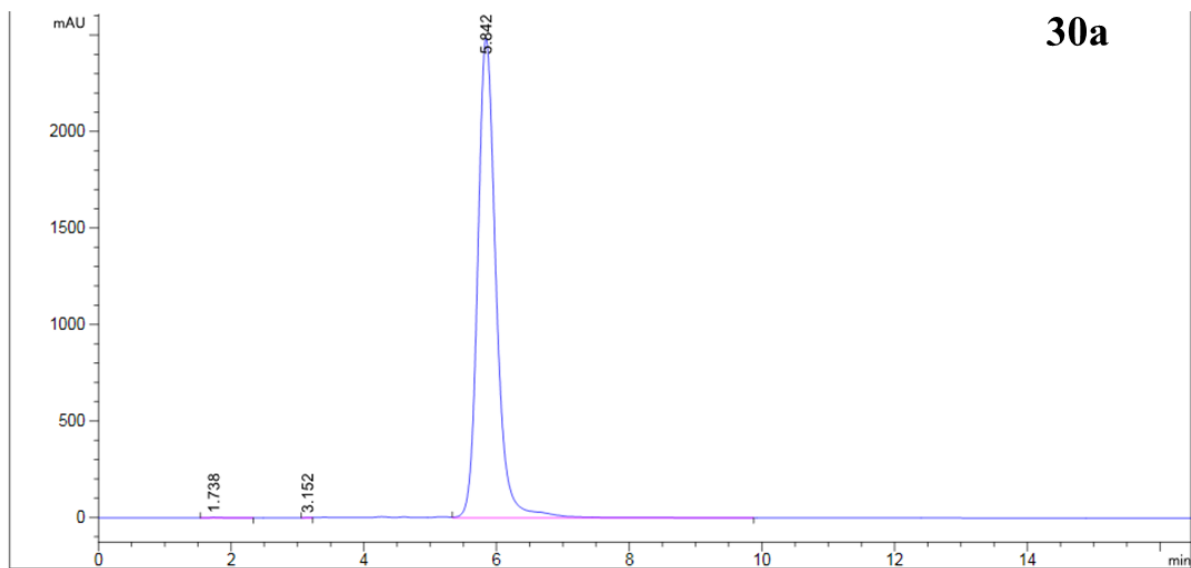
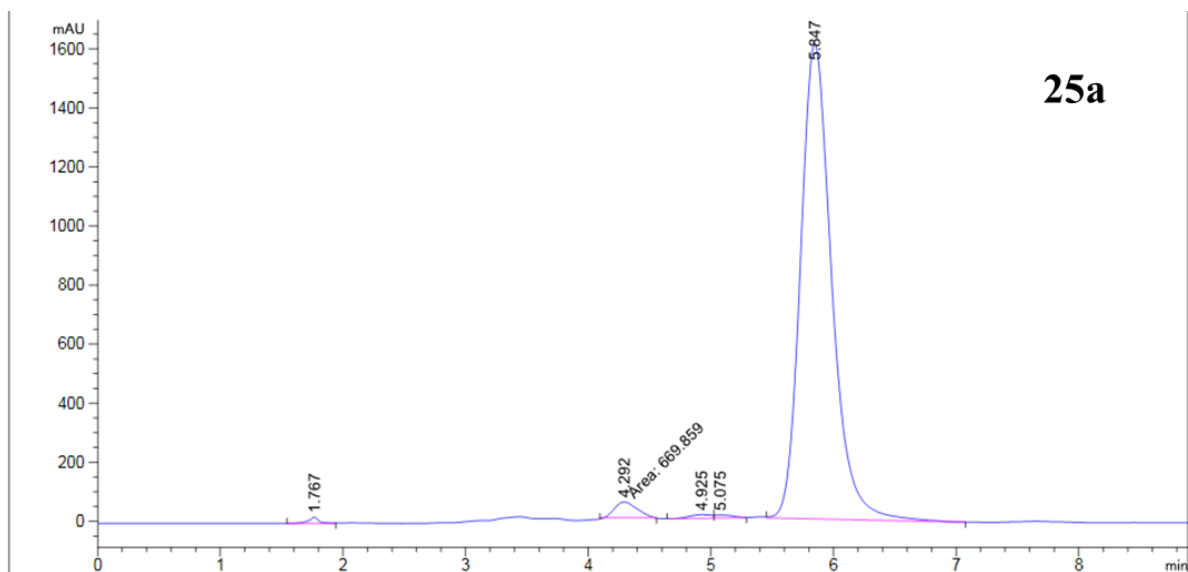


Figure S5 HPLC spectra of compounds **25a** and **30a**.

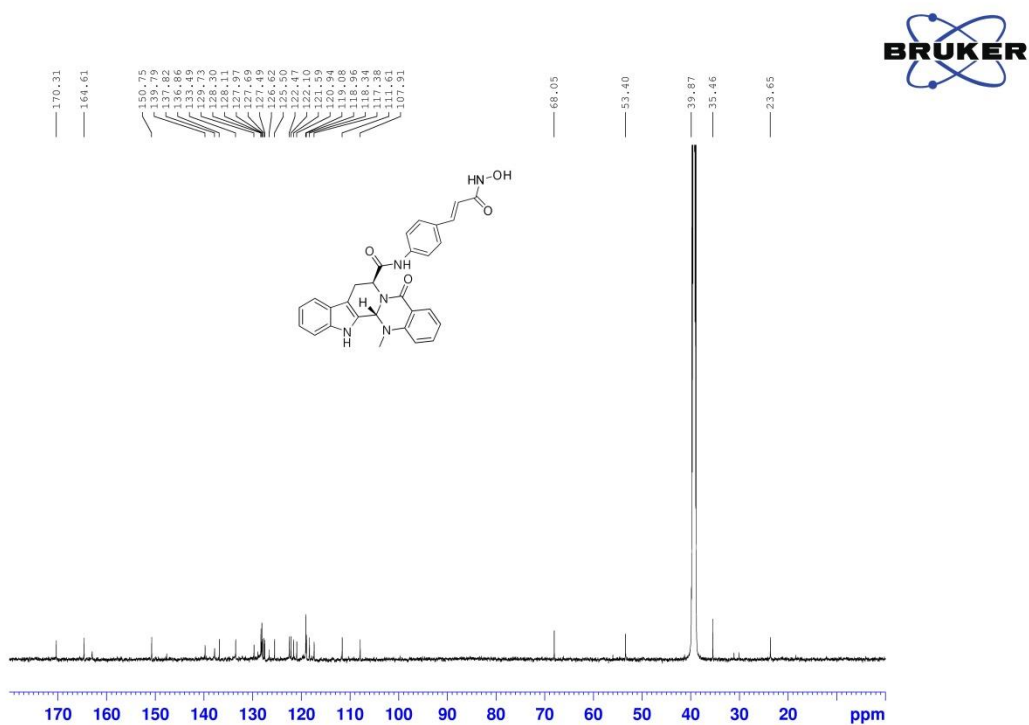
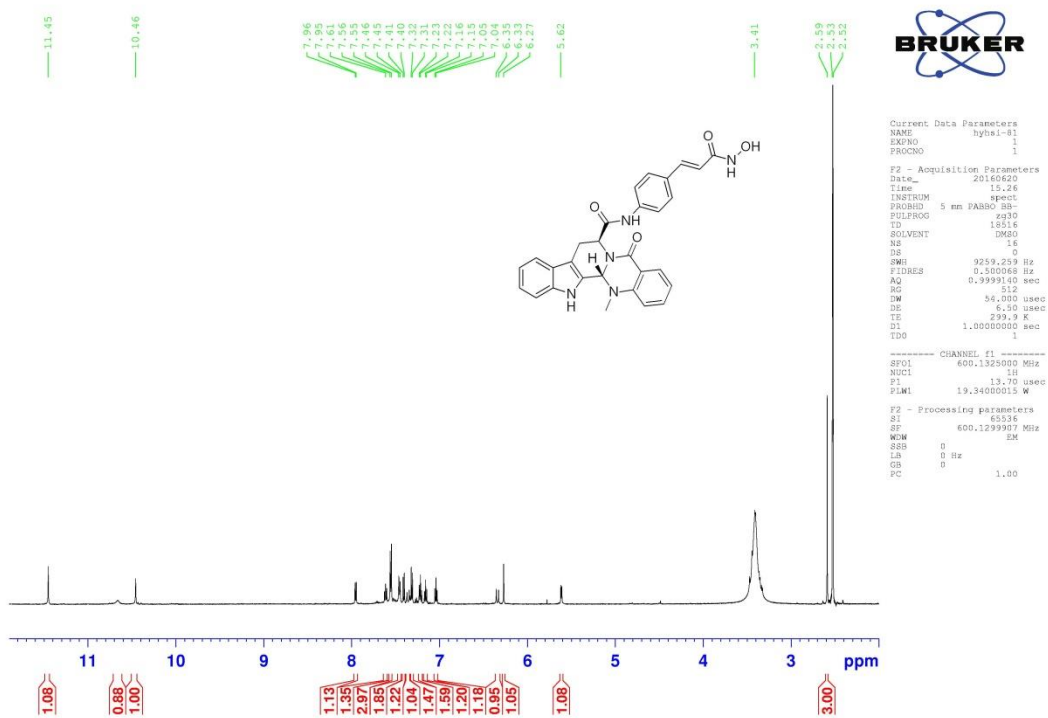


Figure S6 ¹H NMR and ¹³C NMR spectra of compound 25a.

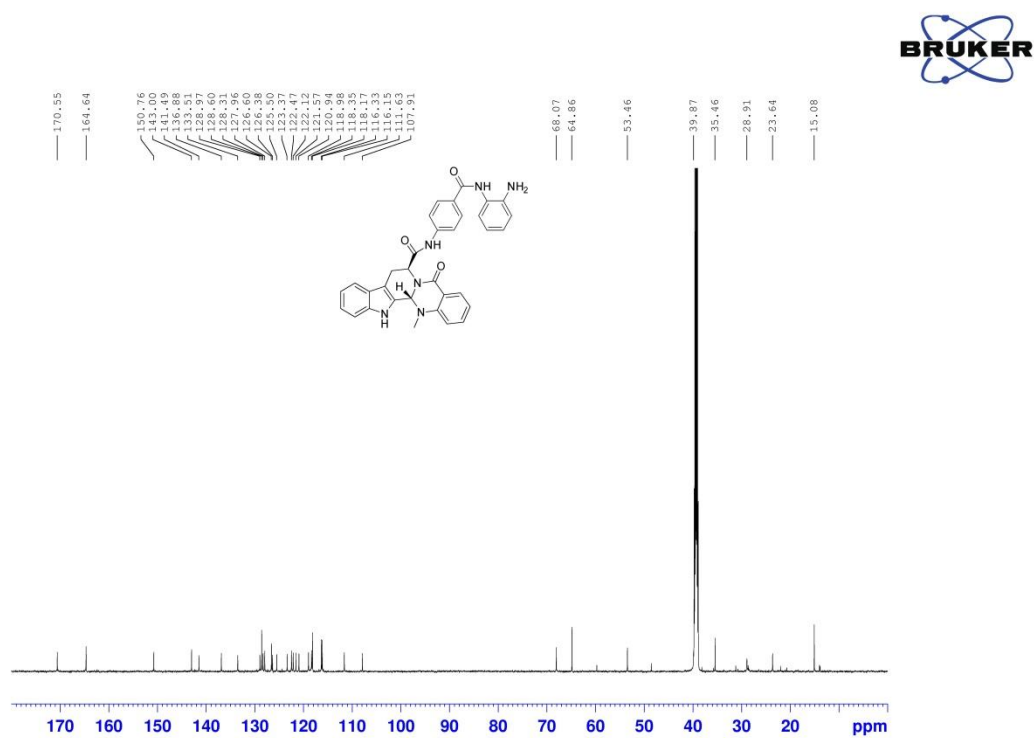
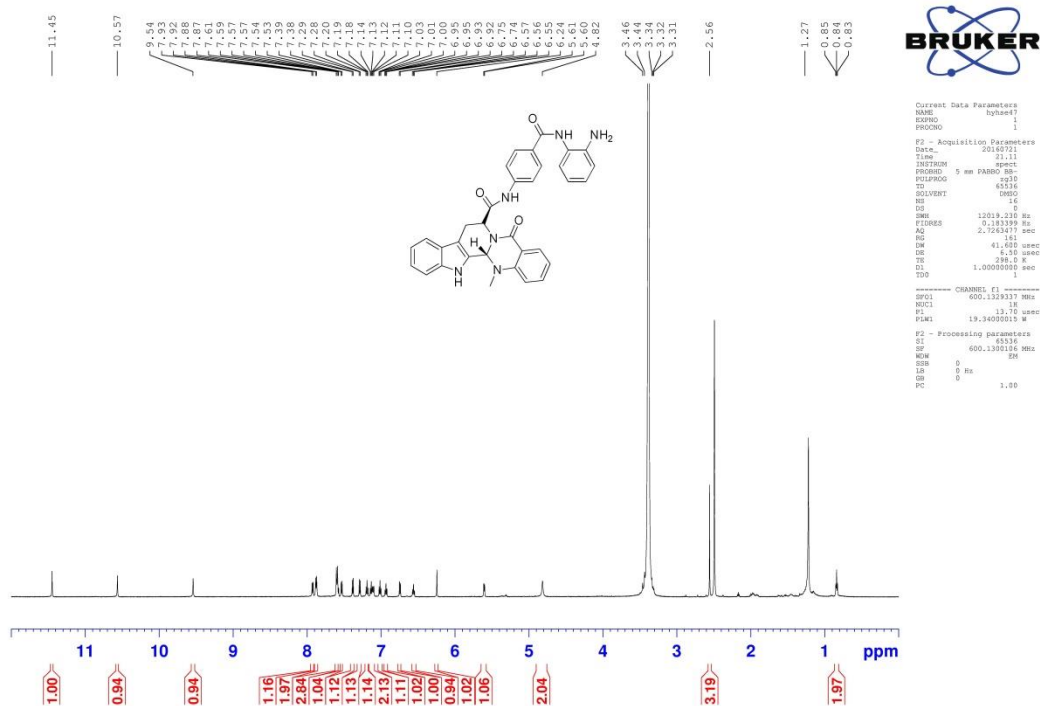


Figure S7 ¹H NMR and ¹³C NMR spectra of compound 30a.

Structural characterization of compounds 37 and 46

Compounds **37** and **46** were obtained from a similar protocol described for compound **20a**.

3-(((3-Fluoro-14-methyl-5-oxo-5,7,8,13,13b,14-hexahydroindolo[2',3':3,4]pyrido[2,1-b]quinazolin-10-yl)oxy)methyl)-N-hydroxybenzamide (37). Light yellow solid (Yield: 74%). ¹H NMR (DMSO-*d*₆, 600 MHz) δ 11.05 (s, 1H), 7.88 (s, 1H), 7.70 (d, *J* = 7.6 Hz, 1H), 7.57 (d, *J* = 6.0 Hz, 1H), 7.53 (dd, *J* = 2.8, 8.5 Hz, 1H), 7.43 (t, *J* = 7.1 Hz, 1H), 7.40–7.35 (m, 1H), 7.27 (d, *J* = 8.9 Hz, 1H), 7.18 (q, *J* = 4.5 Hz, 1H), 7.12 (d, *J* = 2.5 Hz, 1H), 6.87 (dd, *J* = 2.5, 8.6 Hz, 1H), 6.06 (s, 1H), 5.13 (s, 2H), 4.63–4.60 (m, 1H), 3.22–3.17 (m, 1H), 2.83–2.79 (m, 2H), 2.68 (s, 3H). ¹³C NMR (150 MHz, DMSO-*d*₆, TMS) δ 163.51, 158.35, 156.77, 152.80, 146.46, 138.41, 133.93, 132.41, 130.84, 130.26, 128.77, 126.59, 126.46, 126.33, 122.46, 122.41, 122.06, 122.01, 121.09, 120.94, 113.93, 113.77, 113.12, 112.83, 111.89, 102.37, 102.26, 70.07, 69.76, 40.85, 37.06, 20.11. MS (ESI positive): *m/z* [M+H]⁺: 487.18.

N-(4-(Hydroxyamino)-4-oxobutyl)-4-((4-methoxy-5-oxo-7,8-dihydroindolo[2',3':3,4]pyrido[2,1-b]quinazolin-14(5H,13H,13bH)-yl)methyl)benzamide (46). Off-white solid (Yield: 43%). ¹H NMR (DMSO-*d*₆, 600 MHz) δ 11.40 (s, 1H), 8.53 (s, 1H), 7.68 (d, *J* = 7.76 Hz, 2H), 7.49 (d, *J* = 7.88 Hz, 1H), 7.37 (d, *J* = 8.18 Hz, 1H), 7.23 (t, *J* = 8.19 Hz, 1H), 7.17 (d, *J* = 7.74 Hz, 2H), 7.11 (t, *J* = 7.51 Hz, 1H), 7.01 (t, *J* = 7.51 Hz, 1H), 6.59 (d, *J* = 8.65 Hz, 1H), 6.46 (d, *J* = 8.42 Hz, 1H), 6.03 (s, 1H), 4.48 (d, *J* = 15.93 Hz, 1H), 4.36–4.38 (m, 1H), 4.18 (d, *J* = 15.93 Hz, 1H), 3.74 (s, 3H), 3.39–3.41 (m, 3H), 3.18–3.24 (m, 2H), 2.78–2.82 (m, 1H), 2.69–2.73 (m, 1H). ¹³C NMR (DMSO-*d*₆, 600 MHz) δ 167.47, 166.24, 162.74, 161.00, 150.40, 142.32, 137.09, 133.53, 133.30, 129.56, 127.90, 127.50, 126.21, 122.43, 119.33, 118.83, 112.49, 112.10, 111.64, 111.13, 105.60, 68.62, 56.09, 52.12, 36.51, 32.85, 25.41, 19.97.

Molecular docking

The crystal structure of HDAC1 was obtained from protein database bank (PDB ID: 4BKX)¹ and prepared for docking using the protein preparation tool in Discovery Studio 3.0. During this process, the ligands and waters were removed and hydrogens were added to the structure. Staged minimization was performed with default setting. The docking studies were carried out using GOLD 5.0. Binding site was defined as whole residues within a 10 Å radius subset encompassing the ligand. Conformations were generated by genetic algorithm and scored using GoldScore as fitness function. The best conformation was chosen to analyze the ligand–protein interaction. Docking analysis of TOP2 α (PDB ID: 5GWK)² with compound

30a was performed using a similar procedure.

Determination of *in vitro* metabolic stability in female ICR mice liver microsomes

Liver microsomal incubations were conducted in triplicate. Incubation mixtures (0.5 mg/mL microsome protein, pH 7.4 of 100 mmol/L phosphate buffer, 1 μ mol/L compounds in acetonitrile) were first shaken for pre-incubation at 37 °C. The reaction was initiated by adding NADPH to obtain concentration of 1 mmol/L NADPH in the mixture. For metabolic stability studies, aliquots of the incubation sample mixture were collected at 0, 5, 15, 30 and 45 min. After collection of samples, the reaction was terminated with acetonitrile containing the internal standard (200 ng/mL tolbutamide). The mixture was then centrifuged to remove the protein and the supernatant was subsequently applied to LC–MS/MS analysis. Positive control samples were prepared as described above, except the test compound was replaced with the known P450 substrate (1 μ mol/L midazolam).

Pharmacokinetic studies

Male SD rats (6–8 weeks old, body weight 180–220 g) were obtained from Shanghai Sippr-BK laboratory animal Co., Ltd., China. Three animals were given *p.o.* (i.p.) at a dose of 20 mg/kg (20 mg/kg) of compound **30a**. The dose solution was prepared in 5% DMSO + 10% solutol + 85% saline. After administration of compound **30a** ($n = 3$ per time point), 0.25 mL of blood was collected *via* Jugular vein puncture and put on the ice (heparin prevents clotting). Pharmacokinetic time points were 0.083, 0.25, 0.5, 1, 2, 4, 6, 8 and 24 h post-dose. Blood samples were transferred to microcentrifuge tubes and centrifuged (8000 rpm, Eppendorf, Hamburg, SN, Germany, 6 min at 2–8 °C). The separated plasma was stored at approximately –80 °C until thawed for LC–MS/MS. Data acquisition: Analyst version 1.5.1. Pharmacokinetic data analysis was performed on mean plasma concentration-time data and pharmacokinetic parameters were calculated using WinNonlin version 5.2.

References

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2. Wang Y, Chen S, Wu C, Liao Y, Lin T, Liu K, et al. Producing irreversible topoisomerase II-mediated DNA breaks by site-specific Pt(II)-methionine coordination chemistry. *Nucleic Acids Res* 2017;**45**:10920.