Distinct Cdk9-phosphatase switches act at the beginning and end of elongation by RNA polymerase II

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Running title: Cdk9-phosphatase circuits regulate transcription elongation

Supplementary Figures and Legends



Supplementary Fig. 1 PP1 regulates Spt5 phosphorylation in human cells. a Schematic diagram of Spt5 protein. **b** Immunoprecipitated GFP-tagged PP1 isoforms (PP1 α , PP1 β and PP1 γ) dephosphorylated pThr806-containing phosphopeptide and a control (H3pSer10), but not pSer666. Phosphate release was measured colorimetrically. Individual data points are shown in the plot; error bars indicate ± s.d. of three biological replicates (*n* = 3). **c** Anti-GFP immunoblot shows amounts of PP1 isoforms immunoprecipitated. **d** Immunoblot to measure depletion of PP1 isoforms (PP1 α , PP1 β and PP1 γ) using isoform-specific siRNAs (singly or in combinations) and levels of Spt5, and Spt5-pThr806. **e** Immunoblot (top) of chromatin fraction (insoluble) indicates pThr806 was increased, whereas pSer666 was unchanged in cells depleted of all

three PP1 isoforms with an siRNA cocktail. Signals were scanned and quantified (bottom) with ImageJ software (n = 1 biologically independent experiment). Source data are provided as a Source Data file.



Spt5-pThr806

4

Spt5-pThr806

Supplementary Fig. 2 ChIP-seg analysis of Spt5-pThr806 and Pol II-pSer2. a Immunoblot analysis shows that upon phosphorylation by Cdk9, anti-pThr806 antibody recognizes GST-CTR1 (amino acids 720-830) and full-length protein, but not GST-CTR2 (amino acids 844-1087); top (anti-pThr806), bottom left (anti-GST), and bottom right (anti-Spt5). b Ectopically expressed FLAG-tagged Spt5—wild-type (WT) and indicated CTR1-mutant variants (M1-M5) was directly immunoblotted or immunoprecipitated with anti-FLAG (M2) antibody and probed with anti-pThr806 and anti-FLAG antibodies, as indicated (left). Signals were quantified using Image Studio software in Odyssey CLx Imager (right) (n = 1 biologically independent experiment). c Correlation between ChIP-seg samples. Paired-end sequencing reads were mapped to human genome using Bowtie2. Average read coverages were calculated by feeding resulting BAM files into 'multiBamSummary' program (Galaxy Version 3.1.2.0.0). Values in boxes represent Pearson's correlation coefficients between corresponding samples (n = 2biological replicates). d Scatterplots represent correlations between Pol II. Pol II-pSer2, Spt5 and Spt5-pThr806 downstream of the TSS (0 to +200 bp) for genes highly occupied by Pol II (n = 2,572). e Scatterplots represent correlations between Pol II, Pol II-pSer2, Spt5 and Spt5pThr806 at termination zone (TZ; 0 to +2,500 bp of the CPS) for genes highly occupied by Pol II (n = 2,572). Source data are provided as a Source Data file.



Supplementary Fig. 3 Spt5-CTR1 phosphorylation drops in termination zones of replication-dependent histone and noncoding RNA genes transcribed by Pol II. a Browser tracks of representative histone genes, with 3' regions indicated by red arrows, where ChIP-seq signals of Pol II, pSer2, Spt5 and pSer666, but not pThr806, accumulate. **b** Metagene analysis of Pol II, pSer2, Spt5 and pThr806 ChIP-seq signals on snRNA and snoRNA genes, with annotated 3' end indicated by dashed vertical line. The shaded area represents standard deviation (± s.d.) from the mean distribution (solid line).



Supplementary Fig. 4 Cdk9 inhibition diminishes Spt5-pThr806 but increases Pol II CTDpSer2 on GAPDH. a Schematic of the GAPDH gene, indicating positions of primer pairs used in ChIP-qPCR analysis. b ChIP-qPCR analysis of total Pol II and CTD-pSer2 in HCT116 cells treated with NVP-2 (250 nM) or mock treated (DMSO) for 1 hr. c ChIP-qPCR analysis of total Spt5 and Spt5-pThr806 in HCT116 cells treated with NVP-2 (250 nM) or mock treated (DMSO) for 1 hr. Individual data points are shown in the plots; error bars indicate \pm s.d. of three biological replicates (**b** and **c**). Source data are provided as a Source Data file.



Supplementary Fig. 5 Distribution and phosphorylation of Spt5 and Pol II-Ser2 on chromatin. a Immunoblot analysis shows that upon phosphorylation by Cdk9, anti-pSer666 antibody recognizes full-length Spt5 protein, but not GST-CTR1 or GST-CTR2; blots were probed with anti-pSer666 (left), anti-GST (middle), and anti-Spt5 (right). **b** Ectopically expressed FLAG-tagged Spt5—wild-type (WT) and Ser666-mutant variants (Spt5^{S666A} and Spt5^{S666D})—was directly immunoblotted (top, "input") or immunoprecipitated with anti-FLAG antibody (bottom, "FLAG-IP") and probed with anti-pSer666, anti-Spt5, anti-pThr806 antibodies. **c** Metagene analysis and heat map (n = 20,130 genes) of ChIP-seq data for Spt5-pSer666 (n = 2 biological replicates). **d** ChIP-qPCR analysis of Spt5-pSer666 on *GAPDH* in HCT116 cells treated with NVP-2 (250 nM) or mock treated (DMSO) for 1 hr. Individual data points are shown in the plots;

error bars indicate \pm s.d. of three biological replicates (**b** and **c**). Source data are provided as a Source Data file.



Supplementary Fig. 6 Spt5 phosphorylation in response to p53 activation in the absence or presence of a Cdk9 inhibitor. a Time-dependent induction of two p53-responsive genes, *CDKN1A* and *MDM2*, in response to treatment with nutlin-3 (5 μ M), measured by RT-qPCR amplification relative to *GAPDH* control. Individual data points from two biological replicates (*n* = 2) are shown. b Schematic of *CDKN1A* gene, indicating positions of primer pairs used in ChIP-qPCR analysis, and ChIP-qPCR analysis of Spt5-pSer666, Spt5-pThr806, Pol II and Pol II-pSer2 in HCT116 cells mock treated (DMSO) or treated with nutlin-3 (5 μ M) alone or together with NVP-2 (250 nM). Individual data points are shown in the plots; error bars indicate ± s.d. from four biological replicates (*n* = 4). Source data are provided as a Source Data file.



Supplementary Fig. 7 Distribution of Pol II and Pol II-pSer2 on genes induced by p53 activation. a Metagene analysis of Pol II and Pol II-pSer2 occupancy in TSS regions of genes induced by nutlin-3 (n = 75; paused genes). The shaded area represents standard deviation (\pm

s.d.) from the mean distribution (solid line). **b** Box plots comparing ChIP-seq signals in first 100nt interval downstream of TSS, in the absence or presence of nutlin-3, for Pol II and Pol II-pSer2 at genes induced by nutlin-3, divided into those classified as pause-regulated (Pause index \geq 2.0, *n* = 75) or non-paused (pause index < 2.0, *n* = 25). **c** Box plots show distribution of Spt5pSer666 ChIP-seq signals in 100-nt intervals from -100 to +500 bp of TSS (*n* = 75). **d** Box plots show distribution of Spt5-pThr806 ChIP-seq signals in 100-nt intervals from -100 to +500 bp of TSS (*n* = 75). **e** Box plots show enrichment of Spt5 (left), Spt5-pSer666 (middle) and Spt5pThr806 (right) mean ChIP-seq signals in intervals around the CPS (x = -2.5 kb to 0; y = 0 to +2.5 kb) when cells were treated with nutlin-3 or mock-treated (DMSO) (*n* = 75). Indicated *p*values were calculated by two-sided Student's *t*-test (**b**, **c**, **d** and **e**). Source data are provided as a Source Data file.



Supplementary Fig. 8 PP4 is a candidate Spt5 phosphatase. a PP4C and PP4R2 were immunoprecipitated from extracts prepared from HCT116 cells mock treated (DMSO) or treated with NVP-2 (250 nM) or RO-3306 (10 μ M) for 1 hr. PP4C was co-precipitated with anti-PP4R2 but PP4R2 was not detectable in anti-PP4C immunoprecipitate. b Anti-PP4R2 immunoprecipitates were incubated with 5 ng purified, recombinant Cdk9/cyclin T1 and ATP or mock-treated (as indicated), washed and tested for phosphatase activity towards an Spt5-derived phosphopeptide containing pSer666 or pThr806. Error bars indicate + s.d. from three biological replicates (n = 3). **c** Quantification of immunoblot results in Fig. 6d. Error bars indicate \pm s.d. from 4 biological replicates (n = 4). Individual data points from biological replicates are shown in the plots (**b** and **c**). Source data are provided as a Source Data file.



Supplementary Fig. 9 Distinct spatial distributions and substrate specificities of two Cdk9-regulated, phospho-Spt5 phosphatases. a Schematic of the GAPDH and CDKN1A genes, indicating positions of primer pairs used in ChIP-qPCR analysis (top). ChIP-qPCR analysis of PP4C, PP4R2 and PP1y on the GAPDH and CDKN1A gene in unperturbed HCT116 cells (bottom) reveals reciprocal distribution patterns: PP4 subunits are enriched near the 5' end whereas PP1y occupancy peaks at the 3' end. b Schematic of the MYC gene, indicating positions of primer pairs used in ChIP-qPCR analysis (top). Inhibition of Cdk9 with NVP-2 (250 nM, 1 hr) does not affect PP4C occupancy on transcribed chromatin. c ChIP-qPCR analysis of PP4R2 and PP4R2-pThr173 on the MYC gene upon siRNA-mediated depletion of PP4R2 reveals chromatin occupancy of PP4R2 and the Thr173-phosphorylated form is reduced, validating specificity of the anti-pThr173 antibody. d ChIP-qPCR analysis of Pol II on the MYC gene upon siRNA-mediated depletion of PP4R2. e ChIP-qPCR analysis of Pol II on the GAPDH gene upon siRNA-mediated depletion of PP4R2. f ChIP-gPCR analysis of Pol II on the CDKN1A gene upon siRNA-mediated depletion of PP4R2. Individual data points from two independent biological replicates (n = 2) are shown in the plots (**a**-**f**). Source data are provided as a Source Data file.

Supplementary Table 1. Primers for Site-Directed Mutagenesis

Sequence	Key
CTGATCCGGGGAGCCATAGGCGCAAAGCCAC	SUPT5H_S666A_F
GTGGCTTTGCGCCTATGGCTCCCCGGATCAG	SUPT5H_S666A_R
GCTGATCCGGGGGATCCATAGGCGCAAAGCCACC	SUPT5H_S666D_F
GGTGGCTTTGCGCCTATGGATCCCCGGATCAGC	SUPT5H_S666D_R
TCATGCAGGGGCGCCTGTGAGCCGTAG	SUPT5H_T806A-F
CTACGGCTCACAGGCGCCCCTGCATGA	SUPT5H_T806A-R
CTGCCATCATGCAGGGGATCCTGTGAGCCGTAGTGTG	SUPT5H_T806D-F
CACACTACGGCTCACAGGATCCCCTGCATGATGGCAG	SUPT5H_T806D-R
GCCATACATGGGCGCCTGGGAGCCATACA	SUPT5H_T775A-F
TGTATGGCTCCCAGGCGCCCATGTATGGC	SUPT5H_T775A-R
TCCTGGAGGGGTGCCTGTGAGCCGTAC	SUPT5H_T791A-F
GTACGGCTCACAGGCACCCCTCCAGGA	SUPT5H_T791A-R
CCGTAGTGTGGGGGCGCGGCTACCATCC	SUPT5H_T799A-F
GGATGGTAGCCGCGCCCCACACTACGG	SUPT5H_T799A-R
TCTGGGCAGGAGCGCGGCTGCCATC	SUPT5H_T814A-F
GATGGCAGCCGCGCTCCTGCCCAGA	SUPT5H_T814A-R
CCTTGGAGGCGCTACAGGTCTCGTGGCATTTG	PPP1CC_T311A_F
CAAATGCCACGAGACCTGTAGCGCCTCCAAGG	PPP1CC_T311A_R
	Sequence CTGATCCGGGGAGCCATAGGCGCAAAGCCAC GTGGCTTTGCGCCTATGGCTCCCCGGATCAG GCTGATCCGGGGATCCATAGGCGCAAAGCCACC GGTGGCTTTGCGCCTATGGATCCCCGGATCAGC TCATGCAGGGGGCGCCTGTGAGCCGTAG CTACGGCTCACAGGCGCCCTGCATGA CTGCCATCATGCAGGGGATCCTGTGAGCCGTAGTGTG CACACTACGGCTCACAGGATCCCCTGCATGATGGCAG GCCATACATGGGCGCCTGGGAGCCATACA TGTATGGCTCCCAGGCGCCCATGTATGGC TCCTGGAAGGGGTGCCTGTGAGCCGTAC GTACGGCTCACAGGCACCCTCCAGGA CCGTAGTGTGGGGCGCGCGCACCTCC GGATGGTAGCCGCGCCCACATCC GGATGGTAGCCGCGCCCACACTACGG TCTGGGCAGGAGCGCGCCCCACACTACGG TCTGGGCAGGAGCGCGCCCCACATC GATGGCAGCGCGCCCCCACACTACGG CCTTGGAGGCGCCCCCCCACACTACGG CCTTGGAGGCGCTACCAGGCCCCCACACTC GATGGCAGCCGCGCTCCTGCCCAGA CCTTGGAGGCGCTACAGGTCTCGTGGCATTTG CAAATGCCACGAGACCTGTAGCGCCTCCAAGG



Fig. 1b



Fig. 1c



Fig. 1d



Fig 1f



Fig. 6a



Fig. 6d



Fig. 7d





Supp. Fig. 1e









M1: T806A; M2: T775A/T791A/T799A/T814A M3: T806D; M4: T775A/T806A; M5: T791A/T806A

Supp. Fig. 5a



Supp. Fig. 5b





Supp. Fig. 8a

