Yu et al. Supplementary Figure 1



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Fig. S1. Lyz1 deficiency reduces inflammation during experimental colitis, Related to Figure 1.

(A) 15 IBD pathology cases of indicated anatomic region and 6 non-IBD colon tissues were stained for lysozyme to identify lysozyme-positive metaplastic Paneth cells. (B) qPCR analysis of Lyz1 mRNA in WT and $Lyz1^{-/-}$ ileum and colon (N=3-5 mice for each genotype). (C) Representative immunostaining of lysozyme in the ileum and colon of WT and $Lyz1^{-/-}$ mice (N=3 mice per genotype). (D) Differential expression of genes in WT and $Lyz1^{-/-}$ ileum (N=4 mice per genotype). (E) Volcano plot of differentially expressed genes in WT and $Lyz1^{-/-}$ ileum. Blue color: genes with fold change < -2 or > 2; p-value < 0.05 (N= 4 mice per genotype). (F) Immunostaining of Mmp7 in WT and $Lyz1^{-/--}$ ileum (N=3 mice per genotype). (G) GSEA of RNA-seq data revealed an increase in the pathways mentioned in WT ileum. Numbers above the bar graphs represent the number of genes from the WT dataset included within the GSEA gene set (N=4 mice). (H) Gene ontology analysis of RNA-seq data revealed gene sets upregulated in $Lyz1^{-/-}$ ileum. Numbers above the bar graphs represent the number of genes from the $Lyz1^{-/-}$ dataset included within the GSEA gene set (N=4 mice). (I, J) Real-time PCR analysis of Lyz1 (I) and Lyz2 (J) displayed induction of these genes in inflamed colon tissues of WT mice. In KO mice, only Lyz2 was induced in DSS-treated WT and $Lyz1^{-/-}$ mouse colons (N=3-6 mice per genotype). (K) Real-time PCR analysis of Areg in DSS-treated $Lyz1^{-/-}$ colons (N=3 mice per genotype). All bar graphs display mean \pm SEM.

Yu et al. Supplementary Figure 2



p=0.2248



100 µm

Tslp

p=0.65

LY21.

1

N





2.0-

J



Fig. S2. *Lyz1^{-/-}* intestines have increased goblet and tuft cells, Related to Figure 2.

(A) GSEA analysis of RNA-seq data (N=4 mice per genotype) comparing enterocyte gene signature in WT and $Lyz1^{-/-}$ illeum. (B) Immunostaining of Alcian blue and Dclk1 in WT and $Lyz1^{-/-}$ intestines (N=5 mice per genotype; 3 independent experiments). (C, D) Transmission electron microscopy of a cell with granules characteristic of mixed Paneth and goblet cell morphologies at the crypt base in the $Lyz1^{-/-}$ illeum (C) when compared to the Paneth cell observed at the crypt base of WT illeum(D) (N=3 mice per genotype). (E-H) Real-time PCR analysis of genes associated with type 2 (E), type 1 (F), type 3 (G), or regulatory (H) immune response in WT and $Lyz1^{-/-}$ mice (N=3-6 mice per genotype). (I) Tuft cell numbers (counted from 50 villi per field of vision per mouse) in WT or $Lyz1^{-/-}$ mice treated with neutralizing anti-IL-13 antibody, anti-CD90.2, or isotype control (N=2 for each condition per genotype). (J) Representative Alcian blue (goblet) and DCLK1 (tuft) staining in $Lyz1^{-/-}$ and $Lyz1^{-/-}$ mice (N=3 mice per genotype). Data is representative of 50-80 villi per section per mouse per genotype. (K) Alcian blue staining of WT or $Lyz1^{-/-}$ bone marrow chimeras with hematopoietic cells from Stat6^{+/+} or Stat6^{-/-} donors (N=4 mice per genotype per treatment; 2 independent experiments). All bar graphs display mean \pm SEM.

Yu et al. Supplementary Figure 3



UMAP_1

Gene Ontology

(Upregulated in WT Cluster 16)

E Gene Ontology (Upregulated in *Lyz1^{-/-}* Cluster 16)



F

Fig. S3. scRNAseq reveals immune-activated ILC2 in Lyz1-/- ileal lamina propria, Related to Figure 3

(A) Unsupervised separate clustering (t-SNE plot) of 8,011 WT LP and 6,733 $Lyz1^{+/-}$ LP cells identified 23 and 19 distinct clusters, respectively. (**B-D**) Combined clustering of WT and $Lyz1^{+/-}$ ileal LP cells. (**B**) UMAP of 16,529 cells colored by relative expression of Cd4 confirmed ILC2 populations as clusters 1 and 16. (**C**) Heatmap shows differentially expressed genes between WT and $Lyz1^{+/-}$ ILC2 population in cluster 16. (**D**) WT and $Lyz1^{+/-}$ ILC2s were partitioned and clustered. UMAP colored by relative expression of Il17rb in the 9 sub-clusters. (**E**) Bar graphs exhibited the fold enrichment and p-value for Gene ontology (GO) pathways upregulated in $Lyz1^{+/-}$ cluster 16 compared to WT cluster 16. (**F**) Bar graphs exhibited the fold enrichment and p-value for Gene ontology athways upregulated in WT cluster 16 compared to $Lyz1^{+/-}$ cluster 16. (**G**) Representative dot blots showed the cytokine expression profile of *in vitro* cultured WT and $Lyz1^{+/-}$ MLN cells at steady state. Intestinal MLN cells were isolated from separately housed WT and $Lyz1^{+/-}$ MLN cells cultured *in vitro* quantified and represented as bar graphs. (**I**) Quantification of pStat6 and pStat3 levels in WT and $Lyz1^{+/-}$ MLN cell lysates based on western blot analysis. Intestinal MLN cells were isolated from separately housed WT and $Lyz1^{+/-}$ MLN cell lysates based on western blot analysis. Intestinal MLN cells were isolated from separately housed WT and $Lyz1^{+/-}$ MLN cell lysates based on western blot analysis. Intestinal MLN cells were isolated from separately housed WT and $Lyz1^{+/-}$ MLN cell lysates based on western blot analysis. Intestinal MLN cells were isolated from separately housed WT and $Lyz1^{+/-}$ MLN cell lysates based on western blot analysis. Intestinal MLN cells were isolated from separately housed WT and $Lyz1^{+/-}$ MLN cell lysates based on western blot analysis. Intestinal MLN cells were isolated from separately housed WT and $Lyz1^{+/-}$ MLN cell lysates based on western blot



Fig. S4. Type 2 immune response is non-epithelial intrinsic in Lyz1-/- mice, Related to Figure 4

(A) qPCR analysis of mRNA expression of Klf4 in ileal WT and $Lyz1^{-/-}$ enteroids treated with or without recombinant IL-13 (2 independent experiments). (B) Real-time PCR analysis of Dclk1 and Trmp5 in ileal WT and $Lyz1^{-/-}$ enteroids treated with or without recombinant IL-13 (2 independent experiments). (C) Real-time PCR analysis of goblet cell markers (Klf4, Spdef, Tff3, and Muc2) and tuft cell marker (Pou2f3) in untreated and Abs-treated WT and $Lyz1^{-/-}$ mice (N=4 mice for each condition). All bar graphs display mean ± SEM.







Fig. S5. Lyz1-deficiency and –sufficiency alters composition of mucolytic composition, Related to Figure 5 (A) Chao1 index (alpha diversity) of ileal luminal microbiota in WT (N=4 mice) and Lyz1^{-/-} (N=6 mice). (B) Weighted UniFrac analysis of ileal luminal microbiota from WT (in blue, N=4 mice) and Lyz1^{-/-} mice (in green, N=6 mice). (C) LDA of fecal bacteria enriched in WT mice (green bars; N=14 mice) and fecal bacteria enriched in Lyz1^{-/-} mice (red bars; N=16 mice). (D) LDA scores computed by LEfSe found Candidatus Arthromitus differentially enriched in ileal mucosal microbiota of WT mice (N=8 mice) compared to $Lyz1^{-/-}$ mice (N=10 mice). (E) Genotyping PCR confirmed TG alleles. The TG animals marked (*) were validated as founders. (F) qPCR analysis of lysozyme mRNA in WT, Lyz1-/-, and TG ileum and colon. (N=4 for each genotype). (G-H) Immunostaining (N>5) (G) and western blot analysis (N=2 for WT and N=3 for TG) (H) of lysozyme in the colon of TG mice. (I) Immunostaining of lysozyme in WT, Lyz1^{-/-}, and TG ileum (N>5 for each genotype). White arrow indicated presence of lysozyme- positive vesicles. (J) 16S rRNA sequencing of WT (N=6 mice) and TG (N=5 mice) analyzed for Chao1 richness showed no significant difference in alpha-diversity. (K) Body weight change in WT (N=5 mice) and TG (N=10 mice) littermates when treated with 3% DSS and during recovery. Dot represents mouse that died on that day. (L-M) Representative H. & E. images of DSS colons (L) and colitis activities (M) were scored from remaining 4 WT mice and 8 TG mice. (N-P) Phylogenetic distribution and change in relative abundance of species between WT (N=4 mice) and $Lyz1^{-/-}$ (N=6 mice) fecal microbiota (N), WT (N=6 mice) and TG fecal microbiota (N=5 mice) (O), and WT (N=4 mice) and $Lyz1^{-/-}$ (N=6 mice) ileal luminal microbiota (P). Bar size indicates the logarithm of relative abundance fold change. Dorea formicigenerans and Ruminococcus gnavus are marked (*) in Fig S5N-P. (Q) LDA scores computed by LEfSe found 6 species differentially enriched in fecal microbiota of WT mice (N=6) and 5 species differentially enriched in $Lyz1^{-/-}$ mice (N=5). (R) Growth curve of D. formicigenerans decreased on addition of 20 µg/ml or 200 µg/ml of lysozyme during the exponential phase, when compared to the control condition. Red arrow indicated the time point of addition of lysozyme. (S) Growth curve of R. gnavus in L-YBHI.4 medium does not change on addition of lysozyme during exponential or lag phases (2-4 independent) experiments). Red arrow indicated the time of addition of lysozyme.(T) Growth curve of R. gnavus in DP2 medium decreased more on addition of lysozyme 0 or 30 minutes after inoculation of culture compared to addition of lysozyme 1 or 4 hours after inoculation (2-4 independent experiments). All bar graphs display mean ± SEM.









Fig. S6. Transfer of Lyz1^{-/-} microbiota to WT GF mice does not transfer protection, Related to Figure 6

(A-C) Volcano plot of differential gene expression in DSS treatment compared to homeostasis in WT-FMT mice (A), $Lyz1^{-/-}$ -FMT mice (B), and TG-FMT mice (C). Genes in red- fold change > 1; p-value < 0.05. Genes in green- fold change < -1; p-value < 0.05. N=4 mice per group at steady state and N= 5 mice per group when treated with DSS. (D-E) Immunostaining of phosphorylated Stat3 (D) and phosphorylated Stat6 (E) in WT-FMT, $Lyz1^{-/-}$ -FMT, and TG-FMT DSS-treated colon tissues (N= 5 mice per group).(F) KEGG Pathway analysis showed distinct differences in pathways between WT-FMT and TG-FMT, in response to DSS-induced colitis.

Yu et al. Supplementary Figure 7







Genes Downregulated in Lyz1-/- Abx-RG

Fig. S7. Live and lysozyme-processed *R. gnavus* differentially mediates inflammatory tone, Related to Figure 7 (A) Immunostaining of Alcian blue in the small intestinal sections of $Lyz1^{-/-}$ mice with varying abundance of R. gnavus. (B) Representative dot blots showed the cytokine expression profile of WT and Lyz1-/- MLNs stimulated by R. gnavus supernatant, lysozyme-treated R. gnavus supernatant, heat-killed R. gnavus supernatant, and lysozyme- treated LGG supernatant. Intestinal MLN cells were isolated from separately housed WT and Lyz1^{-/-} littermates (N=2 for each genotype) (C-F) Radar plots compared expression levels of cytokines in WT (C) and Lyz1-/- MLNs (D) stimulated with lysozyme-treated R. gnavus sup and heat-killed R. gnavus sup and in WT (E) and Lyz1^{-/-} (F) MLNs treated with lysozyme-treated *R. gnavus* sup and lysozyme-treated LGG sup. Data is representative of 2 independent experiments. (G-H) Quantification of pSTAT6 (G) and pSTAT3 (H) levels in WT (red bars) and Lyz1^{-/-} (blue bars) MLN cells based on western blot analysis. The values are normalized to WT buffer. (I, J) Semi-quantitative PCR using species-specific primer sets confirmed successful colonization of Abx-treated mice with *R. gnavus* (I) or *LGG* (J). (K, L) Real-time PCR analysis of II25 and II13 in the ileum of SPF and R. gnavus colonized WT and Lyz1^{-/-} mice. (N=3 mice per genotype per group). (M-O) Immunostaining of Alcian blue (goblet cells) and Dclk1 (tuft cells) (M) and the quantification of goblet cell (N) and tuft cell (O) numbers in antibiotics-treated, R. gnavus colonized, and LGG-colonized WT and Lyz1^{-/-} mice (N=3) mice per genotype for Abx and R. gnavus group and N=2 mice per genotype for LGG group). Data representative of 90-100 villi per section per mouse per genotype. (P) GSEA of RNA-seq data comparing Th2 signature in R. gnavus colonized $Lyz1^{-/-}$ mice and R. gnavus colonized WT mice (N=3 mice per genotype). All bar graphs display mean \pm SEM.

Name	WT=4 vs. <i>Lyz1^{-/-}</i> =8	WT=6 vs. TG=5
Akkermansia muciniphila	UP in <i>Lyz1^{-/-}</i>	Down in TG
Anaerotruncus colihominis	UP in <i>Lyz1^{-/-}</i>	Down in TG
Clostridium sp	UP in <i>Lyz1^{-/-}</i>	Down in TG
Dorea formicigenerans	UP in <i>Lyz1^{-/-}</i>	Down in TG
Eubacterium plexicaudatum	UP in <i>Lyz1^{-/-}</i>	Down in TG
Lactobacillus reuteri	UP in <i>Lyz1^{-/-}</i>	Down in TG
Lactobacillus vaginalis	UP in <i>Lyz1^{-/-}</i>	Down in TG
Marvinbryantia formatexigens	UP in <i>Lyz1^{-/-}</i>	Down in TG
Moryella indoligenes	UP in <i>Lyz1^{-/-}</i>	Down in TG
Mucispirillum schaedleri	UP in <i>Lyz1^{-/-}</i>	Down in TG
Oscillibacter sp.	UP in <i>Lyz1^{-/-}</i>	Down in TG
Oscillibacter valericigenes	UP in <i>Lyz1^{-/-}</i>	Down in TG
Ruminococcus gnavus	UP in <i>Lyz1^{-/-}</i>	Down in TG
Acetatifactor muris	UP in <i>Lyz1^{-/-}</i>	UP in TG
Bifidobacterium pseudolongum	UP in <i>Lyz1^{-/-}</i>	UP in TG
Parvibacter caecicola	UP in <i>Lyz1^{-/-}</i>	UP in TG
Tyzzerella propionicum	UP in <i>Lyz1^{-/-}</i>	UP in TG
Clostridium viride	UP in <i>Lyz1^{-/-}</i>	UP in TG

Alistipes finegoldii	UP in <i>Lyz1^{-/-}</i>	UNCHANGED in TG
Anaerostipes butyraticus	UP in <i>Lyz1^{-/-}</i>	UNCHANGED in TG
Anaerostipes sp	UP in <i>Lyz1^{-/-}</i>	UNCHANGED in TG
Bacteroides sp	UP in <i>Lyz1^{-/-}</i>	UNCHANGED in TG
Bacteroides uniformis	UP in <i>Lyz1^{-/-}</i>	UNCHANGED in TG
Coprococcus eutactus	UP in <i>Lyz1^{-/-}</i>	UNCHANGED in TG
Eubacterium hadrum	UP in <i>Lyz1^{-/-}</i>	UNCHANGED in TG
Eubacterium ramulus	UP in <i>Lyz1^{-/-}</i>	UNCHANGED in TG
Faecalibacterium prausnitzii	UP in <i>Lyz1^{-/-}</i>	UNCHANGED in TG
Helicobacter cf	UP in <i>Lyz1^{-/-}</i>	UNCHANGED in TG
Helicobacter equorum	UP in <i>Lyz1^{-/-}</i>	UNCHANGED in TG
Helicobacter ganmani	UP in <i>Lyz1^{-/-}</i>	UNCHANGED in TG
Helicobacter sp	UP in <i>Lyz1^{-/-}</i>	UNCHANGED in TG
Lachnoanaerobaculum umeasense	UP in <i>Lyz1^{-/-}</i>	UNCHANGED in TG
Oscillospira guilliermondii	UP in <i>Lyz1^{-/-}</i>	UNCHANGED in TG
Ruminococccus gauvreauii	UP in <i>Lyz1^{-/-}</i>	UNCHANGED in TG
Ruminococcus faecis	UP in <i>Lyz1^{-/-}</i>	UNCHANGED in TG
Stomatobaculum longum	UP in <i>Lyz1^{-/-}</i>	UNCHANGED in TG

Table. S1. Thirty-six species increased in *Lyz1^{-/-}* mouse fecal microbiota. WT (N=4) vs. *Lyz1^{-/-}* (N=8); WT (N=6) vs.TG (N=5), Related to Figure 5.

Ranked by			Lyz1-/-	FC	WT	Lyz1 ^{-/-}	FC (by
Lyz1-/- %	Name	WT (%)	(%)	(by %)	(Counts)	(Counts)	Counts)
1	Ruminococcus gnavus	3.66	6.01	1.64	81498	284916	3.50
2	Clostridium sp.	1.00	1.43	1.43	22233	67840	3.05
3	Lactobacillus vaginalis Dorea	0.11	0.18	1.67	2400	8539	3.56
4	formicigenerans Mucispirillum	0.02	0.14	8.94	351	6680	19.03
5	schaedleri Akkermansia	0.03	0.12	4.14	644	5674	8.81
6	muciniphila	0.05	0.06	1.12	1206	2864	2.37
7	Lactobacillus reuteri	0.03	0.06	1.72	736	2689	3.65
8	Moryella indoligenes	0.02	0.02	1.16	459	1129	2.46
9	Oscillibacter sp.	0.00	0.02	4.73	104	1047	10.07
	Eubacterium						
10	plexicaudatum Oscillibacter	0.00	0.02	45.00	10	958	95.80
11	valericigenes Anaerotruncus	0.00	0.01	3.62	71	547	7.70
12	colihominis Marvinbryantia	0.00	0.00	4.70	22	220	10.00
13	formatexigens	0.00	0.00	NA	0	62	NA

Ranked by WT				FC (by	WT	TG	FC (by
%	Name	WT (%)	TG (%)	%)	(Counts)	(Counts)	Counts)
1	Ruminococcus gnavus	1.72	1.19	0.69	81278	38622	0.48
2	Clostridium sp. Mucispirillum	0.83	0.53	0.64	39367	17278	0.44
3	schaedleri Dorea	0.18	0.02	0.12	8743	733	0.08
4	formicigenerans	0.04	0.01	0.25	1898	321	0.17
5	Lactobacillus vaginalis Oscillibacter	0.03	0.03	0.81	1500	833	0.56
6	valericigenes Akkermansia	0.03	0.01	0.24	1346	219	0.16
7	muciniphila	0.02	0.01	0.22	1128	172	0.15
8	Oscillibacter sp.	0.01	0.01	0.57	496	193	0.39
9	Moryella indoligenes	0.01	0.01	0.74	340	174	0.51
10	Lactobacillus reuteri Anaerotruncus	0.01	0.00	0.48	326	107	0.33
11	colihominis Eubacterium	0.01	0.00	0.46	281	88	0.31
12	plexicaudatum Marvinbryantia	0.00	0.00	0.00	140	0	0.00
13	formatexigens	0.00	0.00	NA	13	0	NA

Table. S2. Thirteen species that increased in *Lyz1^{-/-}* and decreased in TG mice were ranked by their abundance (%) in *Lyz1^{-/-}* fecal microbiota. WT (N=4) vs. *Lyz1^{-/-}* (N=8); WT (N=6) vs. TG (N=5), Related to Figure 5.

			Fecal microbiota					lleum lur	ninal mic	robiota
Ranked by Fecal KO %	Species	WT (%)	Lyz1-/- (%)	FC	WT (%)	TG (%)	FC	WT (%)	Lyz1-/- (%)	FC
1	R. gnavus	3.66	6.01	1.64	1.72	1.19	0.69	0.00	0.39	>39
2	Clostridium sp.	1.00	1.43	1.43	0.83	0.53	0.64	0.031	0.296	9.68
3	L. vaginalis	0.11	0.18	1.67	0.032	0.026	0.81	1.63	0.61	0.38
	D.									
4	formicigenerans	0.02	0.14	8.94	0.04	0.01	0.25	0.004	0.05	14.34
5	M. schaedleri	0.03	0.12	4.14	0.18	0.02	0.12	0.00	1.14	>114
6	A. muciniphila	0.05	0.06	1.12	0.02	0.01	0.22	0.0003	0.0008	2.71
7	L. reuteri	0.03	0.06	1.72	0.01	0.003	0.48	0.40	0.23	0.57
8	M. indoligenes	0.02	0.02	1.16	0.007	0.005	0.74	0.0012	0.0010	0.86
9	Oscillibacter sp.	0.005	0.02	4.73	0.010	0.006	0.57	0.00	0.00	N/D

Table. S3. Bacteria increased in Lyz1^{-/-} and decreased in TG mice, Related to Figure 5.

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Name	Fecal	lleal luminal
Acetatifactor muris	Up in <i>Lyz1^{-/-}</i>	Up in <i>Lyz1^{-/-}</i>
Bifidobacterium pseudolongum	Up in <i>Lyz1^{-/-}</i>	Up in <i>Lyz1^{-/-}</i>
Clostridium viride	Up in <i>Lyz1^{-/-}</i>	Up in <i>Lyz1^{-/-}</i>
Coprococcus eutactus	Up in <i>Lyz1^{-/-}</i>	Up in <i>Lyz1^{-/-}</i>
Dorea formicigenerans	Up in <i>Lyz1^{-/-}</i>	Up in <i>Lyz1^{-/-}</i>
Eubacterium hadrum	Up in <i>Lyz1^{-/-}</i>	Up in <i>Lyz1^{-/-}</i>
Faecalibacterium prausnitzii	Up in <i>Lyz1^{-/-}</i>	Up in <i>Lyz1^{-/-}</i>
Helicobacter cf	Up in <i>Lyz1^{-/-}</i>	Up in <i>Lyz1^{-/-}</i>
Helicobacter ganmani	Up in <i>Lyz1^{-/-}</i>	Up in <i>Lyz1^{-/-}</i>
Mucispirillum schaedleri	Up in <i>Lyz1^{-/-}</i>	Up in <i>Lyz1^{-/-}</i>
Oscillibacter valericigenes	Up in <i>Lyz1^{-/-}</i>	Up in <i>Lyz1^{-/-}</i>
Ruminococcus gnavus	Up in <i>Lyz1^{-/-}</i>	Up in <i>Lyz1^{-/-}</i>
Tyzzerella propionicum	Up in <i>Lyz1^{-/-}</i>	Up in <i>Lyz1^{-/-}</i>
Clostridium sp	Up in <i>Lyz1^{-/-}</i>	Up in <i>Lyz1^{-/-}</i>
Akkermansia muciniphila	Up in <i>Lyz1^{-/-}</i>	Unchanged
Eubacterium ramulus	Up in <i>Lyz1^{-/-}</i>	Unchanged
Helicobacter equorum	Up in <i>Lyz1^{-/-}</i>	Unchanged
Helicobacter sp	Up in <i>Lyz1^{-/-}</i>	Unchanged
Marvinbryantia formatexigens	Up in <i>Lyz1^{-/-}</i>	Unchanged
Oscillibacter sp.	Up in <i>Lyz1^{-/-}</i>	Unchanged
Ruminococccus gauvreauii	Up in <i>Lyz1^{-/-}</i>	Unchanged
Bacteroides uniformis	Up in <i>Lyz1^{-/-}</i>	Unchanged
Lactobacillus reuteri	Up in <i>Lyz1^{-/-}</i>	Unchanged
Lactobacillus vaginalis	Up in <i>Lyz1^{-/-}</i>	Unchanged
Moryella indoligenes	Up in <i>Lyz1^{-/-}</i>	Unchanged
Parvibacter caecicola	Up in <i>Lyz1^{-/-}</i>	Unchanged
Alistipes finegoldii	Up in <i>Lyz1^{-/-}</i>	N/D
Anaerostipes butyraticus	Up in <i>Lyz1^{-/-}</i>	N/D
Anaerostipes sp	Up in <i>Lyz1^{-/-}</i>	N/D
Anaerotruncus colihominis	Up in <i>Lyz1^{-/-}</i>	N/D
Bacteroides sp	Up in <i>Lyz1^{-/-}</i>	N/D
Eubacterium plexicaudatum	Up in <i>Lyz1^{-/-}</i>	N/D
Lachnoanaerobaculum umeasense	Up in <i>Lyz1^{-/-}</i>	N/D
Oscillospira guilliermondii	Up in <i>Lyz1^{-/-}</i>	N/D
Ruminococcus faecis	Up in <i>Lyz1^{-/-}</i>	N/D
Stomatobaculum longum	Up in <i>Lyz1^{-/-}</i>	N/D

Table. S4. Thirty-six species increased in *Lyz1^{-/-}* mouse fecal microbiota. WT (N=4) vs. *Lyz1^{-/-}* (N=8) Fecal microbiota; WT (N=4) vs. *Lyz1^{-/-}* (N=6) ileal luminal content, Related to Figure 5.

Ranked by				FC (by	WT	TG	FC (by
WT %	Species	WT (%)	TG (%)	%)	(Counts)	(Counts)	Counts)
	Lactobacillus						
1	murinus	0.09	0.43	5.05	4079	14144	3.47
	Candidatus						
2	Arthromitus	0.02	0.02	1.19	873	716	0.82
	Lactobacillus						
3	animalis	0.01	0.02	1.64	587	663	1.13
	Clostridium						
4	perfringens	0.00	0.01	8.16	61	342	5.61
	Enterorhabdus						
5	caecimuris	0.00	0.01	4.72	53	172	3.25
6	Roseburia faecis	0.00	0.00	7.22	24	119	4.96
	Enterorhabdus						
7	mucosicola	0.00	0.00	1.19	65	53	0.82
Ranked by				FC (by	WT	Lyz1-/-	FC (by
TG %	Species	WT (%)	Lyz1-/- (%)	%)	(Counts)	(Counts)	Counts)
1	Roseburia faecis	0.17	0.07	0.41	3747	3253	0.87
	Lactobacillus						
2	murinus	0.13	0.05	0.37	3004	2361	0.79
	Clostridium						
3	perfringens	0.10	0.02	0.17	2158	777	0.36

4	caecimuris	0.04	0.02	0.60	881	1126	1.28
5	Candidatus	0.04	0.00	ΝΙΛ	3646	63	0 02
5	Lactobacillus						0.02
6	animalis Enterorhabdus	0.03	0.00	0.15	649	208	0.32
7	mucosicola	0.00	0.00	0.00	13	0	0.00

Enterorhabdus

Table. S5. Seven species increased in TG and decreased in *Lyz1^{-/-}* mice. WT (N=6) vs. TG (N=5); WT (N=4) vs. *Lyz1^{-/-}* (N=8), Related to Figure 5.

	L-YHBHI.4	L-YHBHI.4 + mucin	DP2
	Concentration (g/L)	Concentration (g/L)	Concentration (g/L)
Brain Heart Infusion	37	37	-
Yeast Extract	5	5	-
Maltose	0,50	0,50	-
Cellobiose	0,50	0,50	-
Starch	0,50	0,50	-
Mucin type II	-	1	-
L-Cysteine-HCI	0,5	0,5	0.5
Hemin 0.5%	0.005	0.005	0.005
Resazurin solution 0.1%	0.004	0.004	0.004
Meat extract			5
Sodium Pyruvate			1
Sodium Succinate			5
Bactopeptone			5
CaCl ₂ , 2H ₂ O			0.01
KH ₂ PO ₄			0.04
K ₂ HPO ₄			0.04
NaCl			0.08
ZnSO ₄ , 7H ₂ 0			0.02
NaHCO ₃			0.4
Vitamins			
-K1			1.10 ⁻⁶
-B1			5.10 ⁻⁵
-B2			5.10 ⁻⁵
-B8			2.10 ⁻⁵

Table. S6. Detailed composition of the bacteria culture media used *Ruminococcus gnavus*, Related to STAR Methods.

Gene name	Forward primer	Reverse primer
Dorea formicigenerans	GCAGCTAACGCAATAAGCAG	CTTCCATTACGAAGCGGTC
Ruminococcus gnavus	GGACTGCATTTGGAACTGTCAG	AACGTCAGTCATCGTCCAGAAAG
Bacterial 16s rRNA	ACTACGTGCCAGCAGCC	GGACTACCAGGGTATCTAATCC
Tritrichomonas muris	GCTTTTGCAAGCTAGGTCCC	TTTCTGATGGGGCGTACCAC
Mouse Lyz1	ATGGCTACCGTGGTGTCAAG	CGGTCTCCACGGTTGTAGTT
Mouse Mucin 2 (Muc2)	TGTGTGGGACCTGACAATGT	CACACAATGCCACTTCCACC
Mouse Klf4	CACACCTGCGAACTCACAC	GGTAGTGCCTGGTCAGTTCA
Mouse Spdef	CTATGGCCGCTTCATCCGCT	GTTCTTGCGCACACCCCAC
Mouse II4	AGTGTTCTCATGGAGCTGCA	CAGTGATGTGGACTTGGACTCA
Mouse II13	CGGTGCCAAGATCTGTGTCT	ACACTCCATACCATGCTGCC
Mouse II17a	GTTCCACGTCACCCTGGACT	CTTTCCCTCCGCATTGACAC
Mouse II17f	ACGTGAATTCCAGAACCGCT	TGATGCAGCCTGAGTGTCTG
Mouse II25	ACACCCACCACGCAGAAT	ACCCGATTCAAGTCCCTGTC
Mouse II33	TGGGAAGAAGCTGATGGTGA	CGAGACGTCACCCCTTTGAA
Mouse Tslp	CCTTCACTCCCCGACAAAAC	TGCCATTTCCTGAGTACCGT
Mouse Tnfa	ACAAGCCTGTAGCCCACGTC	CTTTGAGATCCATGCCGTTG
Mouse Pou2f3	ACCCATCTACAACTCCCGGC	CCAGGGGAACAGGATGACGT
Mouse Gata3	GAGGAGGAACGCTAATGGGG	CGGGTCTGGATGCCTTCTTT
Mouse Tbx21 (Tbx)	GGCTTCCAACAATGTGACCC	TGTTAGAAGCACTGCAGGCA
Mouse Rorc (Rorγt)	GTGTGCTGTCCTGGGCTAC	CCCAGATGACTTGTCCCCAC
Mouse Foxp3	TGGAAAAGGAGAAGCTGGGAG	AGTACTGGTGGCTACGATGC
Mouse Tgfb1	CGGAGAGCCCTGGATACCA	CTTCCAACCCAGGTCCTTCC
Mouse Amphiregulin (Areg)	CGGCATCGTTATCACAGTGC	TGCACAGTCCCGTTTTCTTG
Mouse Hprt	TCCCTGGTTAAGCAGTACAGC	TCCAACAAAGTCTGGCCTGT

 Table. S7. Primer list for quantitative RT-PCR, Related to STAR Methods.