Supplementary Information

Supplementary Figure 1. CD31^{low} cells in human angiosarcomas and in angiosarcoma-derived cell lines. (A) Representative CD31 immunohistochemical stainings in four human angiosarcomas cases demonstrating variable numbers of scattered cells with low CD31 expression (dotted lines). (B) Representative CD31 FACS analysis of the newly established HAMON angiosarcoma cell line showing a significant population of CD31^{low} cells (highlighted in blue). (C) Quantification of the CD31-gated fraction of all indicated cell lines. (HUVEC, HDMEC, HAMON n=2; EA.hy926 n=3; ISO-HASc.1 n=6; ASM n=7).

Supplementary Figure 2. CD31^{low} angiosarcoma cells are more resistant against doxorubicininduced cell death. (A) Quantitative densitometric analysis of the apoptosis markers presented in Figure 4B. Values are presented as ratio of representative cleaved form versus unprocessed fulllength protein from PARP, caspase-3 and caspase-7 (n=3). (B) Corresponding isotype-PE controls from FACS analysis presented in Figure 4C. (C) Representative microscopic bright field images demonstrating morphologic differences between parental and doxo-surviving ASM cells. Doxosurviving cells had a more elongated, spidle-like morphology and (D) lost their capacity to form vascular-like tubes even after 18 hours incubation.

Supplementary Figure 3. Pharmacological screen in CD31^{high} angiosarcoma cells identified ROS scavengers retrieving colony formation capacity. (A) Western blot analysis of cell death markers showed no difference in spontaneous apoptosis (Caspase-3) and autophagy (Beclin-1 and LC3) between the two CD31 sublines. However, CD31^{low} cells presented marked differences in basal DNA damage level (indicated by γ -H2AX) compared to CD31^{high} cells (B) CD31^{high} cells were treated for 24 hours with indicated compounds and 1x10³/well were seeded in methylcellulose and incubated for 10 days and assayed for formed colonies (Ctrl: n=22, NAC: n=3, Fer-1: n=6, Trolox: n=5, MG: n=2, all others: n=3). (C) Representative picture of CD31^{high} cells/colonies formed after 10 days in methylcellulose prior treated with or without 1 mM NAC treatment for 24 hours. (D) CD31^{low} cell were treated for 24 hours with indicated oxidative oxidative stressors sodium arsenit (AsN). 1x10³/well were seeded in methylcellulose and assayed for formed colonies (n=6). All data are mean ± SEM and were analyzed using unpaired t-test (D) (*p < 0.05; ***p < 0.001).

Supplementary Figure 4. Validation of the CD31-YAP-redox-axis in normal and malignant endothelial cells. (A) Quantification of flow cytometric determined cytoplamic ROS levels via DCF

fluorescence in parental and doxo-suriving ASM cells. (See also Figure 4C and experimental procedures for further details). (B) Immunoblot analysis revealed increased YAP levels and decreased levels of the DNA damage marker γ -H2AX in doxo-surviving cells compared to parental ASM cells. (C) Gating strategy after ISO-HASc.1 AS cells were sorted according their CD31 levels. (D) Immunoblot analysis demonstrated marked differences in CD31 expression. (E) Freshly sorted CD31^{low} ISO-HASc.1 presented significantly enhanced cell growth and (F) colony formation capacity in methylcellulose (n=3) compared to respective CD31^{high} counterparts (magnification 100x). (G) CD31^{low} cells contain significantly lower levels of cytoplamic ROS as determined by flow cytometric DCF fluorescence (n=3). (H) Immunoblot analysis confirmed higher amounts of YAP and suppressed p-AKT (Ser473) and γ -H2AX levels in CD31^{low} cells compared to CD31^{high} cells. (I) Murine endothelial cells isolated from CD31-knockout mice (CD31^{-/-}) present a more spindle-form fibroblastic morphology compared to the cobblestone morphology of CD31^{+/+} endothelial cells. (J) Immunoblot analysis demonstrated an enrichment of YAP in CD31^{-/-} cells and a suppression of p-Akt (Ser473) and γ -H2AX compared to CD31^{+/+} endothelial cells. (K) Representative FACS analysis demonstrates lower basal level of cytoplasmic ROS (determined by FACS-based DCF fluorescence) in CD31^{-/-} cells compared to wildtype CD31^{+/+} cells. All data are mean \pm SEM and were analyzed using unpaired t-test (*p < 0.05).

Supplementary Figure 5. EndoMT transcription factor Slug is enriched in CD31^{low} cells. Western blot analysis demonstrates increased level of Slug in conjunction with suppressed VE-Cadherin levels in CD31^{low} cells compared to CD31^{high} cells.