

Supplementary Materials for

Three-dimensional structure of 22 uncultured ssRNA bacteriophages: Flexibility of the coat protein fold and variations in particle shapes

Jānis Rūmnieks, Ilva Liekniņa, Gints Kalniņš, Mihails Šišovs, Ināra Akopjana, Jānis Bogans, Kaspars Tārs*

*Corresponding author. Email: kaspars@biomed.lu.lv

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This PDF file includes:

Figs. S1 to S5
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References

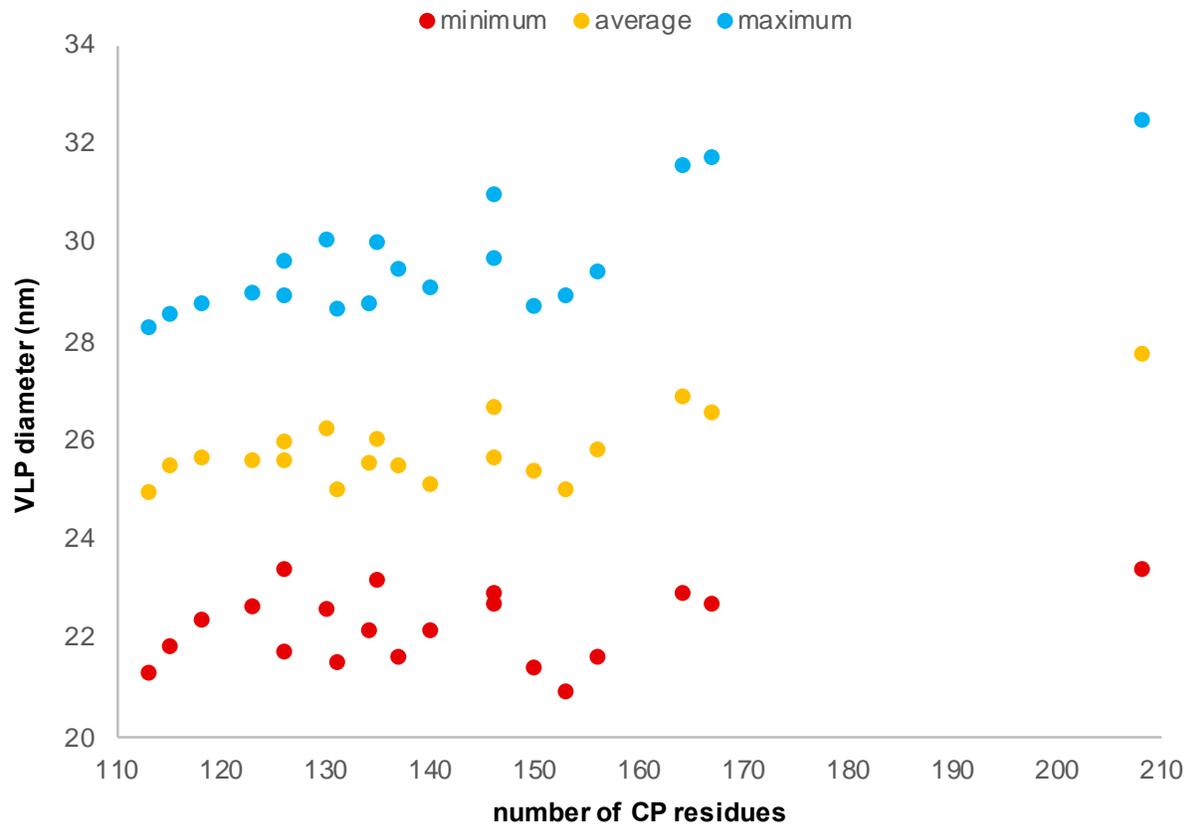


Figure S3. Relationship between coat protein length and particle size in the novel ssRNA phages. Data used for generating the graph are provided in Table S1.

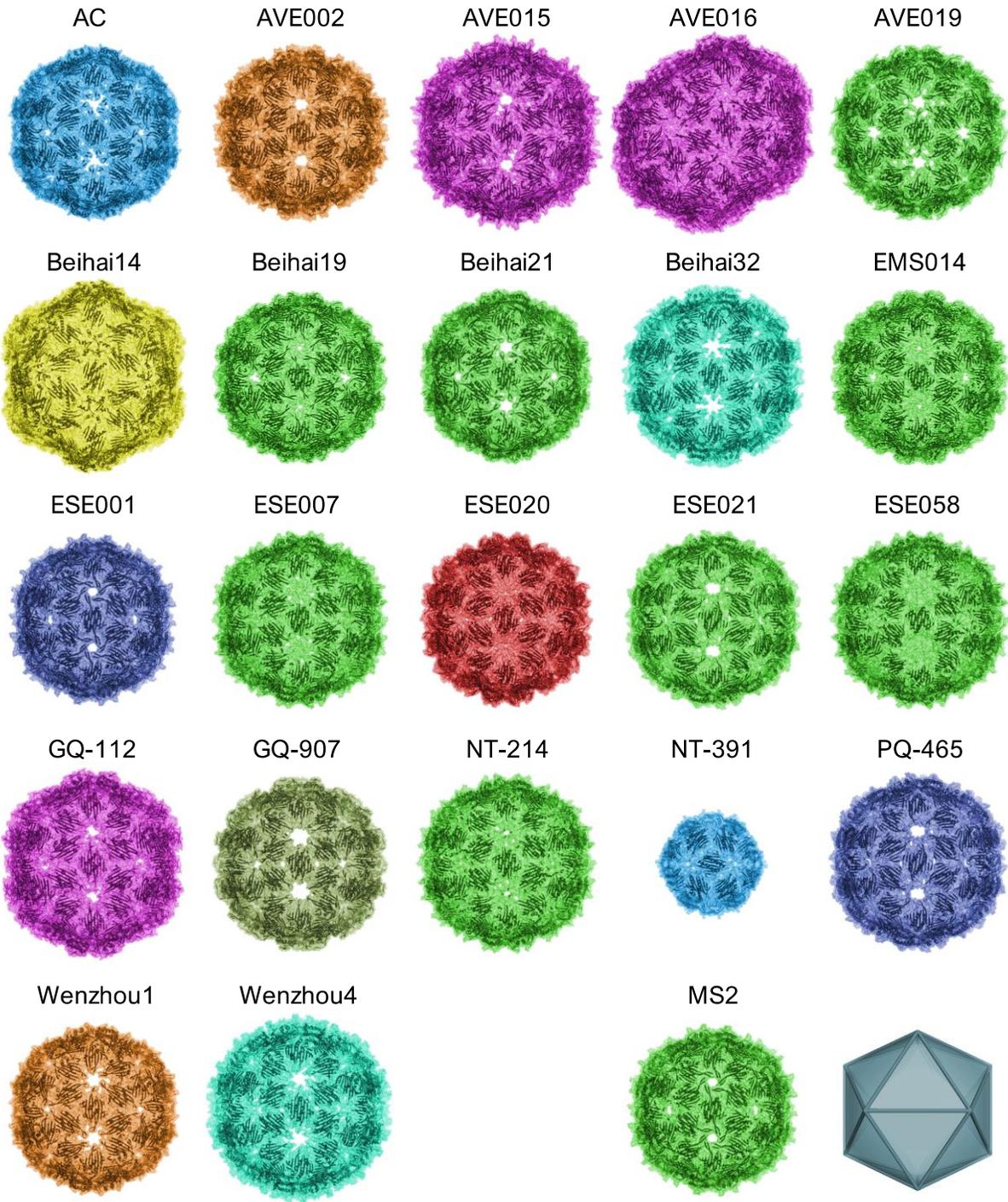
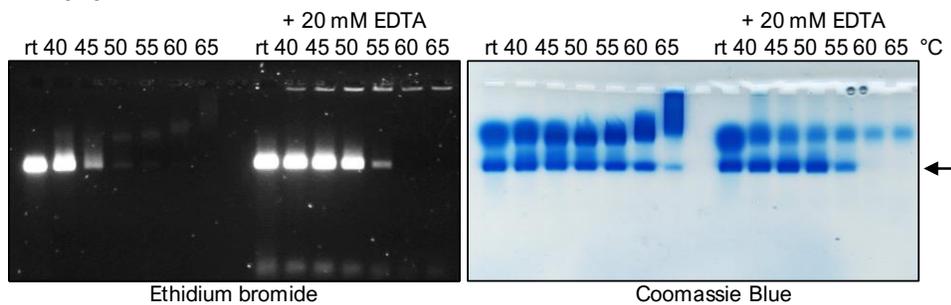
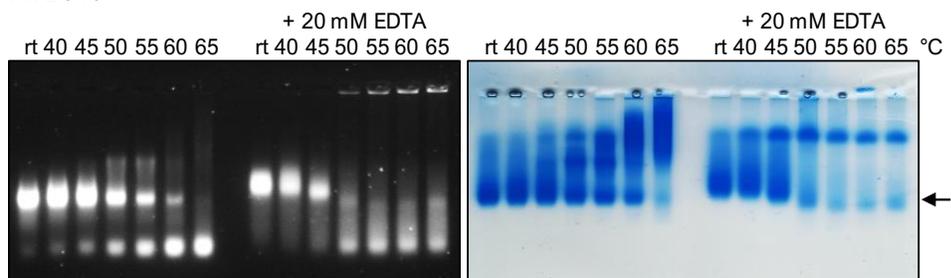


Figure S4. Structure of the novel ssRNA phage VLPs. Coat protein dimers are shown in cartoon representation inside semi-transparent VLP molecular surfaces and are differently colored as per different CP similarity groups. The back sides of the particles are clipped for clarity. All particles are shown on the same scale and in an orientation corresponding to the regular icosahedron on bottom right. Structure of the ssRNA bacteriophage MS2 is included for comparison.

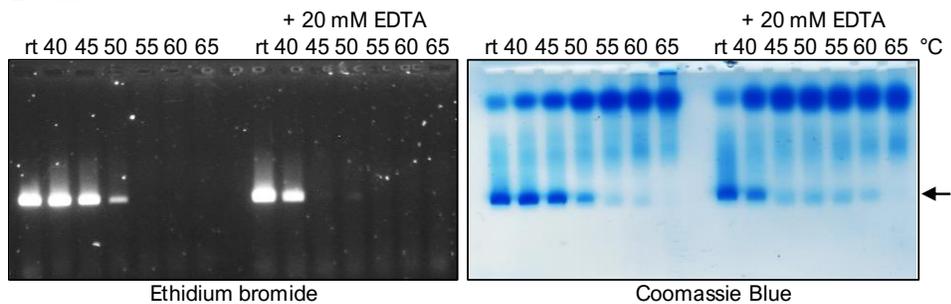
AVE015



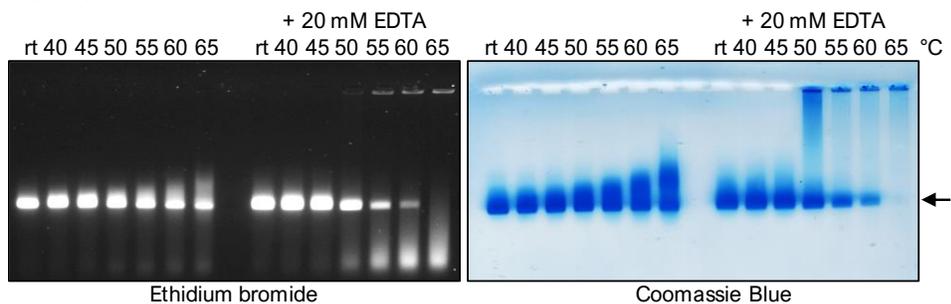
AVE019



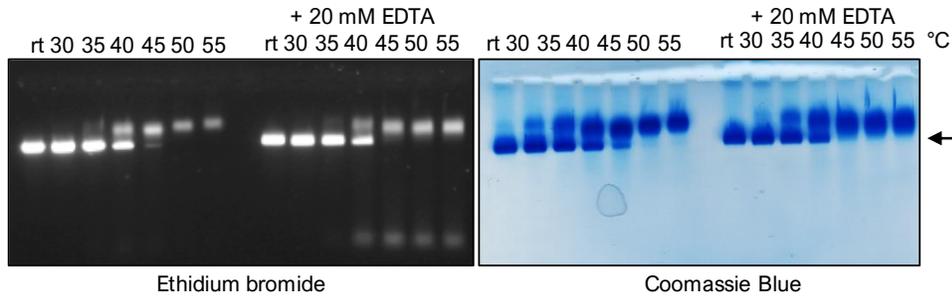
Beihai19



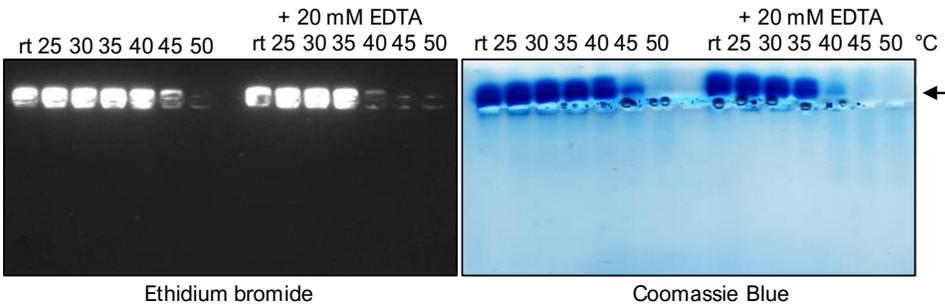
Beihai32



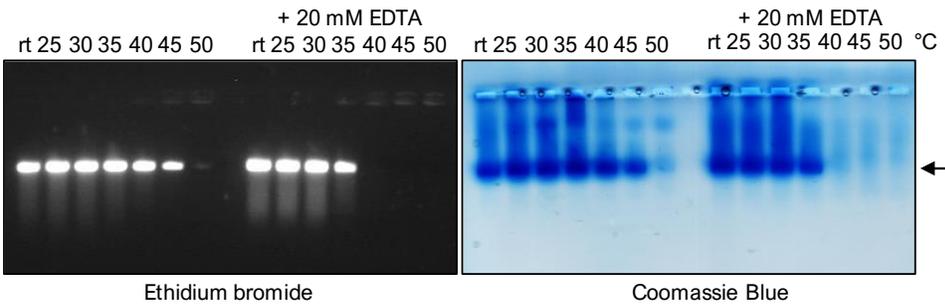
ESE007



ESE021



ESE058



ESE020

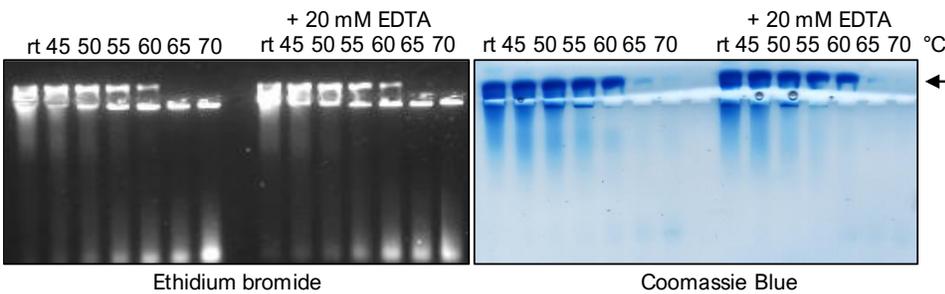


Figure S5. Stabilizing role of divalent metal ions in the novel ssRNA phage particles.

Shown are agarose gels after electrophoresis of heated metal ion-containing virus-like particles in absence and in presence of EDTA. Temperature at which each sample was heated is provided above each track; rt: unheated control at room temperature. The arrow indicates the position of VLPs. ESE020 VLPs were used as a control for particles not containing bound metal ions.

| name | GenBank accession | CP similarity group | CP length | VLP diameter (nm) | | | CP dimer interface area (Å ²) | | |
|----------|-------------------|---------------------|-----------|-------------------|---------|---------|---|--------|--------|
| | | | | minimum | average | maximum | AB | CC | |
| AC | KF616864 | AC | 115 | 21.9 | 25.5 | 28.6 | 1872.7 | 1788.5 | |
| AVE002 | KT462696 | Cb5 | 140 | 22.2 | 25.1 | 29.1 | 2921.7 | 2722.1 | |
| AVE015 | | AVE015 | 167 | 22.7 | 26.6 | 31.8 | 3368.9 | 2689.9 | |
| AVE016 | | AVE015 | 166 | | | | 4371.4 | 4114.0 | |
| AVE019 | KX883502 | MS2 | 123 | 22.7 | 25.6 | 29.0 | 2353.9 | 2203.7 | |
| Beihai14 | | Beihai14 | 208 | 23.4 | 27.8 | 32.5 | 5367.1 | 5293.4 | |
| Beihai19 | | MS2 | 134 | 22.2 | 25.6 | 28.8 | 3093.1 | 3481.1 | |
| Beihai21 | | MS2 | 126 | 21.7 | 25.6 | 29.0 | 2639.0 | 2458.2 | |
| Beihai32 | | Beihai32 | 130 | 22.6 | 26.3 | 30.1 | 1997.6 | 1942.1 | |
| EMS014 | | MS2 | 156 | 21.6 | 25.9 | 29.4 | 3747.3 | 3315.9 | |
| ESE001 | | KT462710 | AP205 | 118 | 22.4 | 25.7 | 28.8 | 2306.3 | 2486.9 |
| ESE007 | | MS2 | 137 | 21.7 | 25.5 | 29.5 | 3971.7 | 3623.3 | |
| ESE020 | | ESE020 | 153 | 20.9 | 25.1 | 29.0 | 2692.3 | 2500.4 | |
| ESE021 | | MS2 | 150 | 21.4 | 25.4 | 28.7 | 4095.0 | 3629.4 | |
| ESE058 | MS2 | 146 | 22.7 | 25.7 | 29.7 | 4669.7 | 4685.6 | | |
| GQ-907 | GALQ01034907 | ESE017 | 131 | 22.9 | 26.9 | 31.6 | 2793.9 | 3026.7 | |
| GQ-112 | GALQ01044112 | AVE015 | 164 | 21.5 | 25.0 | 28.7 | 2634.4 | 2423.0 | |
| NT-214 | NFYT01000214 | MS2 | 135 | 23.2 | 26.0 | 30.0 | 3625.9 | 3468.0 | |
| NT-391 | NFYT01000391 | AC | 123 | 12.1 | 15.2 | 18.3 | 1779.1 | 1778.0 | |
| PQ-465 | PQDQ01001465 | AP205 | 126 | 23.4 | 26.0 | 29.7 | 2582.3 | 2296.4 | |
| Wenzhou1 | KX883612 | Cb5 | 113 | 21.3 | 25.0 | 28.3 | 2342.2 | 2139.1 | |
| Wenzhou4 | KX883621 | Beihai32 | 146 | 22.9 | 26.7 | 31.0 | 1676.8 | 1927.0 | |

Table S1. Properties of the ssRNA phage coat proteins and virus-like particles used in the current study. Accession numbers are provided for the respective genome sequences; entries without an accession number were sourced from Dataset S1 from (12). The VLP diameter was defined by calculating distances to all C α atoms from the particle center and then finding the minimum, average and maximum values. The interface areas for CP dimers were calculated using PISA (51) and were defined as the buried surface between an AB or CC dimer and their four surrounding CP dimers in the particle.

| | AC | AVE002 | AVE015 | AVE016 |
|---------------------------------|--|---|---|--|
| PDB ID | 6YF7 | 6YF9 | 6YFA | 6YFB |
| Crystallization | | | | |
| Conditions | 0.85 M ammonium sulfate | 46% MPD, 0.1 M ammonium nitrate, 0.1 M MES pH 6.5 | 22% MPD, 0.1 M sodium nitrate, 0.1 M sodium acetate pH 4.6 | 30% MPD, 0.2 M sodium citrate, 0.1 M HEPES pH 7.5 |
| Data collection | | | | |
| Date | 2015-11-05 | 2018-05-09 | 2018-05-09 | 2018-01-27 |
| Site | MAX II | BESSY II | BESSY II | BESSY II |
| Beamline | I911-3 | MX 14.1 | MX 14.1 | MX 14.1 |
| Wavelength (Å) | 0.97934 | 0.9184 | 0.9184 | 0.9184 |
| Data reduction | | | | |
| Space group | C 2 | P 21 21 21 | I 2 3 | P 21 |
| Cell parameters | a = 432.29 b = 307.87 c = 679.45 α = 90.00 β = 107.15 γ = 90.00 | a = 278.09 b = 390.21 c = 554.47 α = 90.00 β = 90.00 γ = 90.00 | a = 559.31 b = 559.31 c = 559.31 α = 90.00 β = 90.00 γ = 90.00 | a = 325.47 b = 298.52 c = 417.12 α = 90.00 β = 100.00 γ = 90.00 |
| Total number of observations | 1069421 | 6155935 | 8135712 | 2073077 |
| Number of unique reflections | 591127 | 970502 | 430490 | 855258 |
| Resolution (Å) | 44.26 - 3.40 | 62.21 - 3.20 | 61.77 - 3.30 | 49.32 - 3.59 |
| Highest resolution bin | 3.58 - 3.40 | 3.37 - 3.20 | 3.48 - 3.30 | 3.81 - 3.59 |
| Multiplicity | 1.8 (1.7) | 6.3 (5.9) | 18.9 (18.6) | 2.4 (2.4) |
| Completeness (%) | 51.1 (47.3) | 98.8 (97.0) | 100.0 (100.0) | 93.9 (92.4) |
| R-merge | 0.198 (0.749) | 0.672 (1.727) | 0.702 (2.890) | 0.471 (2.451) |
| Mean I/sig(I) | 3.3 (1.0) | 3.3 (1.1) | 4.6 (1.1) | 2.9 (0.4) |
| CC(1/2) | 0.965 (0.455) | 0.846 (0.190) | 0.970 (0.412) | 0.924 (0.189) |
| Wilson B-factor | 32.2 | 38.5 | 66.5 | 75.7 |
| Refinement | | | | |
| Resolution | 44.262 - 3.400 | 62.205 - 3.196 | 61.765 - 3.300 | 49.317 - 3.591 |
| Highest resolution bin | 3.521 - 3.400 | 3.310 - 3.196 | 3.418 - 3.300 | 3.720 - 3.591 |
| Number of reflections | | | | |
| work set | 590873 | 969144 | 429748 | 845958 |
| free set | 29845 | 6518 | 9978 | 9894 |
| R-work | 0.2681 (0.4047) | 0.2610 (0.3736) | 0.2312 (0.3436) | 0.2779 (0.3795) |
| R-free | 0.2700 (0.4096) | 0.2664 (0.3812) | 0.2331 (0.3361) | 0.2829 (0.3800) |
| Number of atoms | | | | |
| protein | 231390 | 184320 | 78960 | 275100 |
| RNA | | | | |
| water | | | | |
| other | | | 20 | |
| Average B-factor | | | | |
| protein | 69.78 | 33.9 | 66.52 | 103.09 |
| RNA | | | | |
| water | | | | |
| other | | | 59.46 | |
| rmsd from ideal geometry | | | | |
| bonds (Å) | 0.005 | 0.005 | 0.006 | 0.009 |
| angles (°) | 0.697 | 0.812 | 0.865 | 1.09 |
| Ramachandran plot | | | | |
| favored (%) | 99.11 | 98.79 | 98.18 | 98.45 |
| allowed (%) | 100.00 | 100.00 | 100.00 | 100.00 |
| Rotamer outliers (%) | 0.34 | 0.00 | 0.00 | 0.71 |
| Clashscore | 10.67 | 8.47 | 8.65 | 15.62 |

| | AVE019 | Beihai14 | Beihai19 | Beihai21 |
|---------------------------------|--|---|---|---|
| PDB ID | 6YFC | 6YFD | 6YFE | 6YFF |
| Crystallization | | | | |
| Conditions | 46% MPD, 0.1 M HEPES pH 7.5 | 5% PEG 4000, 0.1 M magnesium chloride, 0.1 M MES pH 6.5 | 66% MPD, 100 mM Tris-HCl pH 8.0 | 30% PEG 400, 0.2 M sodium citrate, 0.1 M Tris-HCl pH 8.5 |
| Data collection | | | | |
| Date | 2019-01-24 | 2019-03-22 | 2017-09-01 | 2018-01-27 |
| Site | BESSY II | BESSY II | BESSY II | BESSY II |
| Beamline | MX 14.1 | MX 14.1 | MX 14.1 | MX 14.1 |
| Wavelength (Å) | 0.9184 | 0.9184 | 0.9184 | 0.9184 |
| Data reduction | | | | |
| Space group | P 1 | P 21 21 2 | P 21 21 2 | I 2 2 2 |
| Cell parameters | a = 296.42 b = 277.20 c = 277.44 α = 103.91 β = 117.39 γ = 106.96 | a = 328.67 b = 373.71 c = 310.50 α = 90.00 β = 90.00 γ = 90.00 | a = 298.11 b = 325.05 c = 346.02 α = 90.00 β = 90.00 γ = 90.00 | a = 296.94 b = 306.88 c = 322.56 α = 90.00 β = 90.00 γ = 90.00 |
| Total number of observations | 3314889 | 2846288 | 1102377 | 1499728 |
| Number of unique reflections | 912012 | 536242 | 323252 | 264751 |
| Resolution (Å) | 49.27 - 3.25 | 49.34 - 3.30 | 49.44 - 3.79 | 48.94 - 3.09 |
| Highest resolution bin | 3.44 - 3.25 | 3.50 - 3.30 | 4.02 - 3.79 | 3.28 - 3.09 |
| Multiplicity | 3.6 (3.5) | 5.3 (2.5) | 3.4 (3.4) | 5.7 (5.5) |
| Completeness (%) | 94.0 (88.0) | 94.4 (76.8) | 98.0 (95.3) | 99.1 (95.4) |
| R-merge | 0.372 (2.509) | 0.647 (2.255) | 0.372 (1.495) | 0.358 (2.785) |
| Mean I/sig(I) | 4.3 (0.5) | 2.3 (0.5) | 3.6 (0.8) | 5.6 (0.6) |
| CC(1/2) | 0.988 (0.690) | 0.937 (0.171) | 0.965 (0.243) | 0.985 (0.221) |
| Wilson B-factor | 69.0 | 81.1 | 83.5 | 66.5 |
| Refinement | | | | |
| Resolution | 49.267 - 3.246 | 49.332 - 3.300 | 41.392 - 3.794 | 48.938 - 3.089 |
| Highest resolution bin | 3.362 - 3.246 | 3.418 - 3.300 | 3.930 - 3.794 | 3.200 - 3.089 |
| Number of reflections | | | | |
| work set | 898220 | 525682 | 322856 | 263402 |
| free set | 9890 | 9824 | 9968 | 9929 |
| R-work | 0.2413 (0.3928) | 0.2649 (0.3861) | 0.2609 (0.3706) | 0.2661 (0.3845) |
| R-free | 0.2393 (0.3869) | 0.2670 (0.3925) | 0.2668 (0.3807) | 0.2710 (0.3790) |
| Number of atoms | | | | |
| protein | 169200 | 126960 | 89040 | 43650 |
| RNA | | | | |
| water | | | | |
| other | 60 | | 30 | |
| Average B-factor | | | | |
| protein | 97.39 | 92.15 | 107.17 | 80.85 |
| RNA | | | | |
| water | | | | |
| other | 64.63 | | 53.34 | |
| rmsd from ideal geometry | | | | |
| bonds (Å) | 0.005 | 0.006 | 0.005 | 0.005 |
| angles (°) | 0.786 | 0.848 | 0.74 | 0.794 |
| Ramachandran plot | | | | |
| favored (%) | 98.88 | 98.06 | 98.43 | 98.12 |
| allowed (%) | 100.00 | 100.00 | 100.00 | 100.00 |
| Rotamer outliers (%) | 0.63 | 0.00 | 0.00 | 0.00 |
| Clashscore | 10.22 | 7.93 | 12.21 | 9.82 |

| | Beihai32 | EMS014 (VLP) | EMS014 (subunit) | ESE001 |
|---------------------------------|---|---|--|--|
| PDB ID | 6YFG | 6YFH | 6YFI | 6YFJ |
| Crystallization | | | | |
| Conditions | 26% MPD, 0.1-0.2 M sodium nitrate, 0.1 M HEPES pH 7.5 | 0.55 M sodium malonate, 0.25% Jeffamine ED-2003, 0.05 M HEPES pH 7.0 | 10% PEG 8000, 0.1 M sodium chloride, 0.05 M phosphate/citrate pH 4.2 | 30% MPD, 5 mM DTT, 0.1 M sodium acetate pH 4.6 |
| Data collection | | | | |
| Date | 2018-10-19 | 2018-05-09 | 2018-05-09 | 2017-09-01 |
| Site | MAX IV | BESSY II | BESSY II | BESSY II |
| Beamline | BioMAX | MX 14.1 | MX 14.1 | MX 14.1 |
| Wavelength (Å) | 0.97776 | 0.9184 | 0.9184 | 0.9184 |
| Data reduction | | | | |
| Space group | P 1 | C 2 2 21 | P 21 21 21 | R 3 |
| Cell parameters | a = 291.95 b = 292.53 c = 469.20 α = 75.79 β = 77.92 γ = 69.51 | a = 287.11 b = 492.50 c = 553.71 α = 90.00 β = 90.00 γ = 90.00 | a = 71.56 b = 92.70 c = 94.82 α = 90.00 β = 90.00 γ = 90.00 | a = 283.35 b = 283.35 c = 666.04 α = 90.00 β = 90.00 γ = 120.00 |
| Total number of observations | 2265552 | 1622482 | 1109958 | 1123410 |
| Number of unique reflections | 1227697 | 352052 | 173303 | 312316 |
| Resolution (Å) | 49.84 - 3.90 | 49.41 - 3.89 | 48.63 - 1.25 | 48.95 - 3.23 |
| Highest resolution bin | 4.13 - 3.90 | 4.13 - 3.89 | 1.32 - 1.25 | 3.43 - 3.23 |
| Multiplicity | 1.8 (1.9) | 4.6 (4.3) | 6.4 (6.0) | 3.6 (3.4) |
| Completeness (%) | 96.2 (93.0) | 99.1 (96.7) | 99.1 (94.7) | 97.9 (93.6) |
| R-merge | 0.209 (1.559) | 0.859 (3.144) | 0.085 (1.054) | 0.284 (1.738) |
| Mean I/sig(I) | 3.0 (0.4) | 2.3 (0.5) | 8.5 (1.0) | 4.4 (0.6) |
| CC(1/2) | 0.985 (0.151) | 0.836 (0.136) | 0.998 (0.825) | 0.984 (0.235) |
| Wilson B-factor | 118.2 | 76.7 | 21.6 | 76.2 |
| Refinement | | | | |
| Resolution | 49.833 - 3.897 | 49.357 - 3.893 | 42.210 - 1.248 | 48.944 - 3.233 |
| Highest resolution bin | 4.037 - 3.897 | 4.032 - 3.893 | 1.293 - 1.248 | 3.348 - 3.233 |
| Number of reflections | | | | |
| work set | 1216193 | 346121 | 172802 | 310956 |
| free set | 19851 | 9855 | 8618 | 10007 |
| R-work | 0.2821 (0.3974) | 0.2665 (0.3785) | 0.1753 (0.3683) | 0.2317 (0.3755) |
| R-free | 0.2853 (0.4048) | 0.2699 (0.3696) | 0.1974 (0.3796) | 0.2324 (0.3808) |
| Number of atoms | | | | |
| protein | 362880 | 104400 | 4450 | 52380 |
| RNA | | | | |
| water | | | 1076 | |
| other | 120 | | | |
| Average B-factor | | | | |
| protein | 170.05 | 111.97 | 22.13 | 97.97 |
| RNA | | | | |
| water | | | 33.61 | |
| other | 148.62 | | | |
| rmsd from ideal geometry | | | | |
| bonds (Å) | 0.006 | 0.004 | 0.005 | 0.006 |
| angles (°) | 0.870 | 0.697 | 0.762 | 0.826 |
| Ramachandran plot | | | | |
| favored (%) | 98.18 | 99.13 | 98.09 | 99.14 |
| allowed (%) | 100.00 | 100.00 | 100.00 | 100.00 |
| Rotamer outliers (%) | 0.00 | 0.00 | 0.00 | 0.00 |
| Clashscore | 14.38 | 5.62 | 1.45 | 8.54 |

| | ESE007 | ESE020 | ESE021 | ESE058 |
|---------------------------------|--|--|---|--|
| PDB ID | 6YFK | 6YFL | 6YFM | 6YFN |
| Crystallization | | | | |
| Conditions | 20 % PEG 10000, 0.1 M HEPES pH 7.5 | 36% PEG 300, 0.05 M Bicine pH 9.0 | 5% PEG 3000, 0.08 M zinc acetate, 0.05 M sodium acetate pH 4.6 | 12.5% PEG 1000, 7% PEG 8000 |
| Data collection | | | | |
| Date | 2018-01-27 | 2017-11-04 | 2018-05-09 | 2017-11-04 |
| Site | BESSY II | BESSY II | BESSY II | BESSY II |
| Beamline | MX 14.1 | MX 14.1 | MX 14.1 | MX 14.1 |
| Wavelength (Å) | 0.9184 | 0.9184 | 0.9184 | 0.9184 |
| Data reduction | | | | |
| Space group | R 3 | R 3 2 | I 2 2 2 | P 32 2 1 |
| Cell parameters | a = 278.20 b = 278.20 c = 661.50 α = 90.00 β = 90.00 γ = 120.00 | a = 419.73 b = 419.73 c = 761.14 α = 90.00 β = 90.00 γ = 120.00 | a = 282.02 b = 302.81 c = 352.16 α = 90.00 β = 90.00 γ = 90.00 | a = 500.89 b = 500.89 c = 287.05 α = 90.00 β = 90.00 γ = 120.00 |
| Total number of observations | 338161 | 1819911 | 2023525 | 6181589 |
| Number of unique reflections | 132293 | 380562 | 378707 | 679725 |
| Resolution (Å) | 38.01 - 3.70 | 99.46 - 3.30 | 60.08 - 2.76 | 49.26 - 3.19 |
| Highest resolution bin | 3.90 - 3.70 | 3.48 - 3.30 | 2.91 - 2.76 | 3.38 - 3.19 |
| Multiplicity | 2.6 (1.2) | 4.8 (4.1) | 5.3 (5.6) | 9.1 (9.1) |
| Completeness (%) | 64.9 (28.5) | 99.5 (98.9) | 99.4 (99.8) | 99.8 (98.8) |
| R-merge | 0.218 (0.817) | 0.417 (2.056) | 0.362 (1.532) | 0.796 (3.730) |
| Mean I/sig(I) | 3.2 (0.6) | 4.3 (0.7) | 4.7 (1.1) | 3.7 (0.6) |
| CC(1/2) | 0.968 (0.152) | 0.952 (0.152) | 0.957 (0.293) | 0.958 (0.234) |
| Wilson B-factor | 68.8 | 74.9 | 33.7 | 53.1 |
| Refinement | | | | |
| Resolution | 38.003 - 3.700 | 96.967 - 3.300 | 60.079 - 2.762 | 49.258 - 3.189 |
| Highest resolution bin | 3.832 - 3.700 | 3.418 - 3.300 | 2.861 - 2.762 | 3.303 - 3.189 |
| Number of reflections | | | | |
| work set | 131994 | 379266 | 378514 | 677292 |
| free set | 5027 | 9971 | 9967 | 9955 |
| R-work | 0.2351 (0.3812) | 0.2477 (0.3936) | 0.2445 (0.3695) | 0.2308 (0.3770) |
| R-free | 0.2375 (0.3643) | 0.2506 (0.3825) | 0.2480 (0.3645) | 0.2342 (0.3802) |
| Number of atoms | | | | |
| protein | 62160 | 63520 | 50130 | 101460 |
| RNA | | | | |
| water | | | | |
| other | 20 | | 60 | 30 |
| Average B-factor | | | | |
| protein | 84.62 | 71.17 | 36.12 | 60.45 |
| RNA | | | | |
| water | | | | |
| other | 51.49 | | 154.19 | 78.52 |
| rmsd from ideal geometry | | | | |
| bonds (Å) | 0.006 | 0.005 | 0.004 | 0.005 |
| angles (°) | 0.845 | 0.803 | 0.589 | 0.764 |
| Ramachandran plot | | | | |
| favored (%) | 98.02 | 98.25 | 98.87 | 99.53 |
| allowed (%) | 100.00 | 100.00 | 100.00 | 100.00 |
| Rotamer outliers (%) | 0.30 | 0.58 | 0.27 | 0.00 |
| Clashscore | 13.43 | 11.98 | 4.02 | 7.66 |

| | GQ-907 | GQ-112 | NT-214 | NT-391 |
|---------------------------------|--|--|---|---|
| PDB ID | 6YFO | 6YFP | 6YFQ | 6YFR |
| Crystallization | | | | |
| Conditions | 12.5 % PEG 3350, 0.05 M Bis-tris pH 5.5 | 0.615 M sodium succinate pH 7.0 | 30% MPD, 0.02 M calcium chloride, 0.1 M sodium acetate pH 4.6 | 34% MPD, 0.1 M sodium nitrate, 0.1M sodium acetate pH 4.6 |
| Data collection | | | | |
| Date | 2019-01-24 | 2019-03-22 | 2018-10-19 | 2019-01-24 |
| Site | BESSY II | BESSY II | MAX IV | BESSY II |
| Beamline | MX 14.1 | MX 14.1 | BioMAX | MX 14.1 |
| Wavelength (Å) | 0.9184 | 0.9184 | 0.97776 | 0.9184 |
| Data reduction | | | | |
| Space group | C 2 | C 2 | R 3 2 | P 1 |
| Cell parameters | a = 415.01 b = 335.00 c = 291.78 α = 90.00 β = 134.66 γ = 90.00 | a = 469.42 b = 332.41 c = 293.53 α = 90.00 β = 127.87 γ = 90.00 | a = 341.90 b = 341.90 c = 1253.60 α = 90.00 β = 90.00 γ = 120.00 | a = 161.92 b = 162.07 c = 162.98 α = 71.16 β = 66.67 γ = 66.83 |
| Total number of observations | 1226838 | 1699944 | 5672505 | 690098 |
| Number of unique reflections | 353773 | 508294 | 275564 | 294001 |
| Resolution (Å) | 49.14 - 3.48 | 57.21 - 3.10 | 49.97 - 3.80 | 51.89 - 2.60 |
| Highest resolution bin | 3.69 - 3.48 | 3.27 - 3.10 | 4.01 - 3.80 | 2.74 - 2.60 |
| Multiplicity | 3.5 (3.5) | 3.3 (2.0) | 20.6 (19.2) | 2.3 (2.1) |
| Completeness (%) | 97.9 (94.2) | 79.5 (35.7) | 98.3 (90.0) | 69.7 (56.2) |
| R-merge | 0.449 (2.461) | 0.310 (1.139) | 0.597 (2.764) | 0.239 (1.134) |
| Mean I/sig(I) | 3.2 (0.5) | 3.4 (0.7) | 4.7 (1.0) | 2.5 (0.5) |
| CC(1/2) | 0.966 (0.180) | 0.753 (0.254) | 0.993 (0.467) | 0.927 (0.102) |
| Wilson B-factor | 83.2 | 46.4 | 96.6 | 46.7 |
| Refinement | | | | |
| Resolution | 49.140 - 3.484 | 57.200 - 3.100 | 49.955 - 3.800 | 51.880 - 2.600 |
| Highest resolution bin | 3.609 - 3.484 | 3.211 - 3.100 | 3.936 - 3.800 | 2.693 - 2.600 |
| Number of reflections | | | | |
| work set | 347398 | 502524 | 275362 | 286598 |
| free set | 9832 | 9844 | 4997 | 9703 |
| R-work | 0.2685 (0.3989) | 0.2500 (0.3705) | 0.2199 (0.3775) | 0.3175 (0.4009) |
| R-free | 0.2710 (0.3925) | 0.2537 (0.3828) | 0.2220 (0.3979) | 0.3231 (0.3985) |
| Number of atoms | | | | |
| protein | 87480 | 116370 | 61200 | 56340 |
| RNA | | | | |
| water | | | | |
| other | | | | |
| Average B-factor | | | | |
| protein | 110.95 | 43.41 | 135.8 | 50.25 |
| RNA | | | | |
| water | | | | |
| other | | | | |
| rmsd from ideal geometry | | | | |
| bonds (Å) | 0.005 | 0.006 | 0.007 | 0.008 |
| angles (°) | 0.87 | 0.905 | 0.817 | 0.685 |
| Ramachandran plot | | | | |
| favored (%) | 99.22 | 98.97 | 98.50 | 100.00 |
| allowed (%) | 100.00 | 100.00 | 100.00 | 100.00 |
| Rotamer outliers (%) | 0.00 | 0.00 | 0.00 | 0.00 |
| Clashscore | 11.77 | 7.84 | 12.14 | 2.73 |

| | PQ-465 | Wenzhou1 | Wenzhou4 |
|---------------------------------|--|---|---|
| PDB ID | 6YFS | 6YFT | 6YFU |
| Crystallization | | | |
| Conditions | 38% MPD, 0.1 M sodium acetate pH 4.6 | 10 % v/v ethanol, 0.1 M Tris-HCl pH 8.5 | 30 % MPD, 0.1 M MES pH 6.5 |
| Data collection | | | |
| Date | 2019-06-08 | 2019-01-24 | 2018-10-19 |
| Site | BESSY II | BESSY II | MAX IV |
| Beamline | MX 14.1 | MX 14.1 | BioMAX |
| Wavelength (Å) | 0.9184 | 0.9184 | 0.97776 |
| Data reduction | | | |
| Space group | C 2 | P 1 | I 2 3 |
| Cell parameters | a = 453.15 b = 309.76 c = 294.60 α = 90.00 β = 130.08 γ = 90.00 | a = 277.47 b = 396.64 c = 399.87 α = 69.66 β = 83.65 γ = 83.49 | a = 551.80 b = 551.80 c = 551.80 α = 90.00 β = 90.00 γ = 90.00 |
| Total number of observations | 1350175 | 3182922 | 9650681 |
| Number of unique reflections | 382128 | 1749831 | 229056 |
| Resolution (Å) | 49.35 - 3.50 | 61.16 - 3.50 | 49.96 - 4.02 |
| Highest resolution bin | 3.71 - 3.50 | 3.69 - 3.50 | 4.24 - 4.00 |
| Multiplicity | 3.5 (3.5) | 1.8 (1.8) | 42.1 (40.0) |
| Completeness (%) | 97.5 (96.3) | 87.6 (89.9) | 98.3 (90.1) |
| R-merge | 0.473 (4.064) | 0.307 (1.481) | 0.882 (6.402) |
| Mean I/sig(I) | 3.5 (0.3) | 2.6 (0.5) | 5.2 (0.6) |
| CC(1/2) | 0.970 (0.110) | 0.920 (0.117) | 0.994 (0.201) |
| Wilson B-factor | 98.8 | 71.1 | 127.6 |
| Refinement | | | |
| Resolution | 49.343 - 3.498 | 61.150 - 3.500 | 49.958 - 4.018 |
| Highest resolution bin | 3.623 - 3.498 | 3.625 - 3.500 | 4.162 - 4.018 |
| Number of reflections | | | |
| work set | 357164 | 1741933 | 227552 |
| free set | 9293 | 19893 | 4953 |
| R-work | 0.2522 (0.3874) | 0.2612 (0.3894) | 0.2424 (0.3710) |
| R-free | 0.2593 (0.3856) | 0.2606 (0.3897) | 0.2498 (0.3832) |
| Number of atoms | | | |
| protein | 84900 | 302040 | 66540 |
| RNA | | 25200 | |
| water | | | |
| other | | | |
| Average B-factor | | | |
| protein | 120.8 | 93.43 | 161.39 |
| RNA | | 186.53 | |
| water | | | |
| other | | | |
| rmsd from ideal geometry | | | |
| bonds (Å) | 0.005 | 0.008 | 0.005 |
| angles (°) | 0.755 | 0.955 | 0.994 |
| Ramachandran plot | | | |
| favored (%) | 98.09 | 98.50 | 98.61 |
| allowed (%) | 100.00 | 100.00 | 100.00 |
| Rotamer outliers (%) | 0.00 | 0.00 | 0.00 |
| Clashscore | 11.85 | 4.57 | 15.19 |

Table S2. Crystallographic data collection, reduction and refinement statistics. Values in parentheses correspond to the highest resolution bin.

REFERENCES AND NOTES

1. P. Pumpens, *Single-Stranded RNA Phages: From Molecular Biology to Nanotechnology* (CRC Press, 2020).
2. Y. I. Wolf, D. Kazlauskas, J. Iranzo, A. Lucía-Sanz, J. H. Kuhn, M. Krupovic, V. V. Dolja, E. V. Koonin, Origins and evolution of the global RNA virome. *mBio* **9**, e02329-18 (2018).
3. K. Valegård, L. Liljas, K. Fridborg, T. Unge, The three-dimensional structure of the bacterial virus MS2. *Nature* **345**, 36–41 (1990).
4. X. Dai, Z. Li, M. Lai, S. Shu, Y. Du, Z. H. Zhou, R. Sun, In situ structures of the genome and genome-delivery apparatus in a single-stranded RNA virus. *Nature* **541**, 112–116 (2017).
5. L. Liljas, K. Fridborg, K. Valegård, M. Bundule, P. Pumpens, Crystal structure of bacteriophage fr capsids at 3.5 Å resolution. *J. Mol. Biol.* **244**, 279–290 (1994).
6. R. Golmohammadi, K. Fridborg, M. Bundule, K. Valegård, L. Liljas, The crystal structure of bacteriophage Q β at 3.5 Å resolution. *Structure* **4**, 543–554 (1996).
7. K. Tars, M. Bundule, K. Fridborg, L. Liljas, The crystal structure of bacteriophage GA and a comparison of bacteriophages belonging to the major groups of *Escherichia coli* leviviruses. *J. Mol. Biol.* **271**, 759–773 (1997).
8. K. Tars, K. Fridborg, M. Bundule, L. Liljas, The three-dimensional structure of bacteriophage PP7 from *Pseudomonas aeruginosa* at 3.7-Å resolution. *Virology* **272**, 331–337 (2000).
9. M. Persson, K. Tars, L. Liljas, The capsid of the small RNA phage PRR1 is stabilized by metal ions. *J. Mol. Biol.* **383**, 914–922 (2008).
10. P. Plevka, A. Kazaks, T. Voronkova, S. Kotelovica, A. Dishlers, L. Liljas, K. Tars, The structure of bacteriophage ϕ Cb5 reveals a role of the RNA genome and metal ions in particle stability and assembly. *J. Mol. Biol.* **391**, 635–647 (2009).

11. M. Shishovs, J. Rumnieks, C. Diebolder, K. Jaudzems, L. B. Andreas, J. Stanek, A. Kazaks, S. Kotelovica, I. Akopjana, G. Pintacuda, R. I. Koning, K. Tars, Structure of AP205 coat protein reveals circular permutation in ssRNA bacteriophages. *J. Mol. Biol.* **428**, 4267–4279 (2016).
12. S. R. Krishnamurthy, A. B. Janowski, G. Zhao, D. Barouch, D. Wang, Hyperexpansion of RNA bacteriophage diversity. *PLOS Biol.* **14**, e1002409 (2016).
13. M. Shi, X.-D. Lin, J.-H. Tian, L.-J. Chen, X. Chen, C.-X. Li, X.-C. Qin, J. Li, J.-P. Cao, J.-S. Eden, J. Buchmann, W. Wang, J. Xu, E.C. Holmes, Y.-Z. Zhang, Redefining the invertebrate RNA virosphere. *Nature* **540**, 539–543 (2016).
14. E. P. Starr, E. E. Nuccio, J. Pett-Ridge, J. F. Banfield, M. K. Firestone, Metatranscriptomic reconstruction reveals RNA viruses with the potential to shape carbon cycling in soil. *Proc. Natl. Acad. Sci. U.S.A.* **116**, 25900–25908 (2019).
15. J. Callanan, S. R. Stockdale, A. Shkoporov, L. A. Draper, R. P. Ross, C. Hill, Expansion of known ssRNA phage genomes: From tens to over a thousand. *Sci. Adv.* **6**, eaay5981 (2020).
16. I. Liekniņa, G. Kalniņš, I. Akopjana, J. Bogans, M. Šišovs, J. Jansons, J. Rūmnieks, K. Tārs, Production and characterization of novel ssRNA bacteriophage virus-like particles from metagenomic sequencing data. *J. Nanobiotech.* **17**, 61 (2019).
17. P. Pumpens, R. Renhofa, A. Dishlers, T. Kozlovska, V. Ose, P. Pushko, K. Tars, E. Grens, M.F. Bachmann, The true story and advantages of RNA phage capsids as nanotools. *Intervirology* **59**, 74–110 (2016).
18. S. C. Harrison, A. J. Olson, C. E. Schutt, F. K. Winkler, G. Bricogne, Tomato bushy stunt virus at 2.9 Å resolution. *Nature* **276**, 368–373 (1978).
19. K. Tars, A. Zeltins, L. Liljas, The three-dimensional structure of cocksfoot mottle virus at 2.7 Å resolution. *Virology* **310**, 287–297 (2003).
20. P. Plevka, K. Tars, A. Zeltins, I. Balke, E. Truve, L. Liljas, The three-dimensional structure of ryegrass mottle virus at 2.9 Å resolution. *Virology* **369**, 364–374 (2007).

21. M. A. Asensio, N. M. Morella, C. M. Jakobson, E. C. Hartman, J. E. Glasgow, B. Sankaran, P. H. Zwart, D. Tullman-Ercek, A selection for assembly reveals that a single amino acid mutant of the bacteriophage MS2 coat protein forms a smaller virus-like particle. *Nano Lett.* **16**, 5944–5950 (2016).
22. A. Luque, D. Reguera, The structure of elongated viral capsids. *Biophys. J.* **98**, 2993–3003 (2010).
23. I. Cielens, V. Ose, I. Petrovskis, A. Strelnikova, R. Renhofa, T. Kozlovska, P. Pumpens, Mutilation of RNA phage Q β virus-like particles: From icosahedrons to rods. *FEBS Lett.* **482**, 261–264 (2000).
24. J. Rumnieks, V. Ose, K. Tars, A. Dislers, A. Strods, I. Cielens, R. Renhofa, Assembly of mixed rod-like and spherical particles from group I and II RNA bacteriophage coat proteins. *Virology* **391**, 187–194 (2009).
25. M. F. Moody, The shape of the T-even bacteriophage head. *Virology* **26**, 567–576 (1965).
26. A. Sicard, Y. Michalakakis, S. Gutiérrez, S. Blanc, The strange lifestyle of multipartite viruses. *PLOS Pathog.* **12**, e1005819 (2016).
27. K. Valegård, J. B. Murray, P. G. Stockley, N. J. Stonehouse, L. Liljas, Crystal structure of an RNA bacteriophage coat protein–operator complex. *Nature* **371**, 623–626 (1994).
28. M. Persson, K. Tars, L. Liljas, PRR1 coat protein binding to its RNA translational operator. *Acta Crystallogr. D Biol. Crystallogr.* **69**, 367–372 (2013).
29. J. Rumnieks, K. Tars, Crystal structure of the bacteriophage Q β coat protein in complex with the RNA operator of the replicase gene. *J. Mol. Biol.* **426**, 1039–1049 (2014).
30. J. A. Chao, Y. Patskovsky, S. C. Almo, R. H. Singer, Structural basis for the coevolution of a viral RNA–protein complex. *Nat. Struct. Mol. Biol.* **15**, 103–105 (2008).
31. K. V. Gorzelnik, Z. Cui, C. A. Reed, J. Jakana, R. Young, J. Zhang, Asymmetric cryo-EM structure of the canonical *Allolevivirus* Q β reveals a single maturation protein and the genomic ssRNA in situ. *Proc. Natl. Acad. Sci. U.S.A.* **113**, 11519–11524 (2016).

32. Ó. Rolfsson, S. Middleton, I. W. Manfield, S. J. White, B. Fan, R. Vaughan, N. A. Ranson, E. Dykeman, R. Twarock, J. Ford, C. C. Kao, P. G. Stockley, Direct evidence for packaging signal-mediated assembly of bacteriophage MS2. *J. Mol. Biol.* **428**, 431–448 (2016).
33. D. J. Klein, P. B. Moore, T. A. Steitz, The roles of ribosomal proteins in the structure assembly, and evolution of the large ribosomal subunit. *J. Mol. Biol.* **340**, 141–177 (2004).
34. A. L. N. Rao, Genome packaging by spherical plant RNA viruses. *Annu. Rev. Phytopathol.* **44**, 61–87 (2006).
35. M. Krupovic, E. V. Koonin, Multiple origins of viral capsid proteins from cellular ancestors. *Proc. Natl. Acad. Sci. U.S.A.* **114**, E2401–E2410 (2017).
36. T. G. G. Battye, L. Kontogiannis, O. Johnson, H. R. Powell, A. G. W. Leslie, *iMOSFLM*: A new graphical interface for diffraction-image processing with *MOSFLM*. *Acta Crystallogr. D Biol. Crystallogr.* **67**, 271–281 (2011).
37. P. R. Evans, Scala. Joint CCP4 + ESF-EAMCB. *Newslett. Protein Crystallogr.* **33**, 22–24 (1997).
38. M. D. Winn, C. C. Ballard, K. D. Cowtan, E. J. Dodson, P. Emsley, P. R. Evans, R. M. Keegan, E. B. Krissinel, A. G. W. Leslie, A. McCoy, S. J. McNicholas, G. N. Murshudov, N. S. Pannu, E. A. Potterton, H. R. Powell, R. J. Read, A. Vagin, K. S. Wilson, Overview of the *CCP4* suite and current developments. *Acta Crystallogr. D Biol. Crystallogr.* **67**, 235–242 (2011).
39. W. Kabsch, XDS. *Acta Crystallogr. D Biol. Crystallogr.* **66**, 125–132 (2010).
40. K. M. Sparta, M. Krug, U. Heinemann, U. Mueller, M. S. Weiss, XDSAPP2.0. *J. Appl. Crystallogr.* **49**, 1085–1092 (2016).
41. L. Tong, M. G. Rossmann, Rotation function calculations with GLRF program. *Methods Enzymol.* **276**, 594–611 (1997).
42. A. J. McCoy, R. W. Grosse-Kunstleve, P. D. Adams, M. D. Winn, L. C. Storoni, R. J. Read, *Phaser* crystallographic software. *J. Appl. Cryst.* **40**, 658–674 (2007).

43. G. J. Kleywegt, T. A. Jones, Halloween ... masks and bones in *From First Map to Final model. Proceedings of the CCP4 Study Weekend*, S. Bailey, R. Hubbard, D. Waller, Eds. (SERC Daresbury Laboratory, Daresbury, 1994), pp. 59–66.
44. R. J. Read, Improved Fourier coefficients for maps using phases from partial structures with errors. *Acta Crystallogr. A* **42**, 140–149 (1986).
45. G. J. Kleywegt, R. J. Read, Not your average density. *Structure* **5**, 1557–1569 (1997).
46. P. Emsley, K. Cowtan, *Coot*: Model-building tools for molecular graphics. *Acta Crystallogr. D Biol. Crystallogr.* **60**, 2126–2132 (2004).
47. D. Liebschner, P. V. Afonine, M. L. Baker, G. Bunkóczi, V. B. Chen, T. I. Croll, B. Hintze, L. W. Hung, S. Jain, A. J. McCoy, N. W. Moriarty, R. D. Oeffner, B. K. Poon, M. G. Prisant, R. J. Read, J. S. Richardson, D. C. Richardson, M. D. Sammito, O. V. Sobolev, D. H. Stockwell, T. C. Terwilliger, A. G. Urzhumtsev, L. L. Videau, C. J. Williams, P. D. Adams, Macromolecular structure determination using X-rays, neutrons and electrons: Recent developments in *Phenix*. *Acta Crystallogr. D Struct. Biol.* **75**, 861–877 (2019).
48. V. B. Chen, W. B. Arendall III, J. J. Headd, D. A. Keedy, R. M. Immormino, G. J. Kapral, L. W. Murray, J. S. Richardson, D. C. Richardson, *MolProbity*: All-atom structure validation for macromolecular crystallography. *Acta Crystallogr. D Biol. Crystallogr.* **66**, 12–21 (2010).
49. E. Krissinel, K. Henrick, Secondary-structure matching (SSM), a new tool for fast protein structure alignment in three dimensions. *Acta Crystallogr. D Biol. Crystallogr.* **60**, 2256–2268 (2004).
50. L. Holm, Benchmarking fold detection by DaliLite v.5. *Bioinformatics* **35**, 5326–5327 (2019).
51. E. Krissinel, K. Henrick, Inference of macromolecular assemblies from crystalline state. *J. Mol. Biol.* **372**, 774–797 (2007).