

Supporting Information

SI Text

Calculating the number of symbiont generations in each regime.

In order to put the observed evolutionary changes into context, it was important to calculate the total number of symbiont generations over the course of the experiment as best as possible. This would allow us to compare the number of generations between HT and VT regimes, since a large difference between the two values may improve our understanding of any observed changes.

In the VT treatment, TSY medium was replaced daily and this removed any released EBs present in the medium. Although these EBs were removed from the flask, thus preventing their ability to infect host cells, they still contributed to the total number of symbiont generations. We quantified the amount of EBs present in the medium per day over the course of three successive days with qPCR. We also quantified the amount of symbionts present in a fully infected continuous culture with freshly replaced medium with qPCR. In this way we were able to calculate the number of generations arising from released EBs per day. Apart from this, we measured the growth rate of amoeba infected with symbionts from this VT treatment. Since bacteria are transmitted vertically from parent to daughter cell, the doubling of amoeba cells would be equivalent to doubling of bacteria. We then added the number of generations arising from the released EBs to the number of generations arising from host cell division to calculate the total number of symbiont generations for the VT regime, which equated to 525 generations for the entire duration of the evolution experiment.

In the HT treatment, symbionts isolated from the medium were added weekly to naive amoebae. By quantifying the number of symbionts added at the start of the week and the number of symbionts in the medium at the end of the week, both with qPCR, we were able to calculate the number of symbiont generations arising from released EBs throughout the week. We also measured the growth rate of amoeba infected with

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symbionts from this HT treatment. By adding the two values together we were able to calculate the total number of symbiont generations for the HT regime, which equated to 560 generations for the entire duration of the evolution experiment.

Thus, we can say that the observed evolutionary changes in the HT and VT regimes have not simply been brought about as a result of a large difference between the total numbers of symbiont generations.

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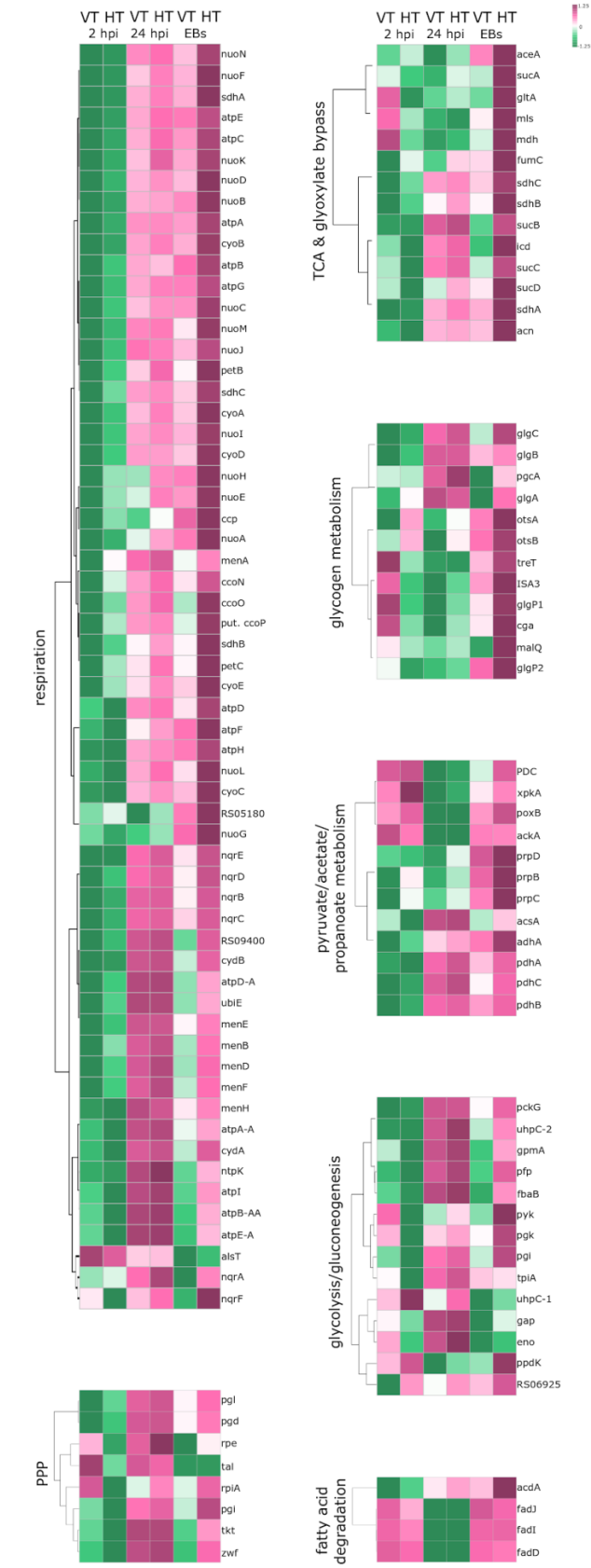


Fig. S1 Heat maps of pathways involved in central carbon metabolism and energy generation.

Heat maps of RNA-Seq data from mean centered gene expression (TPM) of symbiont genes (listed in [Dataset S4](#)) in HT and VT regimes at all three time points during infection. All known loci involved or putatively involved in the pathways shown are depicted. Gene names or locus tags (without the prefix PUV_) are indicated. Highly expressed genes are shown in pink, whereas low gene expression is shown in green. The stronger the color, the stronger the gene expression value.

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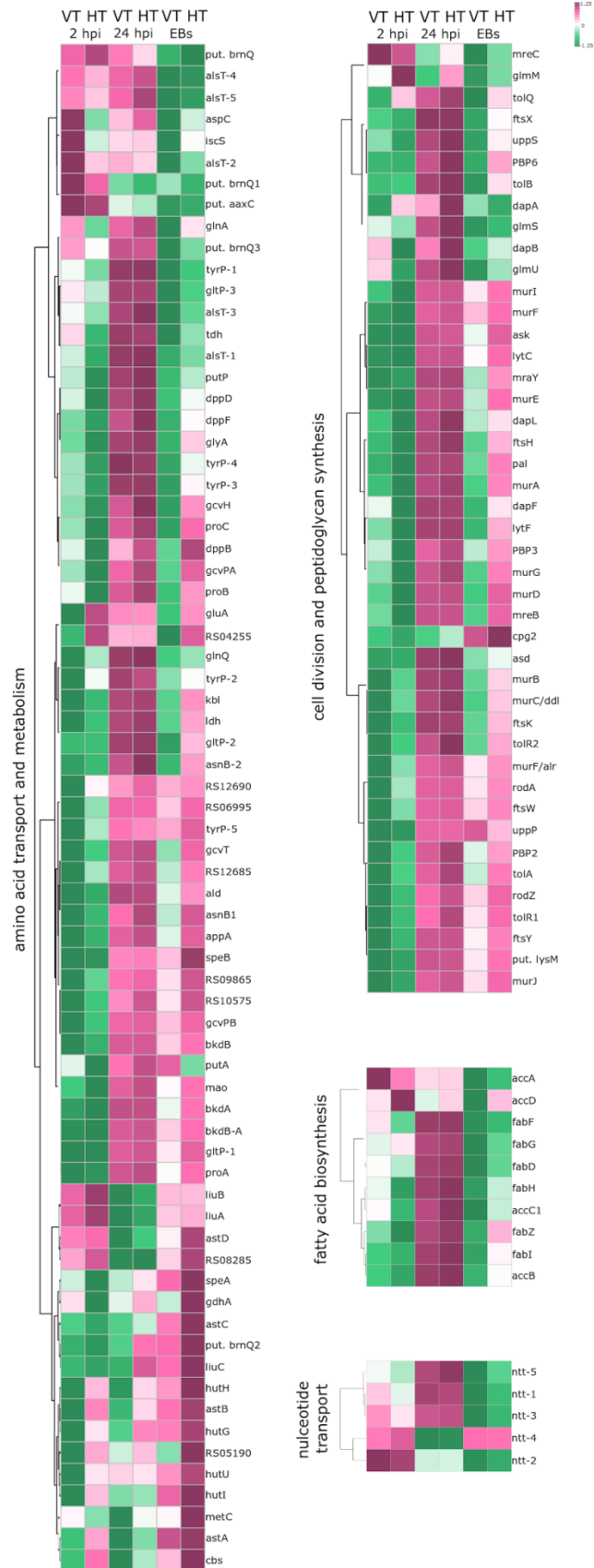


Fig. S2 Heat maps of metabolic pathways strongly upregulated at the peak of RB activity (i.e. at 24 hpi).

Heat maps of RNA-Seq data from mean centered gene expression (TPM) of symbiont genes (listed in [Dataset S4](#)) in HT and VT regimes at all three time points during infection. All known loci involved or putatively involved in the pathways shown are depicted. Gene names or locus tags (without the prefix PUV_) are indicated. Highly expressed genes are shown in pink, whereas low gene expression is shown in green. The stronger the color, the stronger the gene expression value.

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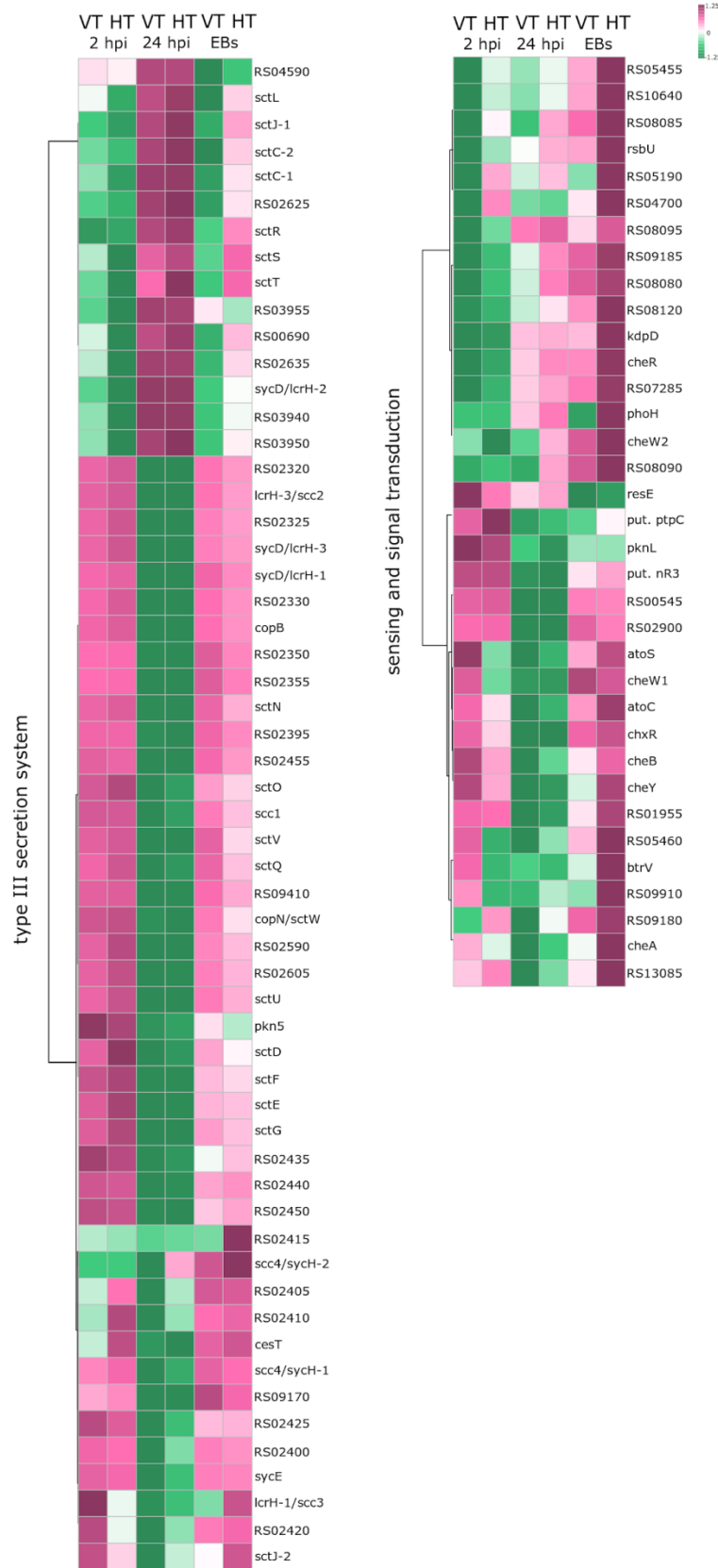


Fig. S3 Heat maps of pathways involved in infection, sensing and signal transduction pathways.

Heat maps of RNA-Seq data from mean centered gene expression (TPM) of symbiont genes (listed in [Dataset S4](#)) in HT and VT regimes at all three time points during infection. All known loci involved or putatively involved in the pathways shown are depicted. Gene names or locus tags (without the prefix PUV_) are indicated. Highly expressed genes are shown in pink, whereas low gene expression is shown in green. The stronger the color, the stronger the gene expression value.

Datasets

Dataset S1.xlsx All values of host and symbiont fitness for individual replicates.

Dataset S2.xlsx Summary of all standing genetic variants and novel variants identified in the HT and VT populations.

Dataset S3.xlsx The percentage of the 112 unique genes differentially expressed in the HT symbiont population affected per functional category.

Dataset S4.xlsx Summary of normalized expression for genes from all replicates and time points of the HT and VT treatments. $TPM = (\text{reads mapping to gene}) / (\text{length of gene} / 1000) / (\text{total reads per kilobase} / 1000000)$.

Dataset S5.xlsx Summary of all differentially expressed genes between two consecutive time points in both treatments. Genes were considered differentially expressed if their expression changed twofold with a false-discovery rate (FDR) smaller or equal 0.05.

Dataset S6.xlsx Summary of all 354 and 248 differentially expressed genes in HT and VT, respectively.

Dataset S7.xlsx Summary of all 112 and 8 uniquely differentially expressed genes in HT and VT, respectively.

Dataset S8.xlsx All individual \log_2 fold changes of genes that were differentially expressed between released EBs and the 2 hpi (false discovery rate of 0.05) for those

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metabolic pathways and T3SS loci with marked differences between the HT and VT treatments.

Dataset S9.xlsx Summary of overrepresented functional categories ($FDR \leq 0.05$) among differentially expressed genes between two consecutive time points for each set-up.

Dataset S10.xlsx RNA-Seq and read mapping statistics.