

Supplementary Materials

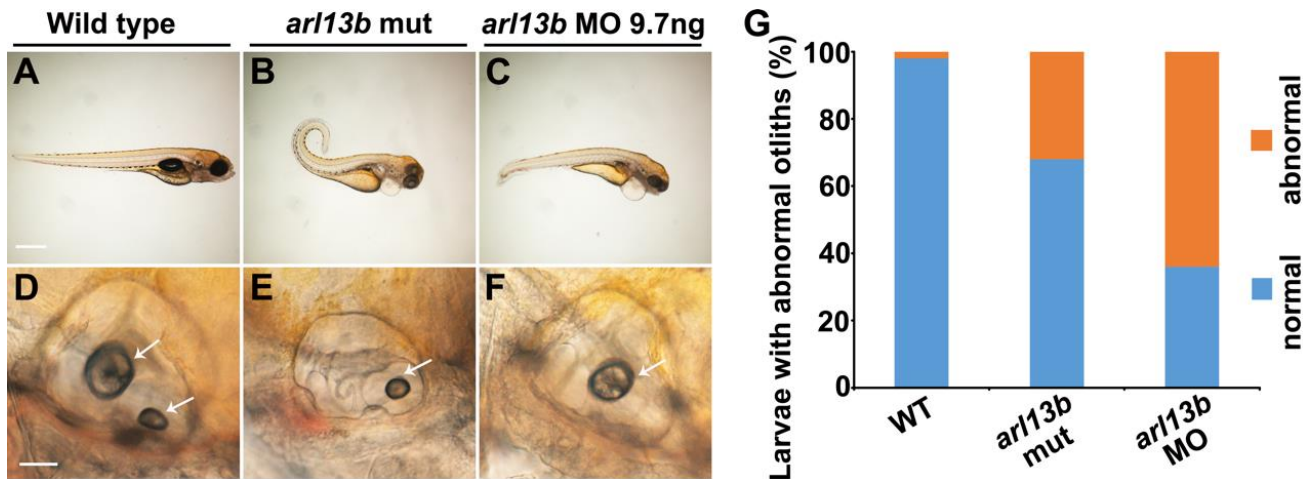


Fig. S1 Disruption of *arl13b* results in morphological defects. **A–C** Wild-type embryos are straight at 4 dpf while the bodies of *arl13b* mutants and morphants are curved. **D–G** There are usually two otoliths in the inner ear; however, the otoliths of *arl13b*-deficient embryos have defects in number, size, and morphology. A-C, Scale bar 400 μ m. F-I'', D-F, Scale bar 100 μ m.

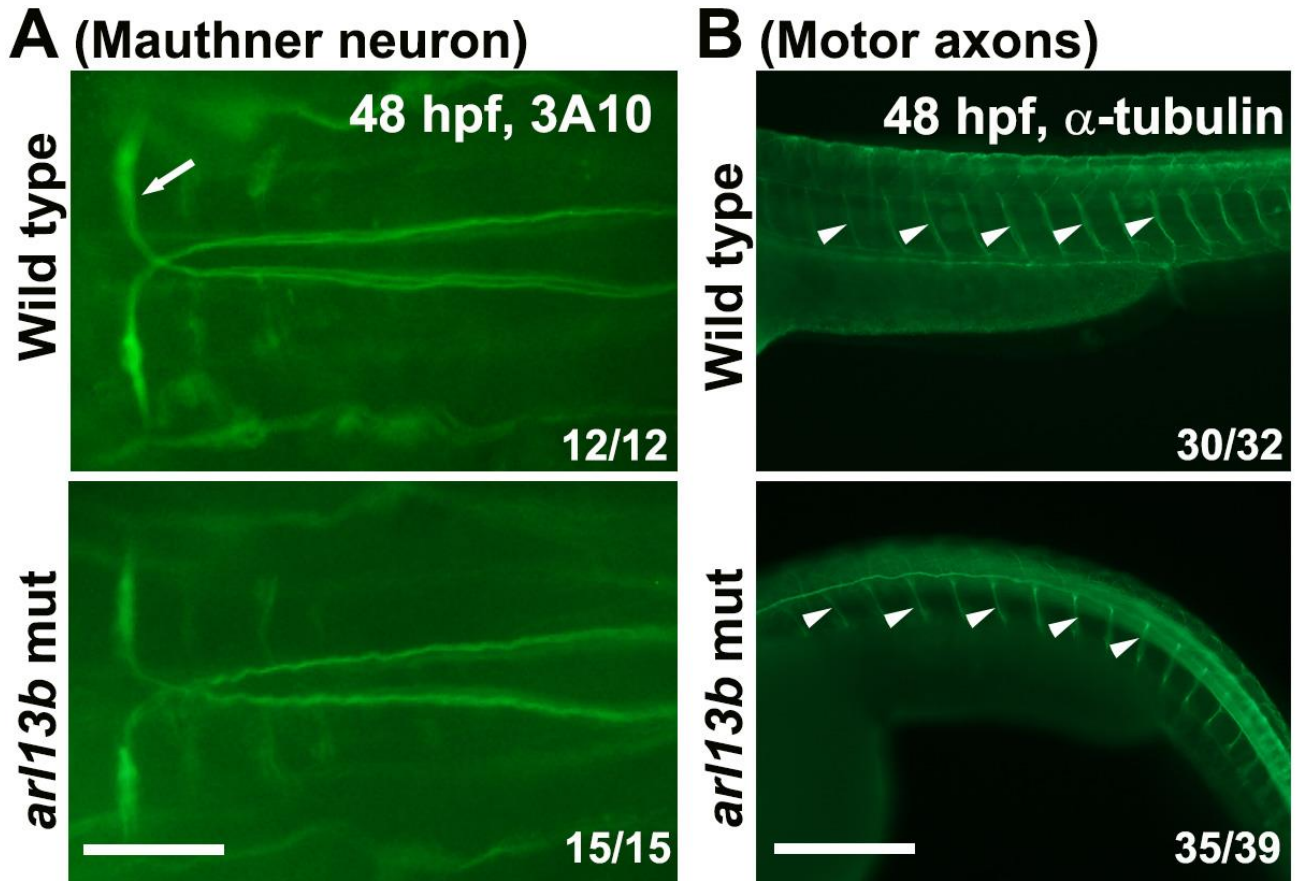


Fig. S2 Locomotion-related neurons remain intact in *arl13b*-deficient embryos. **A** Mauthner neurons still grow out axons, which normally cross the midline and project caudally, in *arl13b*-mutants. **B** No defect of motor axons is detectable in *arl13b*-mutants. A-B, Scale bar 100 μ m.

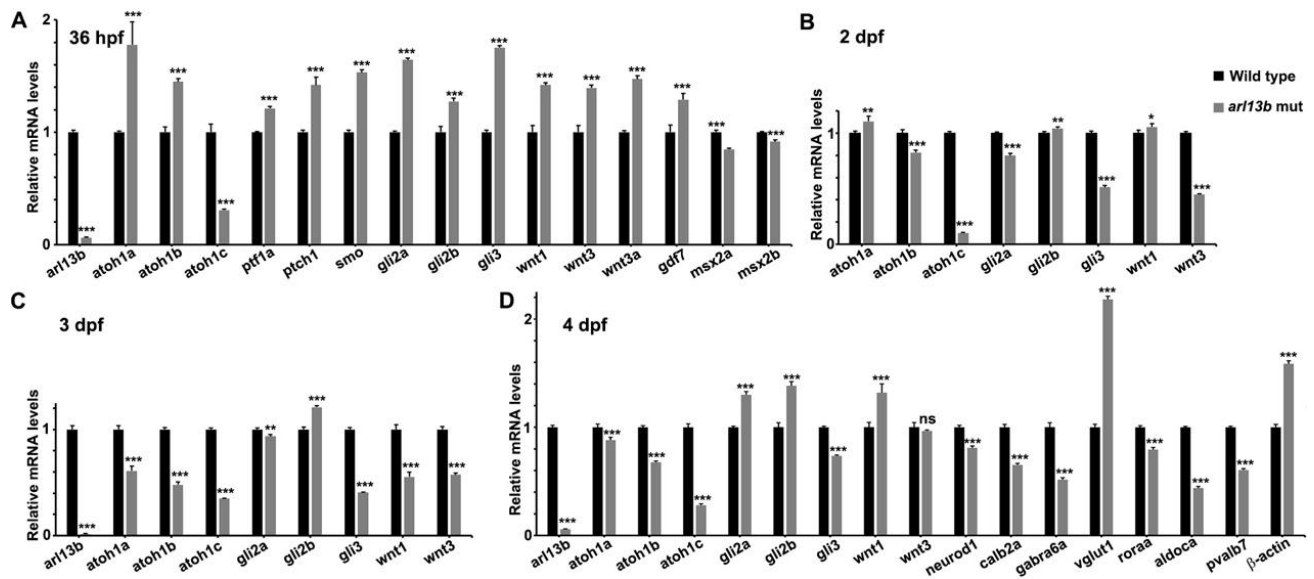


Fig. S3 The expression levels of a network of genes are altered dynamically in *arl13b* mutants. Total RNAs were extracted from wild type and *arl13b* mutant embryos (whole mount) and reversely transcribed. The obtained cDNAs were used as templates for real time quantitative PCR (q-PCR). The expression levels of the genes are normalized to that of elongation factor EF1 α . *arl13b* is significantly reduced in mutants, confirming that the embryos picked on the basis of curved tails are virtually *arl13b* mutants with high penetration, consistent with published data. The expression levels of *atoh1a* and *1b* (granule cell precursors) and *ptf1a* (Purkinje cell precursors) are dynamically regulated in *arl13b* mutants. The expression level of *atoh1c* continually declines in *arl13b* mutants at all developmental stages. The expression levels of Shh signaling (*ptch1*, *smo*, *gli2a*, *gli2b*, and *gli3*), Wnt signaling (*wnt1*, *wnt3*, and *wnt3a*), and BMP signaling (*gdf7*, *msx2a*, and *msx2b*) are altered by *arl13b* deficiency. The molecules expressed in differentiated granule cells (*neurod1*, *calb2a*, *gabra6a*, and *vglut1*) and Purkinje cells (*roraa*, *aldoca*, and *pvalb7*) are also altered. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$.

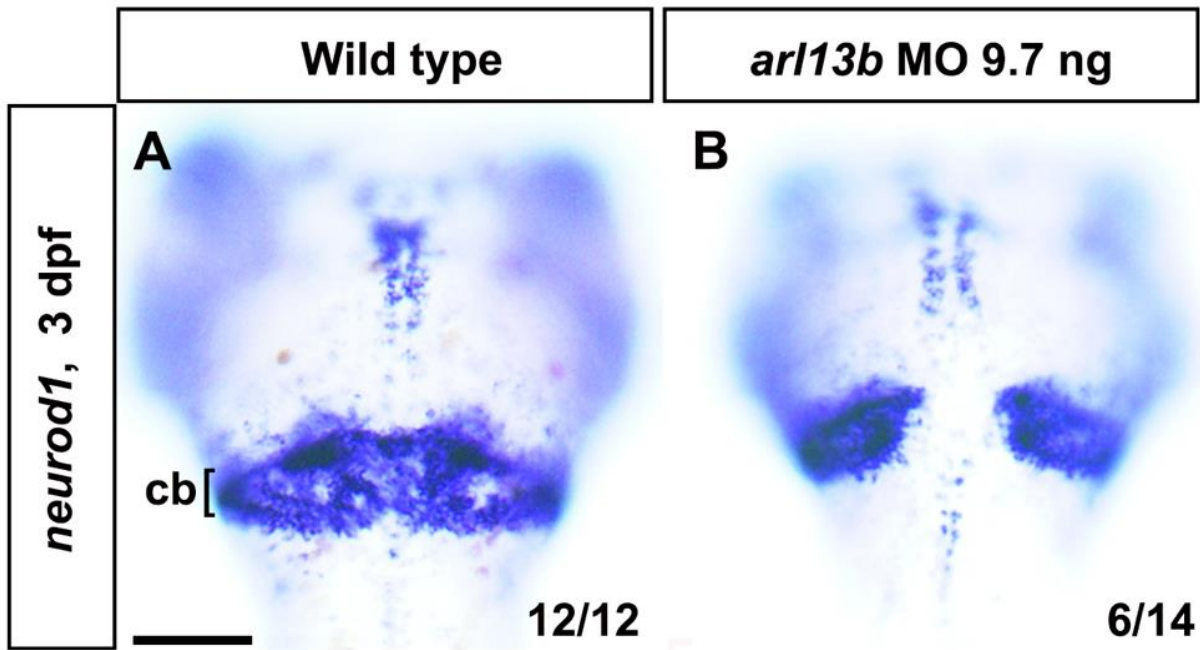


Fig. S4 The expression of granule cell marker *neruod1* is specifically reduced in the dorsomedial clusters. The dorsomedial clusters of cerebellar granule neurons are specifically impaired by *arl13b* disruption. A-B, Scale bar 100 μ m.

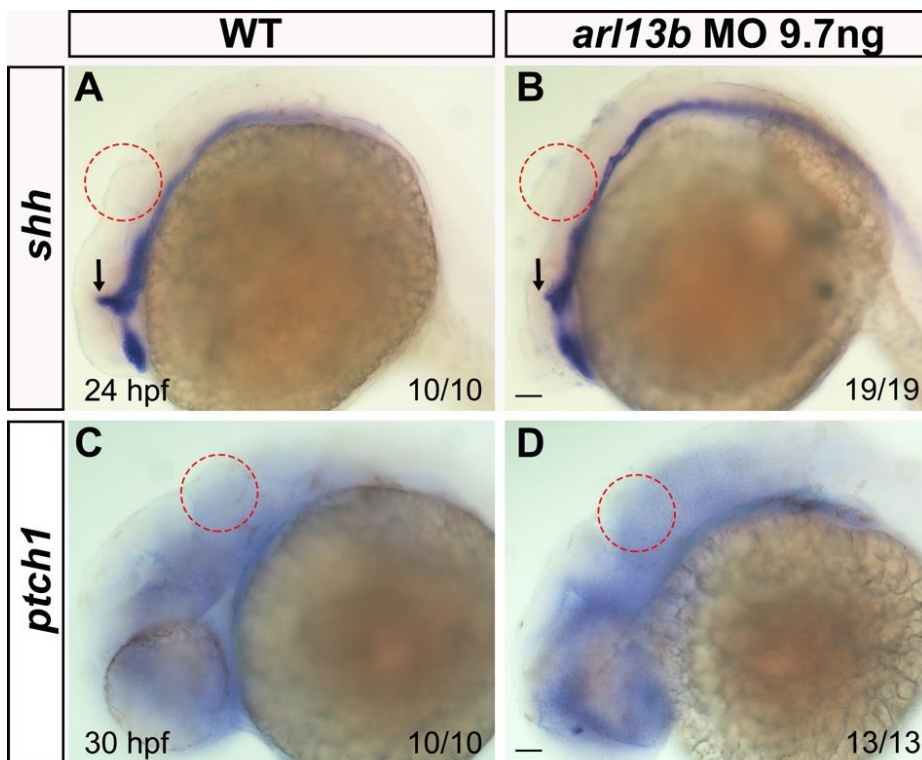


Fig. S5 The expression of *shh* and its receptor, *patched1* (*ptch1*), is not altered by *arl13b* knockdown. **A, C** The expression of Shh signaling pathway components, *shh* and *ptch1*, is barely detectable in the cerebellum (dashed circles) by *in situ* hybridization in wild-type embryos. **B, D** Their expression is not altered by *arl13b* morpholino treatment although the expression of *shh* is changed in the dorsal zona limitans intrathalamica (arrows). A-D, Scale bar 100 μ m.

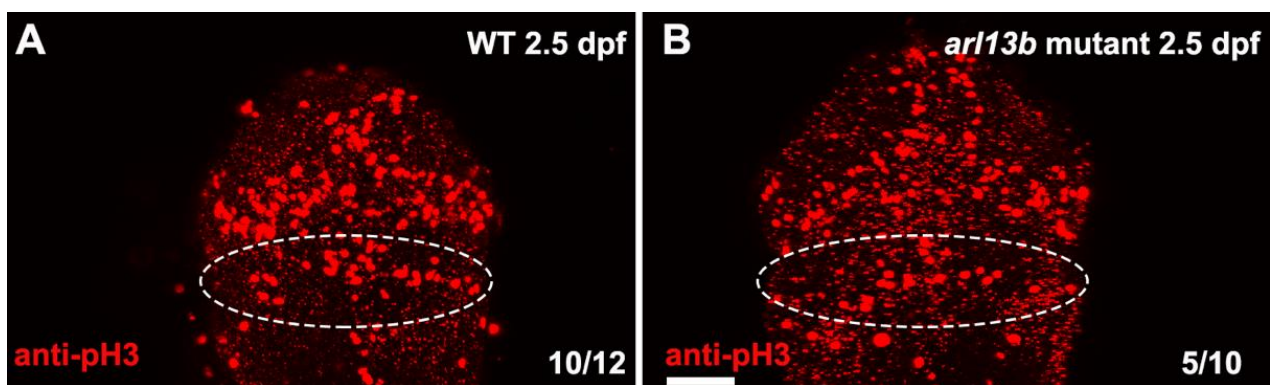


Fig. S6 The proliferation of cells in the cerebellum is impaired in *arl13b* mutants. **A, B** Representative images of wild-type and *arl13b* mutant embryos immunostained with anti-pH3 antibody. The cerebellum is outlined by the dashed oval. A-B, Scale bar 100 μ m.

Supplemental Tables

Table S1 PCR primers for amplifying gene-specific products

Gene	Primers	Size (bp)
<i>atoh1b</i>	AAGCGTCATTCCTTCATTGGAG ACAAGTATTGACGTGTGCTGAG*	402

<i>atoh1c</i>	ATGCCCCATCCGGACACCCCTTTTGG CTATTTTACACCATTGTTCTTTCCA*	615
<i>neurod1</i>	TTTGGGCTCTTTCGGAAATCTTG AGACCCGCTGCCTGATAGTG*	469
<i>wnt1</i>	CCCAAATAACAAGTCAGC AAGCCCTCCCTATTTACCAC*	947
<i>ptch1</i>	ATACCAAGTGTTGCATGGTG AGCTCATTAACGGCAAGAG*	638
<i>zic1</i>	GAAAATCTGAAAATCCACAAAA GGTGCTGTGGCTGCTCGTGT*	365
*	T3 promoter	sequence

(GGATCCATTAACCCTCACTAAAGGGAA) is added to the 5'-end of the reverse primer for transcription of the gene *in vitro*.

Table S2 PCR primers for quantitative real-time PCR.

Gene	Primer
<i>β-actin</i>	CGGTTTTGCTGGAGATGATGC TGGGGTATTTGAGGGTCAGGA
<i>arl13b</i>	GGAGAGAACCTGCAAGGAAGG GATTCCTCTGACTGTCGCCG
<i>atoh1a</i>	CCCAGGCAAAATATCCGTCCCT

CAAGGCTGGTGGGTACTCGCT

atoh1b CCTCAAGCTGCCTCTGAAGA
GGGCCATCTGAAGAGTGTC

atoh1c CCCGAGAGAGGAGGAGGATG
GGGTATGGGGTTTCTGGGTC

neurod1 CGAGCAGAGCCAGGAGAT
AGGGTGGTGTCAAAGAACG

vglut1 CTGTGGAGGGTTTGGAAATGGA
GCGATGTCAAGATGGTTCACA

gabra6b TCTTGTGTTGGACAGGTGGTG
AGGTCGGAGTCTGTTGTCATATC

ptf1a TCGATCTCACATCCCAACA
CCGTAGTCTGGGTCATTTGGA

rora TGGCCCAGAACATTTCCAAGT
TGGCAAACCTCCACCACATACT

pvalb7 ATCCTGAGTGAGACCGAGCT
TGTGGTCGAAACTGTCAGCA

calb2a CCTTACCTTCACTTGGCGGA
GCTCCTCTCCTCGCAATCTC

aldoca GCGTCCCGTTTGTCAAGATG
GGCAAAGTCAGCTCCATCCT

gdf7 AACACGGTCACCAGCTTCAT

GCAGCTCCACTTCCATCAGT

msx2a AGGAGCGAGTCAACTTCACG
CAAGAGCTGAGCAGTGGTGA

msx2b AGAAGGCGAGAGGATGTGTCCAG
GATTGGTCTTGTGCTTGCGGAG

wnt1 ATGTGTCCTCCTGGTGTCTCTC
CTGGCGGCGACTTAGCAGTG

wnt3 AGCAGCAGCAAACCTTGGAG
CCCGAGAGCTGGAGAACCAC

wnt3a CTCCGCTTCTGCCGCAACTAC
CTGATGCTGACACTCCTGGATGC

gli2a CAGCACCAGCAGCAGCACTG
CCTTGGACCGCTTGAAGATCATGG

gli2b GCGGTCAGCAGCACAGTCAAC
ACAGTCGTCCATGTCCTCCTTCAG

gli3 TTGCCATGGACCCTCGTTCT
GGTACTGGCGAGGGTTCGTA

ptch1 TGCGCTGGAGCAGATATCAGAGG
TCCACGTCCGTCTCCAGATTAGC

smo TGGCATTGCTTTGTCGGCT
CAGCCCTGGATGGTTGCTTTTA
