

Appendix

Table I. Primers and probes sequences used for respiratory viral targets.

Target Primers and Probe set	Sequence	Sequence Reference and supplier
<u>FLU A</u>		
11A forward	5'-AAGACCAATCCTGTCACCTCTGA-3'	World Health Organisation, 2007. SA
11B reverse	5'-CAAAGCGTCTACGCTGCAGTCC-3'	
11P probe	5'-FAM-TTTGTGTTACGCTACCGTGCC-BHQ-3'	
<u>FLU A H1</u>		
16A forward	5'-GGAATAGCCCCCCTACAATTG-3'	Personal communication. Anon.
16B reverse	5'-AATTCGCATTCTGGGTTCTA-3'	HPA, Colindale. SOP
16P probe	5'-FAM-CGTTGCCGGATGGA-MGBNFQ-3'	V-5413. LT
<u>FLU A H3</u>		Personal communication. Anon.
17A forward	5'-CCTTTTGTGAAACGCAGCAA-3'	HPA, Colindale. SOP
17B reverse	5'-CGGATGAGGCAACTAGTGACCTA-3'	V-5413. LT
17P probe	5'-FAM-CCTACAGCAACTGTTACC-MGBNFQ-3	Health Protection Agency (2009).
<u>FLU A H5</u>		LT
9A forward	5'-GGATGGCAGGGAATGGTAGA-3'	
9B reverse	5'-TCTATTGCCTTTGAGTGGATTCT T-3'	
9P probe	5'-FAM-TGGGTACCACCATAGCAAYGAGCAGG-MGBNFQ-3'	
<u>FLU B</u>		Van Elden, L J R et al, 2001. SA
4A forward	5'-AAATACGGTGGATTAAATAAAAAGCAA-3'	
4B reverse	5'-CCAGCAATAGCTCCGAAGAAA-3'	
4P probe	FAM-5'-CACCCATATTGGCAATTCCATATGGC-3'-TAMRA	
<u>RSV A</u>		Gunson, R N et al, 2005. SA
4A forward	5'-AGATCAACTCTGTCATCCAGCAA-3'	
4B reverse	5'-TTCTGCACATCATAATTAGGAG TATCAAT-3'	
4P probe	5'-FAM-CACCATCCAACGGAGCACAGGAGAT-BHQ-3'	
<u>RSV B</u>		
5A forward	5'-AAGATGCAAATCATAAATTCACAGGA-3'	Personal Communication, Anon.
5B reverse	5'-TGATATCCAGCATCTTCAGTATCTTATA-3'	Specialist Virology Centre, Gartnavel. SA
5P1 probe	5'-FAM-TTCCCTCCTAACCTGGACATAGCATATAACATACCT-BHQ-3'	
<u>MPV 1A</u>		Kuypers, J et al. 2005. LT
3A forward	5'-GCCGTTAGCTTCAGTCAATTCAA-3'	
3B reverse	5'-TCCAGCATTGTCTGAAAATTGC-3'	
3P probe	5'-FAM-CAACATTAGAACCTTCT-MGBNFQ-3'	
<u>MPV 2B</u>		Kuypers, J et al. 2005. LT
3A forward	5'-GCCGTTAGCTTCAGTCAATTCAA-3'	
3B reverse	5'-TCCAGCATTGTCTGAAAATTGC-3'	
4P probe	5'-FAM-CGCACACATTAGGAATCTTCT-MGBNFQ-3'	
<u>PF1</u>		Personal Communication, Anon.
2A forward	5'-GTGATTAAACCCGTAATTCTCA-3'	Specialist Virology Centre, Gartnavel. LT
2B reverse	5'-CCTGTTCCCTGCAGCTATTACAAGA-3'	
2P probe	5'-FAM-ACCTATGACATCAACGAC-MGBNFQ-3'	
<u>PF2</u>		Personal Communication, Anon.
2A forward	5'-ATGAAAACCATTACCTAACGTGATGGA-3'	Specialist Virology Centre, Gartnavel. LT
2B reverse	5'-CCTCCYGGTATRGCAGTGACTGAAC-3'	
2P probe	5'-FAM-TCAATCGCAAAAGC-MGBNFQ-3'	

PF3		
3A forward	5'-CCAGGGATATAYTAYAAAGGCAAA-3'	Gunson, R N et al, 2005. SA
3B reverse	5'-CCGGGRCACCCAGTTGTG-3'	
3P probe	5'-FAM-TGGRTGTTCAAGACCTCCATAYCCGAGAAA-BHQ-3'	
COV 229E		
6A forward	5'-CAGTCAAATGGGCTGATGCA-3'	Gunson, R N et al, 2005. SA
6B reverse	5'-AAAGGGCTATAAAGAGAATAAGGTATTCT-3'	
6P probe	5'-FAM-CCCTGACGACCACGTTGTGGTTCA-BHQ-3'	
COV OC43		
7A forward	5'-CGATGAGGCTATTCCGACTAGGT-3'	Gunson, R N et al, 2005. SA
7B reverse	5'-CCTCCTGAGCCTTAATATAGTAACC-3'	
7P probe	5'-FAM-TCCGCCTGGCACGGTACTCCCT-BHQ-3'	
COV NL63		
8A forward	5'-ACGTACTTCTATTATGAAGCATGATATTAA-3'	Gunson, R N et al, 2005. SA
8B reverse	5'-AGCAGATCTAATGTTACTTAAACTACG-3'	
8P1 probe	5'-FAM-ATTGCCAAGGCTCCTAACGTACAGGTGTT-BHQ-3'	
COV HKU1		
9A forward	5'-TTGAAGGCTCAGGAAGGTCTGCT-3'	Raymond, F et al, 2009. LT
9B reverse	5'-TGCTTAGTKACTGCTGAGGTTAG -3'	
9P probe	5'-6FAM-TAAAACAAGATTAGCGATCTC-MGBNFQ-3'	
ADV		
3A forward	5'-GCCACGGTGGGTTCTAAACTT-3'	Heim, A et al, 2003. SA
3B reverse	5'-GCCCGAGTGGTCTTACATGCACATC-3'	
3P probe	5'-FAM-TGCACCAGMCCSGGGCTCAGGTACTCCGA-BHQ-3'	
HRV 1		
7A forward	5'-AGCCTGCGTGGCTGCCTG-3'	In-house development. Regional Virus
7A2 forward	5'-CCTGCGTGGCGGCCARC-3'	Laboratory, Belfast.
7B reverse	5'-CCCAAAGTAGTYGGTCCCRTCC-3'	LT
7P probe	5'-FAM-TCCCTCCGGCYCCTGAATGYGGCTAA-3'-MGBNFQ	
HRV 2		
8A forward	5'-GACARGGTGTGAAGAGCC-3'	Scheltinga, S A et al, 2005. SA
8B forward	5'-GACATGGTGTGAAGACYC-3'	
8C reverse	5'-CAAAGTAGTYGGTCCCATCC-3	
8P probe	5'-FAM-TCCCTCCGGCCCTGAATGYGGCTAA-3'-BHQ1	
BOV		
2A forward	5'-GGAAGAGACACTGGCAGACAA-3'	Allander, T et al, 2007. SA
2B reverse	5'-GGGTGTTCTGATGATATGAGC-3'	
2P probe	5'-FAM-CTGCGGCTCCTGCTCCTGTGAT-BHQ-3'	
RNP		
1A forward	5'-AGATTGGACCTGCGAGCG-3'	World Health Organisation, 2009.
1B reverse	5'-GAGCGGCTGTCTCCACAAGT-3'	SA
1P probe	5'-6FAM-TTCTGACCTGAAGGCTCTGCGCG-BHQ-3'	

For each target in the test panel, the forward and reverse primers, probe sequences, reporters and quenchers are presented, along with their initial source references and supplier details.

Abbreviations: "ADV: adenovirus; BHQ: Blackhole Quencher; BOV: bocavirus; FAM: 6-carboxyfluorescein (reporter molecule); COV: coronavirus; FLU A: influenza A; FLU B: influenza B; LT: Life Technologies (Paisley, UK); MGBNFQ: minor groove binding nonfluorescent quencher; MPV: metapneumovirus; PF:

parainfluenza; RHV: rhinovirus; RNP: Ribonuclease P; RSV: Respiratory Syncytial Virus; SA: Sigma-Aldrich Company Ltd(Dorset, UK); TAMRA: Carboxytetramethylrhodamine”.

Table 1. List of reagents used in the preparation of respiratory viral-related amplification mixes.

Each provides instruction for the preparation of 1000 tests when reaction volume used is 8 μ l. All volumes shown are μ l.

All RNA target TaqMan master mixes were prepared using SuperScript® III Platinum® One-Step Quantitative RT-PCR System (Life Technologies, Paisley, UK).

Influenza A Matrix

Nuclease free water	1580
Reaction mix x2	5000
BSA @ 2 μ g/ μ l (final 0.2 μ g/ μ l)	1000
MgSO ₄ @ 50mM (final 4mM)	160
FLA 11A @ 200 μ M (final 0.4 μ M)	20
FLA 11B @ 200 μ M (final 0.4 μ M)	20
FLA 11P @ 100 μ M (final 0.2 μ M) FAM-BHQ	20
Superscript III RT/Platinum® <i>Taq</i> Mix	200

Influenza A H1(seasonal)

NFW	1580
Reaction mix x2	5000
BSA @ 2 μ g/ μ l (final 0.2 μ g/ μ l)	1000
MgSO ₄ @ 50mM (final 4mM)	160
FLA 16A @ 200 μ M (final 0.4 μ M)	20
FLA 16B @ 200 μ M (final 0.4 μ M)	20
FLA 16P @ 100 μ M (final 0.2 μ M) FAM-TAMRA	20
Superscript III RT/Platinum® <i>Taq</i> Mix	200

Influenza A H3

NFW	1580
Reaction Mix x2	5000
BSA @ 2 μ g/ μ l (final 0.2 μ g/ μ l)	1000
MgSO ₄ @ 50mM (final 4mM)	160
FLA 17A @ 200 μ M (final 0.4 μ M)	20
FLA 17B @ 200 μ M (final 0.4 μ M)	20
FLA 17P @ 100 μ M (final 0.2 μ M) FAM-TAMRA	20
Superscript III RT/Platinum® <i>Taq</i> Mix	200

Influenza A H5

NFW	1580
Reaction Mix x2	5000
BSA @ 2 μ g/ μ l (final 0.2 μ g/ μ l)	1000
MgSO ₄ @ 50mM (final 4mM)	160
FLA 9A @ 200 μ M (final 0.4 μ M)	20
FLA 9B @ 200 μ M (final 0.4 μ M)	20
FLA 9P @ 100 μ M (final 0.2 μ M) FAM-MGBNFQ	20
Superscript III RT/Platinum® <i>Taq</i> Mix	200

TaqMan PCR: Influenza B

NFW	1580
Reaction Mix x2	5000
BSA @ 2 μ g/ μ l (final 0.2 μ g/ μ l)	1000
MgSO ₄ @ 50mM (final 4mM)	160

FLB 4A @ 200µM (final 0.4µM)	20
FLB 4B @ 200µM (final 0.4µM)	20
FLB 4P @ 100µM (final 0.2µM) FAM-TAMRA	20
Superscript III RT/Platinum® <i>Taq</i> Mix	200

TaqMan PCR: RSV Types A&B

NFW	1680
Reaction Mix x2	5000
BSA @ 2µg/µl (final 0.2µg/µl)	1000
RSV 4A @ 200µM (final 0.4µM)	20
RSV 4B @ 200µM (final 0.4µM)	20
RSV 4P @ 100µM (final 0.2µM) FAM-BHQ	20
RSV 5A @ 200µM (final 0.4µM)	20
RSV 5B @ 200µM (final 0.4µM)	20
RSV 5P1 @ 100µM (final 0.2µM) FAM-BHQ	20
Superscript III RT/Platinum® <i>Taq</i> Mix	200

TaqMan PCR: Metapneumovirus

NFW	1520
Reaction Mix x2	5000
BSA @ 2µg/µl (final 0.2µg/µl)	1000
MgSO ₄ @ 50mM (final 4mM)	160
MPV 3A @ 200µM (final 0.4µM)	20
MPV 3B @ 200µM (final 0.4µM)	20
MPV 3P @ 100µM (final 0.2µM) FAM-MGB	20
MPV 3A @ 200µM (final 0.4µM)	20
MPV 4B @ 200µM (final 0.4µM)	20
MPV 4P @ 100µM (final 0.2µM) FAM-MGBNFQ	20
Superscript III RT/Platinum® <i>Taq</i> Mix	200

TaqMan PCR: Rhinovirus1

NFW	1560
Reaction mix x2	5000
BSA @ 2µg/µl (final 0.2µg/µl)	1000
MgSO ₄ @ 50mM (final 4mM)	160
HRV 7A @ 200µM (final 0.4µM)	20
HRV 7A2 @ 200µM (final 0.4µM)	20
HRV 7B @ 200µM (final 0.4µM)	20
HRV 7P @ 100µM (final 0.2µM) FAM-MGB	20
Superscript III RT/Platinum® <i>Taq</i> Mix	200

TaqMan PCR: Rhinovirus2

NFW	1660
Reaction mix x2	5000
BSA @ 2µg/µl (final 0.2µg/µl)	1000
MgSO ₄ @ 50mM (final 3.5mM)	80
HRV 8A @ 200µM (final 0.2µM)	10
HRV 8B @ 200µM (final 0.2µM)	10
HRV 8C @ 200µM (final 0.4µM)	20
HRV 8P @ 100µM (final 0.2µM) FAM-BHQ	20

Superscript III RT/Platinum® <i>Taq</i> Mix	200
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RNP

NFW	1700
Reaction mix x2	5000
BSA @ 2µg/µl (final 0.2µg/µl)	1000
RNP 1A @ 200µM (final 0.8µM)	40
RNP 1B @ 200µM (final 0.8µM)	40
RNP 1P @ 100µM (0.2µM) FAM-BHQ	20
Superscript III RT/Platinum® <i>Taq</i> Mix	200

TaqMan PCR: Single Mix: Parainfluenza 1

NFW	1580
Reaction mix x2	5000
BSA @ 2µg/µl (final 0.2µg/µl)	1000
MgSO ₄ @ 50mM (final 4mM)	160
PF1 2A @ 200µM (final 0.4µM)	20
PF1 2B @ 200µM (final 0.4µM)	20
PF1 2P @ 100µM (final 0.2µM) FAM-MGBNFQ	20
Superscript III RT/Platinum® <i>Taq</i> Mix	200

TaqMan PCR: Single Mix: Parainfluenza 2

NFW	1580
Reaction mix x2	5000
BSA @ 2µg/µl (final 0.2µg/µl)	1000
MgSO ₄ @ 50mM (final 4mM)	160
PF2 2A @ 200µM (final 0.4µM)	20
PF2 2B @ 200µM (final 0.4µM)	20
PF2 2P @ 100µM (final 0.2µM) FAM-MGBNFQ	20
Superscript III RT/Platinum® <i>Taq</i> Mix	200

TaqMan PCR: Single Mix: Parainfluenza 3

NFW	1580
Reaction Mix x2	5000
BSA @ 2µg/µl (final 0.2µg/µl)	1000
MgSO ₄ @ 50mM (final 4mM)	160
PF3 3A @ 200µM (final 0.4µM)	20
PF3 3B @ 200µM (final 0.4µM)	20
PF3 3P @ 100µM (final 0.2µM) FAM-BHQ	20
Superscript III RT/Platinum® <i>Taq</i> Mix	200

TaqMan PCR: Parainfluenza MULTIPLEX

NFW	1460
Buffer @ x2	5000
BSA @ 2µg/µl (final 0.2µg/µl)	1000
MgSO ₄ @ 50m (final 4mM)	160
PF1 2A @ 200µM (final 0.4µM)	20
PF1 2B @ 200µM (final 0.4µM)	20
PF1 2P @ 100µM (final 0.2µM) FAM-MGBNFQ	20
PF2 2A @ 200µM (final 0.4µM)	20

PF2 2B @ 200µM (final 0.4µM)	20
PF2 2P @ 100µM (final 0.2µM) FAM-MGBNFQ	20
PF3 3A @ 200µM (final 0.4µM)	20
PF3 3B @ 200µM (final 0.4µM)	20
PF3 3P @ 100µM (final 0.2µM) FAM-BHQ	20
Superscript III RT/Platinum® Taq Mix	200

TaqMan PCR: Coronavirus 229E, OC43, NL63and HKU1 MULTIPLEX

NFW	1400
Buffer @ x2	5000
BSA @ 2µg/µl (final 0.2µg/µl)	1000
MgSO ₄ @ 50m (final 4mM)	160
COV 6A @ 200µM (final 0.4µM)	20
COV 6B @ 200µM (final 0.4µM)	20
COV 6P @ 100µM (final 0.2µM) FAM-MGBNFQ	20
COV 7A @ 200µM (final 0.4µM)	20
COV 7B @ 200µM (final 0.4µM)	20
COV 7P @ 100µM (final 0.2µM) FAM-MGBNFQ	20
COV 8A @ 200µM (final 0.4µM)	20
COV 8B @ 200µM (final 0.4µM)	20
COV 8P1@ 100µM (final 0.2µM) FAM-BHQ	20
COV 9A @ 200µM (final 0.4µM)	20
COV 9B @ 200µM (final 0.4µM)	20
COV 9P@ 100µM (final 0.2µM) FAM-MGB	20
Superscript III RT/Platinum® Taq Mix	200

TaqMan PCR: Single Mix: Coronavirus 229E

NFW	1580
Buffer @ x2	5000
BSA @ 2µg/µl (final 0.2µg/µl)	1000
MgSO ₄ @ 50m (final 4mM)	160
COV 6A @ 200µM (final 0.4µM)	20
COV 6B @ 200µM (final 0.4µM)	20
COV 6P @ 100µM (final 0.2µM) FAM-MGBNFQ	20
Superscript III RT/Platinum® Taq Mix	200

TaqMan PCR: Single Mix: Coronavirus OC43

NFW	1580
Buffer @ x2	5000
BSA @ 2µg/µl (final 0.2µg/µl)	1000
MgSO ₄ @ 50m (final 4mM)	160
COV 7A @ 200µM (final 0.4µM)	20
COV 7B @ 200µM (final 0.4µM)	20
COV 7P @ 100µM (final 0.2µM) FAM-MGBNFQ	20
Superscript III RT/Platinum® Taq Mix	200

TaqMan PCR: Single Mix: Coronavirus NL63

NFW	1580
Buffer @ x2	5000
BSA @ 2µg/µl (final 0.2µg/µl)	1000
MgSO ₄ @ 50m (final 4mM)	160
COV 8A @ 200µM (final 0.4µM)	20
COV 8B @ 200µM (final 0.4µM)	20

COV 8P1 @ 100µM (final 0.2µM) FAM-BHQ1	20
Superscript III RT/Platinum® Taq Mix	200

Taqman PCR: Single Mix: Coronavirus HKU1

NFW	1580
Buffer @ x2	5000
BSA @ 2µg/µl (final 0.2µg/µl)	1000
MgSO ₄ @ 50m (final 4mM)	160
COV 9A @ 200µM (final 0.4µM)	20
COV 9B @ 200µM (final 0.4µM)	20
COV 9P @ 100µM (final 0.2µM) FAM-BHQ1	20
Superscript III RT/Platinum® Taq Mix	200

All DNA target TaqMan amplification mixes were prepared with Platinum® Quantitative PCR SuperMix-UDG (Life Technologies, Paisley, UK).

TaqMan PCR: Bocavirus DNA

NFW	1740
Platinum® qPCR Supermix-UDG	5000
BSA @ 2µg/µl (final 0.2µg/µl)	1000
MgCl ₂ 50mM (final 4mM)	200
BOV 2A @ 200µM (final 0.4µM)	20
BOV 2B @ 200µM (final 0.4µM)	20
BOV 2P @ 100µM (final 0.2µM) FAM-TAMRA	20

TaqMan PCR: Adenovirus DNA

NFW	1740
Platinum® qPCR Supermix-UDG	5000
BSA @ 2µg/µl (final 0.2µg/µl)	1000
MgCl ₂ @ 50mM (final 4mM)	200
ADV 3A @ 200µM (final 0.4µM)	20
ADV 3B @ 200µM (final 0.4µM)	20
ADV 3P @ 100µM (final 0.2µM) FAM-BHQ	20

Abbreviations: "ADV: adenovirus; BHQ: Blackhole Quencher; BOV: bocavirus; BSA: Bovine serum albumin; COV: coronavirus; FAM: 6-carboxyfluorescein; FLU A: influenza A; FLU B: influenza B; MGBNFQ: minor groove binding nonfluorescent quencher; MgCl₂: Magnesium chloride; MgSO₄: Magnesium sulphate; MPV: metapneumovirus; NFW: nuclease free water; PF: parainfluenza; RV: rhinovirus; RSV: Respiratory Syncytial Virus; TAMRA: Carboxytetramethylrhodamine.

Table 3. Primers used in reverse-tagging step (PCR 2).

Name	Sequence	Length
806R_f1	GTGAaGGAGHcAGAcGTGTGCTCCGATCTN NNNNACGGACTACHV GGG TW TCTAAT	61
806R_f2	GTGACTGGAGUCAGACGTGTGCTCUCCGATCTNNTNNNACGGACTACHV GGGTWTCTAA	62
806R_f3	GTGAaGGAGUcAGACGTGTGamCCGATCT NNCTNN NAC GGACTACHV GCGTWTCTAA	63
806R_f4	GTGACTGGAGUCAGACGTGTGCTmCCGATCTNNACTNNNACGGACTACHV GGGTWTCTAA	64
806R_f5	GTGACTGGAGTrCAGACGTGTGCTTrCCGATCTNNGAaNNNACGGAaACHW GGGTWTCTAA	65
806R_f6	GTGACTGGAGTTCAGACGTGTGCTCTCCGATCTNNGACTNNNACGGACTACHV GGGTWTCTAA	66

Table 4. Primers for forward-tagging step (PCR 3).

Name	Sequence	Length
515F_f1	GCCTCCCTCGGCCATCAGAGATGTG TATAAGAGACAG NNNN NNNNGA GTGCCA GCMGCCGCGGTAA	67
515F_f2	GCCTCCCTCGGCCATCAGAGATGTGTATAAGAGACAG NNNNNNNN GAGTGCCA GCM GCCGCGGTAA	68
515F_f3	GCCTCCCTCGGCCATCAGAGATGTG TATAAGAGACAG NNNN T NNNN GA GTGCCAGCMGCCGCGGTAA GCCTCCCTCGGCCATCAGAGATGTG	69
515F_f4	TATAAGAGACAG NNNN CT NNNN GAGTGCCAGCMGCCGC GGTAAGCCTCCCTC GCGC C ATCAGAGATGTG TATAAGAGACAG NNNN ACT NNNN GA	70
515F_f5	GTGCCAGCMGCCGCGGTAA GCCTCCCTCGGCCATC AGAGATGTG TATAAGA GACAG NNNN GACT NNNN GA	71
515F_f6	GTGCCAGCMGCCGCGGTAAAGCCTCCCTCGGCCATCAGAGATGTG TATAAGAG ACAG NNNNTGACT NNNN GA GTGCCAGCMGCCGCGGTAA	72