Supplementary Information

Intratumoral Generation of Photothermal Gold Nanoparticles through a Vectorized Biomineralization of Ionic Gold

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Supplementary Figure 1. Characterization of ionic Au-PEG cluster: (A) UV-Vis absorbance spectra of ionic Au-PEG clusters (200-800 nm) as well as it's individual components at identical

concentrations. TEM images from the extracted solution of ionic Au-PEG treatments to cell culture plates with a magnification of Au-PEG cluster (B), and fresh cell culture media incubated with Au-PEG clusters (C). Additional TEM images of Au-PEG treated MCF-7 cells at various magnifications (D). Enhanced hyperspectral darkfield image of untreated MCF-7 cells used for negative filtering for Figure 2A and 2B (E).



Supplementary Figure 2. Raman spectra of cellular fractions (all from MCF-7 cells): Nucleus and large organelles, 15,000 g (Red); mitochondria, lysosomes, and other medium sized organelles, 100,000 g (Green); membrane fragments, 300,000 g (Blue); and highly soluble/cytosolic molecules (Violet), >300,000 g; and the glass slide without cell fractions (Black).



Supplementary Figure 3. Confirmation of intracellular reduction by SDS-PAGE: SDS-PAGE stained with Coomassie Blue of cellular fractions including (A) nuclear, 15,000 *g*; (B) medium organelle, 100,000 *g*; (C) membrane fragments, 300,000 *g*; and (D) cytosolic, >300,000 *g* regions of MCF-7 cells untreated, treated with Na-PEG, or Au-PEG with either standard SDS-PAGE conditions [(+) β ME], or without β -mercaptoethanol [(∞) β ME]. Arrows indicate direction of protein migration due to electrophoresis.



Supplementary Figure 4. Schematic for separation of Au bound proteins from the non-bounded ones: The process describing the separation of Au-bound proteins from non-bound ones through gel electrophoresis, followed by separation of portions of the gel for LC-MS analysis.



Supplementary Table 1. Proteins identified from the LC-MS analysis and the subcellular regions in which they are located, as annotated on UniProt.org through COMPARTMENTS.¹

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Protein Name	protein binding	hydrolase activity	metal ion binding	protein kinase binding	calcium ion binding	actin filament binding	antigen binding	ATP binding	B cell receptor signaling pathway	carbohydrat e metabolic process	catalytic activity	cation binding	cell adhesion	complemen t activation, classical pathway	defense response to bacterium	immune response	immunoglob ulin receptor binding	muscle contraction	neutrophil degranulati on	nuk b
Zymogen granule protein 16 homolog B																				
Secretoglobin family 1D member 2																				
Immunoglobulin lambda-like polypeptide 5 Alpha-amulase 2B																				
Immunoglobulin heavy constant alpha 1																				
1,4-alpha-glucan-branching enzyme Cadherin-23																				
SRC kinase signaling inhibitor 1 Integrin beta-1-binding protein 1																				
JNK1MAPK8-associated membrane protein																				
Ganglioside GM2 activator Calmodulin-1 Titin																				
Protein FRG1 Probable ATP-dependent RNA helicase																				
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Protein Name	4- hydroxyproli ne metabolic	actin binding	actinin binding	activation of adenylate cyclase activity	activation of protein kinase B activity	adenylate cyclase activator activity	adenylate cyclase binding	alpha- amylase activity	angiogenes is	antioxidant activity	beta-N- acetylgalac tosaminidas e activity	beta-N- acetylhexos aminidase activity	biomineral tissue developme nt	calcium channel inhibitor activity	calcium ion transport	calcium- dependent cell-cell adhesion	calcium- dependent protein binding	calcium- mediated signaling	calmodulin binding	cart e t
Zymogen granule protein 16 homolog B																				
Secretoglobin family 1D member 2																				
Immunoglobulin lambda-like polypeptide 5																				
Immunoglobulin heavy constant alpha 1																				
1,4-alpha-glucan-branching enzyme																				
SRC kinase signaling inhibitor 1 Integrin beta-1-binding protein 1																				
Peroxiredoxin-4 JNK1MAPK8-associated																				
Ganglioside GM2 activator Calmodulin-1																				
Titin Protein FRG1 Probable ATP-dependent RNA																				
helicase																				

			GO Term																
Protein Name	carbohydrat e binding	cardiac muscle contraction	cardiac musole fiber developme nt	oardiac musole hypertrophy	oardiao muscle tissue morphogen	cardiac myofibril assembly	cell differentiati on	cell redox homeostasi s	cellular oxidant detoxificatio n	oellular response to fibroblast growth	cellular response to vascular endothelial	cofactor metabolic process	detection of caloium ion	detection of muscle stretch	disordered domain specific binding	enzyme activator activity	enzyme binding	equilibrioce ption	establishme esta nt of protein nt o localization loc- to
Zymogen granule protein 16 homolog B																			
Secretoglobin family 1D member 2																			
Immunoglobulin lambda-like polypeptide 5																			
Alpha-amylase 25 Immunoglobulin heavy constant alpha 1																			
1,4-alpha-glucan-branching enzyme Cadhario-23																			
SRC kinase signaling inhibitor 1 Integrin beta-1-binding protein 1																			
Peroxiredoxin-4 JNK1/MAPK8-associated membrane protein																			
Ganglioside GM2 activator Calmodulin-1																			
Itin Protein FRG1 Probable ATP-dependent RNA helicase																			

Supplementary Table 2. Proteins identified from the LC-MS analysis and the molecular processes in which they are involved, as annotated on UniProt.org through phylogenetic-

based propagation of functional gene ontology (GO) annotations.²

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										G	o rer	m								
Protein Name	establishme nt of protein localization to	exocytosis	extracellular matrix organizatio n	Fo-epsilon receptor signaling pathway	focal adhesion	G protein- coupled receptor signaling	ganglioside catabolic process	ganglioside metabolic process	GDP- dissociation inhibitor activity	generation of precursor metabolites and energy	glomerular filtration	glycogen biosynthetic process	glycogen metabolic process	glycosphing olipid metabolic process	helicase activity	homophilic cell adhesion via plasma	hydrogen peroxide catabolic process	identical protein binding	I-kappaB phosphoryl ation	in s pr
Zymogen granule protein 16 homolog B																				
Secretoglobin family 1D member 2																				
Immunoglobulin lambda-like polypeptide 5																				
Immunoglobulin heavy constant alpha 1																				
1,4-alpha-glucan-branching enzyme																				
SRC kinase signaling inhibitor 1																				
Integrin beta-1-binding protein 1 Peroxiredoxin-4																				
JNK1/MAPK8-associated membrane protein																				
Ganglioside GM2 activator Calmodulin-1																				
Titin Protein FRG1 Probable ATP-dependent RNA																				
helicase																				_







Supplementary Table 2 contd. Proteins identified from the LC-MS analysis and the molecular processes in which they are involved, as annotated on UniProt.org through phylogenetic-based propagation of functional gene ontology (GO) annotations.²

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Protein Name	Ras guanyl- nucleotide exchange factor	reactive oxygen species metabolic	receptor clustering	receptor- mediated endocytosis	regulation by host of symbiont cAMP-	regulation of blood vessel size	regulation of cardiac muscle contraction	regulation of cardiac muscle contraction	regulation of catalytic activity	regulation of cell adhesion involved in	regulation of cell adhesion mediated by	regulation of cell communica tion by	regulation of cell division	regulation of cell migration involved in	regulation of cell proliferation	regulation of cyclic- nucleotide phosphodie	regulation of cytokinesis	regulation of cytosolic calcium ion concentrati	regulation of dendritic spine morphogen	reç enc
Zymogen granule protein 16 homolog B																				
Secretoglobin family 1D member 2																				
Immunoglobulin lambda-like polypeptide 5 Aloba-amulase 28																				
Immunoglobulin heavy constant alpha 1																				
1,4-alpha-glucan-branching enzyme Cadhetin-23																				
SRC kinase signaling inhibitor 1 Integrin beta-1-binding protein 1																				
Peroxiredoxin-4 JNK1/MAPK8-associated membrane protein																				
Ganglioside GM2 activator Calmodulin-1																				
Titin Protein FRG1 Probable ATP-dependent RNA helioase																				



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Protein Name	sensory perception of sound	skeletal musole thin filament assembly	spermatoge nesis	sphingolipid activator protein activity	sphingolipid metabolic process	striated muscle myosin thick filament	structural constituent of muscle	structural molecule activity conferring	substantia nigra developme nt	substrate adhesion- dependent cell	telethonin binding	thioredoxin peroxidase activity	titin binding	tube formation	type 3 metabotropi c glutamate receptor	ubiquitin protein ligase binding	ubiquitin- dependent ERAD pathway	visual perception	Wnt signaling pathway, calcium	
Zymogen granule protein 16 homolog B																				
Secretoglobin family 1D member 2																				
Immunoglobulin lambda-like polypeptide 5																				
Alpha-amylase 2B Immunoolobulin heavy constant																				
alpha 1																				
enzyme																				
SRC kinase signaling inhibitor 1																				
Integrin beta-1-binding protein 1 Peroxiredoxin-4																				
JNK1/MAPK8-associated membrane protein																				
Ganglioside GM2 activator Calmodulin-1																				
Titin Protein FRG1																				
Probable ATP-dependent RNA helioase																				

Supplementary Table 2 contd. Proteins identified from the LC-MS analysis and the molecular processes in which they are involved, as annotated on UniProt.org through phylogenetic-based propagation of functional gene ontology (GO) annotations.²

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Protein Name	sarcomerog enesis	sensory perception of light stimulus	sensory perception of sound	skeletal muscle thin filament assembly	spermatoge nesis	sphingolipid activator protein activity	sphingolipid metabolic process	striated muscle myosin thick filament	structural constituent of muscle	structural molecule activity conferring	substantia nigra developme nt	substrate adhesion- dependent cell	telethonin binding	thioredoxin peroxidase activity	titin binding	tube formation	type 3 metabotropi c glutamate receptor	ubiquitin protein ligase binding	ubiquitin- dependent ERAD pathway	per
Zymogen granule protein 16 homolog B																				
Secretoglobin family 1D member 2																				
Immunoglobulin lambda-like polypeptide 5																				
Alpha-amylase 2B																				
alpha 1																				
enzyme																				
SRC kinase signaling inhibitor 1																				
Peroxiredoxin-4																				
JNK1/MAPK8-associated membrane protein																				
Ganglioside GM2 activator Calmodulin-1																				
Titin Protein FRG1																				
Probable ATP-dependent RNA helicase																				



Supplementary Table 2 contd. Proteins identified from the LC-MS analysis and the molecular processes in which they are involved, as annotated on UniProt.org through phylogenetic-based propagation of functional gene ontology (GO) annotations.²



Supplementary Figure 5. Microarray based analysis for apoptosis related KEGG pathways for ionic gold treatment: Log-Fold-Change of gene regulation represented through apoptosis related KEGG pathways quantified through microarray analysis comparing Au-PEG treatments against untreated MCF-7 cells.



Supplementary Figure 6. Investigation of gold internalization pathway: Percentage of gold internalization in MCF-7 cells via application of ionic Au-PEG clusters at 0.24 mM Au³⁺(control), or ionic Au-PEG clusters in presence of several endocytic inhibitors is shown. Different endocytic inhibitors employed in this study were sodium azide (NaN₃), 2-deoxyglucose, nystatin (Nys), chlorpromazine (CPM), and dynasore. Error bars are standard deviations of the mean (n=3 biologically independent samples).



Supplementary Figure 7. Microarray based analysis for endocytosis related KEGG pathways for ionic gold treatment: Log-Fold-Change of gene regulation represented through endocytosis related KEGG pathways quantified through microarray analysis comparing Au-PEG treatments against untreated MCF-7 cells.



Supplementary Figure 8. Schematic set up for the photothermal ablation therapy: Computeraided design (CAD) image of laser mount for photothermal ablative therapy (to scale with RTLMRL-635-500-5 Roithner Lasertechnik cw laser), with top, side, and front views (A). Image of complete setup (B). Image of laser setup with positioned mouse (C).



Supplementary Figure 9. Schematic diagram of nanoparticle formation: Artistic representation of how applications of ionic Au-PEG clusters will induce nanoparticle formation through successful intratumoral or intradermal applications with and without photothermal ablation.



Supplementary Figure 10. *In vivo* efficacy of the intratumorally generated gold nanoparticles: (A) Fluorescence contrast afforded via IVIS imaging ($430_{Ex} 840_{Em}$) between on-site and off-site across one week period of treatments for Au-PEG with no photothermal treatment with intratumoral injections (Yellow), Au-PEG with intradermal injections (Orange), or Na-PEG (Blue). Error bars are standard deviation of the mean (n \ge 3 biologically independent animals). *The 1 day Au-PEG (-Laser) / Off site (Intradermal Injection) sample represents 2 biologically independent animals. Bright field (B) and fluorescence images (C) representitive of mice with Au-PEG with intradermal injection (three days post injection) with arrows indicating injection sites for either Na-PEG (Blue) or Au-PEG (Red). Temperature maximums averaged across laser treatments (D) as determined through FLIR IR thermal imaging with representitive thermal image of laser treatments made to mice with Au-PEG with intradermal injection (E). Error bars in for D are standard deviations of the mean (n \ge 5 laser treatments).



Supplementary Figure 11. Raman mapping of gold nanoparticles: Illustration of how peak intensities for regions 1250-1350, 1446-1477, and 2849-2969 cm⁻¹ (A) were used for generating Blue, Green, and Red color maps (B) and were then combined for Raman mapping of unstained histological sections (C).



Supplementary Figure 12. Evaluating *in vivo* toxicity: Representitive H&E stained tissue sections after microtomy from animals having received Au-PEG \pm and/or laser treatments \pm with Au-PEG. Scale bars are 100 µm.

Supplementary References.

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- Gaudet, P., Livstone, M. S., Lewis, S. E. & Thomas, P. D. Phylogenetic-based propagation of functional annotations within the Gene Ontology consortium. *Brief. Bioinform.* 12, 449–62 (2011).