

**A**

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P_GE (-) : UCAAUGUUUUUUU
F_GE (-) : UCAA-UUAUUUUU
NS1_GE (-) : UCAAUUUAUUUUU
G_GE (-) : UCAAUGAAUUUUU
N_GE (-) : UCAAUUUUUUUUU
M_GE (-) : UCAAUUUUUUUUU
SH_GE (-) : UCAAUUAAUUUUU
L_GE (-) : UCAA-UAAUUUUU
NS2_GE (-) : UCAU-UAAAUUUU
M2_GE (-) : UCAA-UAAAUUUU
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Negative-sense gene-end (GE) sequences

**B**

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P_GE (+) : AGUUACAAAAAAA
F_GE (+) : AGUU-AUAUAAAA
NS1_GE (+) : AGUUAAUAUAAAA
G_GE (+) : AGUUACUUAAAAA
N_GE (+) : AGUUAAUAAAAAA
M_GE (+) : AGUUAAUAAAAAA
SH_GE (+) : AGUUAAUUAAAAA
L_GE (+) : AGUU-AUUAAAAA
NS2_GE (+) : AGUA-AUUUAAAA
M2_GE (+) : AGUU-AUUUAAAA
          ***          *****
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Positive-sense gene-end (GE) sequences

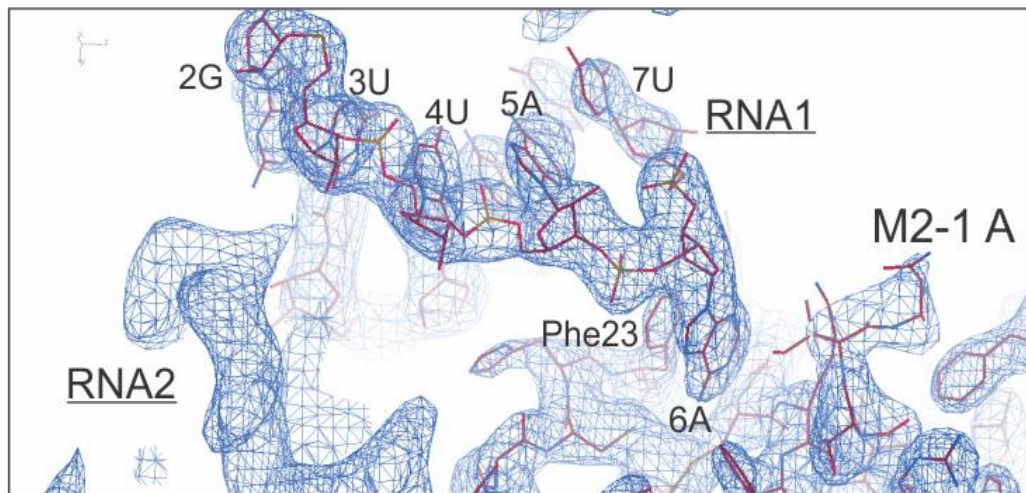
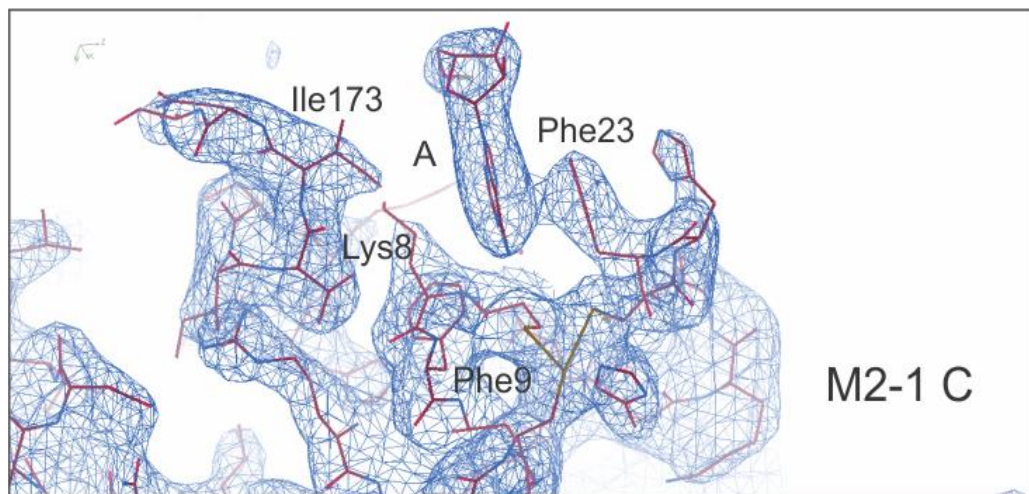
**Figure S1: The sequence alignments of negative-sense and positive-sense gene-end (GE) of the HRSV genes. Related to STAR Methods.**

- A. The sequence alignment of negative sense GE of HRSV genes, namely, P, F, NS1, G, N, M, SH, L, NS2, and M2. \* denotes the conserved positions among GE sequences.
- B. The sequence alignment of positive sense GE of HRSV genes: P, F, NS1, G, N, M, SH, L, NS2, and M2.



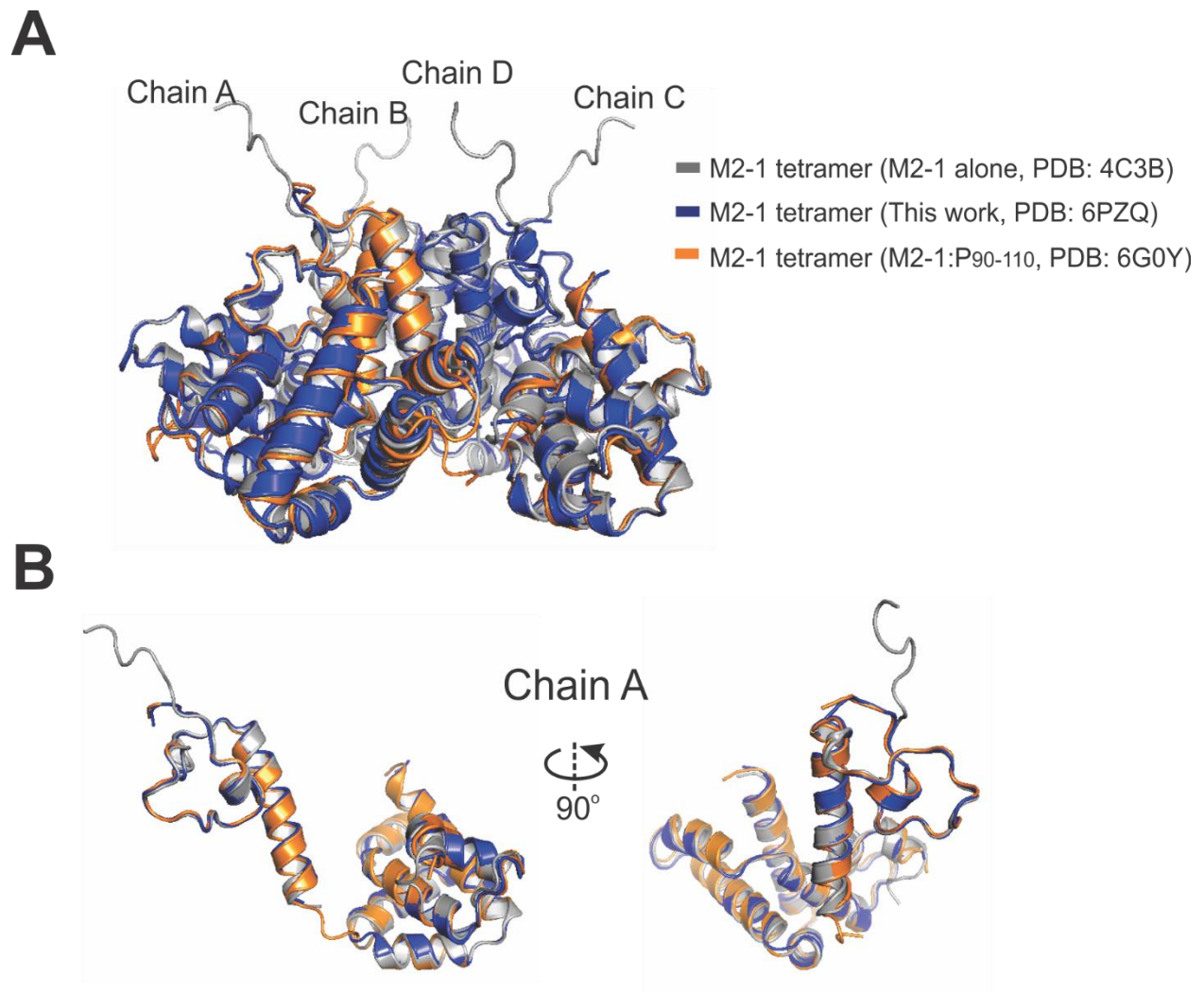
**Figure S2: Multiple sequence alignments of the M2-1 proteins from *Pneumoviridae*. Related to Figures 2 and 4.**

- Multiple sequence alignments of the M2-1 proteins from RSV (ORSV, BRSV, A2 and B1 strains of HRSV) and MPV (A, C, and D strains of AMPV and HMPV). The conservation is highlighted in the degree of the blue color, most conservation with dark blue and least conservation with light blue.
- The Phylogenetic tree of M2-1 from RSV and MPV families. The scores are shown next to each subfamily. A2 and B1 strains of HRSV are distal from the rest of families, including ORSV and BRSV.

**A****B**

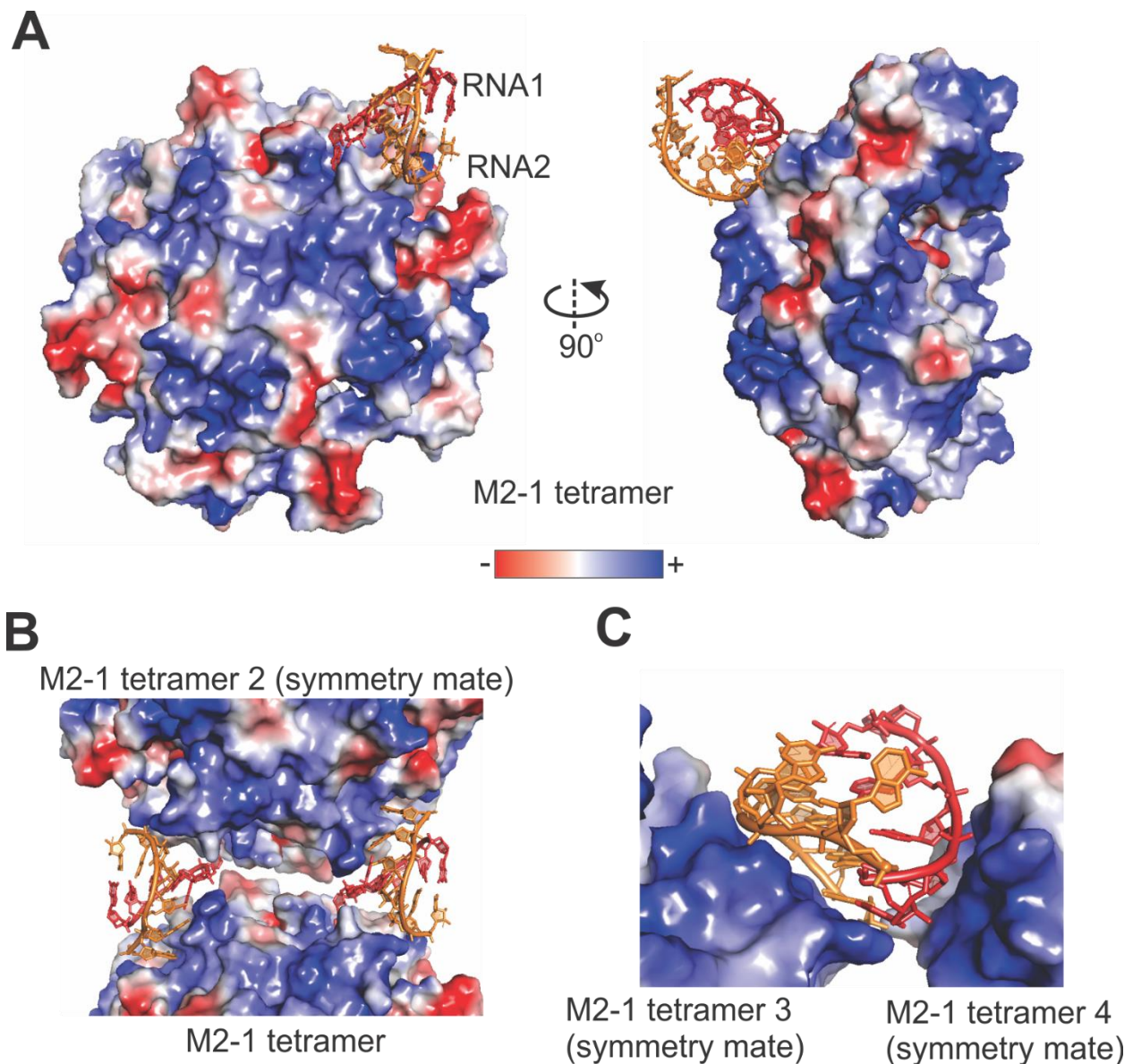
**Figure S3: The representative density maps used for building RNA. Related to Figures 1 and 2.**

- A. The representative electron density  $2F_{\text{obs}} - F_{\text{calc}}$  map (contoured at  $1.5\sigma$ ) shows the electron density of the RNA molecule adjacent to M2-1 protomer A. The meshes show the electron density of the modeled RNA and M2-1.
- B. The representative electron density  $2F_{\text{obs}} - F_{\text{calc}}$  map (contoured at  $1.5\sigma$ ) shows the electron density of the molecule adjacent to M2-1 protomer C. The meshes show the electron density of the modeled RNA and M2-1.



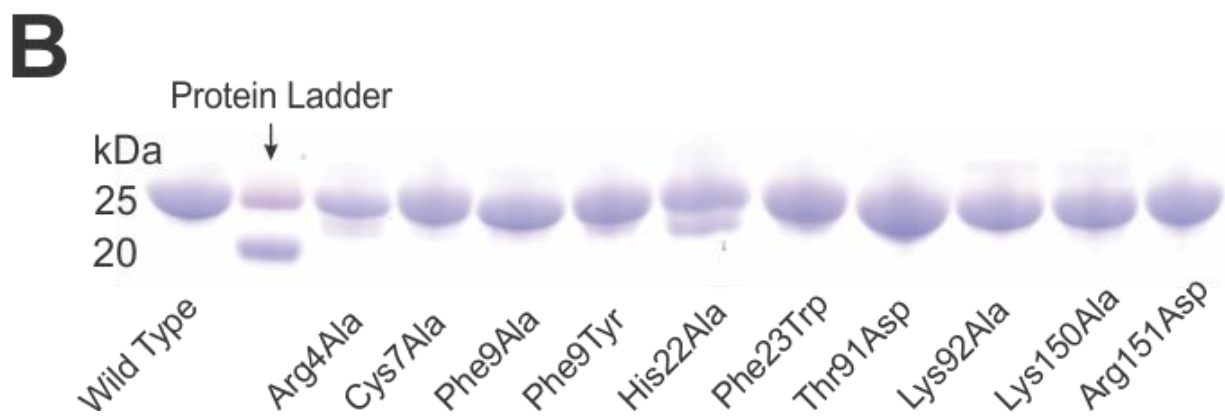
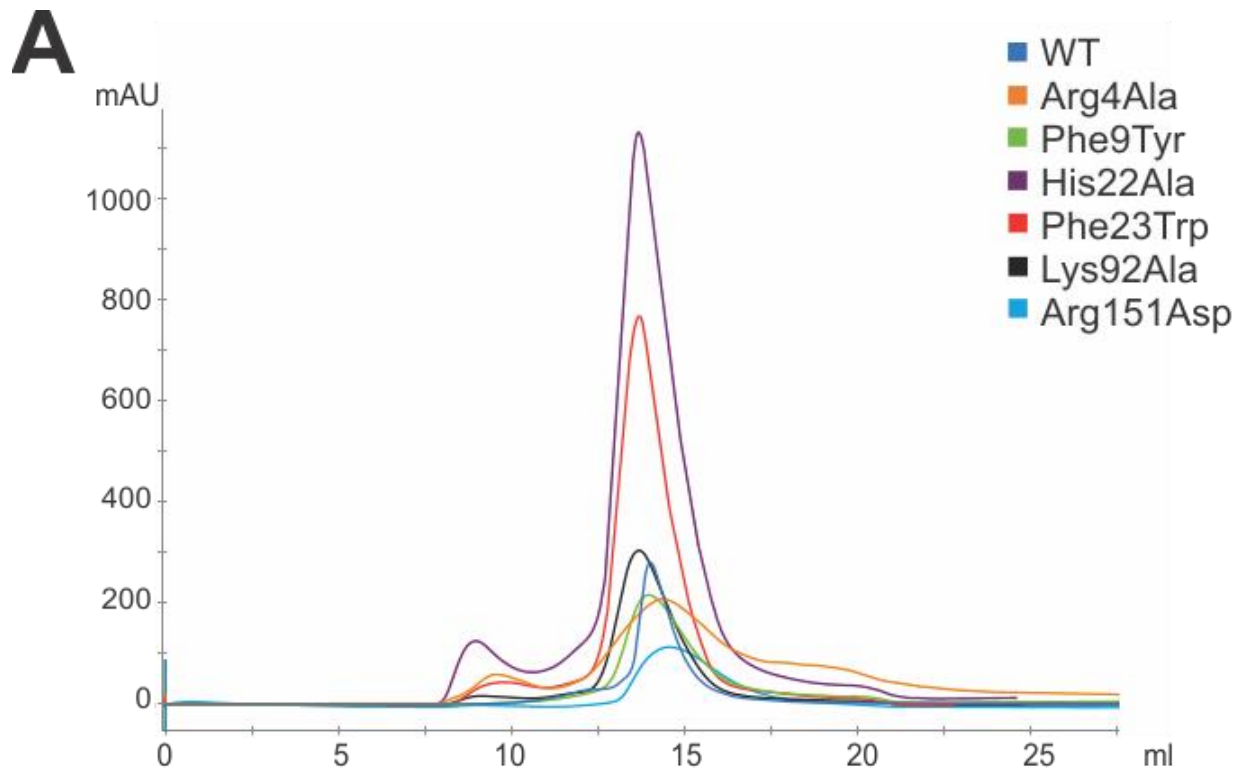
**Figure S4: Superimposition of M2-1 of M2-1:SH7, M2-1 alone, and M2-1:P<sub>90-110</sub>. Related to Figure 1.**

- A. Superimposition of M2-1 of M2-1:SH7 (blue), M2-1 alone (gray, PDB: 4C3B) and M2-1:P90-110 (orange, PDB: 6G0Y).
- B. Superimposition of the chain A of M2-1 from M2-1:SH7, M2-1 alone, and M2-1:P<sub>90-110</sub>. The RMSD are < 0.5 Å.



**Figure S5: The electrostatic surface of the M2-1:SH7 complex. Related to Figures 1, 3, and 4.**

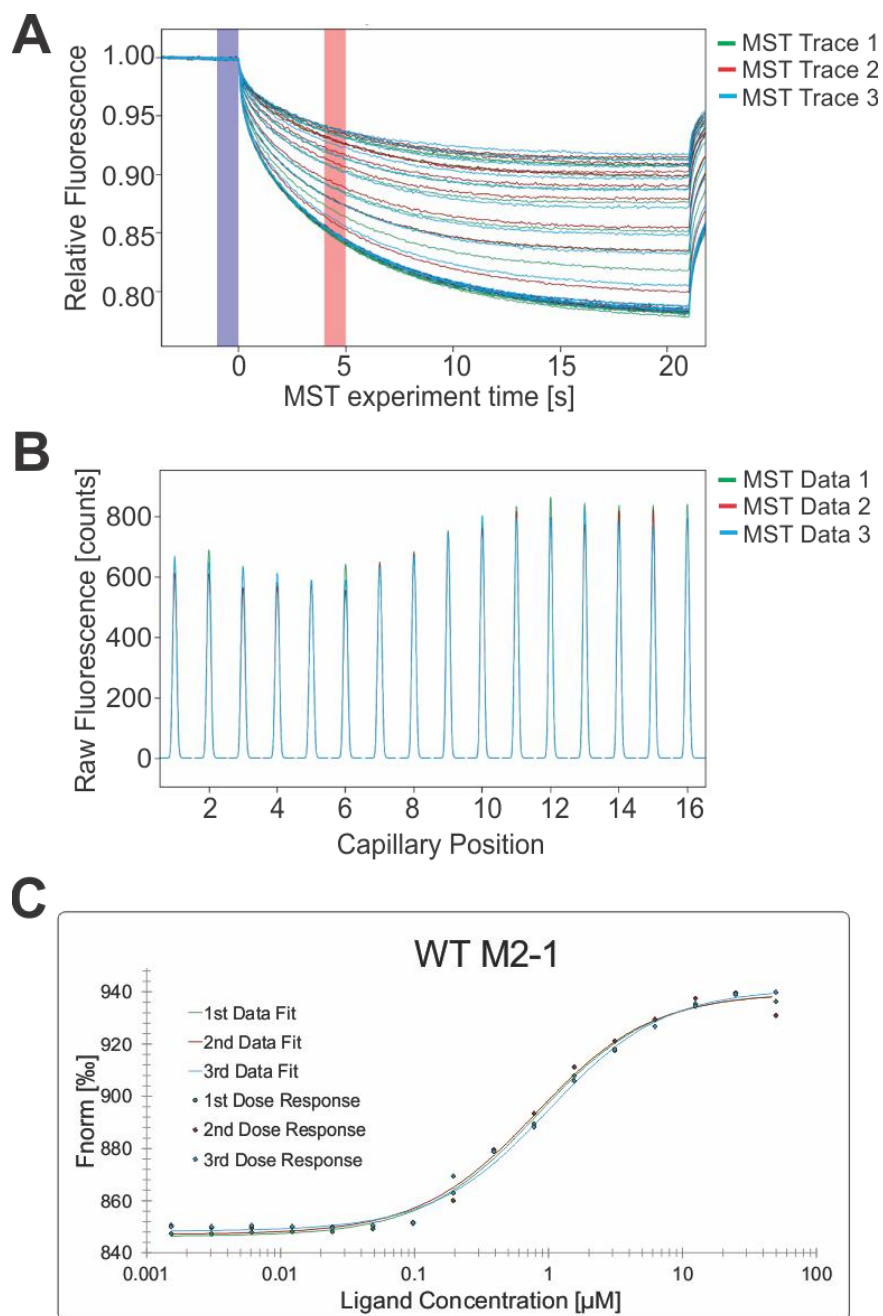
- A. Surface representation of the electrostatic potential of the M2-1 in the front view and side view. The negative charge is in red, and the positive charge is in blue (calculated using APBS).
- B. The zoom-in view of the interactions between the zinc-binding domain (ZBD) of two M2-1 tetramers (the upper is the symmetry mate 2) and RNAs.
- C. The zoom-in view of the interactions between the core domain (CD) of two M2-1 tetramers (both are symmetry mates 3 and 4) and RNAs.



**Figure S6: The wild-type and mutant HRSV M2-1 proteins. Related to Figures 2 and 4.**

A. The representative size exclusion chromatography profiles of wild-type (WT) and mutant HRSV M2-1 proteins, including Arg4Ala, Phe9Tyr, His22Ala, Phe23Trp, Lys92Ala, and Arg151Asp.

B. The SDS-PAGE gel of WT and mutant HRSV M2-1 proteins.



**Figure S7: Affinity analysis of HRSV M2-1 to RNA using MicroScale Thermophoresis (MST) assay. Related to Figures 2 and 4. The data were acquired with a triplicate.**

- A. The representative MicroScale Thermophoresis (MST) experiments show the traces for 16 multiple scans compared using relative fluorescence for WT M2-1 proteins.
- B. The MST data show the raw fluorescence counts for the individual capillary position (a total of 16 capillaries).
- C. The  $K_d$  fitting of the MST data measurements.

M2-1 Proteins	Kd ( $\mu\text{M}$ )	Kd ( $\mu\text{M}$ ) Error	Average ( $\mu\text{M}$ )
Wild Type	0.819	0.070	0.867 $\pm$ 0.090
	0.789	0.095	
	0.993	0.075	
Arg4Ala	2.026	0.149	1.760 $\pm$ 0.208
	1.520	0.119	
	1.732	0.131	
Cys7Ala	9.602	1.234	8.56 $\pm$ 1.018
	7.180	1.013	
	8.907	1.132	
Phe9Ala	5.085	0.383	5.693 $\pm$ 0.511
	6.337	0.369	
	5.659	0.363	
Phe9Tyr	0.739	0.011	0.813 $\pm$ 0.087
	0.766	0.113	
	0.936	0.123	
His22Ala	1.737	0.333	1.592 $\pm$ 0.104
	1.542	0.232	
	1.497	0.215	
Phe23Trp	0.726	1.043	0.711 $\pm$ 0.019
	0.72	1.058	
	0.68	9.132	
Thr91Asp	1.54	0.164	1.716 $\pm$ 0.210
	2.01	0.137	
	1.60	0.158	
Lys92Ala	5.01	0.596	5.175 $\pm$ 0.810
	6.24	0.801	
	4.27	0.917	
Lys150Ala	0.79	0.092	1.111 $\pm$ 0.238
	1.37	0.098	
	1.17	0.096	
Arg151Asp	1.03	0.067	1.212 $\pm$ 0.331
	0.93	0.098	
	1.68	0.125	

**Table S1: The MST measurements of the interactions between M2-1 and SH7 RNA. Related to Figures 2 and 4.** All data were measured as triplicates through NT.115 MST instrument at room temperature.



M2-1_Arg4Ala_F:	TCCAATGCAATGAGCAGGgcGAACCCCT
M2-1_Arg4Ala_R:	CTCGAACTTACAGGGGTTcgcCCTGCT
M2-1_Cys7Ala_F:	ATGAGCAGGAGGAACCCCGCTAAGTTC
M2-1_Cys7Ala_R:	GCCCCTGATCTCGAACTTAGCGGGGTT
M2-1_Phe9Ala_F:	AGGAGGAACCCCTGTAAGgcCGAGATC
M2-1_Phe9Ala_R:	GCAGTGGCCCCTGATCTCGgcCTTACA
M2-1_Phe9Tyr_F:	AACCCCTGTAAGTaCGAGATCAGGGGCCAC
M2-1_Phe9Tyr_R:	GCCCCTGATCTCGtACTTACAGGGGTTCT
M2-1_His22Ala_F:	CTGAACGGCAAGAGGTGCGCCTTCAGC
M2-1_His22Ala_R:	GAAGTAGTTGTGGCTGAAGGCGCACCT
M2-1_Phe23Trp_F:	AAGAGGTGCCActggAGCCACA ACTACTTC
M2-1_Phe23Trp_R:	GTAGTTGTGGCTccaGTGGCACCTCTTGCC
M2-1_Thr91Asp_F:	GGCAGCATCAACAACATCgaCAAGCAG
M2-1_Thr91Asp_R:	CACACAGGCGCTCTGCTTGtcGATGTT
M2-1_Lys92Ala_F:	AACAACATCACCGgcGCAGAGCGCCTGTGTG
M2-1_Lys92Ala_R:	ACAGGCGCTCTGCgcGGTGATGTTGTTGAT
M2-1_Lys150Ala_F:	CAGACCATCCACCTGCTGgcGAGGCTG
M2-1_Lys150Ala_R:	CACGTTCGCGGGCAGCCTCgcCAGCAG
M2-1_Arg151Asp_F:	CACCTGCTGAAGgacCTGCCCGCCGACGTG
M2-1_Arg151Asp_R:	GTCGGCGGGCAGgtcCTTCAGCAGGTGGAT

**Table S2: The primers used in the mutagenesis of the M2-1 proteins. Related to STAR Methods.** The primers are synthesized by Eurofins Genomics.