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Supplementary Materials for

Tailoring of the axon initial segment shapes the conversion of synaptic inputs into spiking output in OFF-α T retinal ganglion cells

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Figs. S1 to S8 Table S1



Fig. S1. Identification of OFF-\alpha T RGCs. (a) Tracings of representative OFF- α T (left) and OFF- α S (right) RGCs. Axons are indicated in red. Scale bar: 100 µm. (b) Light responses of a typical OFF- α T (green) and OFF- α S (black) RGCs. The stimulus was a bright spot 300 µm in diameter on a grey background and was centered over the soma. (c) Distribution of fluorescent voxels of OFF- α T (green) and OFF- α S (black) dendrites relative to the ON and OFF ChAT bands (red peaks). All three traces are normalized to their peaks.



Fig. S2. Passive properties are similar in ventral and dorsal OFF-α T RGCs – (a, left) Each line is the best-fit linear regression for ΔV in response to hyperpolarizing current injections for an individual ventral (red, n=18) or dorsal (blue, n=18) RGC. The slope of each line was used as a measurement for input resistance. (a, right) Mean input resistance was similar in ventral and dorsal cells (44.2 ± 0.4 MΩ vs. 43.0 ± 0.5 MΩ, p=0.6487). (b, left) Each line is the best-fit linear regression for ΔV in response to hyperpolarizing current injections for an individual axotomized ventral (red, n=6) or dorsal (blue, n=3) RGC. The slope of each line was used as a measurement for input resistance. (b, right) Mean input resistance was similar in intact vs. axotomized cells (43.6 ± 0.4 MΩ vs. 45.7 ± 0.8 MΩ, p=0.4494). (c) Each point is a plot of peak firing rate (left) or sustained firing rate (right) vs. input resistance for an individual ventral (red, n=18) or dorsal (blue, n=18) RGC. The grey lines are the best-fit linear regressions (p=0.5397 and p=0.6820, respectively).



Fig. S3. Action potential dynamics are similar in ventral and dorsal OFF- α T RGCs – (a) Overlay of spontaneous spikes from OFF- α T RGCs in the ventral (red) and dorsal (blue) retina. Inset shows the same action potentials on a larger time scale. (b) Phase plots, i.e. time-derivative of membrane potential vs. membrane potential, for the spikes in (a). (c) Comparison of the resting membrane potential (V_{Rest}, -60.5 ± 1.3 mV vs. -59.1 ± 1.0 mV, p=0.4107), the maximum amplitude of depolarization (Δ DepolMax, 56.5 ± 1.4 mV vs. 59.0 ± 1.4 mV, p=0.1917) and the maximum rates during de- (Peak+ Δ V/ Δ t, 386 ± 19 V/s vs. 413 ± 18 V/s, p=0.3038) and repolarization (Peak- Δ V/ Δ t, -241 ± 14 V/s vs. -255 ± 12 V/s, p=0.4311) between action potentials of ventral (red, n=20) and dorsal (blue, n=20) cells.



Fig. S4. Dendritic field sizes are different in ventral vs. dorsal OFF-a T RGCs. (a)

Reconstructions of 8 ventral (top) and 8 dorsal (bottom) OFF- α T RGCs. Axons are indicated in red. Scale bar: 100 µm. (**b**) Soma diameter was compared between ventral (red, n=21) and dorsal (blue, n=22) cells (18.0 ± 0.4 µm vs. 18.5 ± 0.4 µm, p=0.4018). (**c**) Scatter plot of total dendritic length vs. dendritic field diameter in ventral (red, n=8) and dorsal (blue, n=8) reveals a linear correlation (p=0.0005). Total dendritic length was significantly larger in dorsal cells (p=0.0357). (**d**) Scatter plot of dendritic field diameter vs. mean segment length (p=0.0001). Mean segment length was significantly larger in dorsal cells (p=0.0357). (**d**) Scatter plot of dendritic field diameter vs. mean segment length (p=0.0001). Mean segment length was significantly larger in dorsal cells (p=0.0005). (**e**) Sholl analysis of ventral (red, n=15) and dorsal (blue, n=14) cells (Two-way repeated measures ANOVA: interaction p=0.1164, subgroups p=0.9851).



Fig. S5. AIS anatomy scales with cell size. Scatter plots of AIS length (**a**) and AIS distance (**b**) vs. dendritic field diameter for dorsal (blue) and ventral (red) RGCs. The lines are best-fit linear regressions (p=0.0001 and p=0.0001, respectively).



Fig. S6. Nav1.6 channels contribute significantly to spiking responses in OFF- α T RGCs. (a) Average amount of overlap between Nav1.6 and Ankyring staining in ventral (red, n=5) and dorsal (blue, n=9) RGCs (p=0.4969). Error bars represent one standard deviation. (b) Spontaneous firing rate under control conditions (C), during the application of aTTX and after washout of aTTX (W). (c) The response to visual stimulation under control and during the application of aTTX. Scale bar: 100 ms / 10 mV. The inset shows an expanded time scale of the transient burst during application of aTTX. Scale bar: 10 ms / 10 mV. (d) Onset rapidness (black) and threshold depolarization (green) under control (C), during the application of aTTX and after wash.



Fig. S7. Responses from axotomized RGCs – (**a**) Representative current clamp recordings from axotomized ventral (red) and dorsal (blue) OFF- α T RGCs when the axotomy was located between the soma and the AIS. (**b**) Representative current clamp recordings from axotomized ventral (red) and dorsal (blue) OFF- α T RGCs when the axotomy was located distal of the AIS. Pulse duration was 400 ms (grey shading), amplitudes ranged from 100-800 pA. Scale bar: 100 ms / 20 mV. Schematics on top indicate location of axotomy. Horizontal arrows indicate failure threshold.



Fig. S8. Simulated RGC responses to somatic current injections – (a) Representative current clamp recordings from a modeled OFF- α T RGC. Pulse duration was 400 ms (grey shading), amplitudes ranged from 100-1000 pA. Scale bar: 50 ms / 20 mV. Horizontal arrow indicates failure threshold. (b) Membrane potential over time showing one action potential during current injection into the soma. (c) Phase plot, i.e. time-derivative of membrane potential vs. membrane potential, for the spike in (b). (d) The number of elicited spikes is plotted vs. injected current amplitude for the model cell.

	Dendrites	Soma	Soma-AIS	AIS	Axon
gNav1.1	65	65	130	0	70
g Kv1.1	35	35	70	0	70
g Nav1.6	0	0	0	325	0
g Kv1.6	0	0	0	175	0
g Ca	1.5	1.5	1.5	1.5	1.5
g K,Ca	0.15	0.15	0.15	0.15	0.2
gl	0.1	0.1	0.1	0.1	0.1

Table S1. Ion channel densities along the membrane of model neurons. Values are based on Fohlmeister et al. *(39)* with minor modifications. All values are given in mS/cm₂.