

Supplementary Information for

Global variability in seawater Mg:Ca and Sr:Ca ratios in the modern ocean

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Materials and Methods S1

Global sampling program

Sampling efforts from 2009 until 2017 combined ongoing research cruises, official programs and time-series with citizen science initiatives, recreational boats and fellow scientific volunteers to reach globally to as many remote locations and ecosystems as possible (Fig. 1; SI Appendix 1, 3, 4). Agreements were established between the PI and all participants to collect seawater samples following a strict protocol (see SI Appendix Text S1) along with the sample metadata (latitude, longitude, temperature, salinity, depths, and CTD data). Sampling was organized either by direct contact with existing projects/surveys/programs/oceanographic lines/buoys or by posting announcements in group emails/lists and institutions around the world. We accessed samples in a total of 79 programs/cruises/initiatives worldwide, and we were directly involved/present in at least 15 cruises. Sampling effort covered all possible locations at low/temperate/high latitudes, open ocean, and coastal regions with emphasis on the euphotic (0-300 m), mesopelagic (300-1000 m), and abyssal zones (+1000 m) using transects, stations and vertical CTD profiles. The baseline strategy was to sample horizontally (transects) and vertically (CTD rosette) from coastal regions to the open ocean to monitor variability at large, expanding also to the shelves, the neritic zones, river mouths, the intertidal/subtidal and the ice sheets. Number of samples and locations were prearranged with participants but a large number of expeditions changed their plans, thus sampling occurred on opportunity (see SI Appendix Fig. S3 for examples). All participants were provided exactly with the same sampling kit to maximize sampling homogeneity, which consisted in 15/50 ml sterile tubes, 50 ml sterile syringes, 0.20 um sterile filters (to retain particles and bacteria). plastic gloves, and *Parafilm*® (see SI Appendix Text S1 for details).

Seawater sampling procedure

All research cruises had a Niskin-type bottle, a CTD rosette array or a scientific-grade sampling device available to recover seawater samples at surface and depth. Other participants (surface samples) used a bucket or directly collected the sample with a plastic tube. Right after sampling, all samples were gently filtered through a 0.22 µm sterile filter using a sterile syringe and stored into 15/50 ml polyethylene tubes with *Parafilm*® wrapped around the cap, preventing thus contamination and evaporation. All samples were stored in the fridge $(4 \text{ to } 8 \degree C)$ and sent for analysis to the ICP-MS lab at Institute of Geosciences, Christian-Albrechts-Universität zu Kiel, Germany, between 6 to 12 months after collection. After 1 year in storage in a cold place, no

signs of seawater evaporation or leaks were observed $\ll 0.05\%$ Mg/Ca and Sr/Ca ratios variability $n = 30$ (1). All our samples were measured within 12 months after collection to guarantee data quality and comparability, minimizing any % change over time.

Seawater analytical procedures

In the laboratory at Institute of Geosciences, Kiel University, Germany, samples were divided into three splits (A, B, C) and only split A was analyzed. Splits B and C were stored as a back-up in cases of analytical bias, or re-sampling/verification. All split A samples were analyzed in the same laboratory and machine by the same person, for Ca, Mg, Sr with truly simultaneous data acquisition using an inductively coupled plasma – optical emission spectrometer (ICP-OES) SPECTRO CirosTM CCD SOP with radial viewing optics. Samples were 50-fold diluted with 2% (v/v) freshly subboiled in HNO₃ and introduced with a GE SeasprayTM micro-nebulizer and thermostatized CinnabarTM cyclonic spray chamber. All sample preparation work, and storage of sample solutions in the auto-sampler during analysis, was performed under Class-100 clean bench conditions by the same person. Only all-plastic lab ware was used for handling and treatment of the samples, avoiding storage in polypropylene mini-vials having elevated Mg. Only ultrapure reagents and ultrapure water (Elga Labwater) were used. The following emission lines were selected for quantification: Ca: 317.933 (II); Mg: 279.553nm (II); Sr: 407.771nm (II), 421.552nm (II) and measured simultaneously within the same pre-selected acquisition interval ("Phase 3"). In addition, Ar: 597.159nm (I) was used as a monitor line for plasma temperature. Three pixels were summarized for peak area integration of the respective emission line (Smart Analyzer 3.30 Software, Spectro A.I.). Total analysis time per sample with 5 replicate measurements including sample take-up and washout was roughly 6 minutes consuming about 2 ml of measuring solution. A combined intensity calibration and drift correction procedure using IAPSO standards (*2, 3*) was applied for data processing (SI Appendix 4). Calibration solutions were matrix matched with NaCl to 50-fold diluted normal seawater salinity covering the range of seawater Mg:Ca 4.10-5.80 (mol:mol), and Sr:Ca 7.70-8.80 (mmol:mol). Raw data (in cps) was exported from the instrument into spreadsheet software for post-processing. Average raw counts from 5 runs for *Blank-00* (blank for Ca) and *Blank-0* (matrix-matched with 10 mg l^{-1} Ca as a blank for Mg, Sr) were subtracted from all subsequent measurements. From blank-subtracted raw intensities, ratios were calculated for all combinations of selected Mg and Sr wavelengths with the selected Ca line in order to determine the most robust wavelength pairs. A linear least-square regression function was applied for the calculation of molar ratios (in mmol:mol) from intensity ratios of the calibration standards (*2*). Then, the obtained results were normalized to an external standard (IAPSO, ORIL, U.K. using $Mg:Ca = 5.140$ mol:mol, $Sr:Ca = 8.481$ mmol:mol) in a way commonly used in isotope studies. This approach minimizes variability and uncertainties, respectively, resulting from both instrument drift during the day and instrument set-up between different days. Every batch of 6 samples was bracketed by the IAPSO normalization standard that was also used for linear drift correction. For estimating short term reproducibility every $11th$ sample was reanalyzed, and so were the first 5 samples from a session reanalyzed at the end of the session with e.g. 352 samples. Mean uncertainty estimated from duplicate measurements per sample on 33 randomly chosen samples was 0.35 and 0.85 % relative SD (1 SD) for seawater Mg:Ca and Sr:Ca while measurement uncertainty (5 runs) was 0.16 and 0.37 % RSD, respectively (SI Appendix 4). The expanded uncertainties including uncertainty of the true values for IAPSO are 3.30 % for seawater Sr:Ca and 0.5 % for seawater Mg:Ca. Uncertainty as estimated from measurement reproducibility that is important for detecting small differences even in our open ocean sample suite, was well below 0.40 % for seawater Mg:Ca and 0.90 % for seawater Sr:Ca. When averaging only the first three measurements for our IAPSO reference sample right after initial calibration of all three analytical sessions where the samples were run, then the measured

IAPSO average was $Sr:Ca = 8.481 \pm 0.068$ mmol:mol. Comparing data routinely obtained for multi-element analyses of seawater, then we find measured IAPSO averages ranging from Sr:Ca $= 8.273$ to 8.555 mmol:mol. It is always the Ca that is making the difference: we measure around 409 mg L⁻¹ Ca (10.20 mM Ca) while ODP-TAMU reported 422 mg L⁻¹, 10.55 mM Ca. Our Sr results are identical with ODP_TAMU: 7.62 mg L^{-1} Sr (86.97 μ M Sr).

Literature data and information use

Our new measured dataset was complemented with a seawater Mg:Ca and Sr:Ca ratios literature survey (Fig. 4c for comparison with this study data; SI Appendix 2) to expand on the global horizontal and vertical coverage, to consolidate and compare with the work of earlier researchers at a regional scale. Hundreds of publications and reports were screened for data quality and analytical clarity to mine data comparable to our measured dataset (analytical methods varied from ICP-AES/-OES to gravimetric titration and to TIMS). The methods section for each publication/report were also carefully inspected for analytical accuracy. We also put together a table on the historical use of seawater Mg:Ca and Sr:Ca ratios in all marine science disciplines in the last 100 years (Table S3). This served to put in perspective the significance of the new dataset results for the scientific community to foresee if the implications apply to a certain discipline.

Public datasets use

The newly measured seawater Mg:Ca and Sr:Ca ratios dataset (SI Appendix 3) was combined with seawater environmental metadata mined from GLODAPv2 (*4, 5*), and used to analyze and classify seawater Mg:Ca and Sr:Ca ratios as a function of environmental variables (SI Appendix 3). ArcGIS 10.0 (ESRI 2011) and the 3-D analytical tool "NEAR 3-D analysis" were used to link environmental seawater conditions with the sample's seawater ratios. This approach permitted evaluation of the diagonal distance between each ratio input parameter (latitude, longitude, and depth) and the nearest seawater datum (temperature, salinity, Total Alkalinity - TA, dissolved inorganic carbon - DIC, pH_{total} and $pCO₂$) at a specific latitude, longitude, and depth. The prerequisite for data adoption was a matching data set XYZ (including ratio parameter) versus $X_iY_iZ_i$ (including ambient seawater property). Both datasets were plotted on a 2-D field and merged into a single 3-D field, and then the nearest seawater datum to the ratio was determined using a diagonal line in a 3-D matrix. Additionally, each datum was individually checked to verify that the NEAR 3-D approach worked correctly. When the prediction was not sufficiently accurate, the closest value was manually selected. This procedure ensured selection of the most relevant seawater parameters for a given ratio. The GEMS-GLORI database was used to provide a distribution map of seawater Mg:Ca ratios from river mouths/plumes around the world (SI Appendix 6; SI Appendix Fig. S1), but these data were not used in the overall analysis, and are reported merely for comparison purposes. Lastly, various paleo proxies' global databases were mined to provide distribution maps of paleo reconstructions and understand the relevance of our database results for the scientific community (SI Appendix Fig. S2). Our published databases are deposited at the NOAA National Center for Environmental Information (NCEI) under Accession Number 0171017 in<http://accession.nodc.noaa.gov/0171017> (DOI:10.7289/V5571996).

Data analysis and statistics

The final seawater Mg:Ca and Sr:Ca ratios were classified by Ocean, Longhurst Province, Ecosystem type, Latitude, Longitude, Depth (in 5 intervals to facilitate analyses) (SI Appendix 3) and then by all the metadata (chemistry) to facilitate interpretation of results and statistical analyses. Ratios were mapped globally (Fig. 1; SI Appendix S1), versus. Depth, and by ecosystem

(Fig. 1, 3) to understand ratios spatial and vertical distributions. Ratios were also plotted against each property to understand individual processes/mechanisms and to monitor variability (Fig. 2). A Generalized Linear Model (GLM) using forward stepwise selection was applied to the ratios vs. 5 depth intervals to understand complexity and individual parameter significance with depth (SI Appendix 5; SI Appendix Table S2). All analyses and graphical work were performed in Statistica 13.0 (StatSoft), SigmaPlot 12.0 (Systat Software Inc.), ArcGIS (ESRI 2011), SURFER (Golden Software, LLC.), and Corel Draw X3 (Corel Corp.).

Text S1

The following instructions were sent to each project participant and PI along with the same 15 ml falcon tubes, 0.20 μ m filters, Parafilm, and 50 ml syringes and gloves to ensure reproducibility and minimize handling errors. A PDF was emailed and a printed version was included in the boxes with the consumables.

Sampling for the determination of seawater composition in terms of Mg/Ca and Sr/Ca (PI: Mario Lebrato)

Sampling

Note: use gloves during sampling procedure

- 1. Sample along the agreed transect/station using a Niskin-type bottle, a bucket or any means to get seawater in a clean way. Use only the lab gear/consumables we provide.
- 2. Samples must be stored cool and dry place.
- 3. Avoid touching or contaminating the sample before or after the procedure.

Laboratory

- 1. Filter through a fresh sterile $0.20 \mu m$ filter using a syringe for each sampling station/sample.
- 2. Rinse the syringe 3 times with seawater that is going to be filtered (mounting and dismounting the syringe).
- 3. Dismount the back part of the syringe.
- 4. Pour 40 ml of seawater.
- 5. Mount back the syringe.
- 6. Filter gently, but discard the initial 2 to 5 ml of seawater.
- **7.** Store 15 ml of the sample in a Falcon tube**.**
- 8. Close the cap with Parafilm if possible or close very tight.
- 9. Store in a cool and dry place until shipping to us.

Labelling

Samples label and ID should be easily linked with the Excel sheet and metadata.

Metadata

Collect for each sample: Date, Latitude/Longitude, Sample depth, Temperature, and Salinity only with calibrated or trusted sensors. Please include notes on metadata errors, inaccuracies or any relevant issue. If needed discussion, please email the project PI.

Fig. 2. Modern ocean seawater Mg:Ca and Sr:Ca ratios versus environmental parameters. Data horizontally classified for seawater Mg:Ca (a) and Sr:Ca (b) ratios as a function of environmental variables (GLODAPv2; *41, 42*) from the largest to the smallest dimension, and vertically in five depth intervals of 0-5 m, 5-200 m, 100-1000 m, 1000-2500 m, and +2500 m. Data within the red lines represent literature assumed knowledge of modern seawater ratios. Code for "Ocean" is: 1=Pacific, 2=Indian, 3=Atlantic, 4=Arctic, 5=Mediterranean, 6=Southern, 7=Red Sea, 8=China Sea, 9=Arabian Sea, 10=Baltic Sea, 11=IAPSO. Code for "Longhurst Province" is in SI Appendix Table S1 and Vliz (2009): [http://www.marineplan.es/ES/fichas_kml/biogeog_prov.html.](about:blank) Code for "Ecosystem" is: 1=IAPSO (This study), 2=Coastal Sea, 3=Coastal Upwelling, 4=Open Ocean, 5=Open Ocean Upwelling, 6=Shallow Hydrothermal Vent, 7=Coral Reef, 8=Deep Hydrothermal Vent, 9=Glacier Seawater, 10=Estuary, 11=Mudflats, 12=Mangroves, 13=Seagrass. Extended analysis of seawater Mg:Ca ratios data from river mouths to assess coastal waters can be found in SI Appendix Fig. S1.

Supplementary Figures & Captions

Fig. S2. Examples of databases used in paleo research. A good understanding of modern seawater Mg:Ca and Sr:Ca ratios divided by Ocean, Longhurst Province, Ecosystem and region comes into perspective in paleo research depending on the type of reconstruction being conducted. Nowadays, efforts to develop paleo proxies have reached a global scale but still rely on local and regional samples: (a) Geographical distribution by archive type in the proxy records from the PAGES2k initiative (*7*), (b) Census of planktonic foraminifera in surface sediment samples (*8*), and (c) Paleoceanographic sediment cores from the North Pacific, 1951-2016 (*9*). All figures are used as published under a Creative Commons CC-BY license and a Creative Commons Attribution 3.0 License to M. Lebrato.

 32° N-

 30°N
 130°W

 125° W

 120° W

 $-32^{\circ}N$

 $\frac{1}{115^{\circ} \text{W}}$

Fig. S3. Examples of cruises, surveys, and stations where seawater Mg:Ca and Sr:Ca ratios samples were obtained. Maps were provided by PI and collaborators from the corresponding cruise, and have not been modified. (a) Transect in the Peru Oxygen Minimum Zone (OMZ) at Callao (Lima) (PI. A.P.). (b) Transect on the Line P programme (NOAA) off Vancouver (Pacific Ocean) (PI: C.P.). (c) Radials in the Iceland Sea from a permanent programme held by the Marine Research Institute of Iceland (PI: S.O.). (d) Cruise track around the Canadian Archipelago (PI: L.J.). (e) Work done in the Sermilik fjord and the close shelf in the southern Greenland Sea (PI: A.K.). (f) Transects available in the Russian-American Long-term Census of the Arctic (RUSALCA) around the Bering Strait (PI: R.H.). (g) Radials in the INGRES3-02 (200108)/STOCA 2011 08 research cruise in the Strait of Gibraltar (PI: R.S.L.). (h) Radial surveys in a carbonate chemistry cruise along the California Current (PI: R.A.F) (i) Radials in a cruise around Svalbard in the sub-Arctic Ocean (PI: J.B.). (j) Cruise track of stations in a cruise from Woods Hole Oceanographic Institution around the Bering Strait (PI: C. A.). (k) Long-cruise from La Reunion to Spain (PI: P.B.D.B). (l) Long-cruise from South Africa to South America (PI: B.T.). Not all cruises maps and tracks are included here, just a subset for showing this study collaborative approach.

Fig. S4. Global sampling effort of seawater Mg:Ca and Sr:Ca ratios used in this study. Sampling stations used to average seawater Mg:Ca (a) and Sr:Ca (b) ratios at all depths in 5x5 quadrats to show global sampling effort. Note that many stations have also a vertical component with samples down to 6000 m (see Fig. 1, 4)

Supplementary Tables & Captions

Table S1. Mean seawater Mg:Ca and Sr:Ca values divided by Ocean/Sea, Longhurst Province and Ecosystem. Data are classified using mean and standard deviation in the five depth intervals used for the analysis using as major divisions Ocean/Sea, Longhurst Province and Ecosystem. Note that a IAPSO seawater sample is derived from the literature (*10*), and also from our measurements.

Table S2. Generalized Linear Model (GLM) results using forward stepwise regression.

Results of the GLM for seawater Mg:Ca and Sr:Ca ratios using the 5 depth intervals to understand the complexity of the ratios vs. the environmental variables (full results in SI Appendix 5). Significant results are bold. Also included is an indication of the system complexity with depth.

Table S3. Historical use of seawater Mg:Ca and Sr:Ca ratios. Summary of ratios use in various marine science disciplines in the last 100 years.

Supplementary Databases

Appendix 1. Database used to classify and coordinate each individual research cruise/expedition. Excel file providing details of how each cruise and expeditions was planned and organized to recover seawater samples to measure seawater Mg:Ca and Sr:Ca ratios. This file belongs to the start of this project. The final database is in SI Appendix 3.

Appendix 2. Database with the raw literature data. Excel file with all the literature data selected for this study that were found comparable to our samples. The original database is permanently deposited at the NOAA National Center for Environmental Information (NCEI) under Accession Number 0171017 in<http://accession.nodc.noaa.gov/0171017> (DOI:10.7289/V5571996).

Appendix 3. Database with the raw and classified data per depth band. Excel file with all the measured and the literature data compiled in this study. Also included are all environmental metadata mined from GLODAPv2 (*4, 5*), and used to analyze and classify seawater Mg:Ca and Sr:Ca ratios as function of environmental variables. The original database is permanently deposited at the NOAA National Center for Environmental Information (NCEI) under Accession Number 0171017 in<http://accession.nodc.noaa.gov/0171017> (DOI:10.7289/V5571996).

Appendix 4. ICP-OES quality control data and measurements replication. Details on the ICP-OES laboratory quality and replicability that guarantees the high precision results presented in this study.

Appendix 5. General Linear Model (GLM) results using forward stepwise regression. Full results of the GLM for seawater Mg:Ca and Sr:Ca ratios using the 5 depth intervals to understand the complexity of the ratios vs. the environmental variables (summary in SI Appendix Table S2).

Appendix 6. Summarized river mouth seawater Mg:Ca ratios. Compilation of seawater Mg and Ca concentrations along with seawater Mg/Ca ratio of major rivers globally for comparison purposes (the data were not used in this dataset analysis). Original data belong to the GEMS-GLORI database of Meybeck and Alain (2012) which can be accessed at [https://doi.pangaea.de/10.1594/PANGAEA.804574.](https://doi.pangaea.de/10.1594/PANGAEA.804574)

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