Supporting Information

Index	Page
General	S2
Preparation of Ligand L1 to L5 and CC-L1	S3
Procedure for trans-selective Cyanoboration, Ligand Screening (Table 1)	S8
Preparation of Enynes 1 and 2 in Table 2 and Table 3	S9
General Procedure for trans-Selective Cyanoboration Reaction of 1,3-Enynes (Table 2)	S15
Assignment of the trans/cis Configuration of the Products	S20
General Procedure for trans-Selective Cyanoboration Reaction of 1,3-Enynes (Table 3)	S22
Functionalization of Cyanoboration Product (Scheme 4)	S25
Synthesis of Satigrel (Scheme 5)	S27
Crystallographic Data for 4b , 4j-B(pin) and 5g	S31
NMR Spectra Collection	S34
References	S132

General

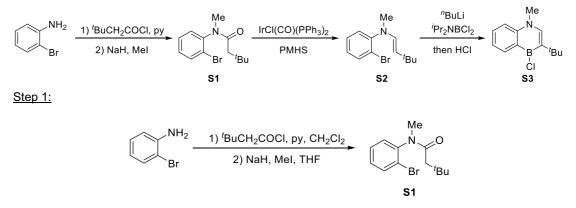
¹H, ¹³C, ³¹P and ¹⁹F spectra were recorded on Varian 400, 500 or 600 MHz spectrometers, and ¹¹B spectra were on an Inova 500 MHz spectrometer at ambient temperature. ¹H NMR spectra were reported with the solvent resonance as internal standard. ¹³C NMR spectra were reported with the solvent resonance as internal standard. ¹¹B NMR spectra were reported with BF₃•Et₂O (δ 0 ppm) as the external reference. ³¹P NMR spectra were reported with H₃PO₄ (δ 0 ppm) as the external reference. IR spectra were recorded on a Bruker FTIP Alpha (ATR mode) spectrometer. High-resolution mass spectroscopy data were obtained at the Mass Spectroscopy Facilities at Chemistry Department of Boston College with DART ion source in positive ion mode.

All oxygen- and moisture-sensitive manipulations were carried out under N_2 atmosphere with standard Schlenk techniques or in N_2 glovebox.

Hexanes were purified by distillation before using in column chromatography. Solvents used under N_2 atmosphere (pentane, THF, benzene and CH_2Cl_2) were purified by passing through a neutral alumina column under argon. 1,8-diaminonaphthlene (dan) was purified by distillation under attenuated pressure followed by recrystallization with hexanes/EtOAc. 1, 2-dichlorobenzene (*o*-DCB) was purified by drying with CaH₂ and subsequent distillation under attenuated pressure. Ligand **L5** was prepared according to literature procedures.¹ All other chemicals and solvents were purchased and used as received.

Preparation of Ligand L1 to L4 and CC-L1

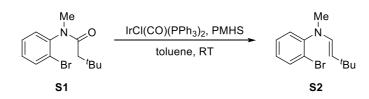
Synthesis of ^tbutyl-substituted 1,4-azaborine precursor S3: Reaction sequence:



To a solution of 2-bromoaniline (5.10 g, 30.0 mmol, 1.00 equiv.) in CH₂Cl₂ (100 mL) and pyridine (2.50 g, 31.5 mmol, 1.05 equiv.) ^{*i*}butylacetyl chloride (4.24 g, 31.5 mmol, 1.05 equiv.) was added in a dropwise fashion at 0 °C, and the mixture was allowed to stir for 4 hours. At the conclusion of reaction, H₂O (60 mL) was added, and the organic layer was separated, which was further washed with brine (60 mL) and dried with Na₂SO₄. Removing of all volatiles *in vacuo* resulted in a white solid, which is used for next step without further purification. Then, THF (80 mL) and NaH (1.80 g, 60% in oil, 45.0 mmol, 1.50 equiv.) was added in 5 portions in 20 minutes. The resulting mixture was allowed to stir for another 30 minutes at room temperature, followed by the addition of MeI (6.40 g, 45.0 mmol, 1.50 equiv.). After allowing the mixture to stir for 2 hours, H₂O (60 mL) along with Et₂O (30 mL) were added to the reaction mixture. The organic layer was separated and washed with brine (50 mL) and dried with Na₂SO₄. After removing all volatiles, **S1** was obtained by distillation under attenuated pressure (130 °C, 250 mmTor) as a white solid (88% yield).

¹H NMR (500 MHz, CDCl₃) δ 7.66 (dt, *J* = 8.0, 1.5 Hz, 1H), 7.37 (tt, *J* = 7.5, 1.3Hz, 1H), 7.25 – 7.19 (m, 2H), 3.16 (d, *J* = 1.6 Hz, 3H), 1.89 (t, *J* = 1.1Hz, 2H), 0.97 (d, *J* = 1.7 Hz, 9H); ¹³C NMR (126 MHz, CDCl₃) δ 171.8, 143.4, 133.9, 130.2, 129.5, 128.9, 123.6, 45.6, 35.8, 31.2, 29.9; IR (ATR) 2952, 2866, 1661, 1475, 1433, 1372, 1361, 1280, 1249, 1117, 1064, 1030, 765, 728, 690, 568, 457 cm⁻¹; HRMS (DART) calcd for $C_{13}H_{19}NOBr$ ([M+H]⁺) 284.06445, found 284.06500.

Step 2:

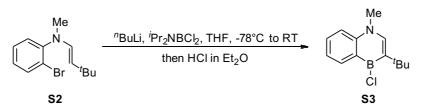


To a 250-mL round bottom flask charged with **S1** (2.80 g, 10.0 mmol, 1.00 equiv.), toluene (30 mL), and PMHS (polymethylhydrosiloxane) (2.60 g, MW 1700-3200, 40.0 mmol, 4.00 equiv.), was added a solution of $IrCl(CO)(PPh_3)_2$ (4 mg, 0.005 mmol, 0.0005 equiv) in toluene (5 mL) in a dropwise fashion under nitrogen at room temperature. Gelation was observed, and the reaction mixture was allowed to stir for another 30 minutes. Et₂O (60 mL) was added, and the mixture was passed through a celite pad. The filter cake was further washed with Et₂O (30 mL) for two times, and the filtrate was combined. All volatiles

were then removed under vacuum and **S2** was obtained by distillation under attenuated pressure (95 °C, 280 mmTor) as a colorless liquid (90% yield).

¹H NMR (600 MHz, CDCl₃) δ 7.57 (dd, *J* = 8.0, 1.5 Hz, 1H), 7.26 (tdd, *J* = 7.2, 1.6, 0.6 Hz, 1H), 7.09 (dd, *J* = 8.0, 1.6 Hz, 1H), 6.97 (tdd, *J* = 7.2, 1.6, 0.6 Hz, 1H), 6.15 (d, *J* = 14.1 Hz, 1H), 4.64 (d, *J* = 14.1 Hz, 1H), 2.98 (s, 3H), 1.08 (s, 9H); ¹³C NMR (151 MHz, CDCl₃) δ 148.8, 134.0, 132.9, 128.3, 126.3, 125.5, 119.9, 115.2, 38.4, 31.9, 31.3; IR (ATR) 2952, 2900, 2864, 1650, 1584, 1481, 1461, 1420, 1356, 1320, 1251, 1238, 1129, 1117, 1026, 981, 937, 753, 723, 656, 446 cm⁻¹; HRMS (DART) calcd for C₁₃H₁₉NBr ([M+H]⁺) 268.06954, found 268.06955.

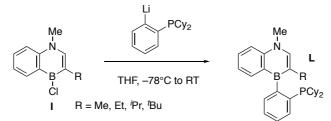
Step 3:



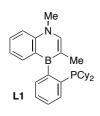
To a 100-mL round bottom flask charged with **S2** (2.42 g, 9.02 mmol, 1.00 equiv.) and THF (30 mL), was added ^{*n*}butyl lithium (3.60 mL, 2.50 M in hexanes, 9.00 mmol, 1.00 equiv.) at -78° C. After allowing the mixture to stir at -78° C for 20 minutes, a solution of ^{*i*}Pr₂NBCl₂ (1.64 g, 9.00 mmol, 1.00 equiv.) in THF (3 mL) was added. The resulting reaction mixture was allowed to warm to room temperature in 2 hours. Then, HCl (4.5 mL, 2.0 M in Et₂O, 9.0 mmol, 1.0 equiv.) was added, and the mixture was allowed to stir for an hour. At the conclusion of the reaction, the formed precipitate was filtered off, and the filtrate was concentrated. The resulting crude material was purified by recrystallization from pentane/CH₂Cl₂ to furnish **S3** as white crystals (36% yield).

¹H NMR (500 MHz, CDCl₃) δ 8.55 (dd, *J* = 7.9, 1.7 Hz, 1H), 7.78 (s, 1H), 7.69 (ddd, *J* = 8.6, 6.9, 1.7 Hz, 1H), 7.46 (d, *J* = 8.6 Hz, 1H), 7.35 (ddd, *J* = 7.9, 6.9, 1.0 Hz, 1H), 3.89 (s, 3H), 1.50 (s, 9H); ¹³C NMR (126 MHz, CDCl₃) δ 145.8, 143.2, 134.0, 133.7 (br), 131.6, 128.4 (br), 121.4, 114.6, 42.4, 34.9, 31.0; ¹¹B NMR (160 MHz, CDCl₃) δ 44.4; HRMS (DART) calcd for $C_{13}H_{18}BNCI$ ([M+H]⁺) 234.12153, found 234.12340.

General Procedure for the Synthesis of Ligand L1 to L4:

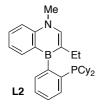


N-Me-*B*-Cl-1,4-Azaborines I where R = Me, Et, ^{*i*}Pr were synthesized according to previous literature.¹ To a 100-mL round bottom flask charged with 2-bromophenyl dicyclohexylphosphine (706 mg, 2.00 mmol, 1.00 equiv.) and Et₂O (15 mL) was added ^{*n*}butyllithium (0.80 mL, 2.5 M in hexanes, 2.0 mmol, 1.0 equiv.) at –78 °C. After allowing the mixture to stir at –78 °C for an hour, the corresponding 1,4-azaborine I (2.0 mmol, 1.0 equiv.) in THF (3 mL) was added, and the resulting mixture was allowed to warm to room temperature in 2 hours. At the conclusion of reaction, all volatiles were removed under vacuum, and the crude residue was purified by silica gel chromatography under inert atmosphere using pentane/Et₂O (20/1 to 10/1 gradient) as the eluent to afford the desired ligand as white solids.



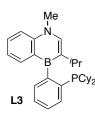
L1, 42% yield. ¹H NMR (500 MHz, CDCl₃) δ 7.74 (dd, J = 7.8, 1.6 Hz, 1H), 7.67 (s, 1H), 7.61 – 7.55 (m, 2H), 7.48 (d, J = 8.6 Hz, 1H), 7.40 – 7.31 (m, 2H), 7.28 – 7.24 (m, 1H), 7.13 (ddd, J = 7.8, 6.8, 0.9 Hz, 1H), 3.90 (s, 3H), 2.06 (d, J = 0.9 Hz, 3H), 2.01 (ddd, J = 11.1, 5.1, 3.0 Hz, 1H), 1.81 – 1.59 (m, 8H), 1.60 – 1.48 (m, 2H), 1.47 – 0.78 (m, 11H); ¹³C NMR (126 MHz, CDCl₃) δ 146.6, 143.2, 137.6, 137.3 (d, J = 11.0 Hz), 131.7 (d, J = 15.4 Hz), 131.7 (d, J = 2.6 Hz), 130.8 (br), 130.4, 127.5, 125.6, 122.9

(br), 120.2, 114.3, 42.0, 35.1 (d, J = 13.7 Hz), 33.6 (d, J = 12.4 Hz), 30.7 (d, J = 14.2 Hz), 30.3 (d, J = 15.9 Hz), 30.0 (d, J = 10.6 Hz), 29.2 (d, J = 6.0 Hz), 27.7, 27.6 (d, J = 6.4 Hz), 27.6, 27.5 (d, J = 3.6 Hz), 26.7, 26.6, 19.6 (d, J = 4.3 Hz) (one signal of aromatic carbon attached to boron is not observed); ¹¹B NMR (160 MHz, CDCl₃) δ 47.3; ³¹P NMR (202 MHz, CDCl₃) δ -3.2; IR (ATR) 3039, 2922, 2849, 1605, 1586, 1491, 1447, 1369, 1269, 1225, 1102, 908, 891, 764, 732, 647 cm⁻¹; HRMS (DART) calcd for C₂₈H₃₈BNP ([M+H]⁺) 430.28294, found 430.28290.



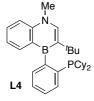
L2, 36% yield. ¹H NMR (500 MHz, CDCl₃) δ 7.71 (dd, J = 7.8, 1.7 Hz, 1H), 7.65 (s, 1H), 7.62 – 7.54 (m, 2H), 7.47 (d, J = 8.6 Hz, 1H), 7.39 – 7.29 (m, 2H), 7.29 (dd, J = 5.5, 3.2 Hz, 1H), 7.11 (t, J = 7.3 Hz, 1H), 3.92 (s, 3H), 2.55 (dq, J = 14.8, 7.5 Hz, 1H), 2.35 (dq, J = 14.7, 7.5 Hz, 1H), 2.08 – 1.94 (m, 1H), 1.79 – 1.59 (m, 8H), 1.57 – 0.79 (m, 16H); ¹³C NMR (101 MHz, CDCl₃) δ 146.0, 143.2, 137.6, 137.2 (d, J = 11.0 Hz), 132.2 (d, J = 15.4 Hz), 131.7 (d, J = 2.5 Hz), 130.5, 127.3, 125.6, 120.1, 114.3, 42.1,

35.3 (d, J = 13.8 Hz), 33.5 (d, J = 12.3 Hz), 30.7 (d, J = 14.0 Hz), 30.3 (d, J = 15.9 Hz), 30.0 (d, J = 11.3 Hz), 29.1 (d, J = 5.7 Hz), 27.7, 27.6 (d, J = 1.5 Hz), 27.6 (d, J = 3.7 Hz), 27.5 (d, J = 8.6 Hz), 26.8, 26.6 (d, J = 4.3 Hz), 26.6, 16.5 (three signals of aromatic carbons attached to boron are not observed); ¹¹B NMR (160 MHz, CDCl₃) δ 47.1; ³¹P NMR (202 MHz, CDCl₃) δ –3.6; IR (ATR) 3040, 2924, 2850, 1604, 1585, 1491, 1448, 1375, 1266, 1223, 1105, 912, 764, 750, 733, 663 cm⁻¹; HRMS (DART) calcd for C₂₉H₄₀BNP ([M+H]⁺) 444.29859, found 444.29726.



L3, 48% yield. ¹H NMR (500 MHz, CDCl₃) δ 7.71 – 7.64 (m, 2H), 7.62 – 7.53 (m, 2H), 7.47 (d, *J* = 8.6 Hz, 1H), 7.40 – 7.29 (m, 3H), 7.10 (t, *J* = 7.3 Hz, 1H), 3.93 (s, 3H), 2.89 (hept, *J* = 7.2 Hz, 1H), 1.96 (tt, *J* = 12.1, 3.5 Hz, 1H), 1.80 – 1.59 (m, 8H), 1.56 (d, *J* = 9.0 Hz, 3H), 1.43 (d, *J* = 13.2 Hz, 1H), 1.34 – 0.79 (m, 15H); ¹³C NMR (126 MHz, CDCl₃) δ 144.3, 143.1, 137.7, 137.5 (d, *J* = 11.0 Hz), 134.9 (br), 132.1 (d, *J* = 15.3 Hz), 131.8 (d, *J* = 2.8 Hz), 131.3 (br), 130.5, 127.3, 125.6, 120.1, 114.3, 42.4,

35.2 (d, J = 14.0 Hz), 34.1 (d, J = 12.9 Hz), 30.6 (d, J = 14.2 Hz), 30.4 (d, J = 16.0 Hz), 29.9 (d, J = 10.3 Hz), 29.7 (d, J = 2.4 Hz), 29.3 (d, J = 6.7 Hz), 27.7, 27.6, 27.5, 27.5, 27.1, 26.7, 26.6, 22.7 (one signal of the aromatic carbon attached to boron is not observed); ¹¹B NMR (160 MHz, CDCl₃) δ 47.5; ³¹P NMR (202 MHz, CDCl₃) δ –4.3; IR (ATR) 3039, 2923, 2849, 1605, 1585, 1491, 1457, 1428, 1383, 1372, 1265, 1223, 908, 765, 750, 733, 667 cm⁻¹; HRMS (DART) calcd for C₃₀H₄₂BNP ([M+H]⁺) 458.31424, found 458.31565.

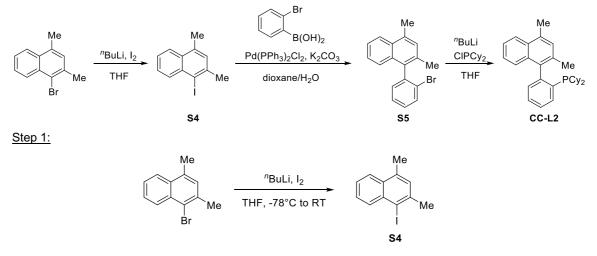


L4, 38% yield. ¹H NMR (600 MHz, CDCl₃) δ 7.83 (s, 1H), 7.57 – 7.49 (m, 2H), 7.46 – 7.37 (m, 3H), 7.36 – 7.28 (m, 2H), 7.04 (ddd, *J* = 7.9, 6.8, 1.0 Hz, 1H), 3.93 (s, 3H), 2.10 (tdd, *J* = 11.1, 6.7, 3.1 Hz, 1H), 1.83 – 0.70 (m, 30H); ¹³C NMR (126 MHz, CDCl₃) δ 145.1, 142.1, 138.8 (d, *J* = 9.7 Hz), 138.1, 132.2 (d, *J* = 14.9 Hz), 131.2 (d, *J* = 2.3 Hz), 130.5, 126.5, 125.2, 119.9, 113.8, 42.5, 36.0 (d, *J* = 14.7 Hz), 35.9, 33.6 (d, *J* =

14.1 Hz), 33.0 (d, J = 1.6 Hz), 32.0 (d, J = 16.6 Hz), 31.1 (d, J = 13.3 Hz), 30.5 (d, J = 15.7 Hz), 28.3 (d, J = 4.9 Hz), 28.0 (d, J = 2.0 Hz), 27.9 (d, J = 3.4 Hz), 27.8, 27.4 (d, J = 10.1 Hz), 26.7, 26.6 (three signals of aromatic carbon attached to boron are not observed); ¹¹B NMR (160 MHz, CDCl₃) δ 47.3; ³¹P NMR (202 MHz, CDCl₃) δ -3.4; IR (ATR) 3039, 2920, 2849, 1604, 1581, 1490, 1459, 1447, 1390, 1376, 1264, 1220, 1086, 918, 765, 742, 669 cm⁻¹; HRMS (DART) calcd for C₃₁H₄₄BNP ([M+H]⁺) 472.32989, found 472.32950.

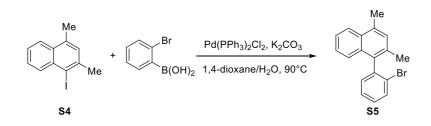
Synthesis of ligand CC-L1

Reaction sequence:



ⁿButyllithium (1.30 mL, 2.50 M in hexanes, 3.24 mmol, 1.20 equiv.) was added in a dropwise fashion into a 20-mL vial charged with a solution 1-bromo-2,4-dimethylnaphthalene (634 mg, 2.70 mmol, 1.00 equiv., synthesized according to previous literature².) in THF (12 mL) at -78 °C. The resulting mixture was allowed to stir for 30 minutes at the same temperature. Then a solution of I₂ (1.35 g, 5.40 mmol, 2.00 equiv.) in THF (3 mL) was added to the reaction mixture, and the resulting mixture was gradually allowed to warm to room temperature in 2 hours. At the conclusion of the reaction, THF was removed, and the residue was taken up in Et₂O (15 mL) and washed with saturated Na₂S₂O₃ (10 mL) and H₂O (10 mL). The organic layer was then concentrated under reduced pressure, and the crude material was purified via silica gel chromatography using hexanes as the eluent to afford **S4** as a colorless liquid (88% yield). ¹H NMR (500 MHz, CDCI₃) δ 8.33 – 8.24 (m, 1H), 7.92 (dd, *J* = 8.3, 1.6 Hz, 1H), 7.58 (ddd, *J* = 8.4, 6.8, 1.3 Hz, 1H), 7.52 (td, *J* = 6.8, 1.4 Hz, 1H), 7.24 (s, 1H), 2.69 (s, 3H), 2.65 (d, *J* = 2.0 Hz, 3H); ¹³C NMR (126 MHz, CDCI₃) δ 140.4, 135.0, 134.6, 132.9, 131.8, 129.2, 127.4, 125.7, 124.4, 103.3, 30.3, 19.2; IR (ATR) 3060, 3007, 2917, 1600, 1510, 1475, 1432, 1387, 1379, 1049, 1026, 875, 752, 731, 709, 669, 655, 453 cm⁻¹; HRMS (DART) calcd for C₁₂H₁₁I ([M]⁺) 281.98999, found 281.99087.

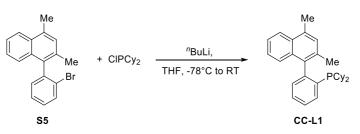
<u>Step 2:</u>



A 20-mL vial was charged with $Pd(PPh_3)_2Cl_2$ (16.5 mg, 0.0235 mmol, 0.0500 equiv.), potassium carbonate (130 mg, 0.940 mmol, 2.00 equiv.) and (2-bromophenyl)boronic acid (104 mg, 0.517 mmol, 1.10 equiv.) under nitrogen. To this mixture 1,4-dioxane (3.5 mL), H₂O (0.5 mL) and **S4** (134 mg, 0.470 mmol, 1.00 equiv.) were added, and the reaction mixture was allowed to stir at 90 °C for 3 hours. At the conclusion of the reaction, Et₂O (5 mL) and H₂O (2 mL) were added to the mixture. The organic layer was separated and the aqueous layer was extracted with Et₂O (10 mL). The combined organic layers were concentrated under reduced pressure. The resulting crude material was purified via silica gel chromatography using hexanes as the eluent to afford **S5** as a colorless liquid (68% yield).

¹H NMR (500 MHz, CDCl₃) δ 8.07 (dt, *J* = 8.5, 1.0 Hz, 1H), 7.81 (dd, *J* = 8.1, 1.3 Hz, 1H), 7.49 (dtd, *J* = 18.1, 7.1, 1.3 Hz, 2H), 7.41 (ddd, *J* = 8.2, 6.7, 1.3 Hz, 1H), 7.37 – 7.32 (m, 2H), 7.29 (td, *J* = 7.2, 1.4 Hz, 2H), 2.78 (d, *J* = 1.0 Hz, 3H), 2.22 (s, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 140.9, 135.5, 134.2, 133.2, 132.9, 132.4, 132.1, 131.1, 129.5, 129.0, 127.6, 126.1, 125.9, 125.1, 124.9, 124.2, 20.3, 19.6; IR (ATR) 2967, 2943, 2913, 2856, 1599, 1555, 1503, 1438, 1326, 1263, 1026, 961, 884, 867, 747, 639, 567, 417 cm⁻¹; HRMS (DART) calcd for C₁₈H₁₆Br ([M+H]⁺) 311.04299, found 311.04305.

Step 3:



To a 20-mL vial charged with **S5** (99 mg, 0.32 mmol, 1.0 equiv.) and THF (2 mL) was added ^{*n*}butyllithium (0.15 mL, 2.5 M in hexanes, 0.38 mmol, 1.2 equiv.) at -78 °C. The mixture was allowed to stir at -78 °C for 30 minutes, then a solution of chlorodicyclohexylphosphine (89 mg, 0.38 mmol, 1.2 equiv.) in THF (1 mL) was added. The resulting mixture was gradually allowed to warm to room temperature in 2 hours. THF was removed *in vacuo*, and the residue was purified via silica gel chromatography using hexanes/EtOAc (100/1) as the eluent to afford **CC-L1** as a white solid (72% yield).

¹H NMR (500 MHz, CDCl₃) δ 7.97 (d, *J* = 8.3 Hz, 1H), 7.67 (dt, *J* = 6.6, 2.3 Hz, 1H), 7.46 – 7.42 (m, 2H), 7.40 (ddd, *J* = 8.2, 6.5, 1.5 Hz, 1H), 7.30 – 7.17 (m, 4H), 2.72 (s, 3H), 2.18 (d, *J* = 2.0 Hz, 3H), 1.91 (ddd, *J* = 15.2, 10.1, 3.2 Hz, 1H), 1.78 – 0.79 (m, 21H); ¹³C NMR (126 MHz, CDCl₃) δ 147.5 (d, *J* = 31.1 Hz), 136.9 (d, *J* = 19.4 Hz), 136.7 (d, *J* = 6.2 Hz), 133.5 (d, *J* = 1.8 Hz), 133.4, 133.0 (d, *J* = 3.2 Hz), 132.7 (d, *J* = 2.0 Hz), 131.6 (d, *J* = 5.9 Hz), 131.0, 129.5, 128.6, 127.7, 126.6, 124.8, 124.4, 124.0, 34.9 (d, *J* = 15.1 Hz), 33.7 (d, *J* = 14.1 Hz), 30.4 (d, *J* = 4.8 Hz), 30.2, 30.1 (d, *J* = 11.9 Hz), 29.8 (d, *J* = 12.9 Hz), 27.6, 27.6 (d, *J* = 18.5 Hz), 27.5 (d, *J* = 4.0 Hz), 27.4 (d, *J* = 6.2 Hz), 26.7, 26.5, 21.4 (d, *J* = 4.3 Hz), 19.7; ³¹P NMR (202 MHz, CDCl₃) δ –9.4; IR (ATR) 3051, 3006, 2921, 2848, 1600, 1511, 1446, 1386, 1034, 1000, 909, 875, 851, 757, 733, 669 cm⁻¹; HRMS (DART) calcd for C₃₀H₃₈P ([M+H]⁺) 429.27056, found 429.26989.

Procedure for *trans*-Selective Cyanoboration, Ligand Screening (Table 1)

To a 4-mL vial charged with Pd(COD)(CH₂TMS)₂ (3.9 mg, 0.010 mmol, 5.0 mol %), corresponding ligand in **Table 1** (0.010 mmol, 5.0 mol %), CuCN (29 mg, 0.32 mmol, 1.6 equiv.), BCatCl (55 mg, 0.36 mmol, 1.8 equiv.), and *o*-DCB (0.30 mL) was added enyne **1a** (28.4 mg, 0.200 mmol, 1.00 equiv) and a solution of hexamethylbenzene in *o*-DCB (5.0 μ L, 3.3 M) as the internal standard. The resulting mixture was allowed to stir for 2.5 hours at 115 °C. At the conclusion of the reaction, the mixture was passed through an acrodisc under an inert atmosphere using CH₂Cl₂ as the solvent, and 1,8-diaminonaphthlene (64 mg, 0.40 mmol, 2.0 equiv) was added to the filtrate. After allowing the mixture to stir for 12 hours, volatiles were removed under reduced pressure, and the *trans:cis* ratios and yields were determined by ¹H NMR of the resulting crude material against hexamethylbenzene as the internal standard.

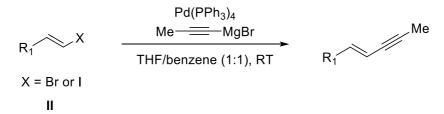
	Ph	Me 1a	1.6 5 mol% l <i>o</i> -D0 1,8-d	equiv. BCatCl equiv. CuCN Pd(COD)(CH ₂ TMS) ₂ 5 mol% L CB, 115 °C, 2.5 h then liaminonaphthlene H ₂ Cl ₂ , RT, 12h	CN Ph → Me B(dan) 4a
entry	L	4a yield (tra	ans:cis)		
1	none	3% (89:	11)	Ме	Me
2	PhPCy ₂	13% (95	:5)		
3	XPhos	10% (90:	10)	BMe	Me
4	dppe	0.3% (N	D)	PPh ₂	PCy ₂
5	L1	92% (96	:4)	<u> </u>	
6	L2	69% (95	:5)	L5	CC-L1
7	L3	48% (95	:5)		
8	L4	24% (95	:5)		
9	L5	46% (96	:4)		
10	CC-L1	6% (89: ⁻	11)		
				-	

o-DCB: ortho-dichlorobenzene

Preparation of Enynes 1 and 2 (Table 2 and Table 3)

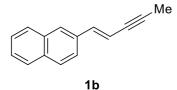
Enynes in Table 2

General Method for Enyne Synthesis by Kumada Coupling:



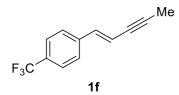
The corresponding (*E*)-styrenyl bromide^{3,4} (to synthesize **1a-1b**, **1d-1h**, **1k**, **1m** and **1s**) or alkenyl iodide^{5,6,7} (to synthesize **1n-1q** and **1t**) precursors are known compounds. (*E*)-3-(2-bromovinyl) thiophene was obtained using reported methods.^{8,9}

To a 20-mL vial charged with Pd(PPh₃)₄ (34 mg, 0.030 mmol, 0.015 equiv.), *(E)*-styrenyl bromide or alkenyl iodide **II** (2.0 mmol, 1.0 equiv.) and benzene (5 mL) was added 1-propynyl magnesium bromide (5.2 mL, 0.50 M in THF, 2.6 mmol, 1.3 equiv.) in a dropwise fashion at room temperature under nitrogen. The mixture was allowed to stir for 14 hours at the same temperature. At the conclusion of reaction, aqueous HCl solution (5.0 mL, 1.0 M) was added slowly along with Et₂O (6 mL). The organic layer was separated, and the aqueous layer was further extracted with Et₂O (5 ml). The organic layers were then combined, and dried with Na₂SO₄. All violates were removed *in vacuo*, and the residue was purified by silica gel chromatography using hexanes as the eluent to afford the internal enynes as colorless oils. Enyne **1s** and **1t** were synthesized with the same procedure using 1-butynyl magnesium bromide and 1-ethynyl magnesium bromide, respectively. The characterization data for enynes are consistent with those reported in the literature: **1a**¹⁰, (**1d**, **1e**, **1g**, **1h**, **1n**, **1p**, **1s**)¹, **1t**¹¹.



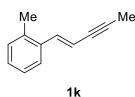
1b, 87% yield. ¹H NMR (600 MHz, CDCl₃) δ 7.86 – 7.76 (m, 3H), 7.73 (d, *J* = 1.6 Hz, 1H), 7.58 (dd, *J* = 8.6, 1.7 Hz, 1H), 7.52 – 7.42 (m, 2H), 7.06 (d, *J* = 16.1 Hz, 1H), 6.29 (dq, *J* = 16.2, 2.4, Hz, 1H), 2.07 (d, *J* = 2.4 Hz, 3H); ¹³C NMR (151 MHz, CDCl₃) δ 140.3, 134.1, 133.6, 133.4, 128.4, 128.2, 127.8, 126.5, 126.5, 126.3, 122.9, 109.3, 88.8, 79.2, 4.7; IR (ATR) 3055, 3029, 2912, 2850, 2216, 1507, 1362, 1185, 959, 867,

828, 816, 740, 486 cm⁻¹; HRMS (DART) calcd for C₁₅H₁₃ ([M+H]⁺) 193.10118, found 193.10037.

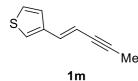


1f, 66% yield. ¹H NMR (500 MHz, CDCl₃) δ 7.56 (d, J = 8.2 Hz, 2H), 7.44 (d, J = 8.2 Hz, 2H), 6.87 (d, J = 16.2 Hz, 1H), 6.22 (dq, J = 16.2, 2.4 Hz, 1H), 2.03 (dd, J = 2.5, 0.8 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 140.1 (d, J = 1.7 Hz), 138.5, 130.0 (d, J = 32.5 Hz), 126.3, 125.7 (q, J= 3.8 Hz), 124.2 (q, J = 271.9 Hz), 111.8, 90.2, 78.7, 4.6; ¹⁹F NMR

 $(470 \text{ MHz}, \text{CDCl}_3) \delta$ –62.6; IR (ATR) 3020, 2924, 2854, 2217, 1612, 1411, 1327, 1165, 1120, 1109, 1070, 954, 860, 820, 599, 517 cm⁻¹; HRMS (DART) calcd for C₁₂H₁₀F₃ ([M+H]⁺) 211.07291, found 211.07317.

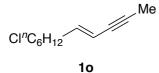


1k, 71% yield. ¹H NMR (600 MHz, CDCl₃) δ 7.47 (dd, J = 7.0, 2.1 Hz, 1H), 7.23 – 7.13 (m, 4H), 6.09 (dq, J = 16.1, 2.4 Hz, 1H), 2.39 (s, 3H), 2.06 (d, J = 2.4 Hz, 3H); ¹³C NMR (151 MHz, CDCl₃) δ 138.0, 135.6, 135.6, 130.5, 128.2, 126.2, 124.9, 109.9, 88.0, 79.4, 19.8, 4.6; IR (ATR) 3014, 3019, 2949, 2913, 2849, 2217, 1482, 1458, 1436, 1377, 1048, 951, 746, 714, 608, 537, 458 cm⁻¹; HRMS (DART) calcd for C₁₂H₁₃ ([M+H]⁺) 157.10118, found 157.10142.



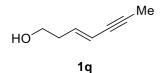
1m, 83% yield. ¹H NMR (500 MHz, CDCl₃) δ 7.28 – 7.25 (m, 1H), 7.20 – 7.14 (m, 2H), 6.86 (d, *J* = 16.1 Hz, 1H), 5.97 (dq, *J* = 16.2, 2.4 Hz, 1H), 2.00 (d, *J* = 2.4 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 139.5, 134.3, 126.4, 124.5, 122.9, 108.8, 88.3, 78.9, 4.7; IR (ATR) 3097, 3033, 2912, 2847, 2218, 1411, 1244, 1183, 947, 865, 834, 766, 623, 687, 472 cm⁻¹; HRMS

(DART) calcd for C_9H_9S ([M+H]⁺) 149.04195, found 149.04177.



10, 87% yield. ¹H NMR (500 MHz, CDCl₃) δ 6.07 – 5.96 (m, 1H), 5.41 (dq, J = 15.8, 1.9 Hz, 1H), 3.51 (t, J = 6.7 Hz, 2H), 2.07 (qd, J = 7.3, 1.5 Hz, 2H), 1.91 (d, J = 2.3 Hz, 3H), 1.75 (dt, J = 14.7, 6.8 Hz, 2H), 1.48 – 1.19 (m, 6H); ¹³C NMR (126 MHz, CDCl₃) δ 143.1, 110.2, 84.2, 78.5, 45.2,

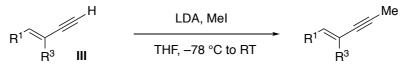
32.9, 32.7, 28.8, 28.4, 26.8, 4.3; IR (ATR) 3019, 2929, 2855, 2225, 1462, 1444, 1309, 1172, 955, 727, 651 cm⁻¹; HRMS (DART) calcd for $C_{11}H_{18}CI$ ([M+H]⁺) 185.10915, found 185.10880.



1q, with 2.6 equiv. 1-propynyl magnesium bromide, 81% yield. ¹H NMR (500 MHz, CDCl₃) δ 5.99 (dtd, *J* = 15.5, 7.3, 0.7 Hz, 1H), 5.60 – 5.42 (m, 1H), 3.63 (t, *J* = 6.4 Hz, 2H), 2.32 (qd, *J* = 6.5, 1.4 Hz, 2H), 1.94 – 1.87 (m, 3H), 1.85 (s, br, 1H); ¹³C NMR (126 MHz, CDCl₃) δ 138.8, 112.9, 85.1,

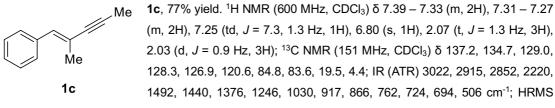
78.1, 61.7, 36.3, 4.2; IR (ATR) 3330, 2917, 2879, 2224, 1428, 1376, 1171, 1043, 954, 636, 543 cm⁻¹; HRMS (DART) calcd for C₇H₁₁O ([M+H]⁺) 111.08044, found 111.08026.

General Method for Enyne Synthesis by Terminal Enyne Methylation:

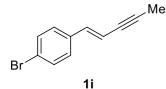


Terminal enynes precursors III for synthesizing 1c, 1i and 1I are known compounds.^{12,1,13}

To a 20-mL vial charged with diisopropylamine (242 mg, 2.40 mmol, 1.20 equiv.) and THF (6 mL), *n*butyl lithium (0.88 mL, 2.5 M in hexanes, 2.2 mmol, 1.1 equiv.) was added in dropwise fashion at -78 °C. The mixture was allowed to stir at the same temperature for 15 minutes. Then the terminal enyne (2.0 mmol, 1.0 equiv.) in THF (2.0 mL) was added to the LDA solution at -78 °C. After allowing the mixture to stir at -78 °C for another 20 minutes, MeI (355 mg, 2.50 mmol, 1.25 equiv.) was added at once. The resulting mixture was allowed to stir at -78 °C for 30 min and then gradually allowed to warm to room temperature in an hour. At the conclusion of the reaction, THF was removed *in vacuo*, and the residue was dissolved in Et₂O (10 mL). The organic layer was further washed by H₂O (5 mL), brine (5 mL) and concentrated. The residue was purified by silica gel column chromatography using hexanes as the eluent to afford the internal enynes as colorless oils.

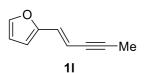


(DART) calcd for $C_{12}H_{13}$ ([M+H]⁺) 157.10118, found 157.10170.



1i, 91% yield. ¹H NMR (500 MHz, CDCl₃) δ 7.46 – 7.39 (m, 2H), 7.24 – 7.16 (m, 2H), 6.78 (d, *J* = 16.2 Hz, 1H), 6.11 (dq, *J* = 16.2, 2.4 Hz, 1H), 2.01 (dd, *J* = 2.4, 0.7 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 138.9, 135.6, 131.9, 127.6, 122.2, 109.8, 89.2, 78.8, 4.7; IR (ATR) 3031, 2910, 2845, 2213, 1584, 1486, 1399, 1074, 1008, 962, 946, 854, 806, 786,

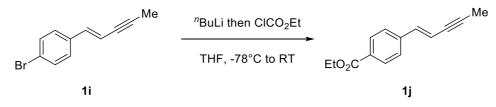
519 cm⁻¹; HRMS (DART) calcd for C₁₁H₁₀Br ([M+H] ⁺) 220.99604, found 220.99502.



1I, 81% yield. ¹H NMR (500 MHz, CDCl₃) δ 7.34 (d, *J* = 1.6 Hz, 1H), 6.62 (d, *J* = 16.0 Hz, 1H), 6.38 (dt, *J* = 3.4, 1.6 Hz, 1H), 6.31 – 6.23 (m, 1H), 6.04 (dp, *J* = 16.1, 2.4 Hz, 1H), 2.01 (dd, *J* = 2.6, 1.6 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 152.5, 142.7, 127.5, 111.8, 109.2, 107.2, 89.3, 79.0, 4.7; IR (ATR)

3397, 3055, 2220, 1600, 1505, 1408, 1373, 1335, 1095, 820, 767 cm⁻¹; HRMS (DART) calcd for C₉H₉O ([M+H]⁺) 133.06479, found 133.06543.

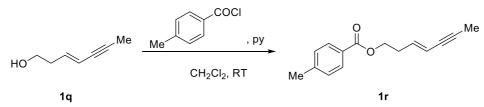
Synthesis of Enyne 1j by Esterification:



A solution of enyne **1i** (236 mg, 1.14 mmol, 1.00 equiv.) in THF (4 mL) was placed in a 20-mL vial. *"*Butyl lithium (0.500 mL, 2.50 M in hexanes, 1.25 mmol, 1.10 equiv.) was added to this solution in a dropwise fashion at -78 °C. After allowing the mixture to stir at the same temperature for 20 minutes, ethyl chloroformate (186 mg, 1.71 mmol, 1.50 equiv.) was added. The mixture was allowed to stir for an hour at -78 °C and then allowed to warm to room temperature in an hour. The resulting slurry was poured into H₂O (10 mL) and then diluted with Et₂O (5 mL). The organic layer was separated and violates were removed *in vacuo*. The resulting residue was purified by silica gel chromatography using hexanes/EtOAc (20/1) as the eluent to afford **1j** a white solid (47%).

¹H NMR (600 MHz, CDCl₃) δ 8.03 – 7.86 (m, 2H), 7.48 – 7.30 (m, 2H), 6.85 (d, *J* = 16.2 Hz, 1H), 6.21 (dq, *J* = 16.3, 2.4 Hz, 1H), 4.35 (q, *J* = 7.2 Hz, 2H), 2.08 – 1.89 (m, 3H), 1.37 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (151 MHz, CDCl₃) δ 166.3, 140.8, 139.0, 130.0, 129.9, 125.9, 111.6, 90.1, 78.8, 61.0, 14.4, 4.6; IR (ATR) 3035, 2983, 2912, 2212, 1711, 1607, 1411, 1363, 1279, 1266, 1178, 1127, 1105, 1026, 944, 864, 759, 694, 519 cm⁻¹; HRMS (DART) calcd for C₁₄H₁₅O₂ ([M+H]⁺) 215.10666, found 215.10591.

Synthesis of Enyne 1r by Ester Coupling:

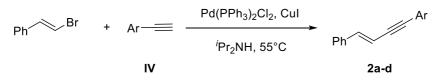


Under nitrogen, *p*-toluoyl chloride (240 mg, 1.56 mmol, 1.20 equiv.) and pyridine (123 mg, 1.56 mmol, 1.20 equiv.) were added to a 20-mL vial charged with a solution of enyne **1q** (143 mg, 1.30 mmol, 1.00 equiv.) in CH_2Cl_2 (8 mL). The resulting mixture was allowed to stir at room temperature for 14 hours. Then, the solvent was removed under reduced pressure, and the residue was purified by silica gel chromatography with hexans/EtOAc (40/1) as the eluent to afford **1r** as a white solid (68%).

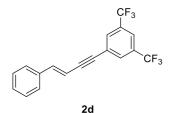
¹H NMR (500 MHz, CDCl₃) δ 7.98 – 7.80 (m, 2H), 7.23 (d, *J* = 8.2 Hz, 2H), 6.08 (dt, *J* = 15.8, 7.1 Hz, 1H), 5.58 (dp, *J* = 15.8, 1.9 Hz, 1H), 4.32 (t, *J* = 6.6 Hz, 2H), 2.54 (qd, *J* = 6.8, 1.5 Hz, 2H), 2.40 (s, 3H), 1.92 (d, *J* = 2.3 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 166.6, 143.7, 138.0, 129.7, 129.1, 127.6, 112.8, 85.3, 78.1, 63.6, 32.4, 21.7, 4.3; IR (ATR) 2955, 2916, 2226, 1715, 1612, 1454, 1379, 1271, 1177, 1105, 1020, 955, 841, 753, 691 cm⁻¹; HRMS (DART) calcd for C₁₅H₁₇O₂ ([M+H]⁺) 229.12231, found 229.12131.

Enynes in Table 3

General Method for Enyne Synthesis by Sonogashira Coupling:



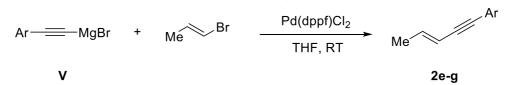
A 50-mL round bottom flask was charged with $PdCl_2(PPh_3)_2$ (84 mg, 0.12 mmol, 0.030 equiv.) and Cul (38 mg, 0.20 mol, 0.050 equiv.) under nitrogen. To this flask ^{*i*}Pr₂NH (20 mL), (*E*)-styrenyl bromide (0.73 g, 4.0 mmol, 1.0 equiv.) and the corresponding terminal alkyne **IV** (4.8 mmol, 1.2 equiv.) were added. The mixture was allowed to stir at 55 °C for 16 h. At the conclusion of the reaction, H₂O (15 mL) and Et₂O (15 mL) were added to the reaction mixture, and the organic layer was separated. The aqueous layer was extracted with another portion of Et₂O (10 mL). The organic layers were combined, dried, and concentrated. The resulting crude residue was purified by silica gel chromatography using hexanes as the eluent to obtain the product as white solids. Characterization data of enyne **2a** to **2c** are consistent with reported in the literature.¹⁴



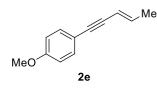
2d, 76% yield. ¹H NMR (600 MHz, CDCl₃) δ 7.91 (d, J = 1.7 Hz, 2H), 7.83 (s, 1H), 7.52 – 7.44 (m, 2H), 7.44 – 7.33 (m, 3H), 7.17 (d, J = 16.3 Hz, 1H), 6.41 (d, J = 16.2 Hz, 1H); ¹³C NMR (151 MHz, CDCl₃) δ 143.6, 135.9, 132.1 (q, J = 33.8 Hz), 131.4 (q, J = 4.0 Hz), 129.4, 129.0, 126.7, 126.0, 123.2 (q, J = 272.8 Hz), 121.5 (dq, J = 7.7, 3.9 Hz), 106.9, 92.6, 88.6; ¹⁹F NMR (564 MHz, CDCl₃) δ –65.8; IR (ATR) 3033, 2201, 1464,

1385, 1277, 1244, 1174, 1132, 1107, 1048, 953, 896, 848, 748, 684 cm⁻¹; HRMS (DART) calcd for $C_{18}H_{11}F_6$ ([M+H]⁺) 341.07595, found 341.07569.

General Method for Enyne Synthesis by Kumada Coupling:

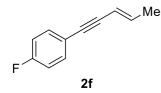


The protocol for was adapted from literature procedure.¹⁵ Under nitrogen to a 20-mL vial charged with $Pd(dppf)Cl_2$ (44 mg, 0.060 mmol, 0.020 equiv.) and (*E*)-1-bromopropene (436 mg, 3.60 mmol, 1.20 equiv.) was added alkynyl magnesium bromide **V** (3.0 mmol, 1.0 equiv.) in THF (12 mL) at room temperature. The mixture was allowed to stir for another 14 hours at room temperature. Then the solvent was removed *in vacuo*, and the residue was dissolved in Et₂O (10 mL). This organic solution was further washed by H₂O (10 mL) and brine (10 mL) and dried with Na₂SO₄. Volatiles were removed under reduced pressure, and the resulting crude residue was purified by silica gel column chromatography using hexane as the eluent to afford the product as colorless liquids.



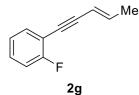
2e, with 1-bromopropene mixture (4.6 equiv., E/Z = 26:74). Crude ¹H NMR indicates the ratio of E/Z enyne is 95:5. Purification provides pure **2e**, 56% yield. ¹H NMR (500 MHz, CDCl₃) δ 7.40 – 7.31 (m, 2H), 6.87 – 6.76 (m, 2H), 6.21 (dd, J = 15.8, 6.8 Hz, 1H), 5.70 (dq, J = 15.7, 1.8 Hz, 1H), 3.80 (s, 3H), 1.83 (dd, J = 6.8, 1.8 Hz, 3H); ¹³C NMR (126 MHz,

CDCl₃) δ 159.4, 139.1, 132.9, 115.9, 114.0, 111.1, 87.7, 87.0, 55.4, 18.8; IR (ATR) 3001, 2959, 2934, 2912, 2837, 1603, 1506, 1441, 1286, 1244, 1171, 1036, 950, 828, 803, 533 cm⁻¹; HRMS (DART) calcd for C₁₂H₁₃O ([M+H]⁺) 173.09609, found 173.09544.



2f, 74% yield. ¹H NMR (500 MHz, CDCl₃) δ 7.43 – 7.34 (m, 2H), 7.07 – 6.90 (m, 2H), 6.24 (dqd, *J* = 14.8, 6.8, 1.2 Hz, 1H), 5.69 (dp, *J* = 15.8, 1.8 Hz, 1H), 1.84 (dt, *J* = 6.9, 1.5 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 162.4 (d, *J* = 248.9 Hz), 140.1, 133.4 (d, *J* = 8.2 Hz), 119.9 (d, *J* = 3.4 Hz), 115.7 (d, *J* = 22.0 Hz), 110.8, 88.0 (d, *J* = 1.4 Hz), 86.7, 18.9; ¹⁹F

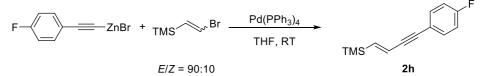
NMR (470 MHz, CDCl₃) δ –111.7 (dp, *J* = 13.6, 4.8 Hz); IR (ATR) 2914, 1599, 1504, 1229, 1155, 1092, 950, 909, 831, 678, 527, 503 cm⁻¹; HRMS (DART) calcd for C₁₁H₁₀F ([M+H]⁺) 161.07610, found 161.07598.



2g, 80% yield. ¹H NMR (500 MHz, CDCl₃) δ 7.41 (td, J = 7.5, 1.8 Hz, 1H), 7.26 (dddd, J = 8.1, 7.2, 5.2, 1.8 Hz, 1H), 7.15 – 6.98 (m, 2H), 6.31 (dq, J = 15.7, 6.8 Hz, 1H), 5.75 (dq, J = 15.8, 1.8 Hz, 1H), 1.85 (dd, J = 6.9, 1.9 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 162.6 (d, J = 250.9 Hz), 140.8, 133.4 (d, J= 1.4 Hz), 129.6 (d, J = 8.0 Hz), 124.0 (d, J = 3.8 Hz), 115.5 (d, J = 21.1 Hz),

112.4 (d, J = 15.8 Hz), 110.7, 93.5 (d, J = 3.3 Hz), 81.1, 18.9 (d, J = 2.2 Hz); ¹⁹F NMR (470 MHz, CDCl₃) δ –110.4 (dt, J = 8.7, 6.0 Hz); IR (ATR) 3031, 2914, 2207, 1572, 1491, 1447, 1256, 1218, 1106, 951, 912, 823, 754, 577 cm⁻¹; HRMS (DART) calcd for C₁₁H₁₀F ([M+H]⁺) 161.07610, found 161.07542.

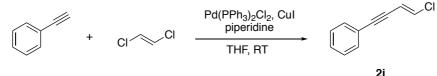
Synthesis of Enyne 2h by Negishi Coupling:



The protocol was adapted from literature procedure.¹⁶ Under nitrogen to a 20-mL vial charged with $Pd(PPh_3)_4$ (92 mg, 0.080 mmol, 0.020 equiv.) was added a solution of ((4-fluorophenyl)ethynyl)zinc bromide (3.42 mmol, 0.850 equiv.) in THF (15 mL) followed by (2-bromovinyl)trimethylsilane (716 mg, 4.00 mmol, 1.00 equiv., E/Z = 90/10) at room temperature. The resulting mixture was allowed to stir for 14 hours at room temperature. Then the solvent was removed *in vacuo* and the residue was dissolved in Et_2O (10 mL). This organic solution was further washed by H₂O (10 mL) and concentrated. The crude product was carefully purified by silica gel chromatography using hexanes as the eluent to afford **2h** as a colorless liquid (57% yield).

¹H NMR (500 MHz, CDCl₃) δ 7.46 – 7.36 (m, 2H), 7.01 (t, *J* = 8.7 Hz, 2H), 6.54 (d, *J* = 19.2 Hz, 1H), 6.15 (d, *J* = 19.2 Hz, 1H), 0.13 (s, 9H); ¹³C NMR (126 MHz, CDCl₃) δ 162.6 (d, *J* = 249.6 Hz), 146.1, 133.6 (d, *J* = 8.2 Hz), 123.3, 119.6 (d, *J* = 3.6 Hz), 115.7 (d, *J* = 22.0 Hz), 89.5 (d, *J* = 1.5 Hz), 88.8, -1.5; ¹⁹F NMR (470 MHz, CDCl₃) δ –116. 3 (m); IR (ATR) 2956, 1601, 1569, 1505, 1249, 1231, 1155, 975, 863, 833, 740, 727, 694, 658, 529 cm⁻¹; HRMS (DART) calcd for C₁₃H₁₆FSi ([M+H]⁺) 219.09998, found 219.09980.

Synthesis of Enyne 2i by Sonogashira Coupling:

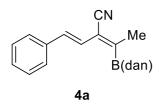


Under nitrogen a 50-mL round bottom flask was charged with $PdCl_2(PPh_3)_2$ (42 mg, 0.060 mmol, 0.020 equiv.) and Cul (23 mg, 0.12 mol, 0.040 equiv.). To this flask Et_2O (6 mL), piperidine (1.02 g, 12.0 mmol, 4.00 equiv.), phenylacetylene (306 mg, 3.00 mmol, 1.00 equiv.) and *trans*-dichloroethylene (437 mg, 4.50 mmol, 1.50 equiv.) were added. The mixture was allowed to stir at room temperature for 16 h. At the conclusion of the reaction H_2O (15 mL) and Et_2O (15 mL) were added, and the organic layer was separated. The aqueous layer was further washed by Et_2O (10 mL). The organic layers were combined, concentrated, and purified by silica gel chromatography using pentane as the eluent to obtain **2i** as a colorless liquid (87%).

¹H NMR (500 MHz, CDCl₃) δ 7.49 – 7.39 (m, 2H), 7.33 (dd, J = 5.0, 1.9 Hz, 3H), 6.63 (d, J = 13.6 Hz, 1H), 6.16 (d, J = 13.6 Hz, 1H); ¹³C NMR (126 MHz, CDCl₃) δ 131.7, 130.3, 128.8, 128.5, 122.8, 114.0, 92.1, 84.5; IR (ATR) 3067, 3031, 2204, 1581, 1488, 1442, 1272, 1224, 912, 843, 752, 687, 528, 498 cm⁻¹; HRMS (DART) calcd for C₁₀H₇Cl ([M+H]⁺) 162.02308, found 162.02267.

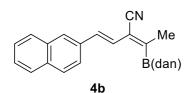
General Procedure for *trans*-Selective Cyanoboration Reaction of 1,3-Enynes (Table 2)

To a 4-mL vial charged with $Pd(COD)(CH_2TMS)_2$ (3.9 mg, 0.010 mmol, 5.0 mol %), **L1** or **L5** (mol % is indicated in the corresponding entries), CuCN (29 mg, 0.32 mmol, 1.6 equiv), BCatCl (55 mg, 0.36 mmol, 1.8 equiv.), and *o*-DCB (0.30 mL) was added the corresponding enyne substrate (0.20 mmol, 1.0 equiv). The resulting mixture was allowed to stir for 2.5 hours at the temperature indicated in the entries. At the conclusion of reaction, the mixture was passed through an acrodisc with CH_2Cl_2 under nitrogen. 1,8-Diaminonaphthlene (64 mg, 0.40 mmol, 2.0 equiv) was then added to the filtrate and the mixture was allowed to stir for another 12 hours. Then the mixture was passed through an acrodisc and concentrated under reduced pressure. The resulting crude residue was purified by silica gel chromatography using hexanes/ CH_2Cl_2 (4/1 to 1/1 gradient) as the eluent to afford the pure *trans*-selective cyanoboration products as yellow solids.



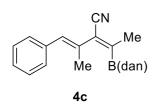
4a, with 5 mol % L1 at 115 °C, 86% yield (run1: 87%, run2: 85%), 96:4. ¹H MR (500 MHz, CDCl₃) δ 7.40 (d, *J* = 7.0 Hz, 2H), 7.31 (t, *J* = 7.2 Hz, 2H), 7.27 (d, *J* = 7.0 Hz, 1H), 7.18 – 7.09 (m, 4H), 7.03 (d, *J* = 15.9 Hz, 1H), 6.93 (d, *J* = 15.8 Hz, 1H), 6.39 (dd, *J* = 7.1, 1.3 Hz, 2H), 5.70 (s, 2H), 2.33 (s, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 140.1, 136.4, 135.8, 134.0, 129.0, 128.8, 127.8, 127.1, 123.9, 120.8, 120.1, 118.8, 115.6, 106.7, 22.5

(*B*-alkenyl carbon signal not observed); ¹¹B NMR (160 MHz, CDCl₃) δ 28.8; IR (ATR) 3397, 3055, 2220, 1600, 1505, 1408, 1373, 1335, 1095, 820, 767 cm⁻¹; HRMS (DART) calcd for C₂₂H₁₉BN₃ ([M+H]⁺) 336.16665, found 336.16979.



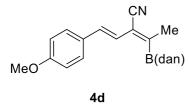
4b, with 5 mol % **L1** at 115 °C, 77% yield (run1: 75%, run2: 79%), 98:2. Crystals for single crystal X-ray diffraction analysis were obtained by recrystallization from hexane/CH₂Cl₂ at room temperature. ¹H NMR (500 MHz, CDCl₃) δ 7.77 (dt, *J* = 7.0, 2.6 Hz, 3H), 7.72 (d, *J* = 8.6 Hz, 1H), 7.53 (dd, *J* = 8.6, 1.7 Hz, 1H), 7.49 –

7.41 (m, 2H), 7.21 – 7.10 (m, 5H), 7.03 (d, J = 15.8 Hz, 1H), 6.41 (dd, J = 7.2, 1.3 Hz, 2H), 5.77 (s, 2H), 2.34 (s, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 152.5 (br), 140.2, 136.4, 134.0, 133.6, 133.5, 133.3, 128.7, 128.4, 127.9, 127.8, 127.8, 126.6, 126.6, 124.1, 123.3, 120.8, 120.1, 118.8, 115.7, 106.7, 22.6; ¹¹B NMR (160 MHz, CDCl₃) δ 28.9; IR (ATR) 3378, 3053, 2924, 2217, 1627, 1600, 1504, 1407, 1373, 1336, 1094, 953, 819, 765 cm⁻¹; HRMS (DART) calcd for C₂₆H₂₁BN₃ ([M+H]⁺) 386.18230, found 386.18179.



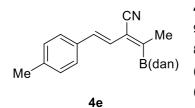
4c, with 5 mol % **L1** at 115 °C, 63% yield (run1: 65%, run2: 61%), >98:2. ¹H NMR (500 MHz, CDCl₃) δ 7.32 (t, J = 7.5 Hz, 2H), 7.27 – 7.19 (m, 3H), 7.11 (t, J = 7.8 Hz, 2H), 7.05 (d, J = 8.2 Hz, 2H), 6.86 (s, 1H), 6.32 (d, J = 7.2 Hz, 2H), 5.68 (s, 2H), 2.31 (s, 3H), 2.16 (d, J = 1.4 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 140.1, 136.5, 136.4, 133.0, 132.9, 129.2, 128.5, 127.7, 127.6, 125.4, 119.8, 118.6, 117.0, 106.5, 22.9, 18.1 (*B*-alkenyl

carbon signal not observed); ¹¹B NMR (160 MHz, CDCl₃) δ 29.0; IR (ATR) 3396, 3054, 2925, 2211, 1599, 1504, 1373, 1332, 1109, 820, 765, 701 cm⁻¹; HRMS (DART) calcd for C₂₃H₂₁BN₃ ([M+H]⁺) 350.18230, found 350.18304.



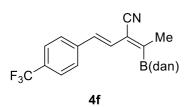
4d, with 1.5 equiv. BcatCl and 5 mol % **L1** at 115 °C, 64% yield (run1: 65%, run2: 62%), 96:4. ¹H NMR (600 MHz, CDCl₃) δ 7.35 – 7.31 (m, 2H), 7.15 (dd, *J* = 8.3, 7.2 Hz, 2H), 7.11 (dd, *J* = 8.4, 1.1 Hz, 2H), 6.96 (d, *J* = 15.8 Hz, 1H), 6.86 – 6.82 (m, 2H), 6.80 (d, *J* = 15.8 Hz, 1H), 6.39 (dd, *J* = 7.2, 1.1 Hz, 2H), 5.73 (s, 2H), 3.79 (s, 3H), 2.31 (s, 3H); ¹³C NMR (151 MHz, CDCl₃) δ 160.2, 150.9 (br), 140.2, 136.4, 133.4,

128.6, 128.4, 127.8, 121.8, 120.9, 120.1, 118.7, 115.8, 114.4, 106.6, 55.4, 22.4; ¹¹B NMR (160 MHz, CDCl₃) δ 28.8; IR (ATR) 3358, 3053, 2932, 2219, 1598, 1508, 1407, 1373, 1333, 1247, 1173, 1032, 1095, 820, 767 cm⁻¹; HRMS (DART) calcd for C₂₃H₂₁BN₃O ([M+H]⁺) 366.17722, found 366.17843.



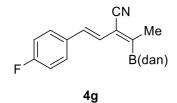
4e, with 5 mol % **L1** at 115 °C, 87% yield (run1: 88%, run2: 85%), 97:3. ¹H NMR (500 MHz, CDCl₃) δ 7.32 – 7.28 (m, 2H), 7.15 (dd, *J* = 8.3, 7.1 Hz, 2H), 7.13 – 7.09 (m, 4H), 7.00 (d, *J* = 15.8 Hz, 1H), 6.88 (d, *J* = 15.8 Hz, 1H), 6.38 (dd, *J* = 7.1, 1.2 Hz, 2H), 5.70 (s, 2H), 2.33 (s, 3H), 2.32 (s, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 140.2, 138.9, 136.4, 133.9, 133.1, 129.7, 127.8, 127.0, 122.9, 120.9, 120.1, 118.8,

115.7, 106.7, 22.4, 21.4 (*B*-alkenyl carbon signal not observed); ¹¹B NMR (160 MHz, CDCl₃) δ 28.6; IR (ATR) 3396, 3052, 2917, 2220, 1629, 1600, 1505, 1408, 1373, 1334, 1095, 820, 766 cm⁻¹; HRMS (DART) calcd for C₂₃H₂₁BN₃ ([M+H]⁺) 350.18230, found 350.18279.



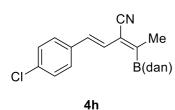
4f, with 5 mol % **L1** at 115 °C, 80% yield (run1: 77%, run2: 82%), >98:2. ¹H NMR (600 MHz, THF- d_8) δ 7.65 (d, J = 8.2 Hz, 2H), 7.58 (d, J = 8.2 Hz, 2H), 7.48 (s, 2H), 7.21 (d, J = 15.9 Hz, 1H), 7.06 – 6.99 (m, 3H), 6.96 (dd, J = 8.4, 0.9 Hz, 2H), 6.37 (dd, J = 7.3, 1.0 Hz, 2H), 2.34 (s, 3H); ¹³C NMR (126 MHz, THF- d_8) δ 158.0 (br), 142.6, 141.2, 137.7, 131.6, 130.49 (q, J = 32.0 Hz), 128.6, 128.4, 128.2,

126.63 (q, *J* = 3.9 Hz), 125.5 (q, *J* = 272.2 Hz), 121.7, 120.2, 118.6, 115.8, 107.0, 22.9; ¹¹B NMR (160 MHz, THF-*d*₈) δ 29.5; ¹⁹F NMR (564 MHz, THF-*d*₈) δ –63.4; IR (ATR) 3411, 3371, 2922, 2852, 2217, 1602, 1506, 1408, 1329, 1177, 1105, 1068, 820, 764, 690, 455 cm⁻¹; HRMS (DART) calcd for $C_{23}H_{18}BN_3F_3$ ([M+H]⁺) 404.15404, found 404.15382.



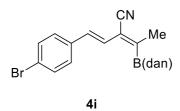
4g, with 5 mol % **L1** at 115 °C, 66% yield (run1: 66%, run2: 65%), 96:4. ¹H NMR (600 MHz, CDCl₃) δ 7.35 (dd, J = 8.6, 5.5 Hz, 2H), 7.16 (\mathbf{t} , J = 7.7 Hz, 2H), 7.12 (d, J = 8.2 Hz, 2H), 7.01 – 6.93 (m, 3H), 6.84 (d, J = 15.8 Hz, 1H), 6.40 (d, J = 7.2 Hz, 2H), 5.74 (s, 2H), 2.33 (s, 3H); ¹³C NMR (151 MHz, CDCl₃) δ 163.0 (d, J = 249.1 Hz), 152.7 (br), 140.1,

136.4, 132.6, 132.03 (d, J = 3.4 Hz), 128.7 (d, J = 8.1 Hz), 127.8, 123.63 (d, J = 2.6 Hz), 120.4, 120.1, 118.8, 116.0 (d, J = 21.9 Hz), 115.6, 106.7, 22.5; ¹¹B NMR (160 MHz, CDCl₃) δ 28.7; ¹⁹F NMR (564 MHz, CDCl₃) δ -112.3 (tt, J = 8.6, 5.3 Hz); IR (ATR) 3404, 3385, 3052, 2216, 1601, 1505, 1407, 1376, 1334, 1227, 1092, 819, 763, 681 cm⁻¹; HRMS (DART) calcd for C₂₂H₁₈BN₃F ([M+H]⁺) 354.15723, found 354.15886.



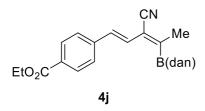
4h, with 5 mol % **L1** at 115 °C, 84% yield (run1: 82%, run2: 86%), >98:2. ¹H NMR (600 MHz, CDCl₃) δ 7.32 (d, *J* = 8.5 Hz, 2H), 7.27 (d, *J* = 8.4 Hz, 2H), 7.18 – 7.08 (m, 4H), 6.96 (d, *J* = 15.8 Hz, 1H), 6.89 (d, *J* = 15.8 Hz, 1H), 6.39 (d, *J* = 7.1 Hz, 2H), 5.70 (s, 2H), 2.33 (s, 3H); ¹³C NMR (151 MHz, CDCl₃) δ 153.2 (br), 140.0, 136.4, 134.5, 134.3, 132.6, 129.2, 128.2, 127.8, 124.4, 120.5, 120.1, 118.9, 115.4,

106.7, 22.6; ¹¹B NMR (160 MHz, CDCl₃) δ 28.8; IR (ATR) 3403, 3379, 2217, 1604, 1508, 1411, 1337, 1095, 820, 765 cm⁻¹; HRMS (DART) calcd for C₂₂H₁₈BN₃Cl ([M+H]⁺) 370.12768, found 370.12759.



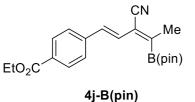
4i, with 5 mol % **L1** at 115 °C, 55% yield (run1: 53%, run2: 57%), 95:5. ¹H NMR (500 MHz, THF- d_8) δ 7.49 – 7.34 (m, 6H), 7.10 (dd, J = 15.9, 1.0 Hz, 1H), 7.03 (ddd, J = 8.3, 7.2, 1.0 Hz, 2H), 6.97 – 6.88 (m, 3H), 6.36 (dd, J = 7.3, 1.1 Hz, 2H), 2.31 (s, 3H); ¹³C NMR (126 MHz, THF- d_8) δ 142.7, 137.7, 136.5, 132.9, 132.0, 129.5, 128.4, 126.8, 122.9, 121.6, 120.3, 118.5, 115.9, 107.0, 22.8 (*B*-alkenyl carbon

signal not observed); ¹¹B NMR (160 MHz, THF- d_8) δ 29.0; IR (ATR) 3403, 3374, 2216, 1603, 1506, 1408, 1376, 1335, 1094, 1072, 1007, 819, 763, 754, 686 cm⁻¹; HRMS (DART) calcd for C₂₂H₁₈BN₃Br ([M+H]⁺) 414.07717, found 414.07770.



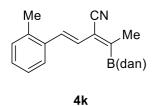
4j, with 5 mol % **L1** at 115 °C, 69% yield, (run1: 71%, run2: 67%), >98:2. ¹H NMR (600 MHz, CDCl₃) δ 7.99 – 7.94 (m, 2H), 7.47 – 7.41 (m, 2H), 7.18 – 7.09 (m, 4H), 7.02 (d, *J* = 4.1 Hz, 2H), 6.39 (dd, *J* = 7.0, 1.2 Hz, 2H), 5.73 (s, 2H), 4.35 (q, *J* = 7.1 Hz, 2H), 2.35 (s, 3H), 1.37 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (151 MHz, CDCl₃) δ 166.3, 140.1, 140.0, 136.4, 132.8, 130.3, 130.2, 127.8, 126.9,

126.0, 120.5, 120.1, 119.0, 115.3, 106.8, 61.2, 22.7, 14.5 (*B*-alkenyl carbon signal not observed); ¹¹B NMR (160 MHz, CDCl₃) δ 28.9; IR (ATR) 3372, 2223, 1691, 1603, 1510, 1409, 1335, 1290, 1272, 1182, 1097, 765, 698 cm⁻¹; HRMS (DART) calcd for C₂₅H₂₃BN₃O₂ ([M+H]⁺) 408.18778, found 408.18712.



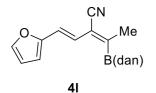
4j-B(pin), with 5 mol % **L1** at 115 °C, pinacol (236 mg, 2.00 mmol, 10.0 equiv.) in CH_2Cl_2 (5 mL) was used rather than 1,8-diaminonaphthlene, purified by silica gel chromatography using hexanes/EtOAc (10/1) as the eluent, 56% yield (run1: 58%, run2: 53%), >98:2. Crystals for single crystal X-ray diffraction analysis were obtained by recrystallization from hexane/CH₂Cl₂ at room

temperature. ¹H NMR (600 MHz, CDCl₃) δ 8.06 – 8.00 (m, 2H), 7.76 (d, *J* = 16.0 Hz, 1H), 7.51 – 7.45 (m, 2H), 7.03 (d, *J* = 16.0 Hz, 1H), 4.38 (q, *J* = 7.1 Hz, 2H), 2.25 (s, 3H), 1.40 (t, *J* = 7.1 Hz, 3H), 1.35 (s, 12H); ¹³C NMR (151 MHz, CDCl₃) δ 166.4, 140.9, 133.1, 130.2 (2 signals), 127.1, 126.9, 125.7, 115.5, 84.7, 61.1, 25.0, 21.9, 14.5 (*B*-alkenyl carbon signal not observed); ¹¹B NMR (160 MHz, CDCl₃) δ 29.4; IR (ATR) 2979, 2929, 2221, 1712, 1605, 1361, 1269, 1141, 1102, 1019, 764, 847, 679, 627 cm⁻¹; HRMS (DART) calcd for C₂₁H₂₇BNO₄ ([M+H]⁺) 368.20277, found 368.20267.



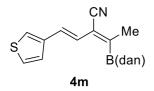
4k, with 5 mol % **L1** at 115 °C, 58% yield (run1: 60%, run2: 56%), 94:6. ¹H NMR (600 MHz, CDCl₃) δ 7.40 (d, J = 7.7 Hz, 1H), 7.32 – 7.23 (m, 1H), 7.20 – 7.08 (m, 7H), 6.86 (d, J = 15.7 Hz, 1H), 6.39 (dd, J = 7.2, 1.2 Hz, 2H), 5.74 (s, 2H), 2.42 (s, 3H), 2.34 (s, 3H); ¹³C NMR (151 MHz, CDCl₃) δ 152.6 (br), 140.1, 136.5, 136.3, 134.8, 131.6, 130.7, 128.6, 127.8, 126.4, 125.6, 125.0, 120.9, 120.0, 118.7, 115.7, 106.7, 22.5, 19.9; ¹¹B NMR (160 MHz,

CDCl₃) δ 28.9; IR (ATR) 3395, 3055, 2926, 2219, 1629, 1600, 1505, 1408, 1373, 1335, 1093, 957, 908, 820, 766, 732 cm⁻¹; HRMS (DART) calcd for C₂₃H₂₁BN₃ ([M+H]⁺) 350.18230, found 350.18173.



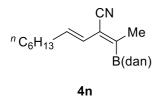
4I, with 5 mol% **L5** at 105 °C, 71% yield (run1: 69%, run2: 73%), 98:2. ¹H NMR (500 MHz, CDCl₃) δ 7.34 (d, *J* = 1.6 Hz, 1H), 7.20 – 7.05 (m, 4H), 6.91 – 6.74 (m, 2H), 6.39 (m, 4H), 5.73 (s, 2H), 2.31 (s, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 151.8, 143.3, 140.2, 136.3, 127.8, 122.2, 121.2, 120.4, 120.1, 118.7, 115.5, 112.1, 111.3, 106.7, 22.5 (*B*-alkenyl carbon signal not

observed); ¹¹B NMR (160 MHz, CDCl₃) δ 28.8; IR (ATR) 3389, 3053, 2924, 2220, 1629, 1598, 1503, 1407, 1334, 1263, 1094, 1013, 820, 766, 736 cm⁻¹; HRMS (DART) calcd for C₂₀H₁₇BN₃O ([M+H]⁺) 326.14592, found 326.14575.



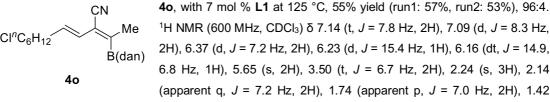
4m, with 5 mol % **L1** at 115 °C, 72% yield (run1: 70%, run2: 74%), 97:3. ¹H NMR (500 MHz, CDCl₃) δ 7.28 (dd, *J* = 3.1, 1.2 Hz, 1H), 7.24 (dd, *J* = 3.0, 0.6 Hz, 1H), 7.19 – 7.09 (m, 5H), 7.03 (d, *J* = 15.8 Hz, 1H), 6.77 (d, *J* = 15.8 Hz, 1H), 6.38 (dd, *J* = 7.1, 1.2 Hz, 2H), 5.71 (s, 2H), 2.31 (s, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 140.1, 138.6, 136.4, 128.0, 127.8, 126.8, 125.0, 124.6,

123.8, 120.7, 120.1, 118.8, 115.6, 106.7, 22.4 (*B*-alkenyl carbon signal not observed); ¹¹B NMR (160 MHz, CDCl₃) δ 28.9; IR (ATR) 3394, 3053, 2219, 1628, 1599, 1504, 1407, 1373, 1335, 1095, 820, 766, 650 cm⁻¹; HRMS (DART) calcd for C₂₀H₁₇BN₃S ([M+H]⁺) 342.12308, found 342.12416.



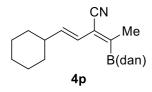
4n, with 7 mol % **L1** at 125 °C, 60% yield (run1: 62%, run2: 57%), 96:4. ¹H NMR (600 MHz, CDCl₃) δ 7.14 (dd, *J* = 8.3, 7.3 Hz, 2H), 7.09 (dd, *J* = 8.3, 1.0 Hz, 2H), 6.37 (dd, *J* = 7.2, 1.0 Hz, 2H), 6.24 (d, *J* = 15.4 Hz, 1H), 6.18 (dt, *J* = 15.4, 6.7 Hz, 1H), 5.69 (d, *J* = 4.4 Hz, 2H), 2.24 (s, 3H), 2.14 (q, *J* = 7.2 Hz, 2H), 1.41 (h, *J* = 6.8 Hz, 2H), 1.34 – 1.20 (m, 6H), 0.88 (t, *J* = 6.9 Hz, 3H); ¹³C NMR (151 MHz, CDCl₃) δ 149.8 (br), 140.2, 137.1,

136.3, 127.7, 125.5, 120.4, 120.0, 118.6, 115.9, 106.5, 32.7, 31.7, 29.1, 29.0, 22.7, 22.0, 14.2; ¹¹B NMR (160 MHz, CDCl₃) δ 28.8; IR (ATR) 3367, 3054, 2954, 2853, 2218, 1598, 1505, 1406, 1372, 1361, 1333, 1095, 957, 820, 764, 664 cm⁻¹; HRMS (DART) calcd for C₂₂H₂₇BN₃ ([M+H]⁺) 344.22925, found 344.22994.



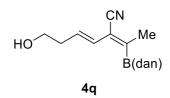
(apparent h, J = 7.5 Hz, 4H), 1.32 (apparent p, J = 7.6 Hz, 2H); ¹³C NMR (151 MHz, CDCl₃) δ 140.2,

136.8, 136.4, 127.7, 125.8, 120.4, 120.0, 118.7, 115.8, 106.5, 45.2, 32.6, 32.6, 29.0, 28.5, 26.8, 22.0 (*B*-alkenyl carbon signal not observed); ¹¹B NMR (160 MHz, CDCl₃) δ 29.0; IR (ATR) 3393, 3054, 2927, 2854, 2218, 1628, 1598, 1504, 1406, 1373, 1333, 1095, 820, 765, 730, 665 cm⁻¹; HRMS (DART) calcd for C₂₂H₂₆BN₃Cl ([M+H]⁺) 378.19028, found 378.18893.



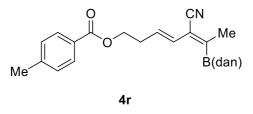
4p, with 7 mol % **L1** at 125 °C, 60% yield (run1: 61%, run2: 59%), 93:7. ¹H NMR (600 MHz, CDCl₃) δ 7.14 (dd, J = 8.3, 7.2 Hz, 2H), 7.09 (dd, J = 8.3, 1.0 Hz, 2H), 6.37 (dd, J = 7.3, 1.0 Hz, 2H), 6.20 (dd, J = 15.5, 1.0 Hz, 1H), 6.12 (dd, J = 15.5, 7.1 Hz, 1H), 5.66 (s, 2H), 2.24 (s, 3H), 2.06 (tdt, J = 10.9, 7.0, 3.4 Hz, 1H), 1.71 (ddq, J = 13.6, 7.2, 3.4 Hz, 4H), 1.64 (dtd, J = 12.4,

3.6, 1.7 Hz, 1H), 1.25 (m, 2H), 1.20 – 1.06 (m, 3H); ¹³C NMR (151 MHz, CDCl₃) δ 149.8 (br), 142.6, 140.2, 136.4, 127.7, 123.3, 120.7, 120.0, 118.6, 115.9, 106.5, 41.1, 32.8, 26.0, 25.9, 22.1; ¹¹B NMR (160 MHz, CDCl₃) δ 28.7; IR (ATR) 3429, 3393, 2926, 2852, 2218, 1600, 1504, 1408, 1335, 1264, 1093, 963, 820, 732, 703 cm⁻¹; HRMS (DART) calcd for C₂₂H₂₅BN₃ ([M+H]⁺) 342.21360, found 342.21457.



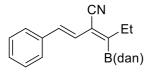
1q was pre-treated with HBCat (29 mg, 0.24 mmol, 1.2 equiv.) in CH_2CI_2 (1.0 mL) followed by removal of volatiles under high vacuum (150 mmTor) for 30 min at room temperature. This material was then used for the *trans*-selective cyanoboration reaction with 7 mol % **L1** at 125 °C, 35% yield (run1: 32%, run2: 37%), 98:2. **4q** appears to decompose

slowly on silica gel column. ¹H NMR (500 MHz, CDCl₃) δ 7.13 (dd, *J* = 8.3, 7.2 Hz, 2H), 7.08 (dd, *J* = 8.4, 1.2 Hz, 2H), 6.39 – 6.31 (m, 3H), 6.17 (dt, *J* = 15.0, 7.1 Hz, 1H), 5.66 (s, 2H), 3.73 (t, *J* = 6.4 Hz, 2H), 2.46 – 2.35 (m, 2H), 2.25 (s, 3H), 1.39 (s, 1H); ¹³C NMR (151 MHz, CDCl₃) δ 151.2 (br), 140.2, 136.3, 132.5, 128.1, 127.7, 120.1, 120.0, 118.6, 115.7, 106.6, 61.7, 35.9, 22.1; ¹¹B NMR (160 MHz, CDCl₃) δ 28.9; IR (ATR) 3387, 3053, 2926, 2219, 1599, 1509, 1408, 1373, 1335, 1096, 1036, 821, 768 cm⁻¹; HRMS (DART) calcd for C₁₈H₁₉BN₃O ([M+H]⁺) 304.16157, found 304.16183.



4r, with 7 mol % **L1** at 135 °C, 73% yield (run1: 71%, run2: 75%), 96:4. ¹H NMR (600 MHz, CDCl₃) δ 7.93 – 7.83 (m, 2H), 7.18 (d, *J* = 8.0 Hz, 2H), 7.15 – 7.04 (m, 4H), 6.44 – 6.36 (m, 1H), 6.32 (dd, *J* = 7.2, 1.1 Hz, 2H), 6.23 (dt, *J* = 14.9, 7.1 Hz, 1H), 5.69 (s, 2H), 4.36 (t, *J* = 6.5 Hz, 2H), 2.60 (q, *J* = 6.7 Hz, 2H), 2.39 (s, 3H), 2.24 (s,

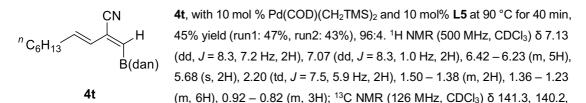
3H); ¹³C NMR (151 MHz, CDCl₃) δ 166.6, 151.6 (br), 143.8, 140.1, 136.3, 131.5, 129.7, 129.2, 128.1, 127.7, 127.4, 120.0, 120.0, 118.7, 115.6, 106.6, 63.6, 32.0, 22.2, 21.8; ¹¹B NMR (160 MHz, CDCl₃) δ 28.5; IR (ATR) 3367, 2957, 2219, 1702, 1599, 1508, 1407, 1373, 1335, 1274, 1178, 1096, 908, 820, 768, 754, 731 cm⁻¹; HRMS (DART) calcd for C₂₆H₂₅BN₃O₂ ([M+H]⁺) 422.20343, found 422.20384.



4s

4s, with 5 mol % **L1** at 115 °C, 66% yield (run1: 63%, run2: 68%), 93:7. ¹H NMR (500 MHz, CDCl₃) δ 7.40 (d, J = 7.4 Hz, 2H), 7.31 (t, J = 7.3 Hz, 2H), 7.27 (d, J = 6.9 Hz, 1H), 7.17 (t, J = 7.7 Hz, 2H), 7.12 (d, J = 8.2 Hz, 2H), 7.04 (d, J = 15.9 Hz, 1H), 6.91 (d, J = 15.8 Hz, 1H), 6.40 (d, J = 7.1 Hz, 2H), 5.74 (s, 2H), 2.71 (q, J = 7.6 Hz, 2H), 1.22 (t, J = 7.6 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 159.2 (br), 140.1, 136.4, 135.9, 134.1, 128.9, 128.7, 127.8,

127.1, 124.1, 120.1, 119.5, 118.8, 115.5, 106.7, 30.1, 14.0; ^{11}B NMR (160 MHz, CDCl₃) δ 29.0; IR (ATR) 3382, 3052, 2966, 2214, 1599, 1504, 1406, 1373, 1333, 1166, 1103, 951, 818, 761, 750, 686 cm $^{-1}$; HRMS (DART) calcd for $C_{23}H_{21}\text{BN}_3$ ([M+H]+) 350.18230, found 350.18183.

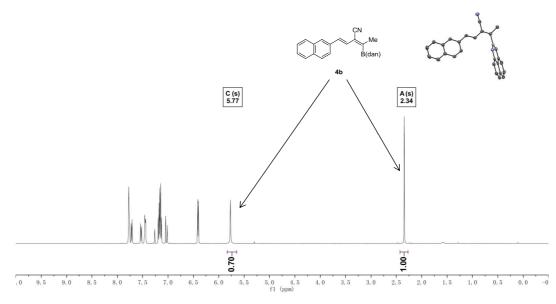


137.7 (br), 136.4, 127.7, 126.6, 125.0, 120.1, 118.7, 117.9, 106.5, 32.9, 31.8, 29.0, 28.9, 22.7, 14.2; ^{11}B NMR (160 MHz, CDCI₃) δ 27.5; IR (ATR) 3387, 2923, 2853, 2223, 1629, 1598, 1502, 1406, 1373, 1331, 1165, 1143, 1047, 962, 819, 763 cm⁻¹; HRMS (DART) calcd for C₂₁H₂₅BN₃ ([M+H]⁺) 330.21360, found 330.21405.

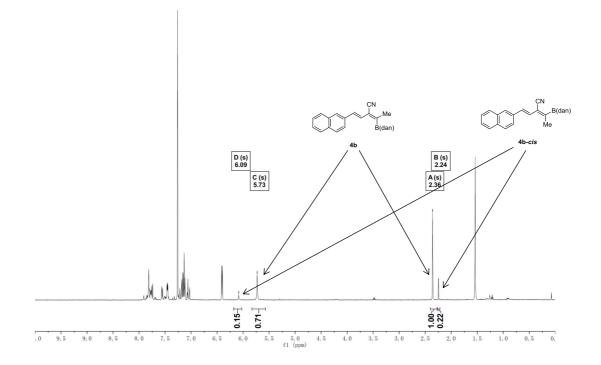
Assignment of the trans/cis Configuration of the Products

The trans/cis configuration of **4b** and **4j-B(pin)** was unambiguously determined by single-crystal X-ray diffraction analysis of the corresponding major pure isolated product. The assignment of the trans/cis configuration of the other products shown in Table 2 and Table 3 are made by analogy based on characteristic ¹H NMR signature of the Me and NH(Bdan) signals: the Me signal of the *trans* product (signal A) appears more downfield than the Me signal (signal B) of the *cis* product, and the NH(Bdan) signal of the *trans* product (signal C) appears more upfield than the NH(Bdan) signal (signal D) of the *cis* product (see below spectra). A mixture of *trans/cis* products can oftentimes be obtained in small quantities during the silica gel chromatography purification process. The labeled ¹H NMR spectra are shown using compound **4b** as the representative example.

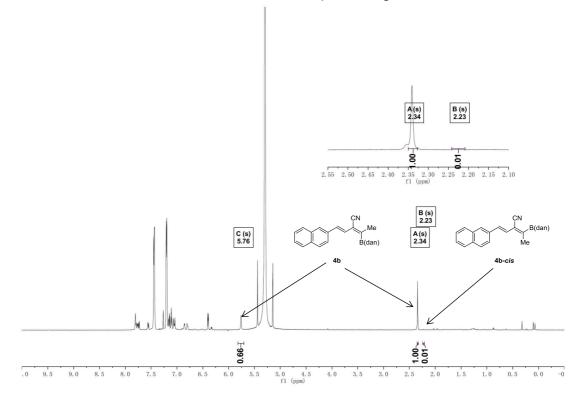
Fraction 2 of the crude mixture after column purification, pure 4b.



Fraction 1 of the crude mixture after column purification, 4b and 4b-cis mixture.

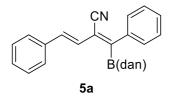


Crude mixture of **4b** and **4b-cis** after quenching with dan.



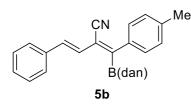
General Procedure for *trans*-Selective Cyanoboration Reaction of 1,3-Enynes (Table 3)

To a 4-mL vial charged with $Pd(COD)(CH_2TMS)_2$ (3.9 mg, 0.010 mmol, 10 mol %), L5 (4.3 mg, 0.010 mmol, 10 mol %), CuCN (16 mg, 0.18 mmol, 1.8 equiv), BCatCl (28 mg, 0.18 mmol, 1.8 equiv.) and *o*-DCB (0.10 mL) was added the corresponding enyne (0.10 mmol, 1.0 equiv). The resulting mixture was allowed to stir for 1 hour at 105 °C. At the conclusion of the reaction, the mixture was passed through an acrodisc with CH₂Cl₂ under nitrogen. 1,8-diaminonaphthlene (32 mg, 0.20 mmol, 2.0 equiv) was then added to the filtrate, and the mixture was allowed to stir for another 12 hours. The reaction mixture was then again passed through an acrodisc and concentrated under reduced pressure. The resulting crude residue was purified by silica gel chromatography using hexanes/CH₂Cl₂ (4/1 to 1/1 gradient) as the eluent to afford the pure *trans*-selective cyanoboration product as yellow solids.



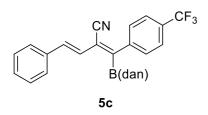
5a, 60% yield (run1: 62%, run2: 57%), 86:14. ¹H NMR (500 MHz, CDCl₃) δ 7.59 (d, J = 7.5 Hz, 2H), 7.49 – 7.38 (m, 5H), 7.32 (m, 3H), 7.24 (d, J = 15.4 Hz, 1H), 7.14 (m, 5H), 6.37 (d, J = 6.8 Hz, 2H), 5.79 (s, 2H); ¹³C NMR (151 MHz, CDCl₃) δ 140.2, 138.9, 136.4, 135.7, 129.5, 129.1, 129.0, 129.0, 128.8, 127.8, 127.6, 127.3, 124.9, 120.1, 119.1, 118.9,

116.6, 106.8 (*B*-alkenyl carbon signal not observed); ¹¹B NMR (160 MHz, CDCl₃) δ 29.1; IR (ATR) 3396, 3055, 2219, 1628, 1599, 1504, 1407, 1373, 1336, 1149, 820, 766, 695 cm⁻¹; HRMS (DART) calcd for C₂₇H₂₁BN₃ ([M+H]⁺) 398.18230, found 398.18286.



5b, 48% yield (run1: 46%, run2: 50%), 75:25. ¹H NMR (500 MHz, CDCl₃) δ 7.51 (d, J = 7.9 Hz, 2H), 7.45 (d, J = 7.0 Hz, 2H), 7.37 – 7.18 (m, 6H), 7.19 – 7.09 (m, 5H), 6.36 (dd, J = 7.0, 1.4 Hz, 2H), 5.79 (s, 2H), 2.40 (s, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 140.2, 139.8, 136.4, 136.0, 135.9, 135.3, 129.7, 129.0, 129.0, 128.9, 127.8, 127.3, 125.0, 120.1, 118.8, 118.3, 116.9, 106.8, 21.5

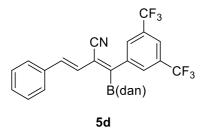
(*B*-alkenyl carbon signal not observed); ¹¹B NMR (160 MHz, CDCl₃) δ 28.8; IR (ATR) 3419, 3055, 2923, 2219, 1599, 1507, 1406, 1374, 1338, 1260, 1147, 1052, 961, 908, 819, 765, 732 cm⁻¹; HRMS (DART) calcd for C₂₈H₂₃BN₃ ([M+H]⁺) 412.19795, found 412.19878.



5c, 84% yield (run1: 82%, run2: 86%), 93:7. ¹H NMR (600 MHz, CDCl₃) δ 7.70 (d, J = 8.5 Hz, 2H), 7.67 (d, J = 8.3 Hz, 2H), 7.49 – 7.42 (m, 2H), 7.37 – 7.30 (m, 3H), 7.27 (d, J = 15.6 Hz, 1H), 7.18 – 7.12 (m, 5H), 6.38 (dd, J = 6.7, 1.6 Hz, 2H), 5.78 (s, 2H); ¹³C NMR (151 MHz, CDCl₃) δ 152.3 (br), 142.6, 139.9, 137.0, 136.3, 135.4, 131.1 (q, J = 32.9 Hz), 129.5, 129.1, 129.1, 127.8, 127.5,

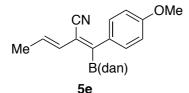
126.0 (q, *J* = 3.8 Hz), 124.4, 124.0 (q, *J* = 272.4 Hz), 120.9, 120.1, 119.1, 116.1, 106.9; ¹¹B NMR (160 MHz, CDCl₃) δ 29.0; ¹⁹F NMR (564 MHz, CDCl₃) δ –62.8; IR (ATR) 3394, 3055, 2921, 2217, 1628, 1598, 1502, 1408, 1373, 1334, 1148, 907, 820, 767, 730, 692, 648 cm⁻¹; HRMS (DART) calcd for C₂₈H₂₀BN₃F₃ ([M+H]⁺) 466.16969, found 466.16856.

5d, 86% yield (run1: 88%, run2: 83%), >98:2. ¹H NMR (600 MHz, CDCl₃) δ 7.96 (d, *J* = 1.5 Hz, 2H), 7.93 (s, 1H), 7.52 – 7.45 (m, 2H), 7.39 – 7.34 (m, 3H), 7.32 (d, *J* = 15.8 Hz, 1H), 7.20 – 7.13 (m, 5H), 6.41 (dd,



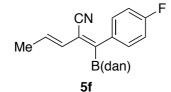
J = 5.9, 2.4 Hz, 2H), 5.79 (s, 2H); ¹³C NMR (151 MHz, CDCl₃) δ 150.1 (br), 141.2, 139.7, 138.2, 136.3, 135.2, 132.5 (q, *J* = 33.7 Hz), 129.8, 129.2, 128.8 (d, *J* = 3.5 Hz), 128.7 (d, *J* = 4.1 Hz), 127.8, 127.6, 123.8, 123.1 (q, *J* = 273.0 Hz), 122.9 (p, *J* = 3.8 Hz), 122.7, 119.4, 115.5, 107.1; ¹¹B NMR (160 MHz, CDCl₃) δ 28.7; ¹⁹F NMR (564 MHz, CDCl₃) δ –62.8; IR (ATR) 3394, 3057, 2224, 1600, 1503, 1407, 1372, 1275, 1176, 1129, 820, 736, 680 cm⁻¹;

HRMS (DART) calcd for C₂₉H₁₉BN₃F₆ ([M+H]⁺) 534.15707, found 534.15663.



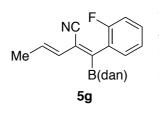
5e, obtained by using hexane/Et₂O (5/1) as eluent, 71% yield (run1: 69%, run2: 73%), 94:6. ¹H NMR (500 MHz, CDCl₃) δ 7.58 – 7.48 (m, 2H), 7.18 – 7.07 (m, 4H), 6.96 – 6.89 (m, 2H), 6.47 – 6.31 (m, 4H), 5.73 (s, 2H), 3.83 (s, 3H), 1.88 (dd, J = 6.3, 1.1 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 160.4, 151.5 (br), 140.3, 136.3, 132.9, 131.0,

130.4, 128.1, 127.7, 120.0, 118.6, 117.3, 116.8, 114.3, 106.6, 55.4, 18.4; ¹¹B NMR (160 MHz, CDCl₃) δ 29.0; IR (ATR) 3366, 2930, 2214, 1629, 1598, 1507, 1409, 1373, 1333, 1248, 1180, 1154, 1031, 953, 908, 821, 768, 732 cm⁻¹; HRMS (DART) calcd for C₂₃H₂₁BN₃O ([M+H]⁺) 366.17722, found 366.17607.



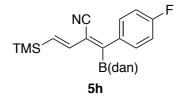
5f, 70% yield (run1: 67%, run2: 72%), 95:5. ¹H NMR (500 MHz, CDCl₃) δ 7.52 (dd, *J* = 8.4, 5.3 Hz, 2H), 7.17 – 7.04 (m, 6H), 6.47 – 6.39 (m, 2H), 6.35 (dd, *J* = 7.0, 1.2 Hz, 2H), 5.70 (s, 2H), 1.90 (d, *J* = 4.9 Hz, 3H); ¹³C NMR (151 MHz, CDCl₃) δ 163.1 (d, *J* = 250.1 Hz), 150.6 (br), 140.1, 136.3, 134.9 (d, *J* = 3.5 Hz), 134.3 (two signals),

130.6 (d, J = 8.3 Hz), 127.8, 120.0, 119.0, 118.9, 116.7, 116.0 (d, J = 21.8 Hz), 106.7, 18.5; ¹¹B NMR (160 MHz, CDCl₃) δ 29.0; ¹⁹F NMR (470 MHz, CDCl₃) δ –111.5 (tt, J = 9.1, 5.2 Hz); IR (ATR) 3369, 2919, 2213,1600, 1504, 1413, 1374, 1335, 1231, 1159, 959, 820, 768 cm⁻¹; HRMS (DART) calcd for C₂₂H₁₈BN₃F ([M+H]⁺) 354.15723, found 354.15671.



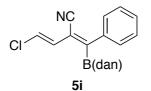
5g, 63% yield (run1: 64%, run2: 61%), 98:2. Crystals for single crystal X-ray diffraction analysis were obtained by recrystallization from hexane/CH₂Cl₂ at -30 °C. ¹H NMR (500 MHz, THF-*d*₈) δ 7.51 – 7.35 (m, 3H), 7.30 (tdd, *J* = 7.3, 5.1, 1.8 Hz, 1H), 7.16 (td, *J* = 7.6, 1.1 Hz, 1H), 7.10 (ddd, *J* = 9.7, 8.4, 1.1 Hz, 1H), 6.97 (t, *J* = 7.8 Hz, 2H), 6.89 (dd, *J* = 8.3, 1.0 Hz, 2H), 6.59 (dq, *J* = 15.3, 1.6 Hz, 1H), 6.37 – 6.23

(m, 3H), 1.84 (dd, J = 6.8, 1.6 Hz, 3H); ¹³C NMR (126 MHz, THF- d_8) δ 160.6 (d, J = 247.6 Hz), 142.6, 137.5, 133.8, 131.5 (d, J = 2.8 Hz), 131.1 (d, J = 8.2 Hz), 129.2, 128.8 (d, J = 14.8 Hz), 128.2, 125.1 (d, J = 3.5 Hz), 122.6, 121.5, 118.3, 116.6 (d, J = 22.0 Hz), 116.3, 106.8, 18.2 (*B*-alkenyl carbon signal not observed); ¹¹B NMR (160 MHz, THF- d_8) δ 25.4; ¹⁹F NMR (470 MHz, THF- d_8) δ -110.2 (m); IR (ATR) 3060, 2977, 2923, 2218, 1638, 1555, 1447, 1394, 1360, 1334, 1266, 1229, 1139, 976, 953, 850, 801, 755 cm⁻¹; HRMS (DART) calcd for C₂₂H₁₈BN₃F ([M+H]⁺) 354.15723, found 354.15738.



5h, 38% yield (run1: 40%, run2: 36%), 94:6. ¹H NMR (500 MHz, CDCl₃) δ 7.56 – 7.48 (m, 2H), 7.19 – 7.06 (m, 6H), 6.89 (d, *J* = 18.5 Hz, 1H), 6.67 (d, *J* = 18.5 Hz, 1H), 6.34 (dd, *J* = 7.0, 1.4 Hz, 2H), 5.70 (s, 2H), 0.13 (s, 9H); ¹³C NMR (126 MHz, CDCl₃) δ 163.3 (d, *J* = 250.7 Hz), 153.5 (br), 140.0, 139.2, 138.6, 136.3, 134.9 (d, *J* = 3.6 Hz),

130.8 (d, J = 8.5 Hz), 127.8, 121.5, 120.0, 118.9, 116.5, 116.1 (d, J = 21.8 Hz), 106.6, -1.3; ¹¹B NMR (160 MHz, CDCl₃) δ 29.8; ¹⁹F NMR (470 MHz, CDCl₃) δ –110.9 (m); IR (ATR) 3360, 3056, 2955, 2897, 2218, 1599, 1505, 1409, 1373, 1336, 1247, 1235, 1160, 1145, 839, 820, 766 cm⁻¹; HRMS (DART) calcd for C₂₄H₂₄BN₃FSi ([M+H]⁺) 412.18111, found 412.18054.

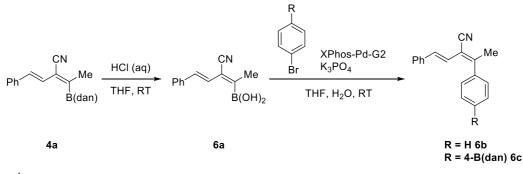


5i, with 15 mol % **L5**, 1.2 equiv. CuCN and 1.6 equiv. BCatCl at 90 °C for 25 min, obtained by using hexane/Et₂O (8/1) as the eluent, 28% yield (run1: 27%, run2: 28%), 96:4. ¹H NMR (600 MHz, CDCl₃) δ 7.59 – 7.49 (m, 2H), 7.49 – 7.35 (m, 3H), 7.18 – 7.04 (m, 4H), 6.89 (d, *J* = 1.1 Hz, 2H), 6.36 (dt, *J* = 6.9, 1.3 Hz, 2H), 5.71 (s, 2H); ¹³C NMR (151 MHz, CDCl₃) δ 139.8, 138.1,

136.3, 130.1, 129.9, 129.1, 128.7, 127.8, 126.2, 120.1, 119.1, 115.8, 115.2, 106.9 (*B*-alkenyl carbon signal not observed); ¹¹B NMR (160 MHz, CDCl₃) δ 29.0; IR (ATR) 3397, 3365, 3062, 2220, 1629, 1599, 1505, 1407, 1374, 1336, 1151, 1051, 820, 766, 732, 696 cm⁻¹; HRMS (DART) calcd for C₂₁H₁₆BN₃Cl ([M+H]⁺) 356.11203, found 356.11198.

Functionalization of Cyanoboration Product 4a (Scheme 4)

Reaction sequence:

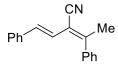


Step 1:

Aqueous HCI (1.4 mL, 3.0 M) was added into a 20-mL vial charged with 4a (33 mg, 0.10 mmol, 1.0 equiv.) and THF (2.8 mL). The resulting mixture was allowed to stir at room temperature for 30 minutes. At the conclusion of reaction, the mixture was diluted with Et₂O (4 mL), and the organic layer was separated. The aqueous layer was extracted with Et₂O (5 mL). The organic layers were combined and concentrated to afford **6a** as a mixture of boronic acid and the trimer boroxine. **6a** was used for the next step without further purification (quant. yield).

Step 2:

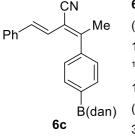
Under nitrogen to a 4-mL vial charged with 6a (from Step 1), XPhos-Pd-G2 (4 mg, 0.005 mmol, 0.05 equiv.), THF (0.3 mL), and aqueous K₃PO₄ (0.3 mL, 0.5 M) the corresponding aryl bromide (0.15 mmol, 1.5 equiv.) was added. The resulting mixture was allowed to stir for 3 hours at room temperature. Then, H₂O (1 mL) was added, and the organic layer was separated. The aqueous layer was further extracted with Et₂O (3 mL), and the organic layers were combined and concentrated. The resulting crude material was purified by silica gel chromatography using hexanes/EtOAc (10/1) as the eluent to afford 6b-c.



6b

6b, 86% yield. ¹H NMR (500 MHz, CDCl₃) δ 7.48 – 7.36 (m, 3H), 7.31 – 7.16 (m, 7H), 7.01 (d, J = 16.0 Hz, 1H), 6.71 (d, J = 16.0 Hz, 1H), 2.53 (s, 3H); ¹³C NMR $(126 \text{ MHz}, \text{ CDCl}_3) \delta$ 153.7, 139.3, 136.3, 133.2, 129.1, 128.8, 128.8, 128.4, 128.1, 126.9, 122.2, 117.0, 112.2, 25.3; IR (ATR) 3058, 2922, 2219, 1492, 1441,

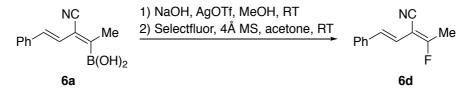
1274, 1025, 960, 765, 754, 699, 693 cm⁻¹; HRMS (DART) calcd for C₁₈H₁₆N ([M+H]⁺) 246.12773, found 246.12774.



6c, 85% yield. ¹H NMR (500 MHz, CDCl₃) δ 7.72 (d, J = 7.8 Hz, 2H), 7.38 – 7.22 (m, 7H), 7.18 (t, J = 7.8 Hz, 2H), 7.10 (d, J = 8.3 Hz, 2H), 7.05 (d, J = 16.0 Hz, 1H), 6.74 (d, J = 16.0 Hz, 1H), 6.46 (d, J = 7.2 Hz, 2H), 6.09 (s, 2H), 2.56 (s, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 153.1, 141.0, 140.9, 136.5, 136.2, 133.6, 131.9, 128.8, 128.5, 127.9, 127.8, 127.0, 122.0, 120.0, 118.2, 116.9, 112.4, 106.3, 25.2 (B-aryl carbon signal not observed); ¹¹B NMR (160 MHz, CDCl₃) δ 29.0; IR (ATR) 3416, 3054, 2218, 1599, 1524, 1493, 1408, 1374, 1332, 1088, 820, 764, 692

cm⁻¹; HRMS (DART) calcd for C₂₈H₂₃BN₃ ([M+H]⁺) 412.19795, found 412.19759.

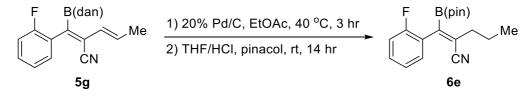
Synthesis of Compound 6d



Under nitrogen to a 4-mL vial charged with a solution of NaOH (9.6 mg, 0.24 mmol, 1.2 equiv.) in MeOH (2.0 mL) was added **6a** (0.20 mmol, 1.0 equiv.) at room temperature. The resulting mixture was allowed to stir for 15 minutes, and then the mixture was cooled to 0 °C. AgOTf (154 mg, 0.600 mmol, 3.00 equiv.) was added in the absence of light, and the resulting mixture was allowed to stir for 30 minutes at 0°C. Then, methanol was removed *in vacuo*, and anhydrous acetone (1 mL) was added and then removed under reduced pressure to help remove methanol. After repeating for four times, 4Å Molecular sieve (100 mg), Selectfluor (85 mg, 0.24 mmol, 1.2 equiv.), and acetone (2 mL) were added to the residue. The resulting slurry was allowed to stir for 2 hours at room temperature. At the conclusion of reaction, the mixture was passed through an acrodisc and washed with acetone. The filtrate was concentrated, dissolved in Et₂O (5 mL) and further washed with H₂O (5 mL). The organic layer was dried with Na₂SO₄, and **6d** was obtained via silica gel chromatography using hexanes/EtOAc (15/1) as the eluent as a white solid (67% yield).

¹H NMR (500 MHz, CDCl₃) δ 7.45 (d, *J* = 7.2 Hz, 2H), 7.35 (t, *J* = 7.4 Hz, 2H), 7.33 – 7.27 (m, 1H), 6.88 (s, 2H), 2.39 (d, *J* = 17.0 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 168.7 (d, *J* = 280.5 Hz), 135.9 (d, *J* = 1.3 Hz), 132.7 (d, *J* = 4.2 Hz), 128.9, 128.8, 127.0, 115.6 (d, *J* = 3.6 Hz), 115.3 (d, *J* = 13.7 Hz), 98.8 (d, *J* = 28.6 Hz), 18.3 (d, *J* = 24.7 Hz); ¹⁹F NMR (564 MHz, CDCl₃) δ –75.6 (q, *J* = 17.0 Hz); IR (ATR) 3027, 2960, 2923, 2229, 1651, 1625, 1496, 1430, 1388, 1312, 1303, 1216, 1184, 1059, 1028, 953, 802, 688 cm⁻¹; HRMS (DART) calcd for C₁₂H₁₁NF ([M+H]⁺) 188.08700, found 188.08652.

Synthesis of Compound 6e (eq1)



Palladium on carbon (12 mg, 0.011 mmol, 10% wt, 0.20 equiv.), **5g** (20 mg, 0.057 mmol, 1.0 equiv.), and EtOAc (0.60 mL) were added into a 4-mL vial. The vial was quickly placed under vacuum and then back filled with H_2 (1 atm) twice, and the mixture was allowed to heat at 40 °C for 3 hours under a H_2 balloon pressue. At the conclusion of reaction, the reaction mixture was passed through an acrodisc, and all volaties were removed *in vacuo*. Pinacol (47 mg, 0.40 mmol, 7.0 equiv.) was added to this residue along with THF (1.5 mL) and aqueous HCI (0.75 mL, 4.3 M). The resulting mixture was allowed to stir for 14 hours at room temperature followed by addition of Et_2O (5 mL) and H_2O (5 mL). The organic layer was separated and the aqueous layer was further washed with Et_2O (5 mL). The combined organic layers were concentrated, and the crude material was purified via silica gel chromatography using hexanes/EtOAc (8/1) as the eluent to afford **6e** as a white solid (54% yield).

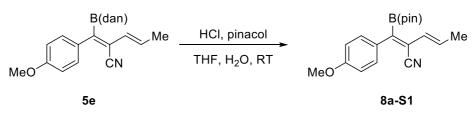
¹H NMR (500 MHz, CDCl₃) δ 7.39 (td, *J* = 7.7, 1.8 Hz, 1H), 7.35 – 7.29 (m, 1H), 7.17 (td, *J* = 7.6, 1.1 Hz, 1H), 7.06 (ddd, *J* = 9.6, 8.3, 1.1 Hz, 1H), 2.68 – 2.61 (m, 2H), 1.71 (h, *J* = 7.4 Hz, 2H), 1.28 (s, 12H), 1.01 (t, *J* = 7.4 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 159.6 (d, *J* = 246.5 Hz), 130.4, 130.4 (d, *J* = 10.6 Hz),

128.0, 127.5 (d, *J* = 15.1 Hz), 124.4 (d, *J* = 3.4 Hz), 118.4, 115.5 (d, *J* = 21.9 Hz), 84.7, 36.0, 24.8, 22.3, 13.4 (*B*-alkenyl carbon signal not observed); ¹¹B NMR (160 MHz, CDCl₃) δ 29.9; ¹⁹F NMR (470 MHz, CDCl₃) δ –112.8 (m); IR (ATR) 2963, 2929, 2873, 2213, 1486, 1450, 1357, 1327, 1268, 1140, 1081, 971, 846, 753 cm⁻¹; HRMS (DART) calcd for $C_{18}H_{24}BNO_2F$ ([M+H]⁺) 316.18786, found 316.18866.

Synthesis of Satigrel (Scheme 5)

Reaction sequence:

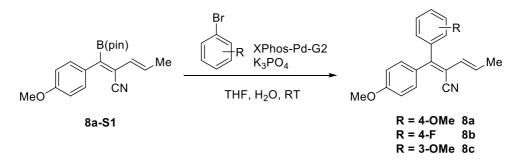
Step 1:



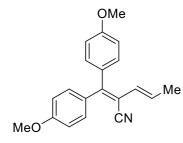
To a 20-mL vial charged with **5e** (146 mg, 0.400 mmol, 1.00 equiv.), pinacol (236 mg, 2.00 mmol, 5.00 equiv.) and THF (8 mL) was added followed by aqueous HCI (6.0 mL, 4.3 M). The resulting mixture was allowed to stir at room temperature for 20 hours. At the conclusion of reaction, Et_2O (4 mL) was added to the mixture, and the organic layer was separated. The aqueous layer was further extracted with additional Et_2O (5 mL). The combined organic layers were concentrated under reduce pressure, and the resulting crude material was further purified by silica gel chromatography using hexanes/EtOAc (5/1) as the eluent to afford **8a-S1** as a white solid (84% yield).

¹H NMR (500 MHz, CDCl₃) δ 7.40 – 7.34 (m, 2H), 6.94 – 6.86 (m, 2H), 6.69 (dq, *J* = 15.4, 1.6 Hz, 1H), 6.38 (dq, *J* = 15.3, 6.8 Hz, 1H), 3.82 (s, 3H), 1.90 (dd, *J* = 6.8, 1.7 Hz, 3H), 1.33 (s, 12H); ¹³C NMR (126 MHz, CDCl₃) δ 159.9, 133.7, 131.4, 130.1, 127.9, 121.9, 116.9, 113.9, 84.8, 55.4, 24.9, 18.6 (*B*-alkenyl carbon signal not observed); ¹¹B NMR (160 MHz, CDCl₃) δ 30.0; IR (ATR) 2978, 2934, 2838, 2218, 1604, 1509, 1357, 1328, 1291, 1250, 1178, 1137, 1032, 972, 849, 832, 732 cm⁻¹; HRMS (DART) calcd for C₁₉H₂₅BNO₃ ([M+H]⁺) 326.19220, found 326.19142.

Step 2:

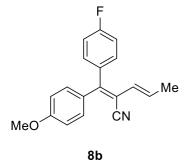


Under nitrogen to a 20-mL vial charged with **8a-S1** (109 mg, 0.330 mmol, 1.00 equiv.) and XPhos-Pd-G2 (13 mg, 0.016 mmol, 0.050 equiv.) was added THF (2.7 mL), aqueous K_3PO_4 (0.9 mL, 0.6 M) and the corresponding aryl bromide (0.50 mmol, 1.5 equiv.). The resulting mixture was allowed to stir for 15 hours at room temperature. Then, H₂O (1 mL) was added and the organic layer was separated. The aqueous layer was further extracted with Et₂O (3 mL), and the organic layers were combined and concentrated. This crude material was purified by silica gel chromatography using hexanes/EtOAc (15/1) as the eluent to afford **8a-8c** as white solids.



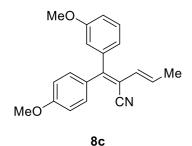
8a

8a with 4-bromo-anisole, 88% yield. ¹H NMR (500 MHz, CDCl₃) δ 7.33 – 7.28 (m, 2H), 7.13 – 7.07 (m, 2H), 6.93 – 6.85 (m, 4H), 6.32 (dq, *J* = 15.4, 6.6 Hz, 1H), 6.21 (dq, *J* = 15.4, 1.5 Hz, 1H), 3.85 (s, 3H), 3.84 (s, 3H), 1.82 (dd, *J* = 6.7, 1.5 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 160.7, 160.3, 153.3, 132.7, 132.1, 131.7, 131.4, 131.3, 126.6, 118.8, 113.8, 113.7, 108.8, 55.4, 55.4, 18.4; IR (ATR) 3005, 2930, 2838, 2214, 1602, 1573, 1506, 1461, 1442, 1302, 1282, 1247, 1172, 1029, 960, 829, 737, 603, 572 cm⁻¹; HRMS (DART) calcd for $C_{20}H_{20}NO_2$ ([M+H]⁺) 306.14886, found 306.14963.



8b with 4-bromo-fluorobenzene, 80% yield. ¹H NMR (500 MHz, CDCl₃) δ 7.35 – 7.27 (m, 2H), 7.15 (ddt, *J* = 8.2, 5.1, 2.5 Hz, 2H), 7.12 – 7.04 (m, 2H), 6.92 – 6.84 (m, 2H), 6.35 (dq, *J* = 15.4, 6.8 Hz, 1H), 6.10 (dq, *J* = 15.4, 1.6 Hz, 1H), 3.83 (s, 3H), 1.82 (dd, *J* = 6.8, 1.6 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 163.1 (d, *J* = 249.9 Hz), 160.9, 152.2, 135.2 (d, *J* = 3.4 Hz), 132.5, 132.4, 132.2, 131.6, 126.2, 118.4, 115.6 (d, *J* = 21.5 Hz), 113.9, 110.0, 55.5, 18.5; ¹⁹F NMR (470 MHz, CDCl₃) δ –111.6 (ddd, *J* = 13.5, 8.4, 5.1 Hz); IR (ATR) 2960, 2926, 2850, 2216, 1602, 1575, 1505, 1461, 1444,

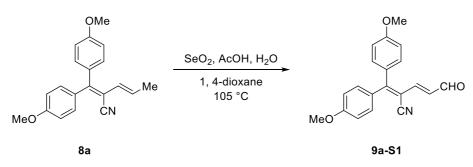
1289, 1251, 1228, 1173, 1158, 1095,1028, 960 831, 804, 736, 566 cm⁻¹; HRMS (DART) calcd for $C_{19}H_{16}NOF$ ([M+H]⁺) 293.12104, found 293.11995.



8c with 3-bromo-anisole, 90% yield. ¹H NMR (600 MHz, CDCl₃) δ 7.35 – 7.24 (m, 3H), 6.93 (dd, J = 8.4, 2.5 Hz, 1H), 6.91 – 6.84 (m, 2H), 6.76 (dt, J = 7.6, 1.2 Hz, 1H), 6.68 (dd, J = 2.7, 1.5 Hz, 1H), 6.38 – 6.25 (m, 1H), 6.15 (dq, J = 15.4, 1.5 Hz, 1H), 3.83 (d, J = 0.9 Hz, 3H), 3.78 (d, J = 0.9 Hz, 3H), 1.81 (dt, J = 6.8, 1.2 Hz, 3H); ¹³C NMR (151 MHz, CDCl₃) δ 160.7, 159.6, 153.2, 140.5, 132.2, 132.0, 131.5, 129.5, 126.4, 122.8, 118.5, 115.9, 114.5, 113.8, 109.9, 55.5, 55.5, 18.5; IR (ATR) 3004, 2935, 2913, 2837, 2216, 1604, 1576,

1509, 1462, 1446, 1287, 1253, 1178, 1034, 961, 837, 802, 787, 700 cm⁻¹; HRMS (DART) calcd for $C_{20}H_{20}NO_2$ ([M+H]⁺) 306.14886, found 306.14950.

Step 3:

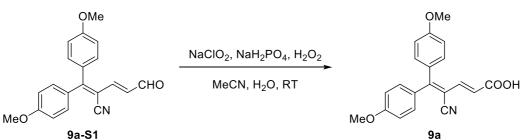


SeO₂ (162 mg, 1.45 mmol, 5.00 equiv.) and **8a** (89 mg, 0.29 mmol, 1.0 equiv.) were placed in a 20-mL vial. To this vial 1,4-dioxane (1.8 mL), H_2O (0.10 mL) and AcOH (0.050 mL) were added successively.

The resulting mixture was allowed to stir at 105 °C for 12 hours. Then, another portion of SeO₂ (64 mg, 0.58 mmol, 2.0 equiv.) was added, and the mixture was allowed to stir for another 12 hours at 105 °C. At the conclusion of reaction, the reaction mixture was passed through celite, and the filtrate was concentrated. This crude residue was purified by silica gel chromatography using hexanes/EtOAc (4/1) as the eluent to afford **9a-S1** as a yellow solid (77% yield).

¹H NMR (500 MHz, CDCl₃) δ 9.55 (d, *J* = 7.5 Hz, 1H), 7.46 – 7.36 (m, 2H), 7.25 (d, *J* = 15.5 Hz, 1H), 7.18 – 7.12 (m, 2H), 7.00 – 6.95 (m, 2H), 6.95 – 6.91 (m, 2H), 6.67 (dd, *J* = 15.5, 7.5 Hz, 1H), 3.89 (s, 3H), 3.87 (s, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 192.6, 164.5, 162.4, 162.0, 147.1, 133.1, 132.9, 131.3, 131.0 (two signals), 130.1, 114.2, 114.0, 105.7, 55.6, 55.6; IR (ATR) 3007, 2934, 2839, 2730, 2214, 1672, 1593, 1573, 1501, 1253, 1171, 1116, 1027, 835, 732 cm⁻¹; HRMS (DART) calcd for C₂₀H₁₈NO₃ ([M+H]⁺) 320.12812, found 320.12696.

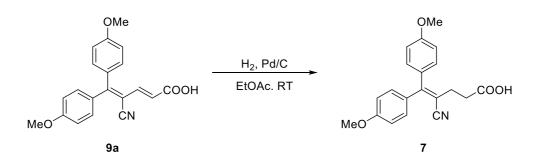
Step 4:



 H_2O (0.50 mL), MeCN (0.13 mL) and aqueous H_2O_2 (35% v/v, 0.25 mmol, 25 µl, 2.0 equiv.) were added to a 4-mL vial charged with **9a-S1** (40.4 mg, 0.125 mmol, 1.00 equiv.) and NaH₂PO₄ (7.8 mg, 0.065 mmol, 0.52 equiv.). The mixture was cooled to 0 °C and then a pre-cooled solution (0 °C) of NaClO₂ (35 mg, 0.31 mmol, 2.5 equiv.) in H₂O (0.31 mL) was added in a dropwise fashion. The mixture was allowed to stir for 30 min and then allowed to warm to room temperature. At the conclusion of reaction, Et₂O (5 mL) and H₂O (3 mL) were added to this mixture. The organic layer was separated, and the concentrated organic layer was purified by silica gel chromatography using hexanes/EtOAc (1/1) as the eluent to afford **9a** as a yellow solid (69% yield).

¹H NMR (500 MHz, CDCl3) δ 11.79 (s, br, 1H), 7.53 (d, *J* = 15.4 Hz, 1H), 7.41 – 7.35 (m, 2H), 7.14 – 7.07 (m, 2H), 6.98 – 6.88 (m, 4H), 6.42 (d, *J* = 15.5 Hz, 1H), 3.88 (s, 3H), 3.86 (s, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 171.8, 164.0, 162.2, 161.9, 142.4, 133.2, 132.8, 131.6, 130.3, 120.6, 117.8, 114.2, 114.0, 105.5, 55.6, 55.6; IR (ATR) 2934, 2839, 2213, 1685, 1598, 1508, 1286, 1255, 1210, 1173, 1029, 836, 732 cm⁻¹; HRMS (DART) calcd for C₂₀H₁₈NO₄ ([M+H]⁺) 336.12303, found 336.12345.

Step 5:



A 4-mL vial was charged with **9a** (10.2 mg, 0.0300 mmol, 1.00 equiv), Pd/C (3.1 mg, 0.0030 mmol, 10% wt, 0.10 equiv) and EtOAc (0.4 mL). The vial was quickly placed under vacuum and then and back filled with H_2 (1 atm) twice. The resulting mixture was allowed to stir at room temperature for 2 hours under a H_2 balloon pressure. At the conclusion of reaction, all violates were removed *in vacuo*, and the residue was purified by silica gel chromatography using EtOAc as the eluent to afford **7** as a white solid (91% yield, 97:3 hydrogenation selectivity (**7** *vs.* fully hydrogenated byproduct)).

¹H NMR (600 MHz, CDCl₃) δ 7.27 (d, *J* = 8.8 Hz, 2H), 7.07 – 7.01 (m, 2H), 6.92 – 6.83 (m, 4H), 3.83 (s, 3H), 3.82 (s, 3H), 2.71 (apparent s, 4H), (COOH signal not observed); ¹³C NMR (151 MHz, CDCl₃) δ 173.6 (br), 160.7, 160.3, 158.2, 132.4, 131.3, 131.1, 131.0, 120.0, 114.0, 113.8, 107.0, 55.5, 55.5, 32.8, 27.4; IR (ATR) 2931, 2839, 2204, 1709, 1605, 1509, 1283, 1249, 1174, 1030, 833 cm⁻¹; HRMS (DART) calcd for $C_{20}H_{20}NO_4$ ([M+H]⁺) 338.13868, found 338.13898.

Crystallographic Data for 4b, 4j-B(pin) and 5g

Crystal Data for 4b:

Identification code	lentification code C26H20BN3	
Empirical formula	C26 H20 B N3	
Formula weight	385.26	
Temperature	123(2) K	
Wavelength	1.54178 Å	
Crystal system	Tetragonal	
Space group	$P4_2/n$	
Unit cell dimensions	a = 23.0525(5) Å	α= 90°.
	b = 23.0525(5) Å	β= 90°.
	c = 7.6338(2) Å	$\gamma = 90^{\circ}$.
Volume	4056.7(2) Å ³	
Ζ	8	
Density (calculated)	1.262 Mg/m ³	
Absorption coefficient	0.574 mm ⁻¹	
F(000)	1616	
Crystal size	$0.560 \ge 0.220 \ge 0.160 \text{ mm}^3$	
Theta range for data collection	2.711 to 66.840°.	
Index ranges	-27<=h<=22, -27<=k<=27, -9<=l<=9	
Reflections collected	22309	
Independent reflections	ons $3569 [R(int) = 0.0276]$	
Completeness to theta = 66.840°	99.2 %	
Absorption correction	Semi-empirical from equivalents	
Max. and min. transmission	0.7528 and 0.6580	
Refinement method	Full-matrix least-squares on F ²	
Data / restraints / parameters	3569 / 2 / 278	
Goodness-of-fit on F ²	1.060	
Final R indices [I>2sigma(I)]	R1 = 0.0338, wR2 = 0.0869	
R indices (all data)	R1 = 0.0345, wR2 = 0.0874	
Extinction coefficient	n/a	
Largest diff. peak and hole	0.206 and -0.156 e.Å ⁻³	

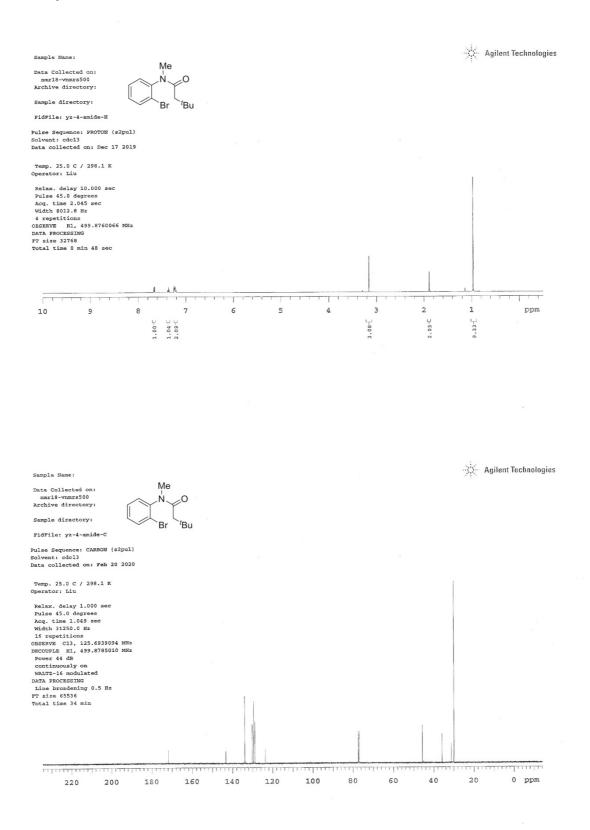
Crystal Data for 4j-B(pin):

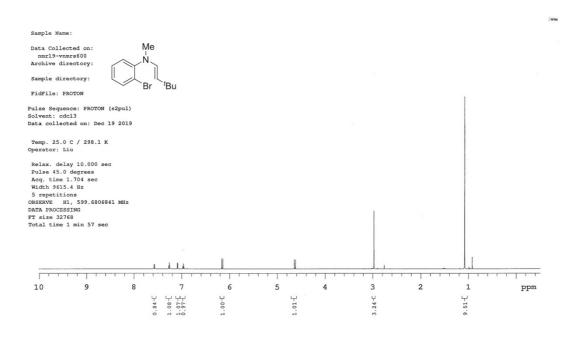
Identification code	C21H26BNO4	
Empirical formula	C21 H26 B N O4	
Formula weight	367.24	
Temperature	123(2) K	
Wavelength	1.54178 Å	
Crystal system	Orthorhombic	
Space group	Fdd2	
Unit cell dimensions	a = 31.1427(6) Å	α= 90°.
	b = 37.9695(8) Å	β= 90°.
	c = 13.9845(3) Å	$\gamma = 90^{\circ}$.
Volume	16536.3(6) Å ³	
Ζ	32	
Density (calculated)	1.180 Mg/m ³	
Absorption coefficient	0.645 mm ⁻¹	
F(000)	6272	
Crystal size	$0.580 \ge 0.280 \ge 0.150 \text{ mm}^3$	
Theta range for data collection	3.655 to 66.608°.	
Index ranges	-37<=h<=36, -41<=k<=45, -16<=l<=16	
Reflections collected	30171	
Independent reflections	ons $7254 [R(int) = 0.0267]$	
Completeness to theta = 66.608°	99.7 %	
Absorption correction	Semi-empirical from equivalents	
Max. and min. transmission	0.7528 and 0.6531	
Refinement method	Full-matrix least-squares on F ²	
Data / restraints / parameters	7254 / 571 / 575	
Goodness-of-fit on F ²	1.035	
Final R indices [I>2sigma(I)]	R1 = 0.0369, wR2 = 0.1073	
R indices (all data)	R1 = 0.0383, wR2 = 0.1091	
Extinction coefficient	n/a	
Largest diff. peak and hole	0.258 and -0.215 e.Å ⁻³	

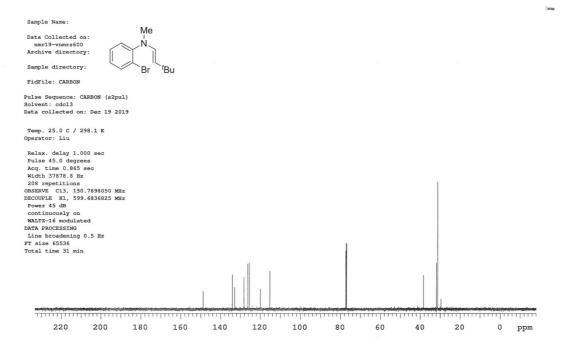
Crystal Data for 5g:

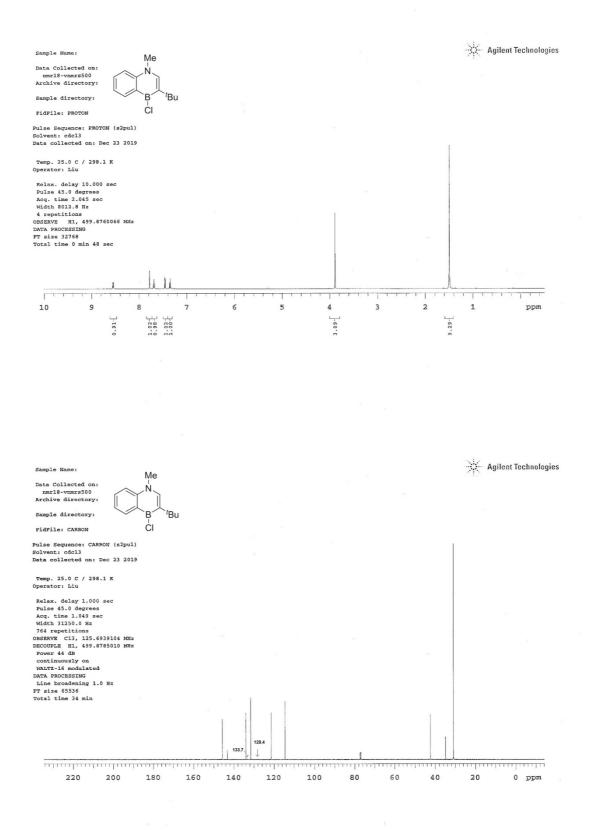
Identification code	C22H17BFN3	
Empirical formula	C22 H17 B F N3	
Formula weight	353.19	
Temperature	173(2) K	
Wavelength	1.54178 Å	
Crystal system	Monoclinic	
Space group	P21/n	
Unit cell dimensions	a = 8.7227(4) Å	<i>α</i> = 90°.
	b = 7.3770(3) Å	β= 98.3760(10)°.
	c = 28.3779(11) Å	$\gamma = 90^{\circ}$.
Volume	1806.57(13) Å ³	
Z	4	
Density (calculated)	1.299 Mg/m ³	
Absorption coefficient	0.675 mm ⁻¹	
F(000)	736	
Crystal size	$0.420 \ x \ 0.280 \ x \ 0.260 \ mm^3$	
Theta range for data collection	3.148 to 66.424°.	
Index ranges	-10<=h<=10, -8<=k<=8, -33<=l<=32	
Reflections collected 19722		
ndependent reflections $3171 [R(int) = 0.0228]$		
Completeness to theta = 66.424°	99.4 %	
Absorption correction	Semi-empirical from equivale	nts
Max. and min. transmission	0.7528 and 0.6787	
Refinement method	Full-matrix least-squares on F	2
Data / restraints / parameters	3171 / 2 / 251	
Goodness-of-fit on F ²	1.036	
Final R indices [I>2sigma(I)]	R1 = 0.0438, wR2 = 0.1190	
R indices (all data)	R1 = 0.0477, wR2 = 0.1236	
Extinction coefficient	n/a	
Largest diff. peak and hole	0.733 and -0.331 e.Å ⁻³	

NMR Spectra Collection

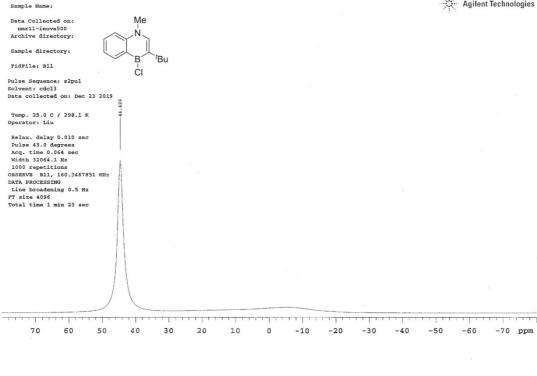


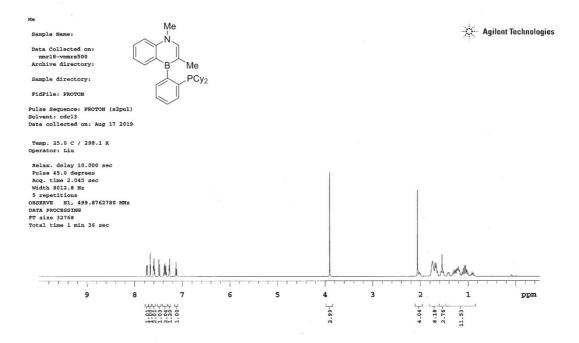


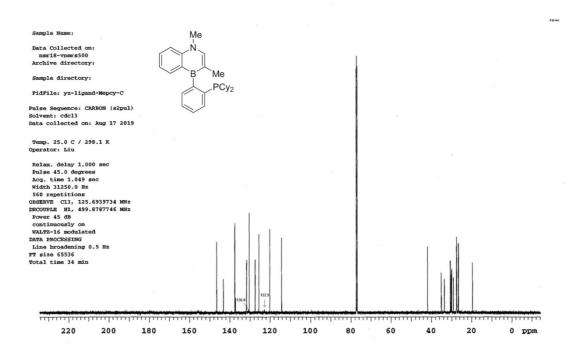


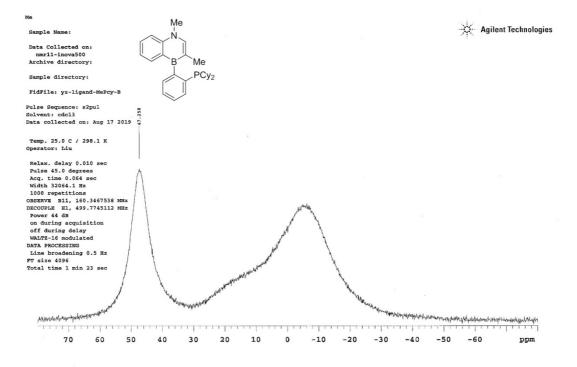


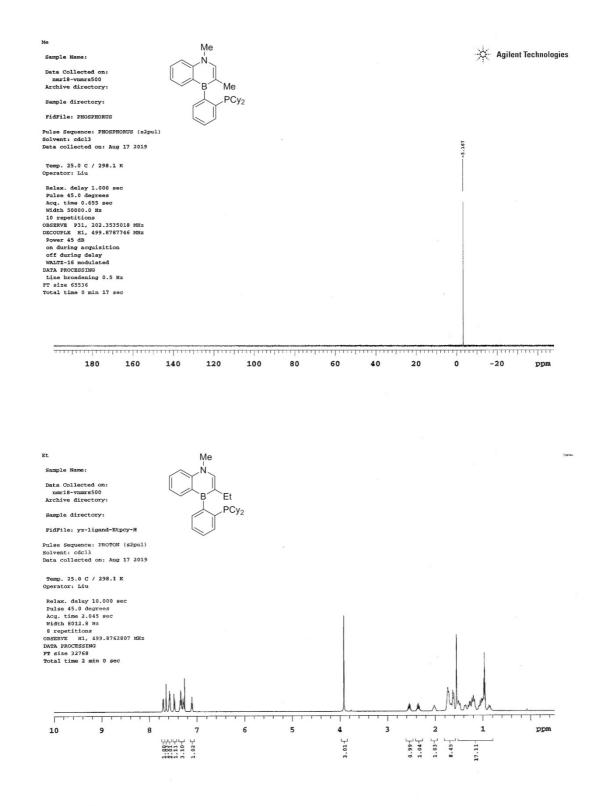
Agilent Technologies

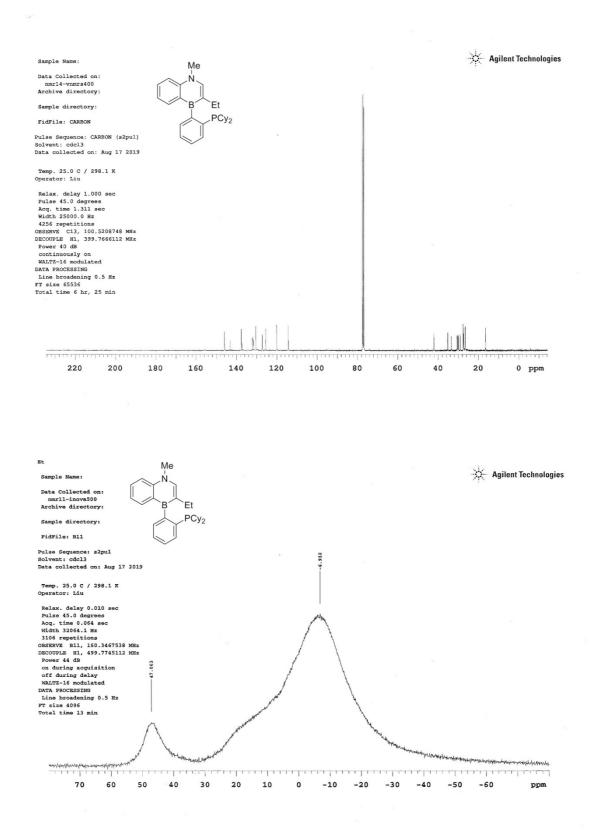


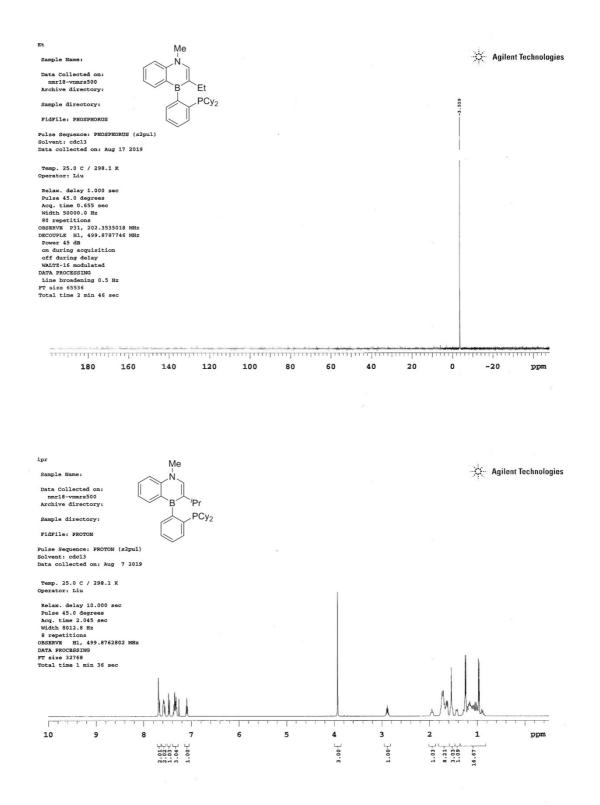


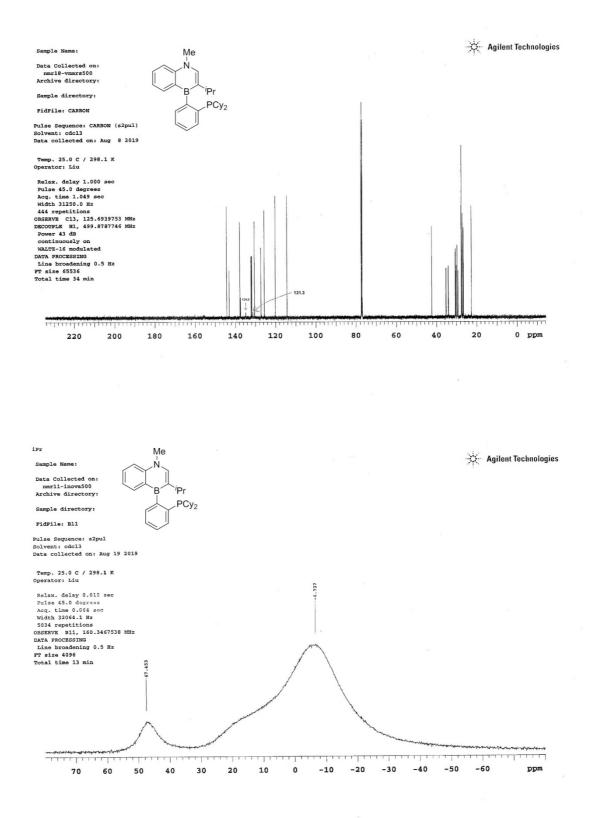


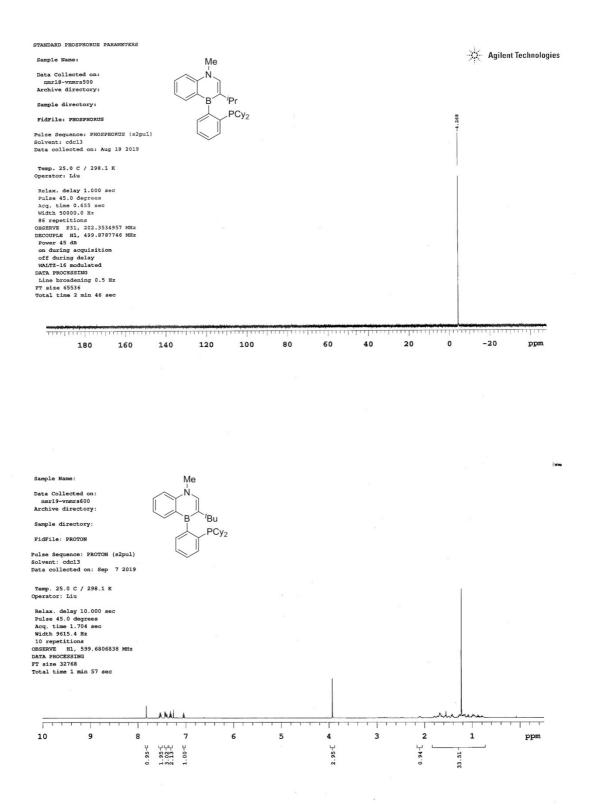


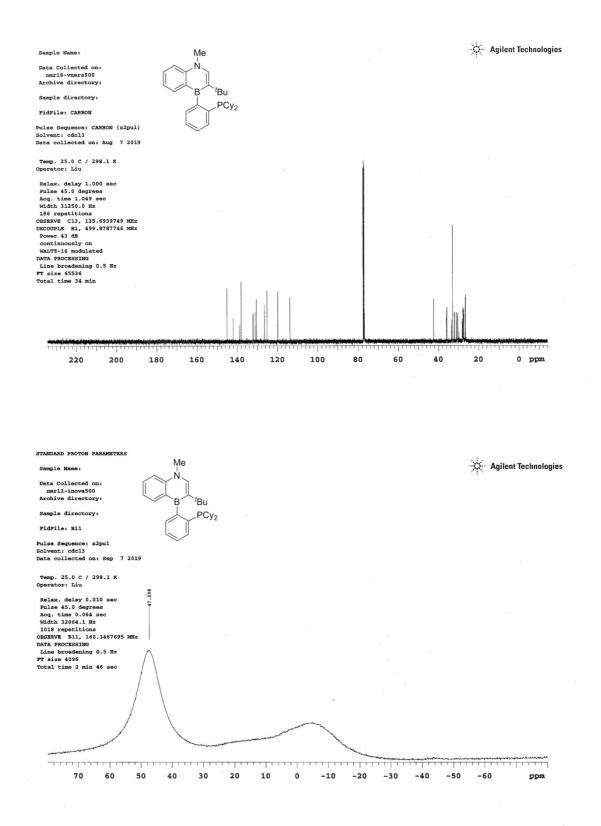


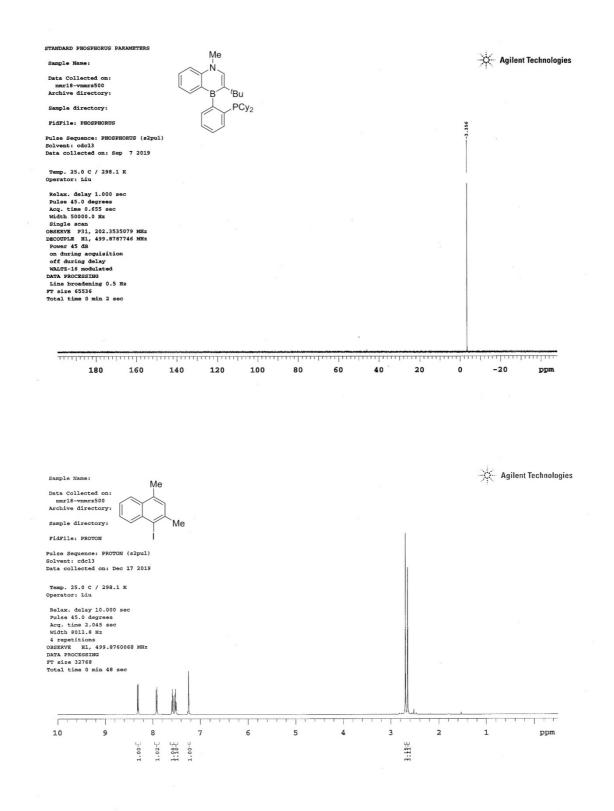


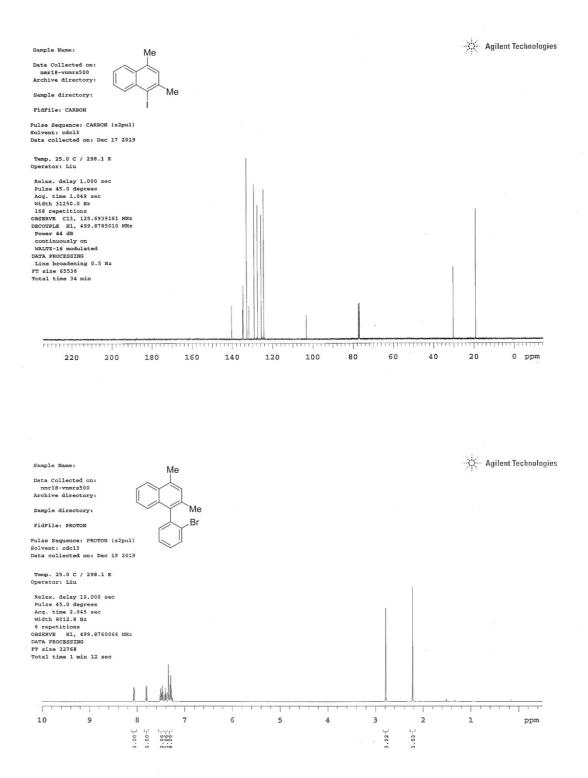


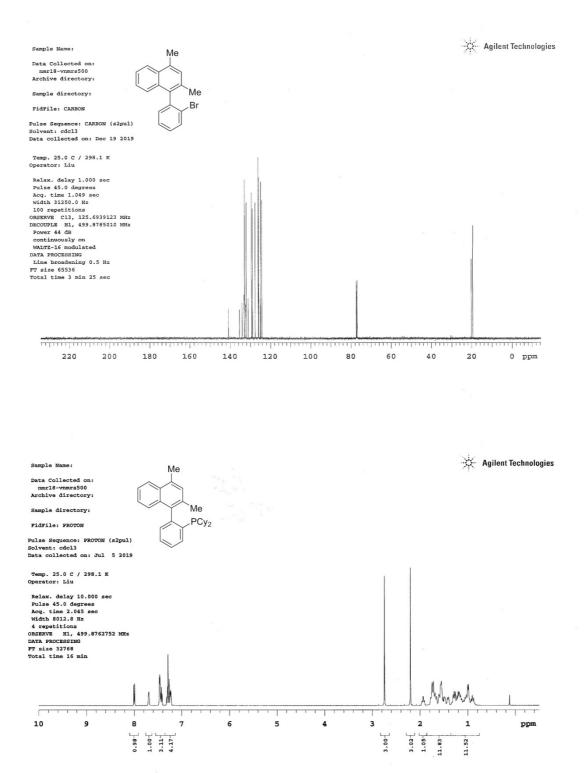


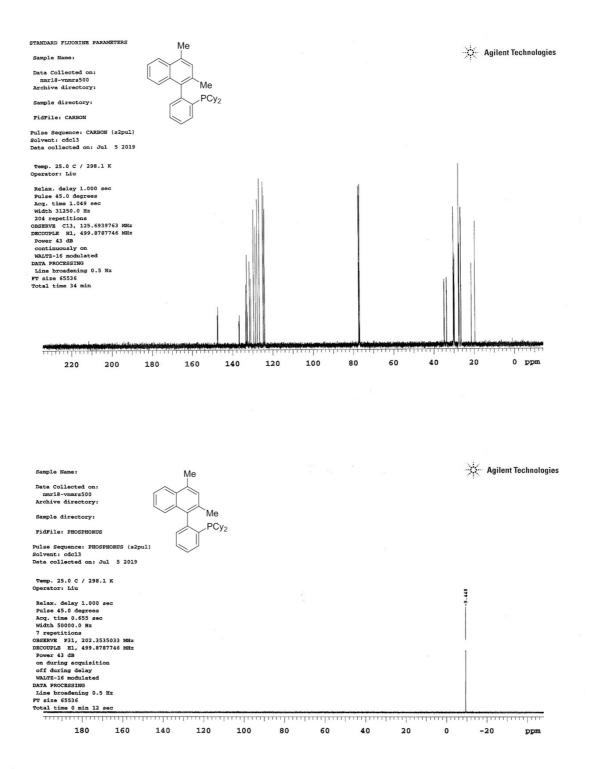


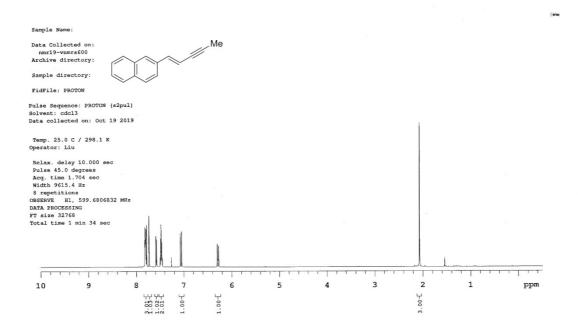


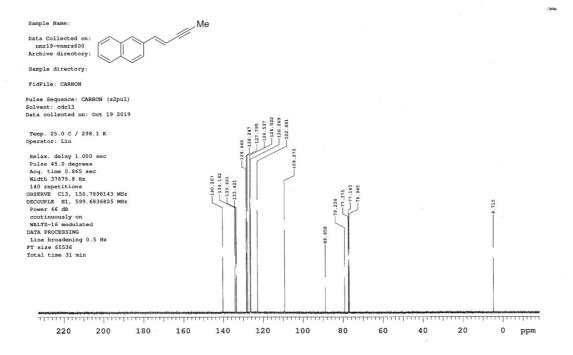


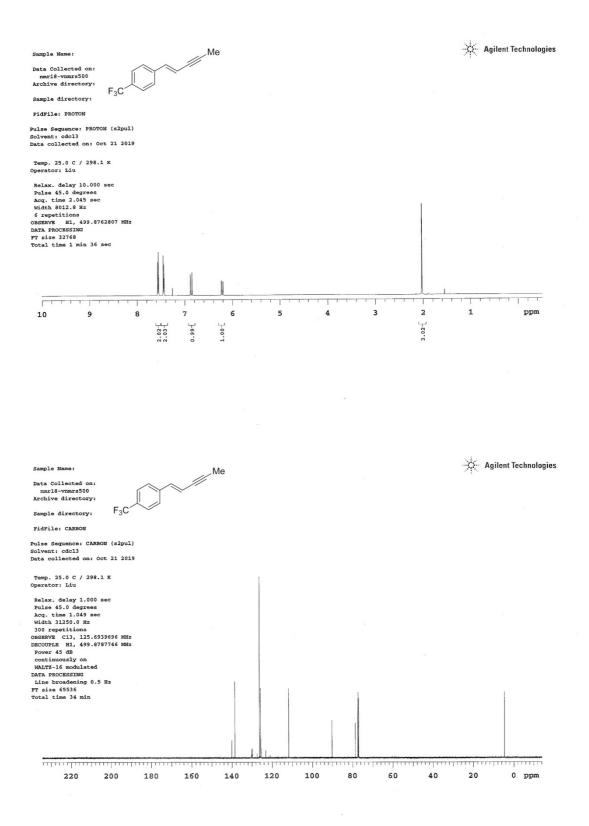


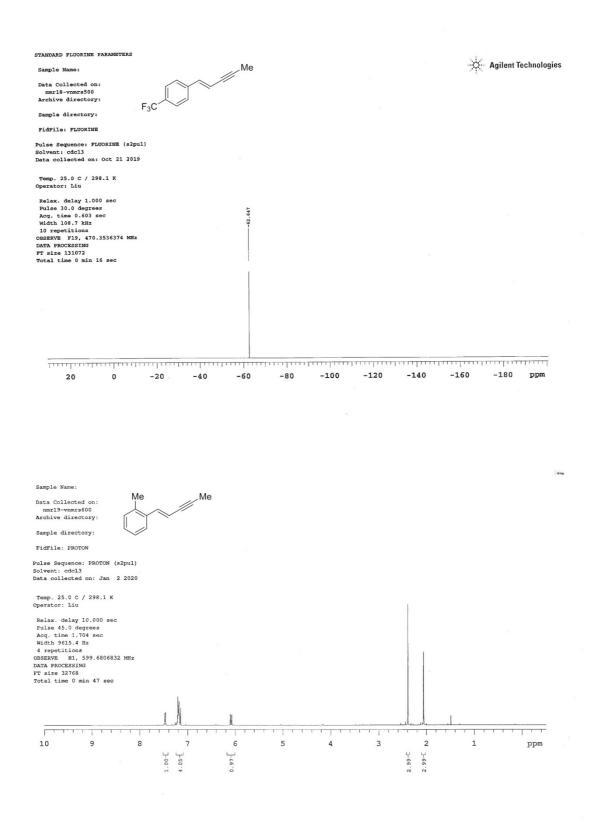


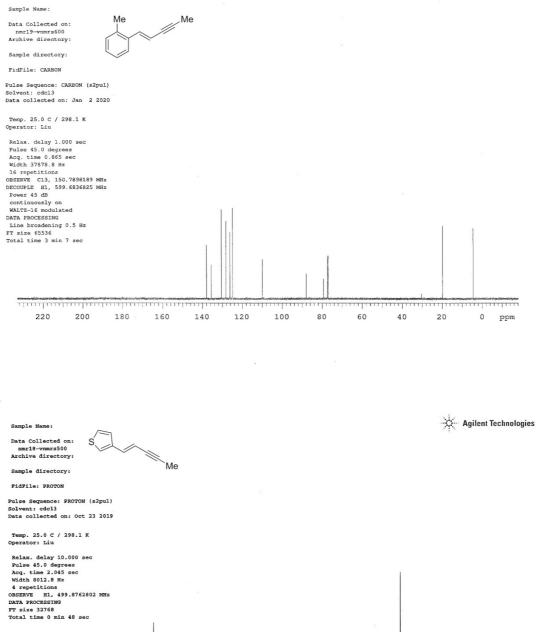






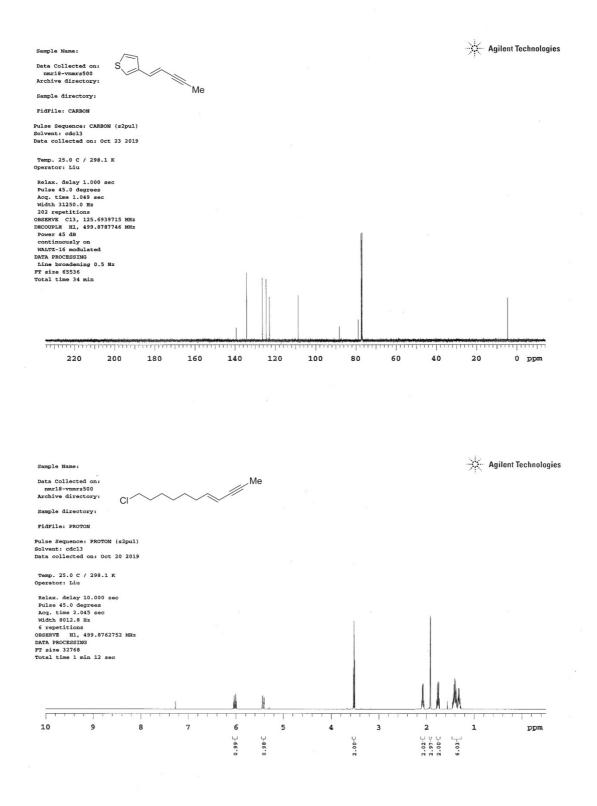


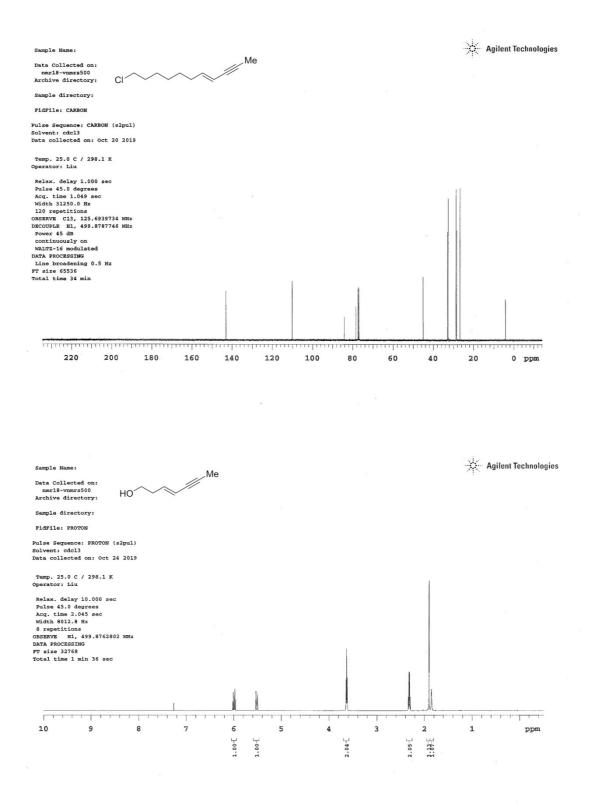


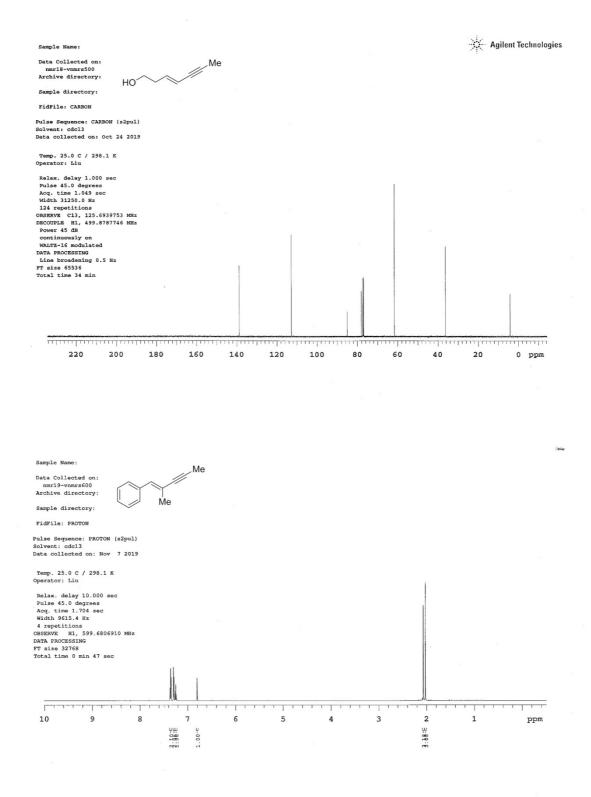


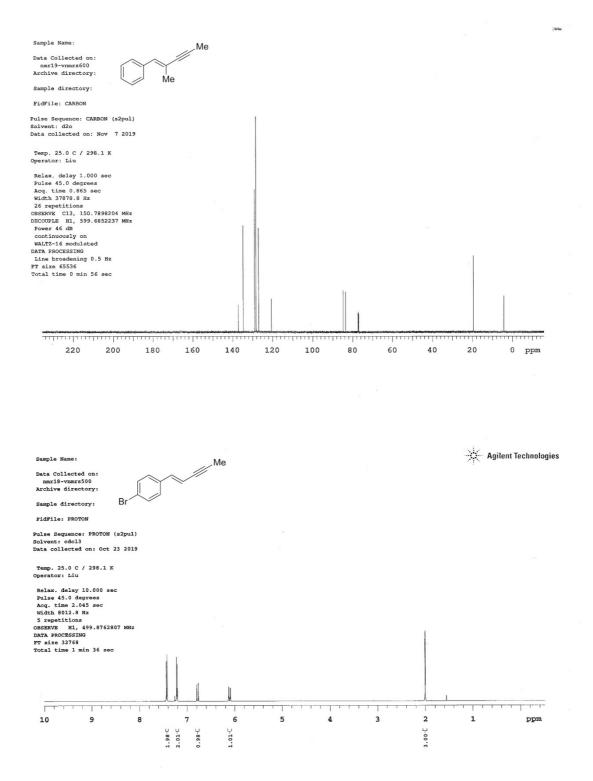
ang

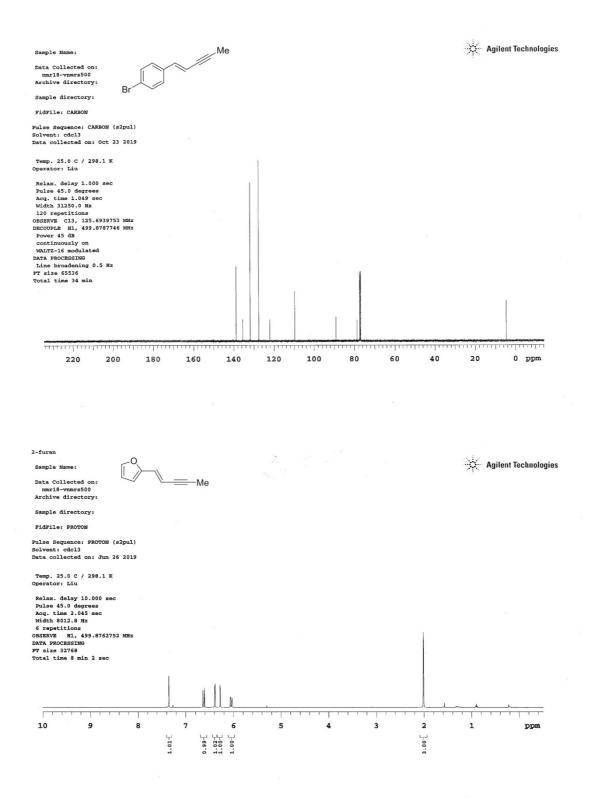
J T-1-TT 7 6 2 10 9 8 5 4 3 1 ppm 2.00-0 1.00-[3.06-1.00-[

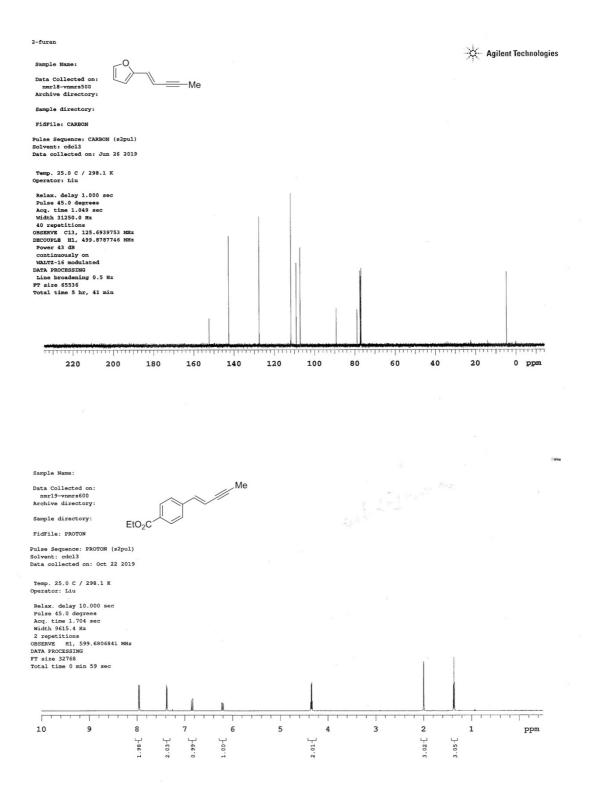


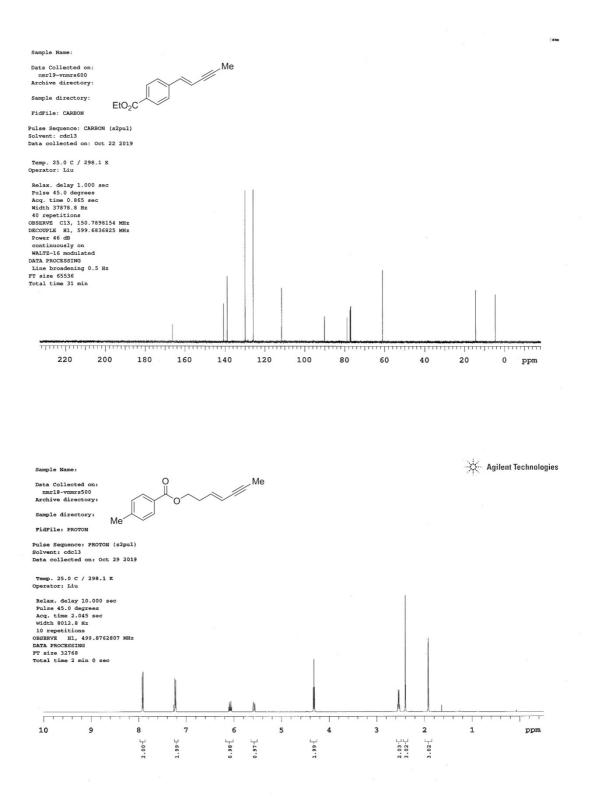


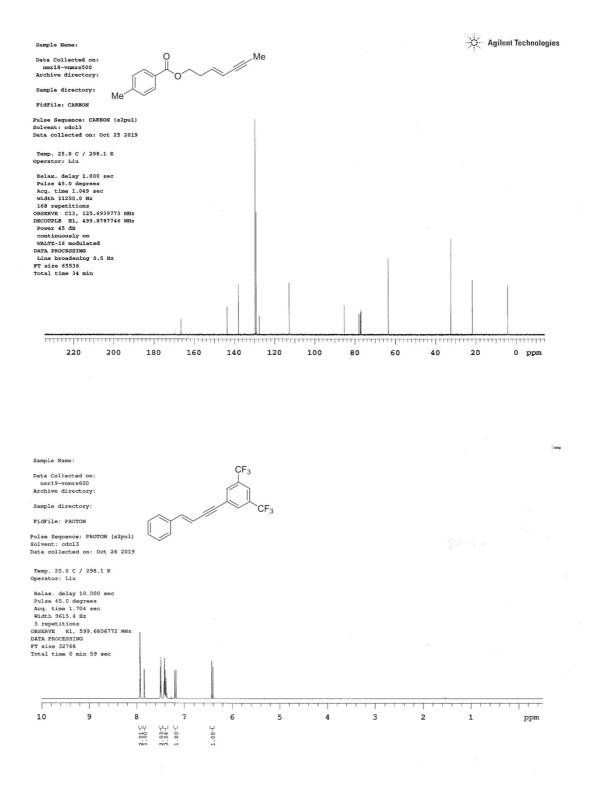


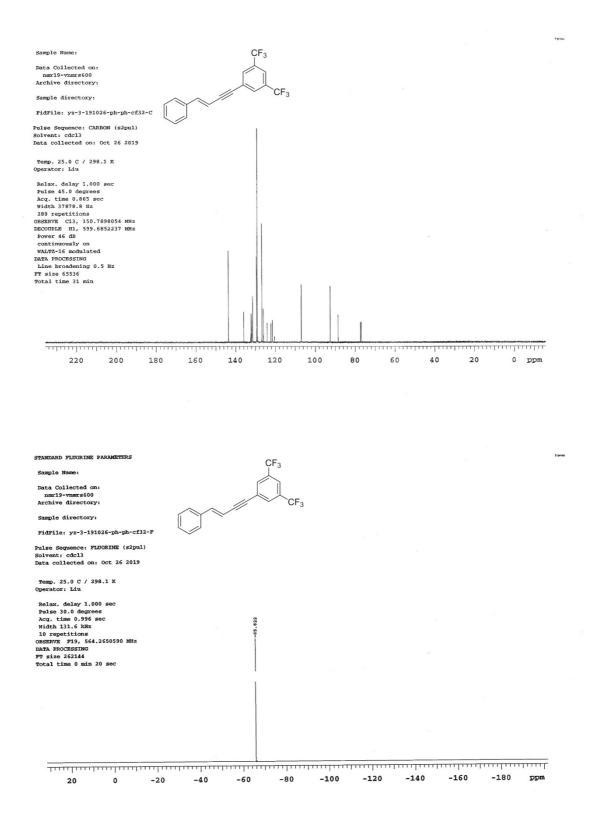


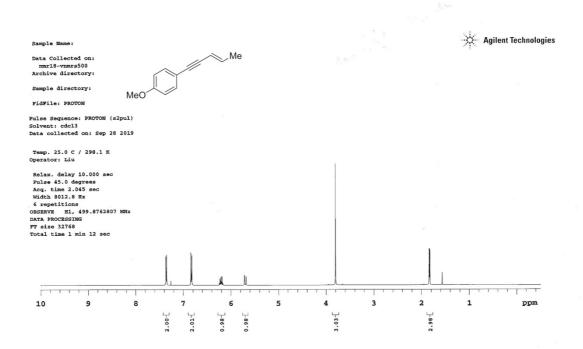


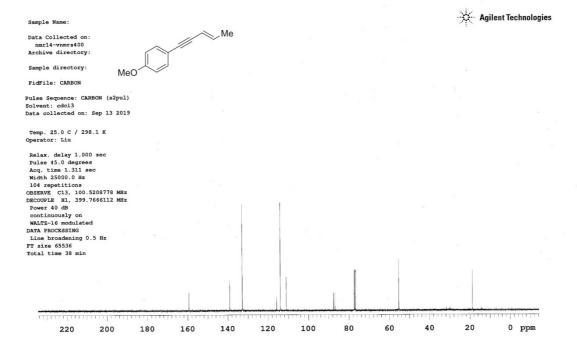


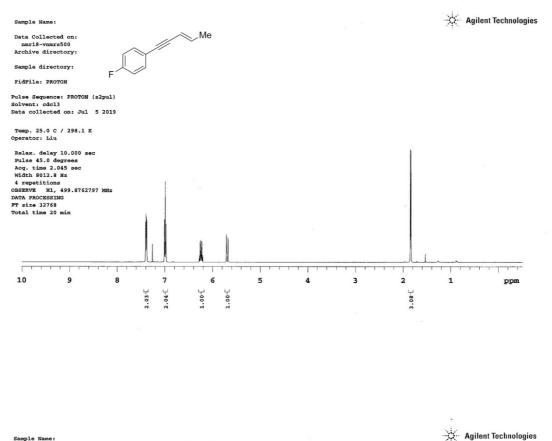


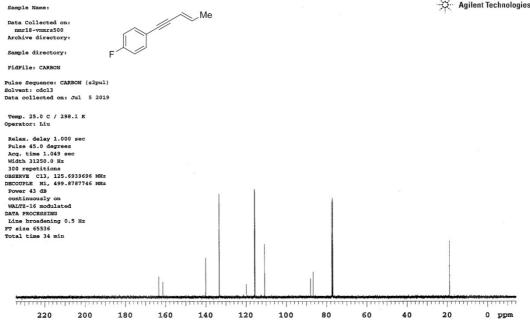


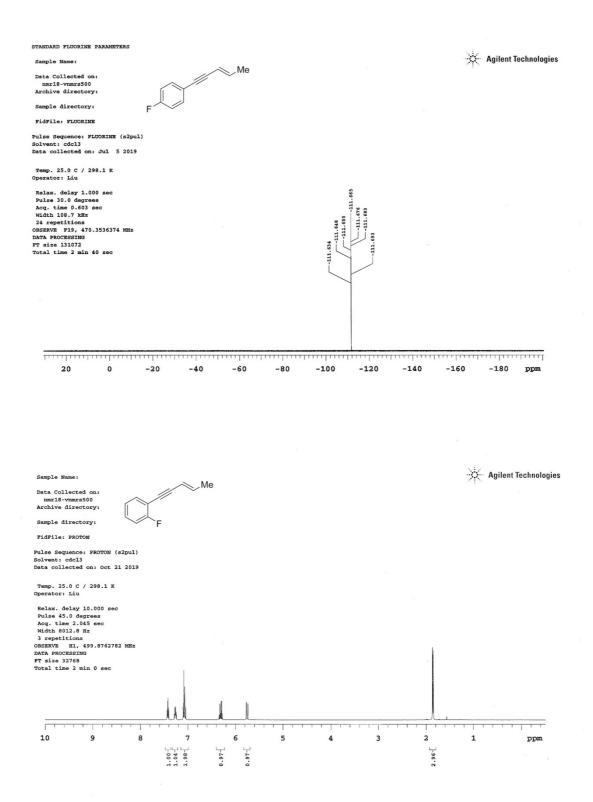


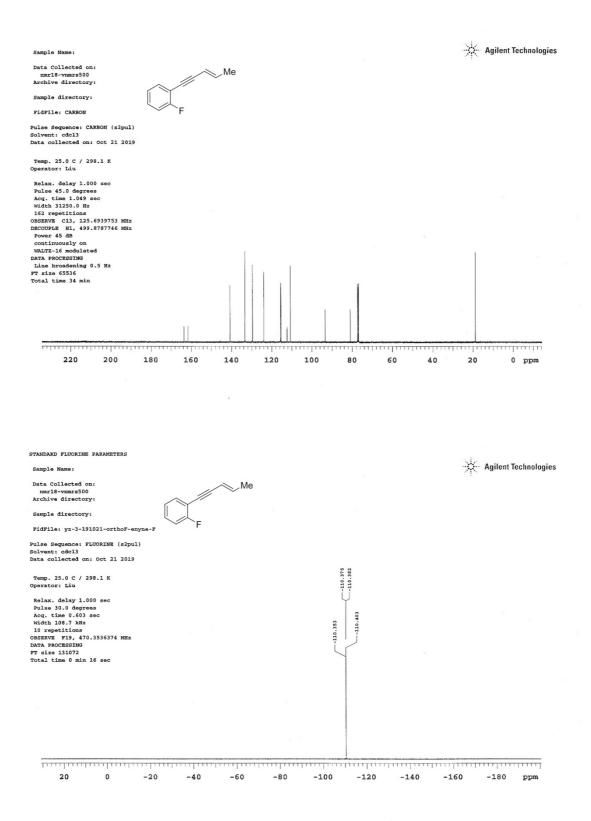


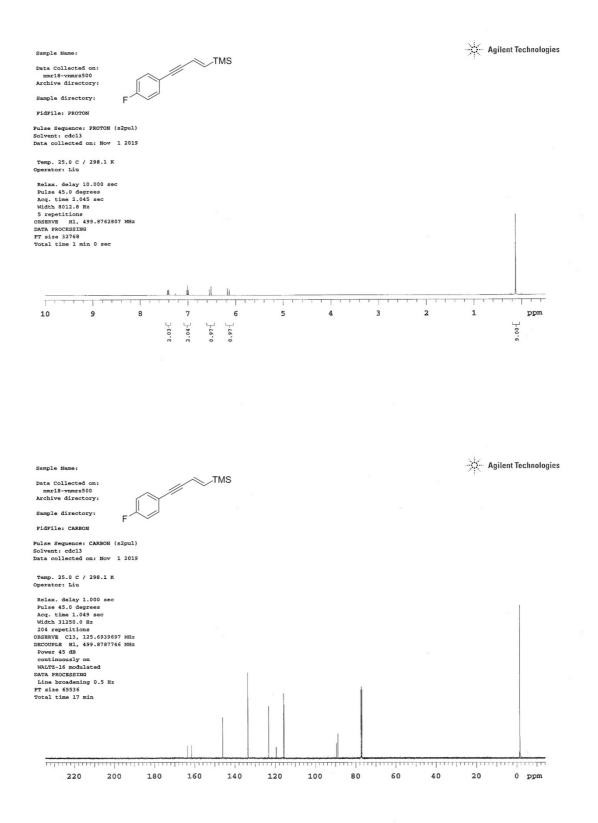


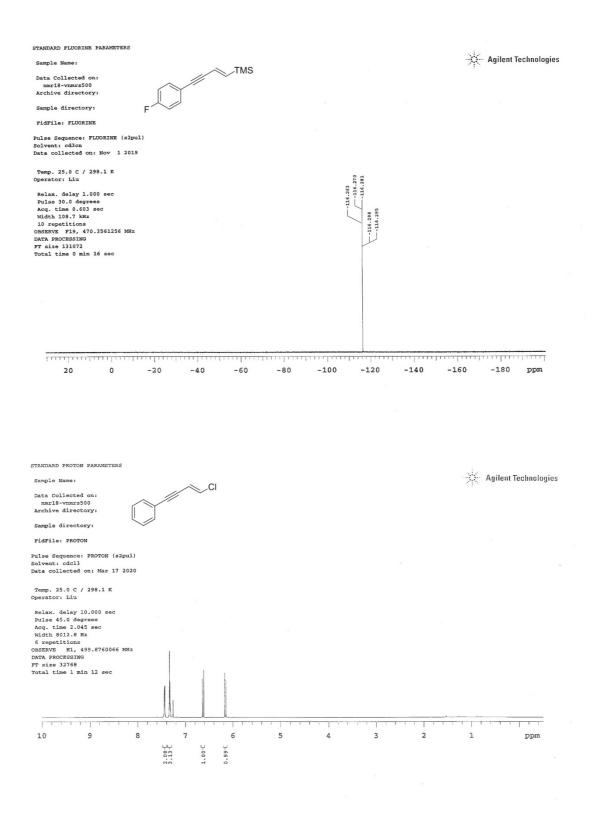


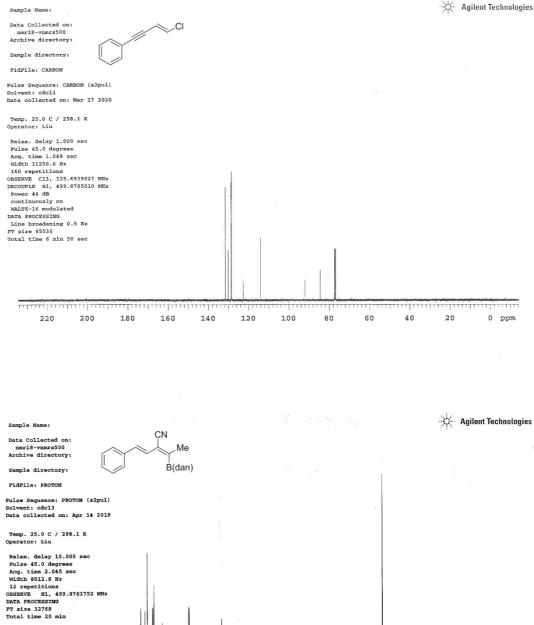


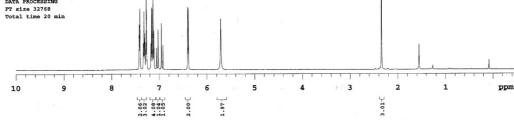


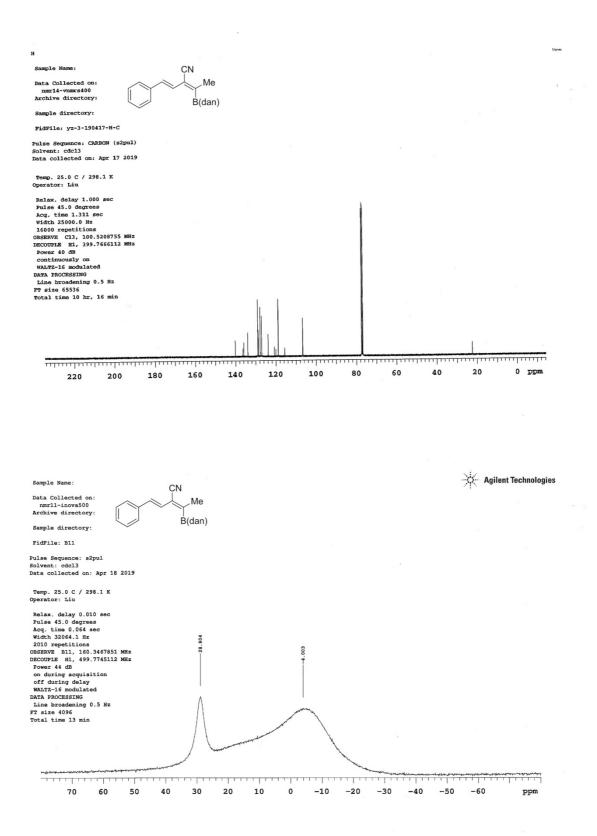


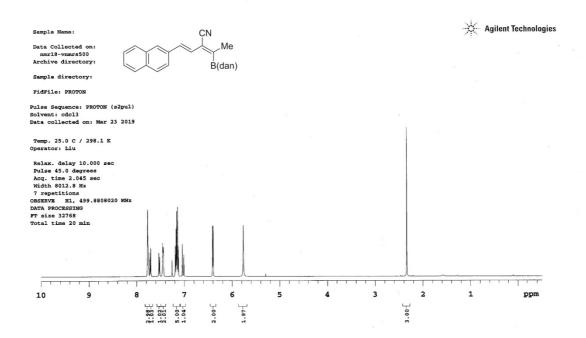


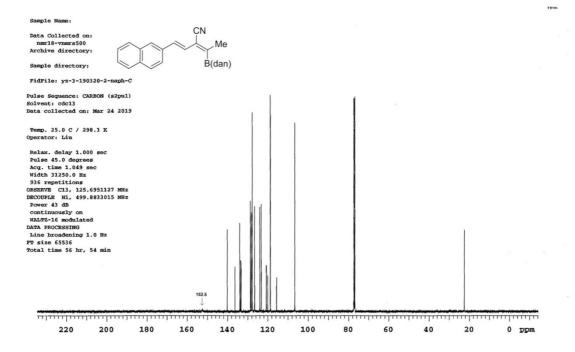


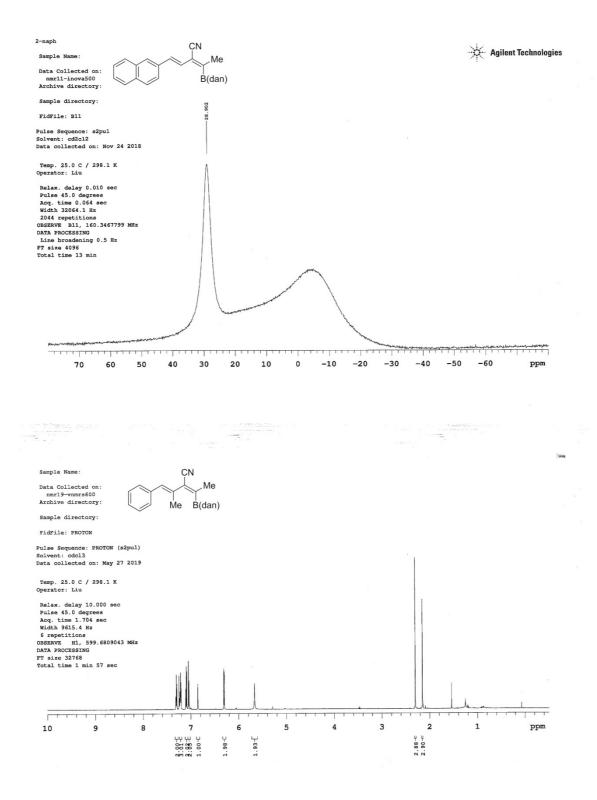


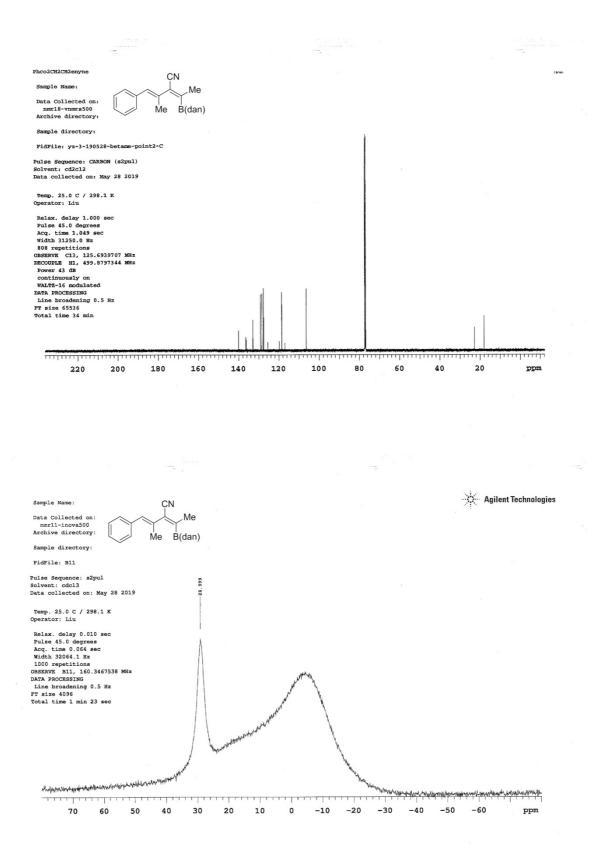


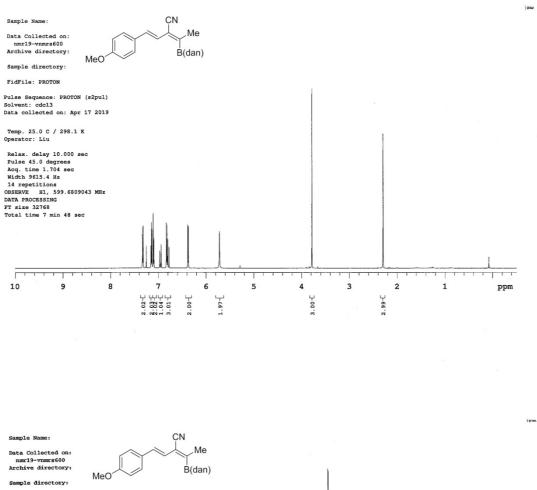


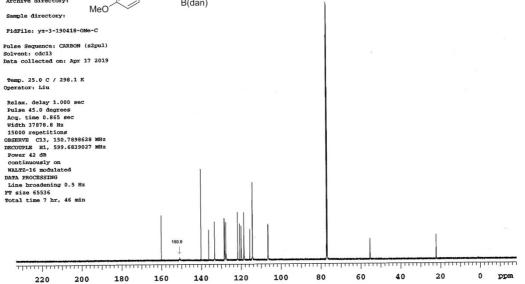


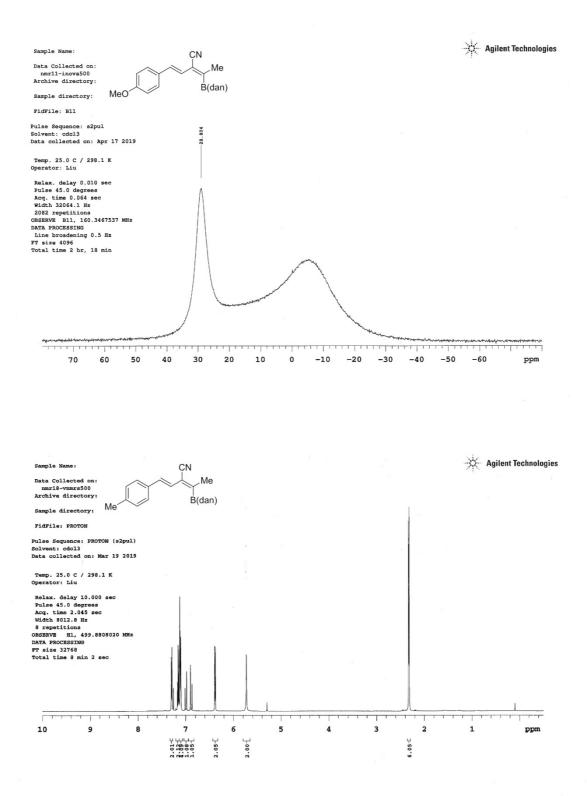


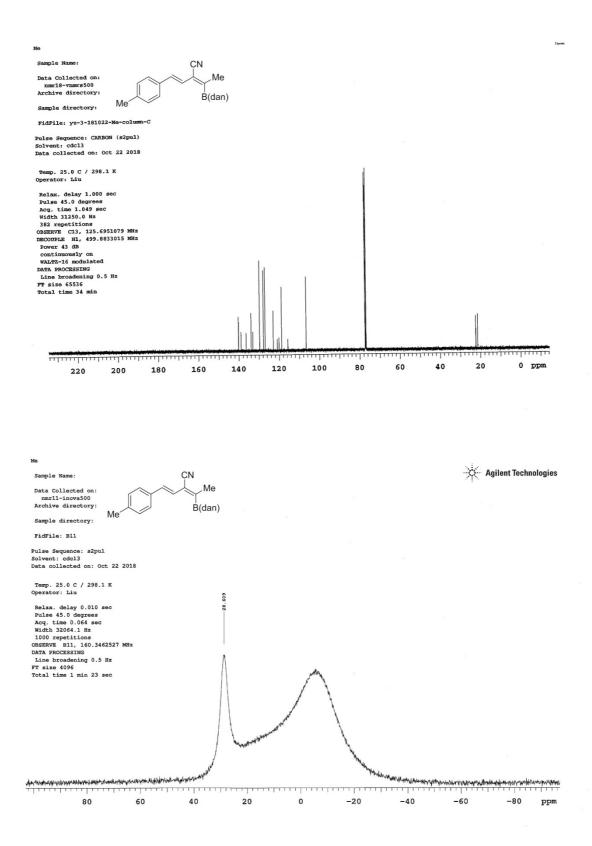


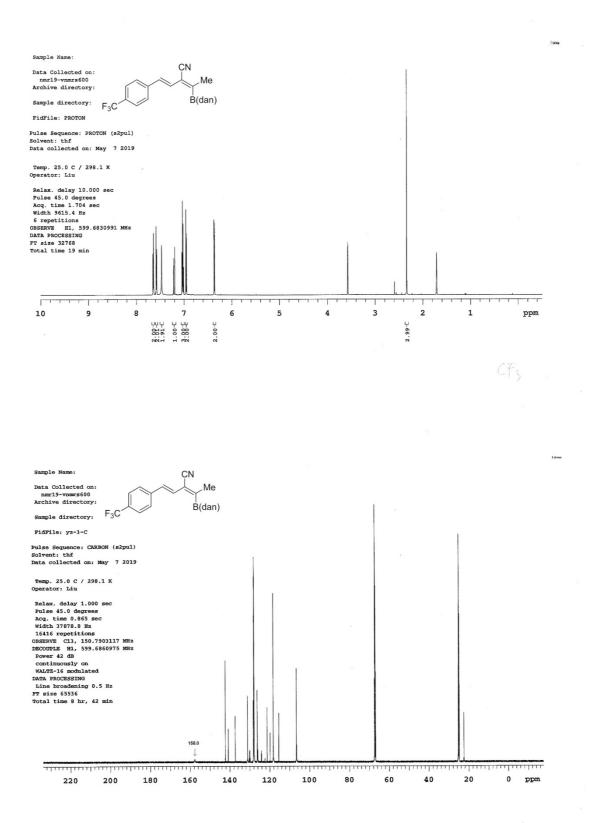


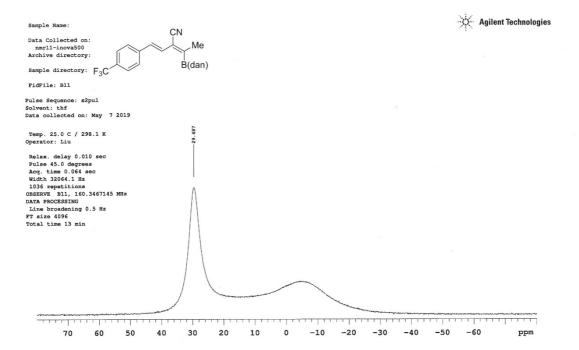


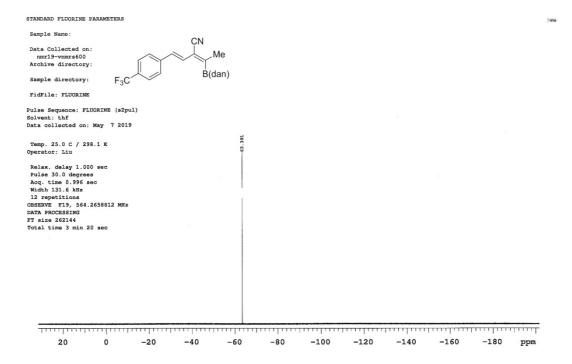


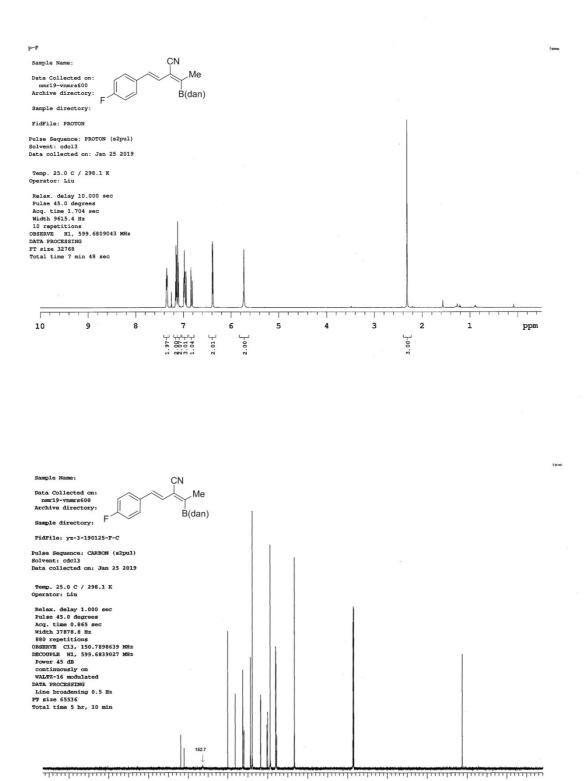




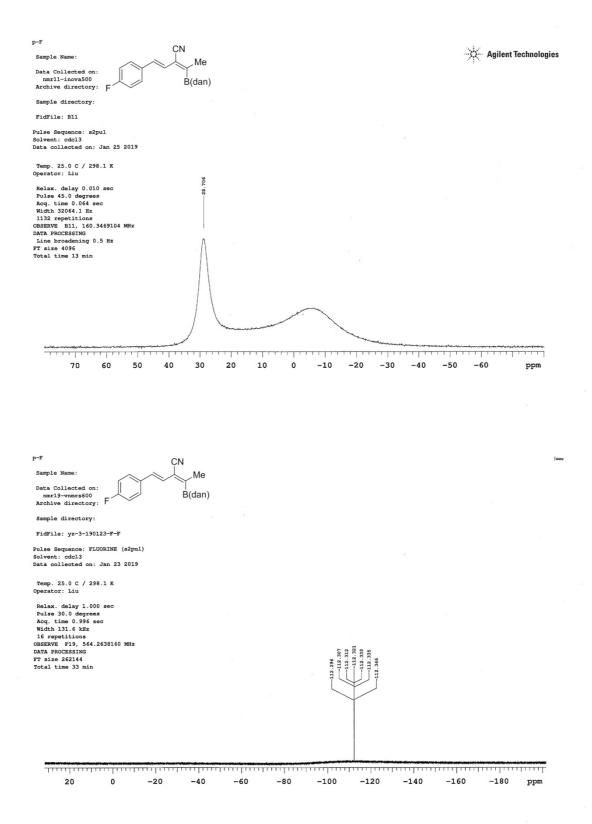


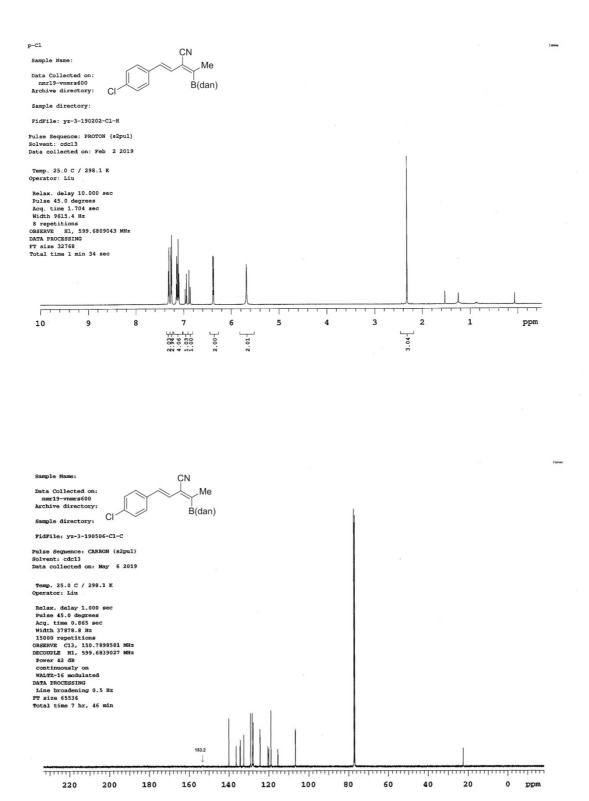


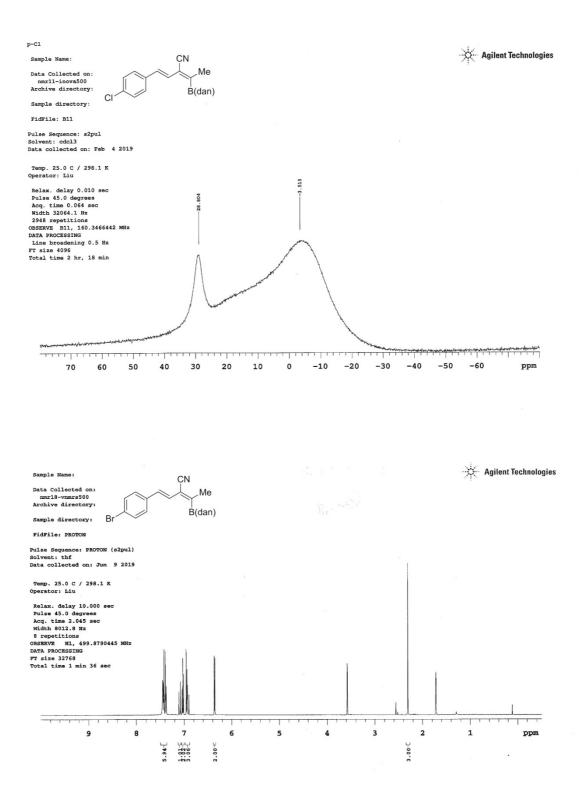


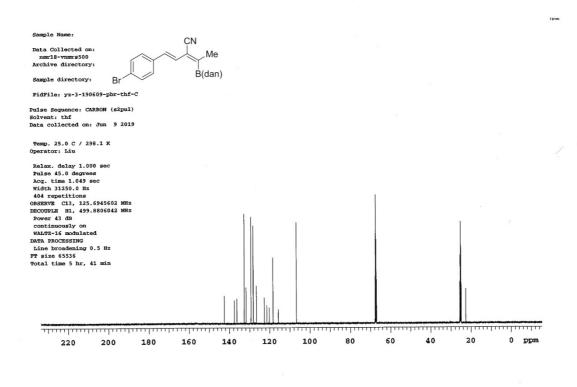


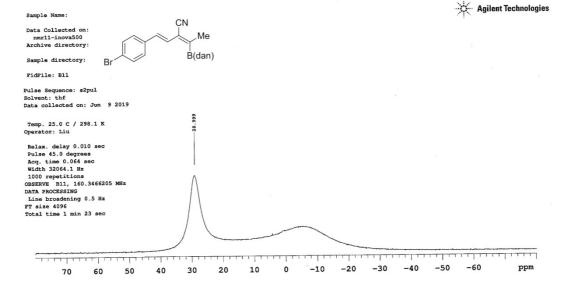
0 ppm

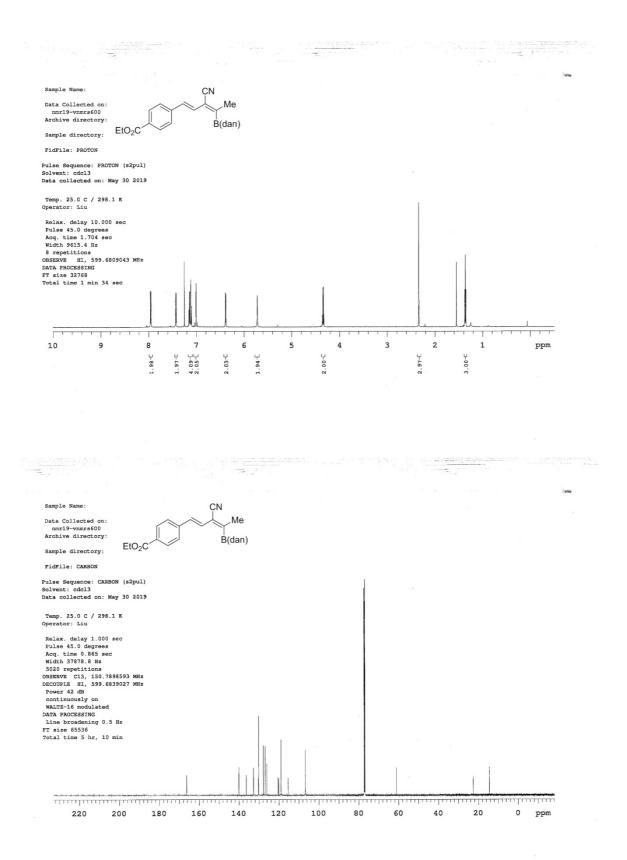


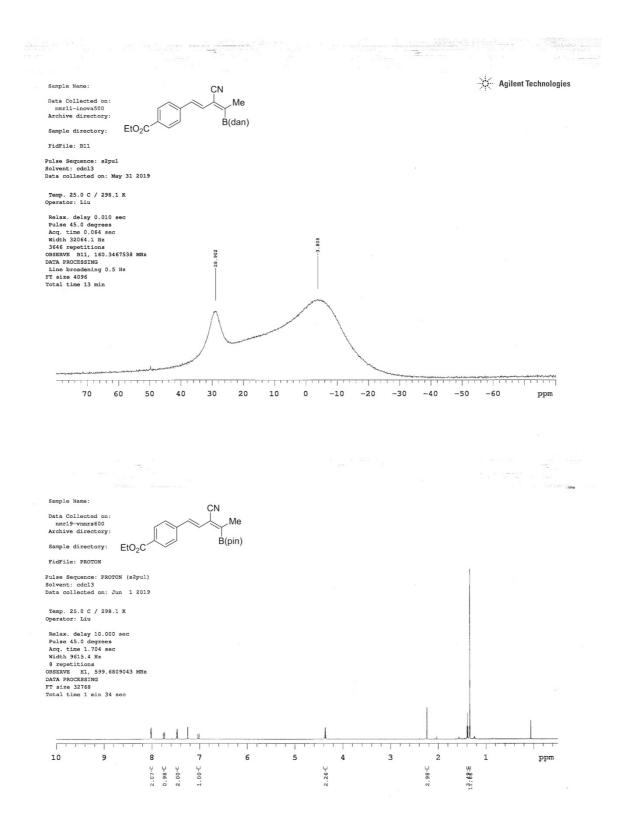


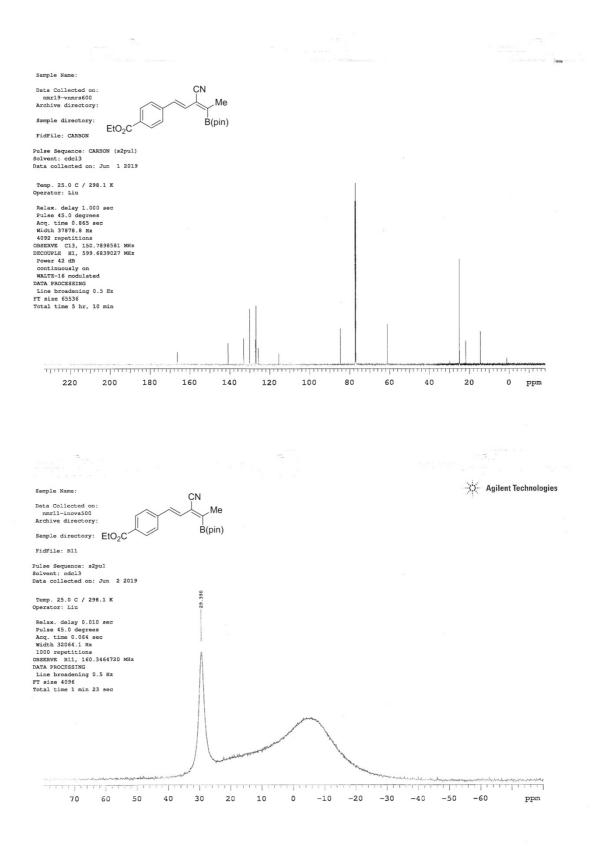




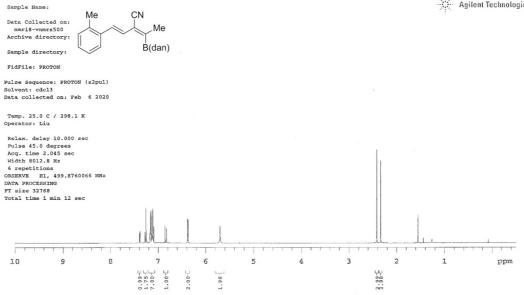


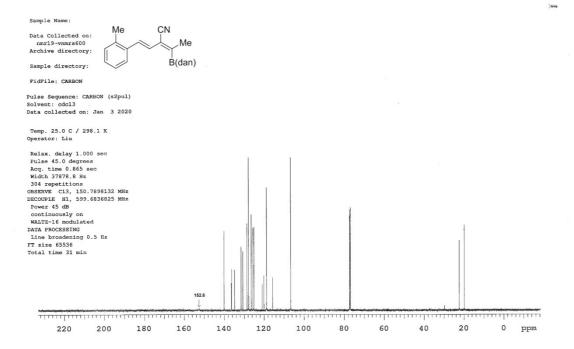


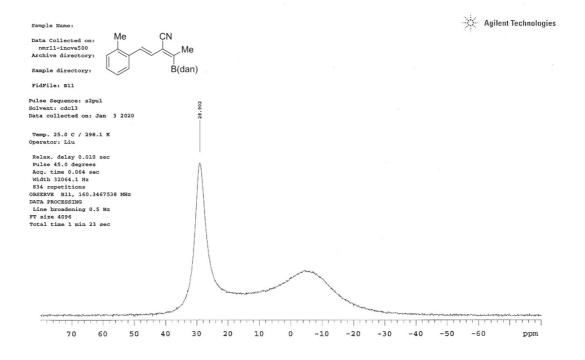


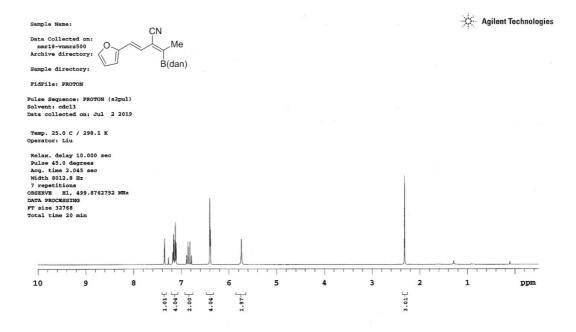


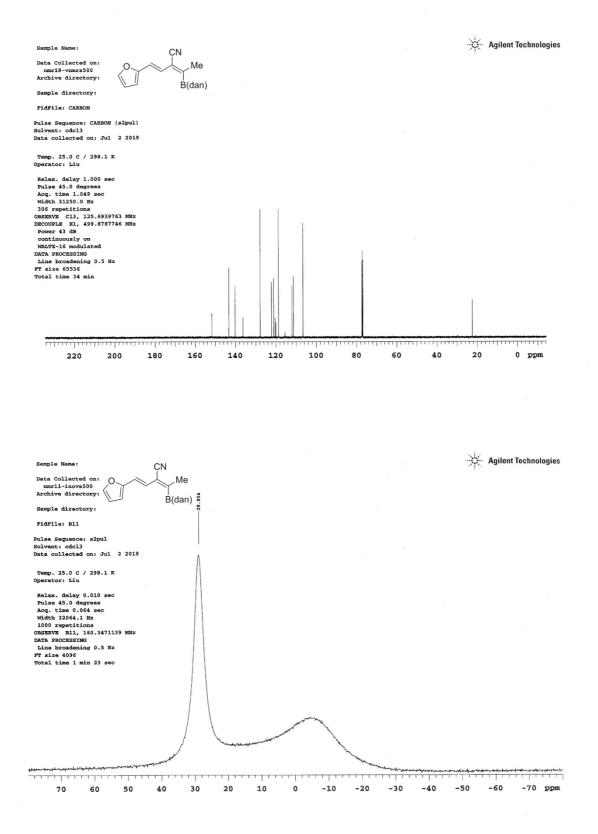
Agilent Technologies

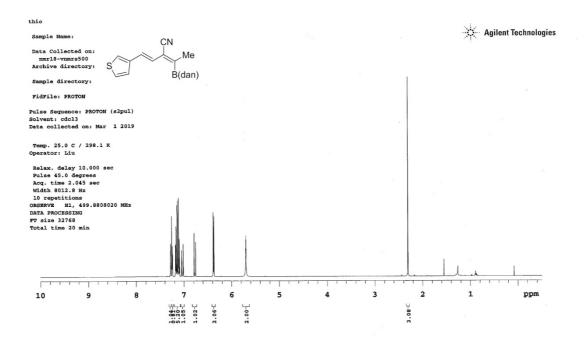


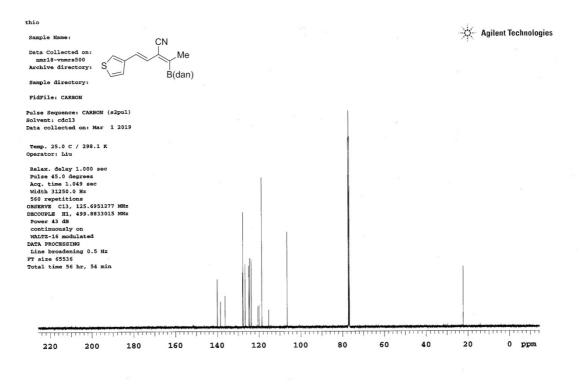


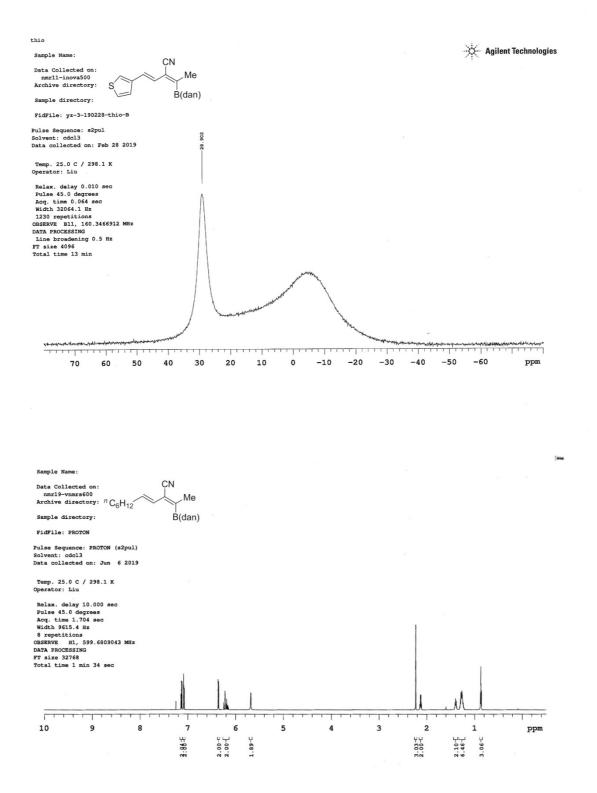


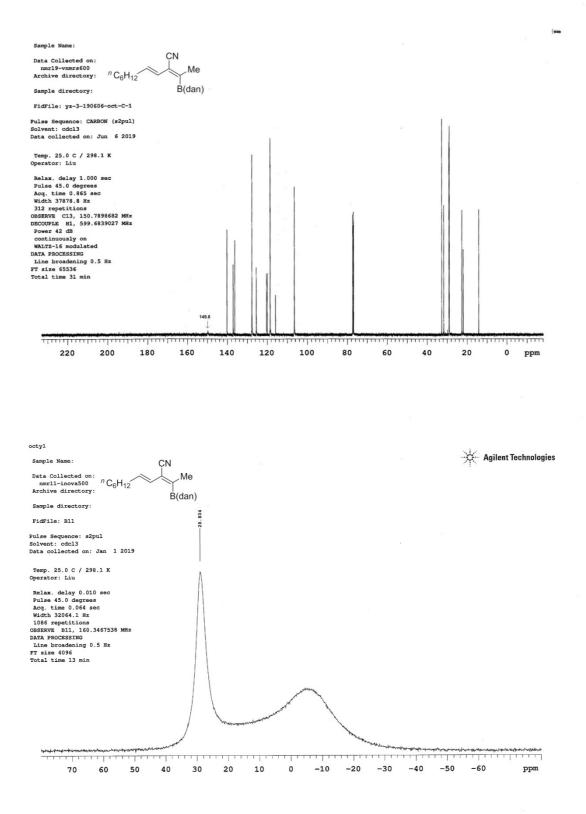


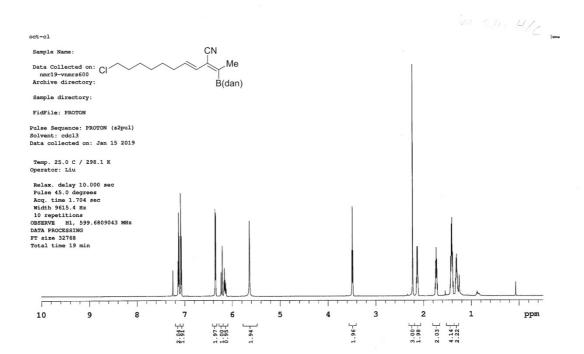


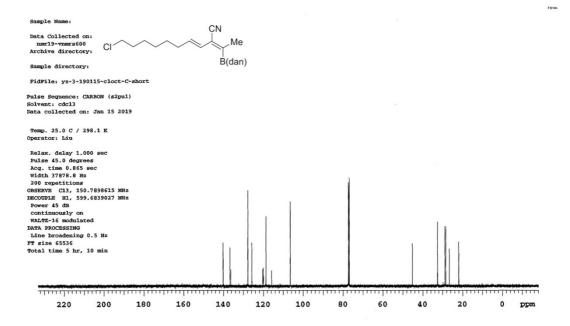


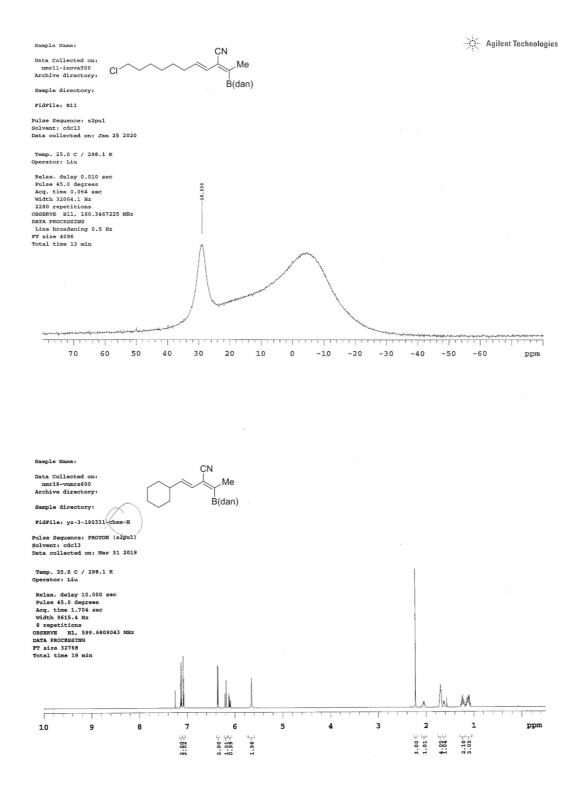


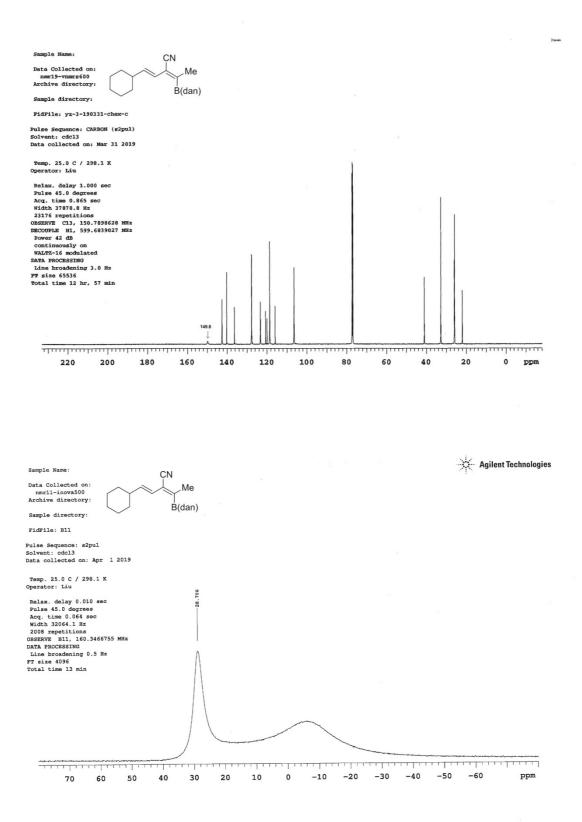


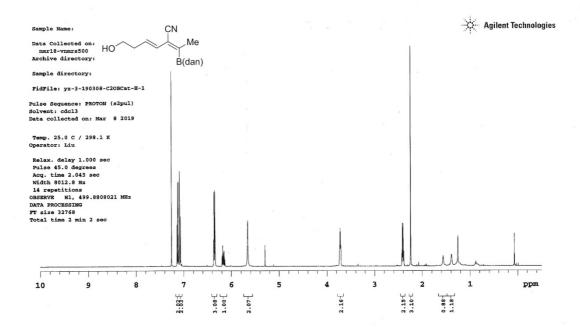


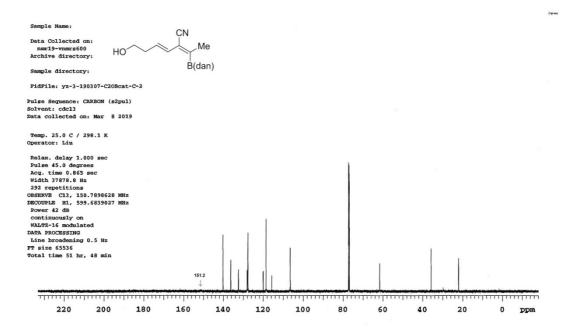




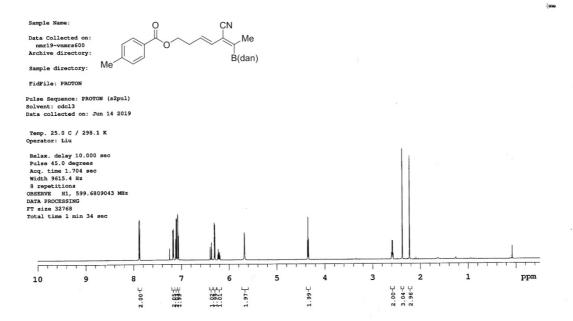


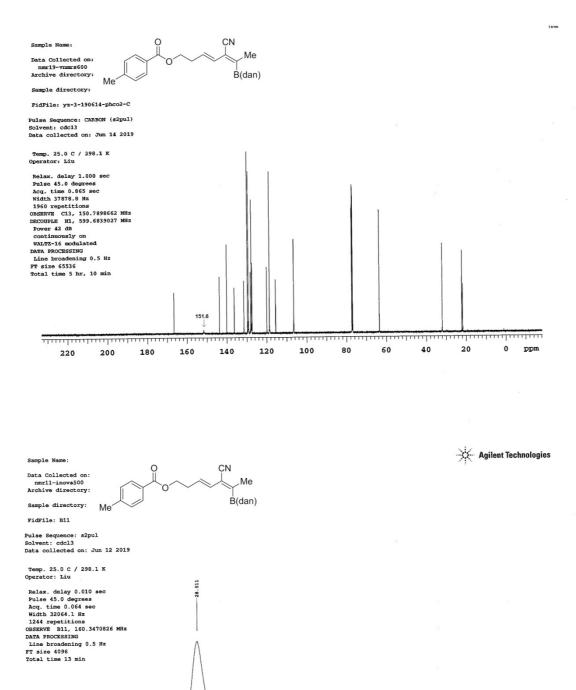






Agilent Technologies Sample Name: ÇN Data Collected on: nmr11-inova500 Archive directory: HO Me B(dan) Sample directory: FidFile: B11 Pulse Sequence: s2pul Solvent: cdcl3 Data collected on: Mar 8 2019 Temp. 25.0 C / 298.1 K Operator: Liu Relax. delay 0.010 sec Fule 45.0 degrees Acq. time 0.064 sec Width 32064.1 Hz 1022 repetitions 005ERVE Bil, 160.3467382 MHz DATA PROCESSING Line broadening 0.5 Hz FT size 4056 Total time 2 hr, 18 min -28.902 70 60 50 40 30 20 10 0 -10 -20 -30 -40 -50 -60 ppm





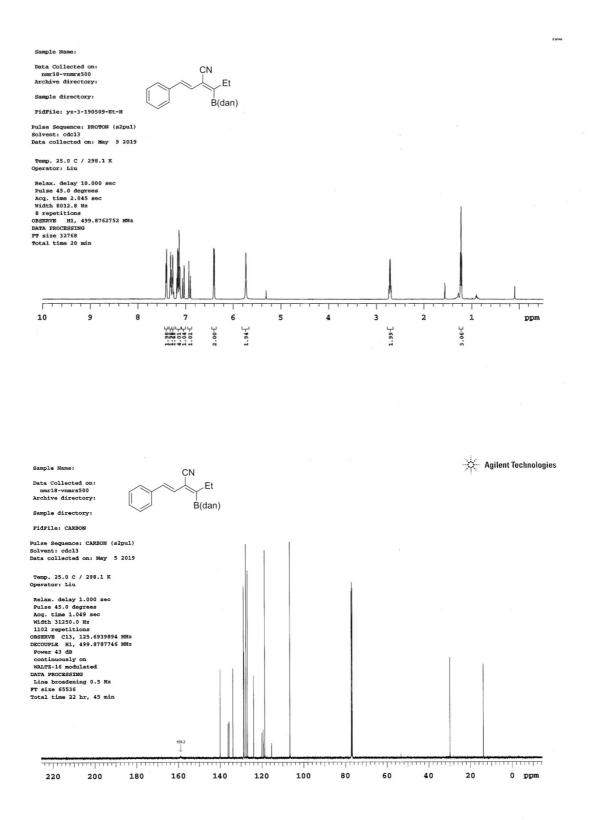
and the second s

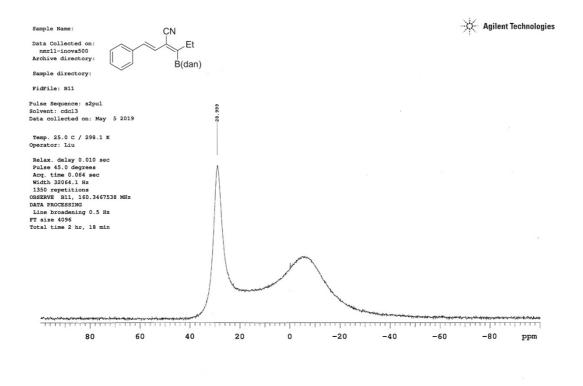
-20

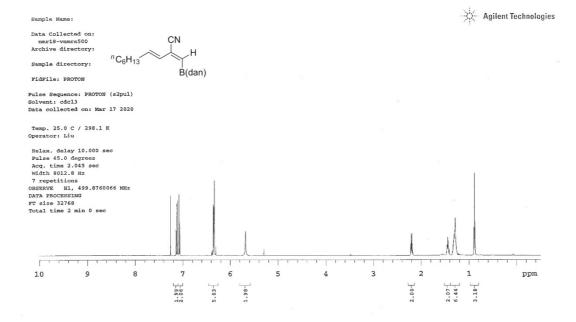
ppm

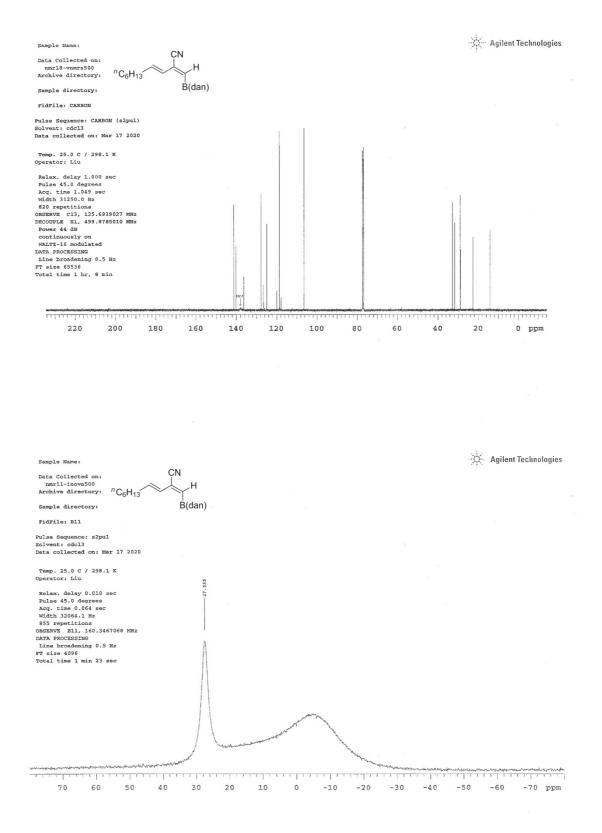
-30 -40 -50 -60

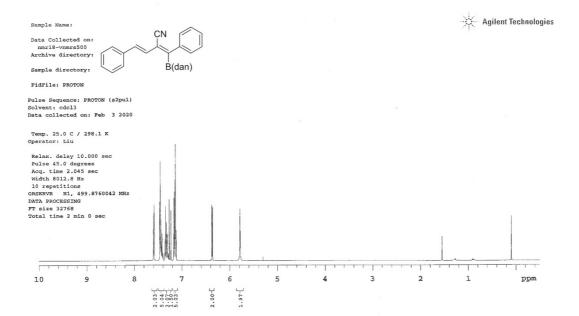
-10

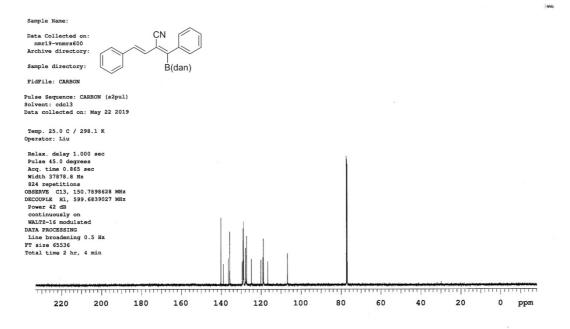


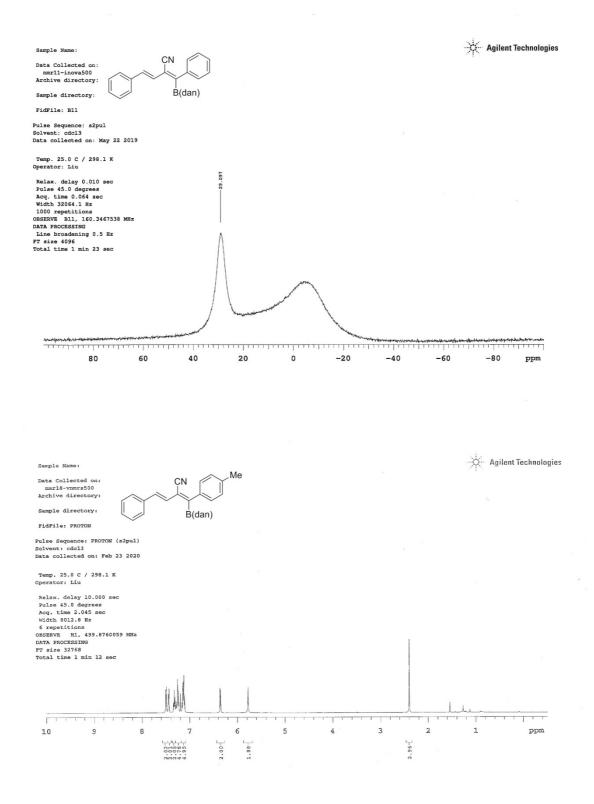


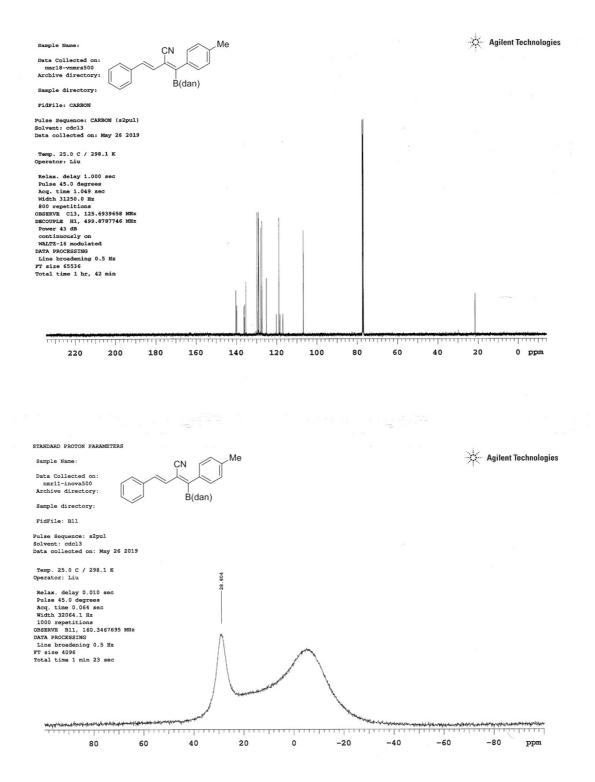


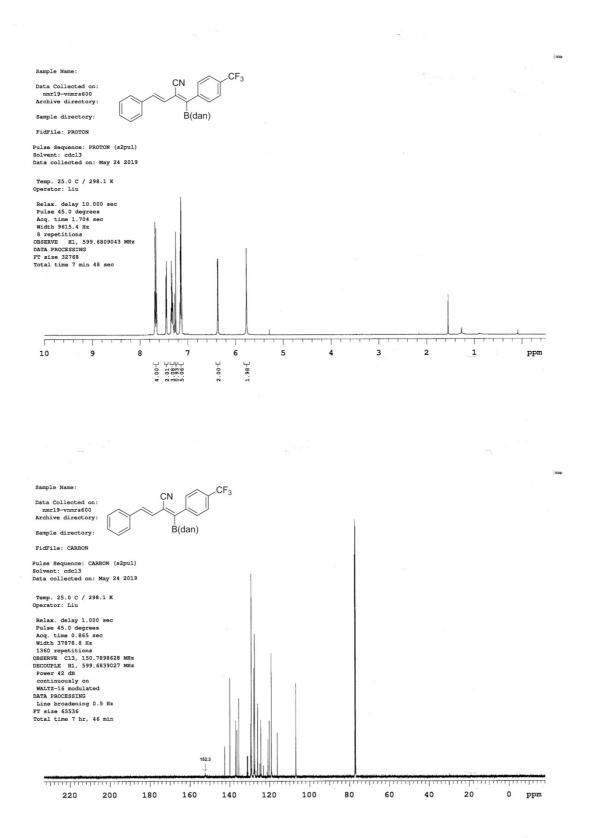


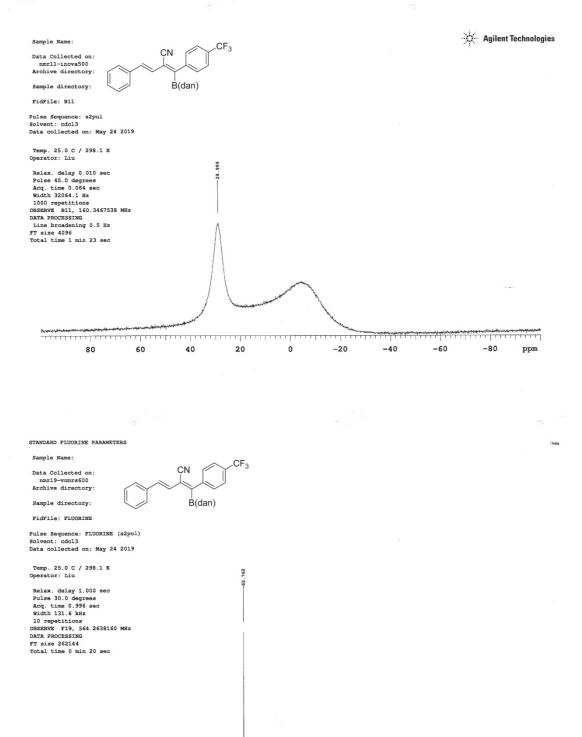




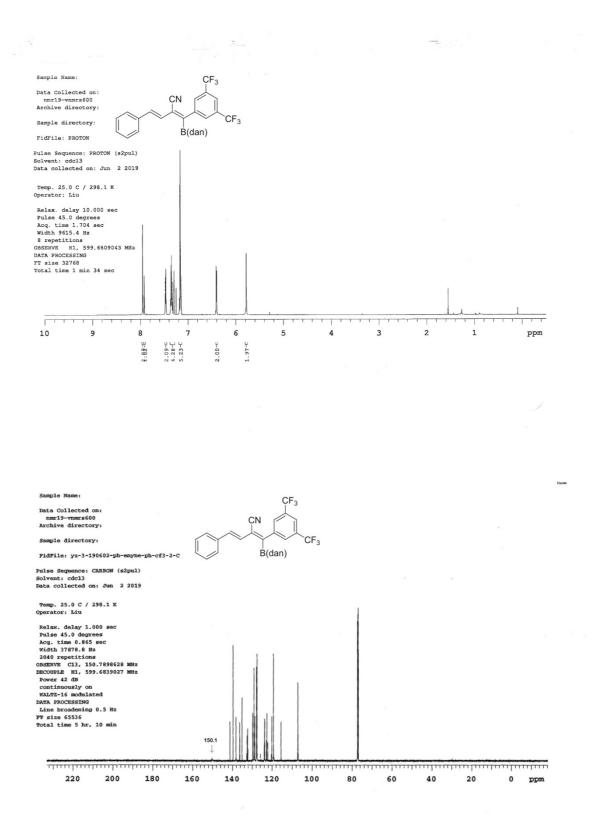


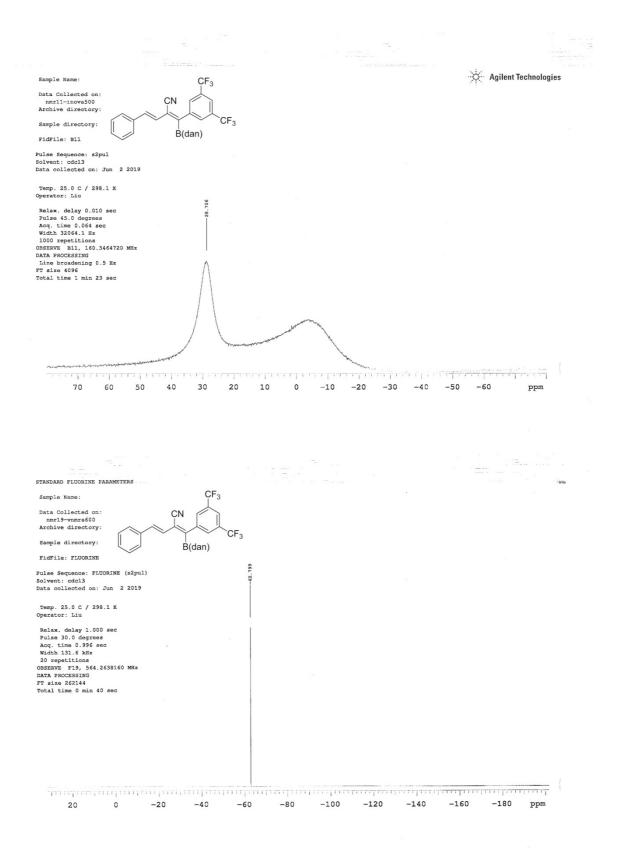


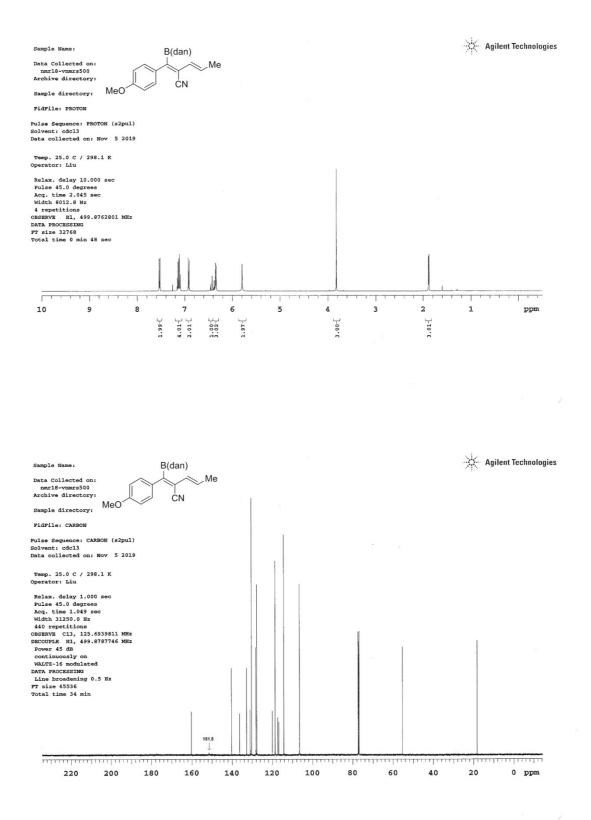


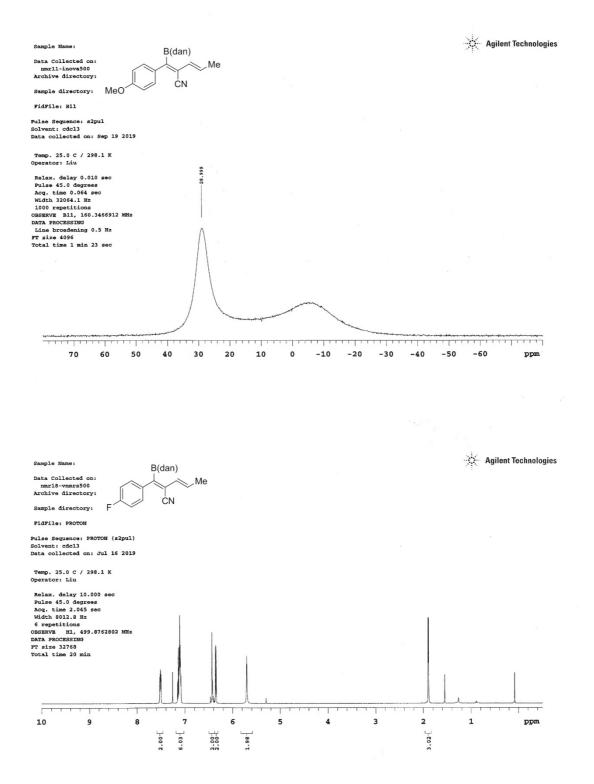


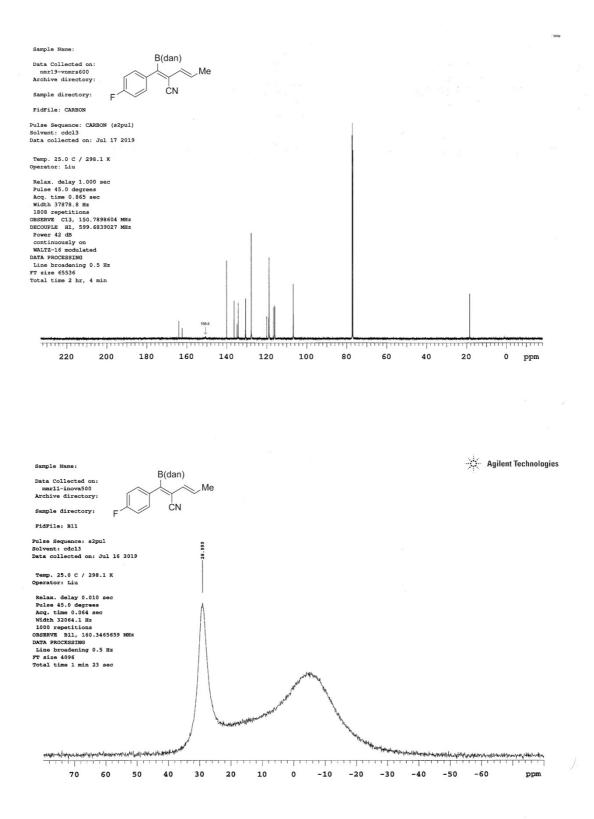
20 0 -20 -40 -60 -80 -100 -120 -140 -160 -180 ppm

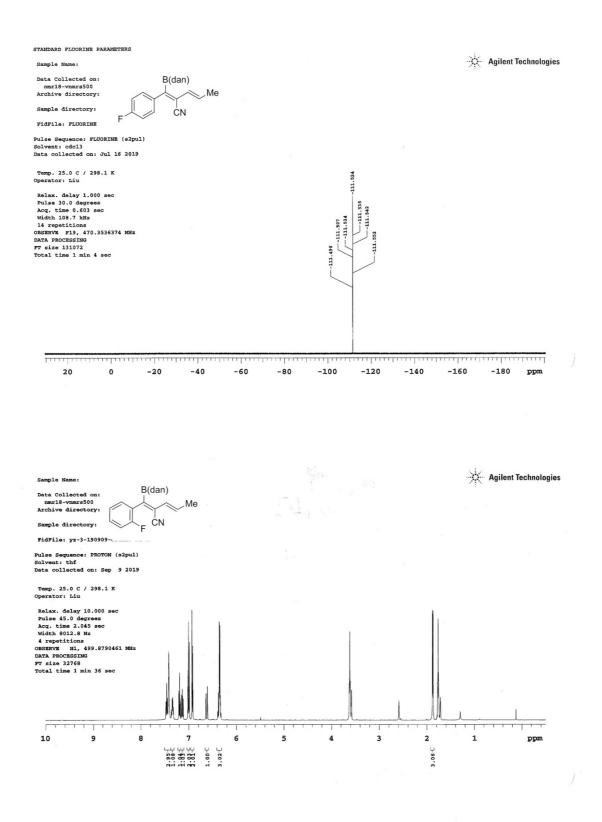


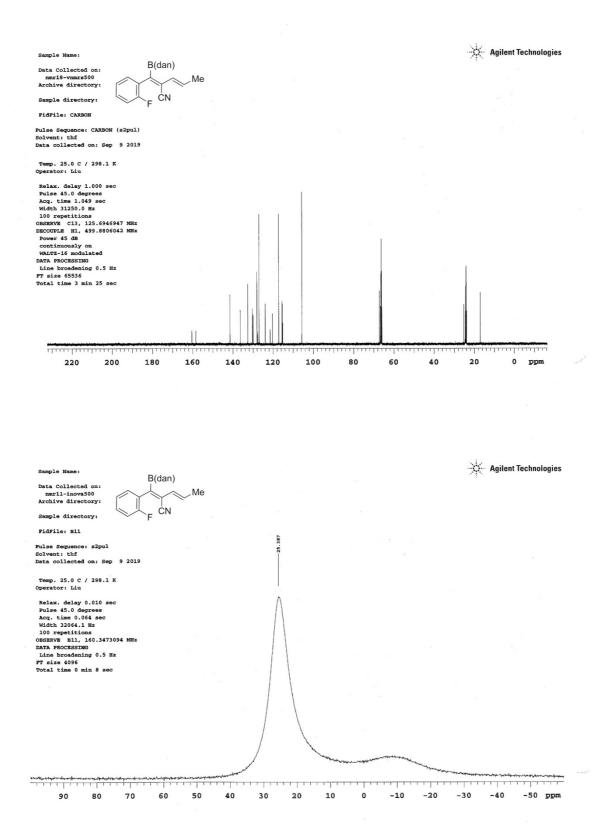


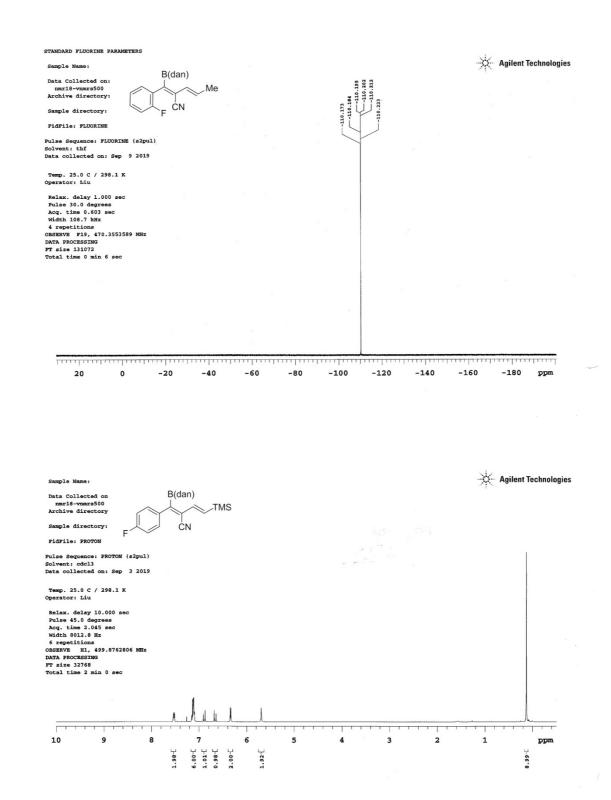


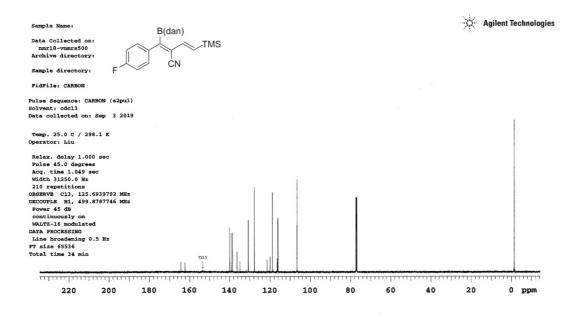


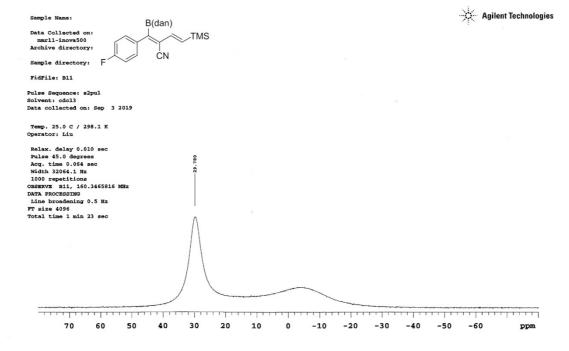


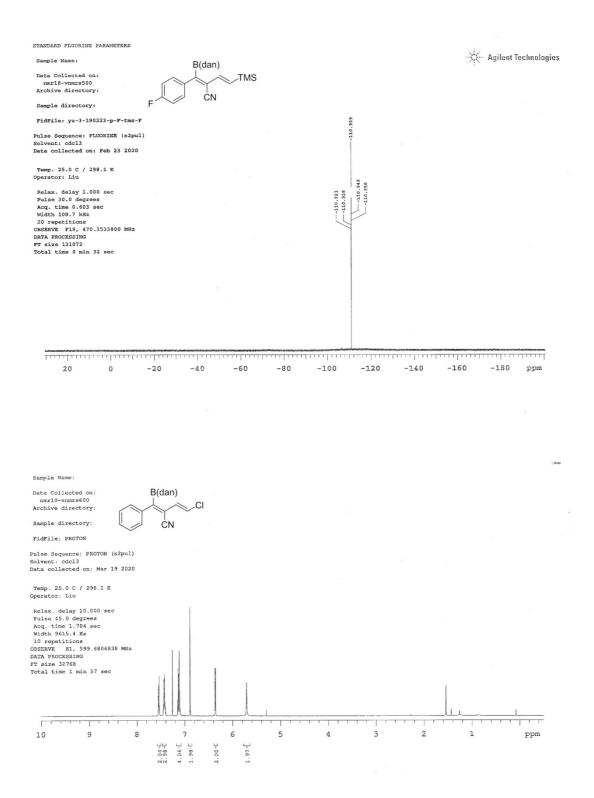


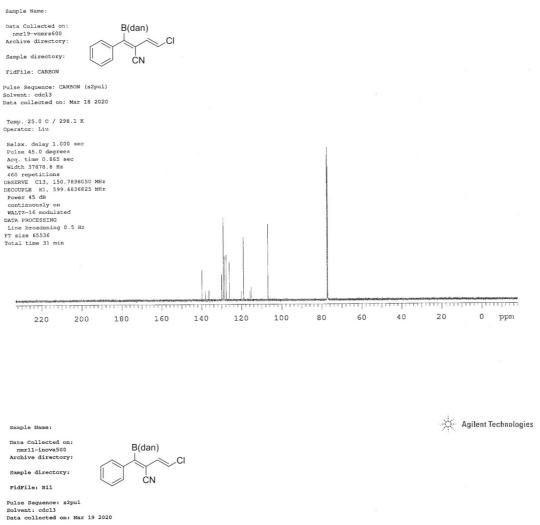




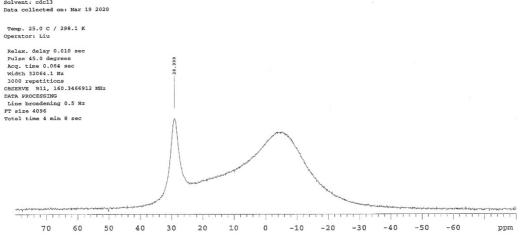


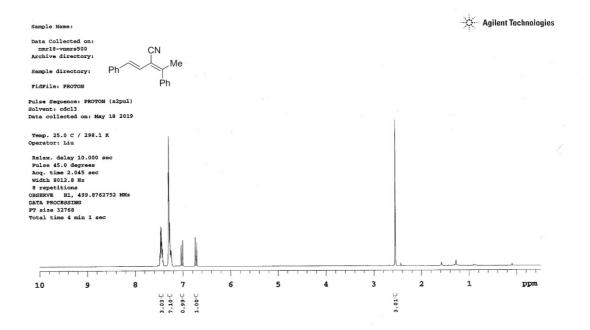


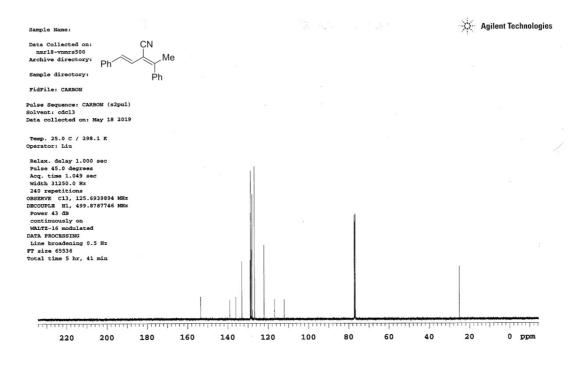


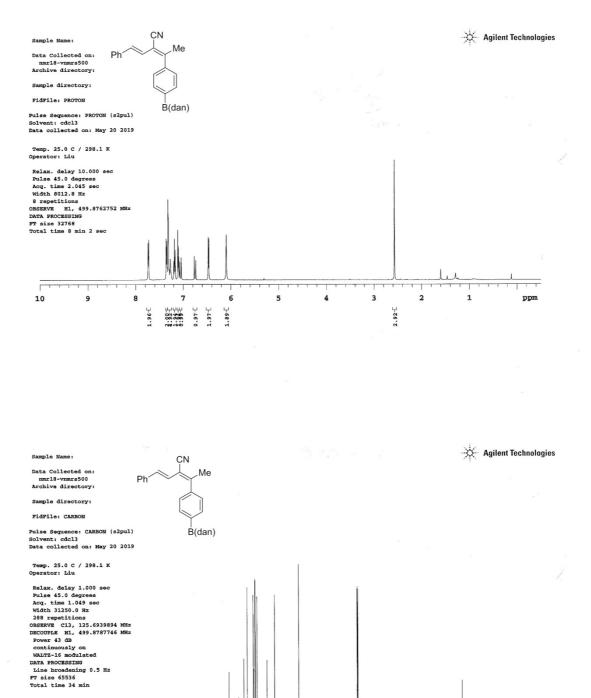


aw.







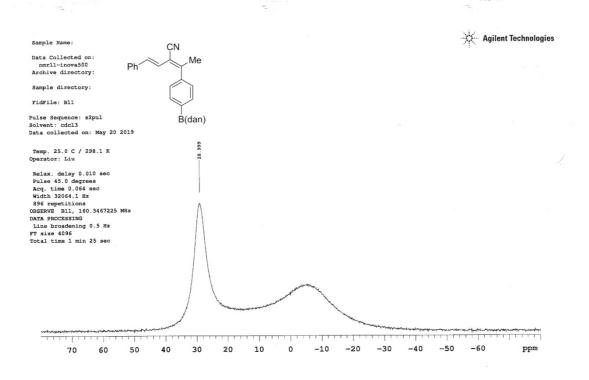


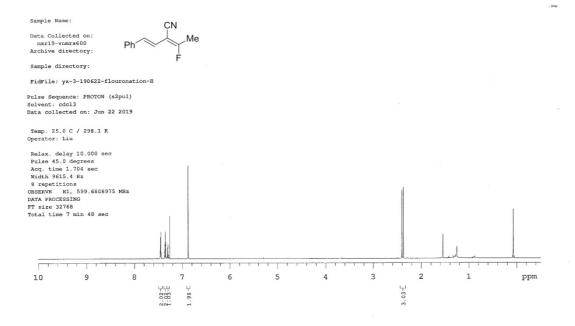


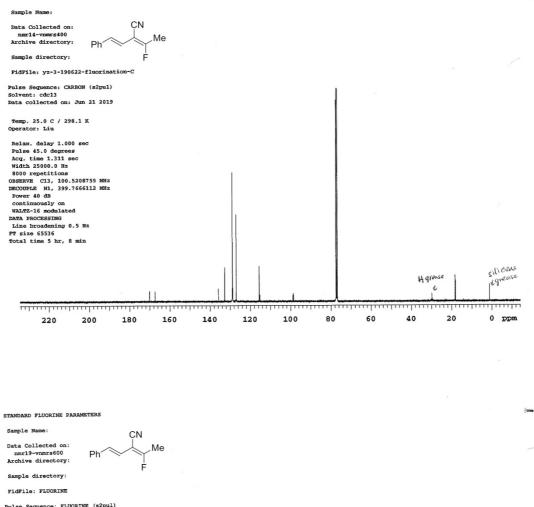
THE

0 ppm

......







Pulse Sequence: FLUORINE (s2pul) Solvent: cdcl3 Data collected on: Jun 22 2019

Temp. 25.0 C / 298.1 K Operator: Liu

20

Ralax. delay 1.000 sec Pulse 30.0 degrees Acq. time 0.996 sec Width 131.6 kiz 20 repetitions OBSERVE F19, 564.2638160 MHz DATA PROCESSING FT size 262144 Total time 3 min 20 sec

0

-20

-80

75.610

580 -75.671

-60

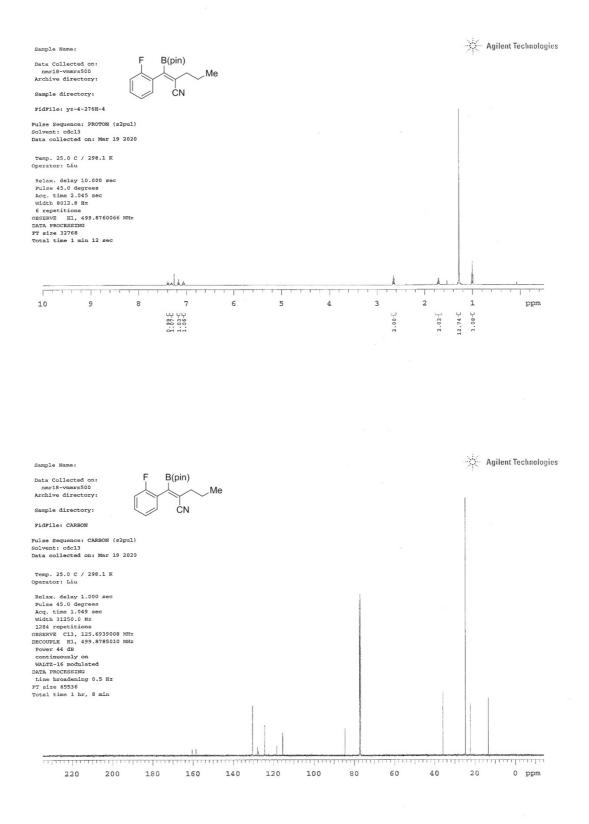
-40

-160 -140 -100 -120

-180

ppm

3 grintle



Agilent Technologies Sample Name: Data Collected on: nmr11-inova500 Archive directory: F B(pin) Me Sample directory: ćΝ FidFile: B11 Pulse Sequence: s2pul Solvent: cdcl3 Data collected on: Mar 19 2020 Temp. 25.0 C / 298.1 K Operator: Liu 878 Relax. delay 0.010 sec Pulse 45.0 degrees Acq. time 0.064 sec Width 32064.1 Hz 1000 repetitions OSSERVE E11, 160.3467068 MHz DXTA FROCESSING Line broadening 0.5 Hz FT size 4096 Total time 1 min 23 sec 29. -----50 10 -20 -30 -40 -50 -60 -70 ppm 70 60 40 30 20 0 -10 STANDARD FLUORINE PARAMETERS Agilent Technologies Sample Name: B(pin) F Data Collected on: nmr18-vnmrs500 Archive directory: Me ĊΝ Sample directory: FidFile: yz-4-276F-4 Pulse Sequence: FLUORINE (s2pul) Solvent: cdcl3 Data collected on: Mar 19 2020 Temp. 25.0 C / 298.1 K Operator: Liu -112.744 -112.755 -112.765 -112.765 Relax. delay 1.000 sec Pulme 30.0 degrees Acq. time 0.603 sec Width 108.7 kHz 4 repetitions ONSERVE F19, 470.3533800 MHz DATA PHOCESSING FT size 13.072 Total time 0 min 6 sec 727 112. transfera transfera fransfera fransfera fransfera fransfera fransfera fransfera fransfera fransfera fransfera f

-80 -100

-120

-140

-160

-180 ppm

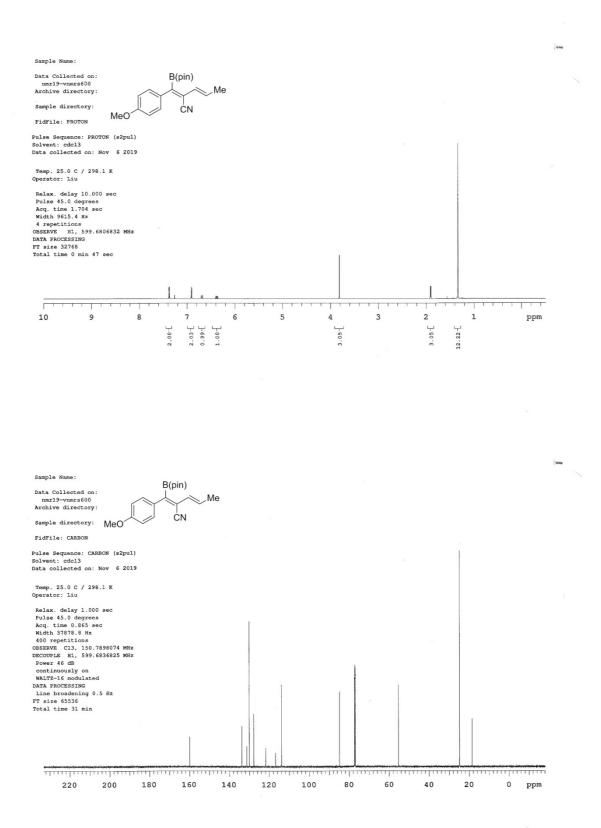
20

0

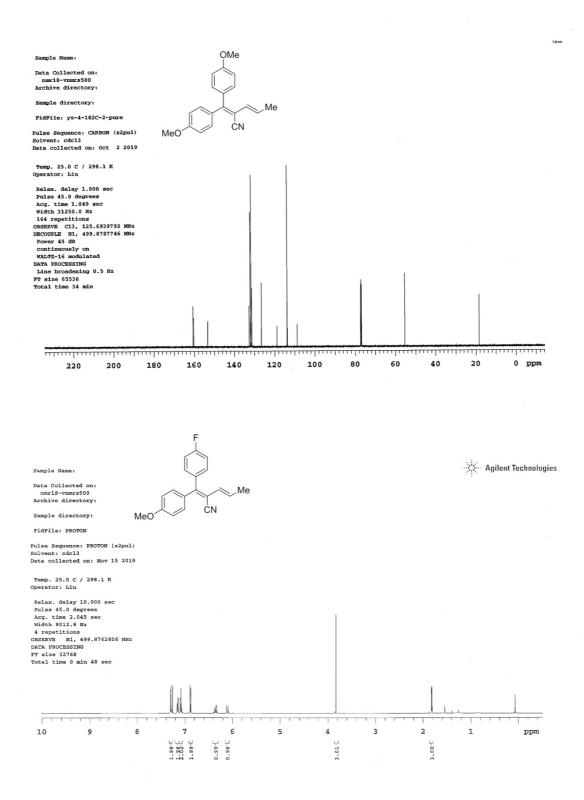
-20

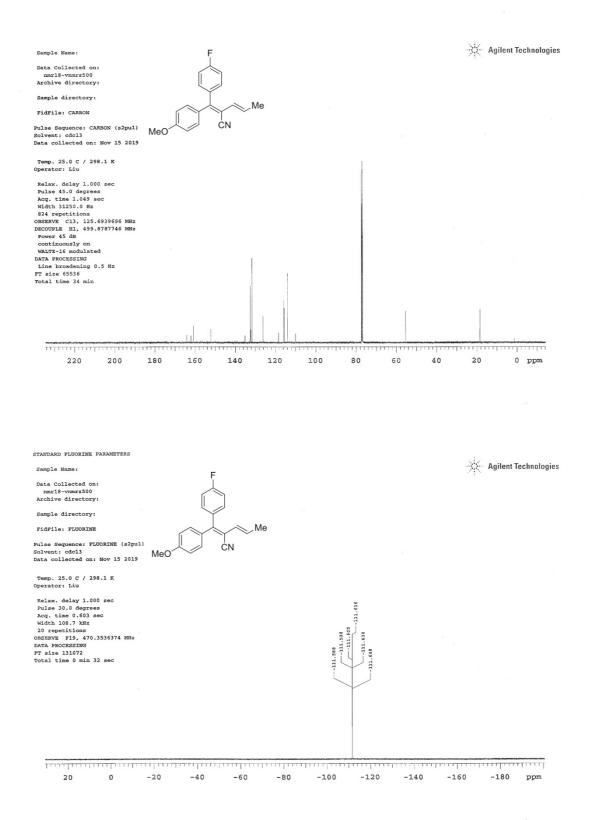
-40

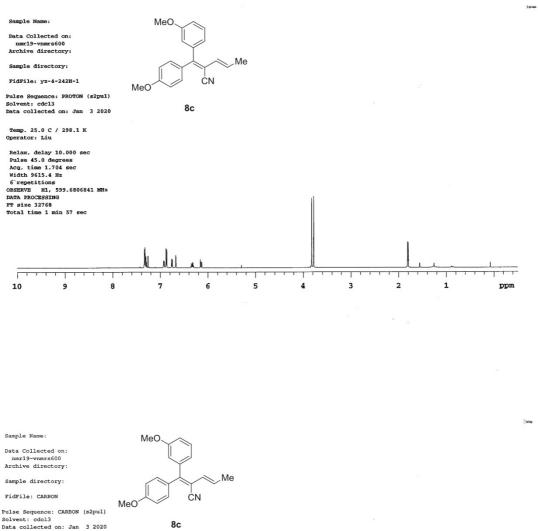
-60

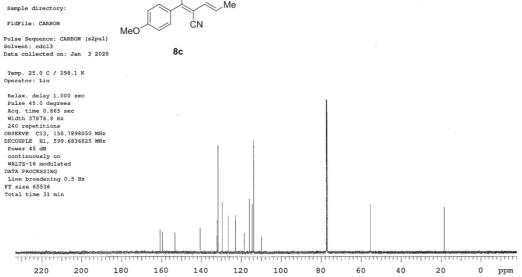


Agilent Technologies Sample Name: Data Collected on: nmr11-inova500 Archive directory: B(pin) _Me Sample directory: ćΝ MeO FidFile: B11 Pulse Sequence: s2pul Solvent: cdcl3 Data collected on: Sep 21 2019 Temp. 25.0 C / 298.1 K Operator: Liu Relax. delay 0.010 sec Fulae 45.0 degrees Acq. time 0.064 sec Width 32064.1 Hz 1000 repetitions OSERWE BIL, 160.3466912 MHz DATA PROCESSING Line broadening 0.5 Hz FT size 4096 Total time 1 min 23 sec -29.976 70 60 50 0 -20 -30 -40 -50 -60 ppm 40 30 20 10 -10 Agilent Technologies Sample Name: ОМе Data Collected on: nmr18-vnmrs500 Archive directory: Sample directory: FidFile: PROTON Me Fulse Sequence: PROTON (s2pul) Solvent: cdcl3 Data collected on: Oct 2 2019 MeO ćΝ Temp. 25.0 C / 298.1 K Operator: Liu Relax. delay 10.000 sec Fulae 45.0 degrees Acq. time 2.045 sec Width 8012.8 Hz 8 repetitions OBSERVE HI, 499.8762802 MHz DATA PROCESSING FT size 32768 Total time 1 min 36 sec 1 -------1 Т 10 7 4 2 9 8 6 5 3 1 ppm 2.01-C 2.02-C 4.03-C 1.01-3:93∉ 3.00-

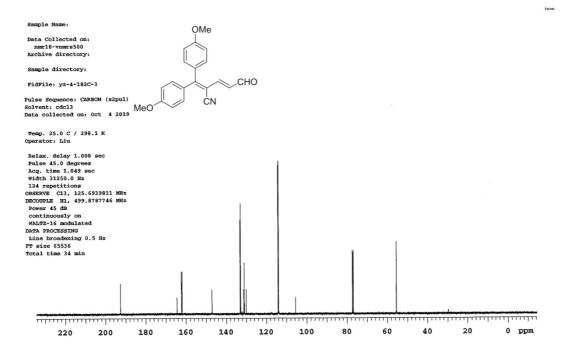


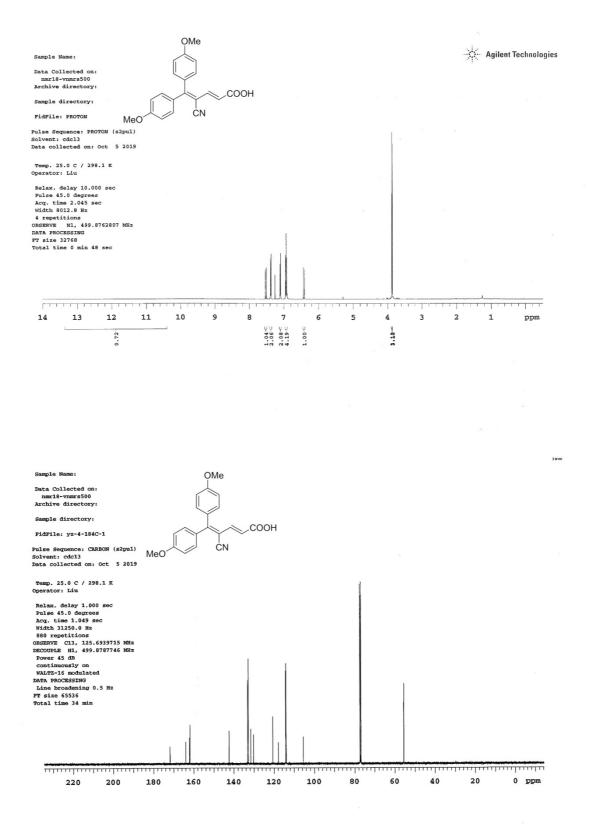


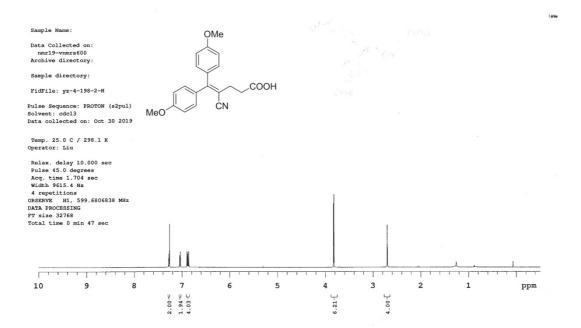


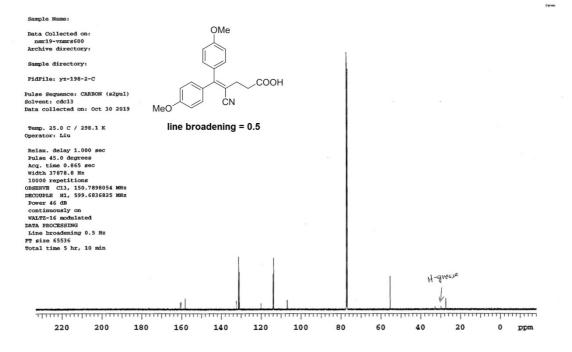


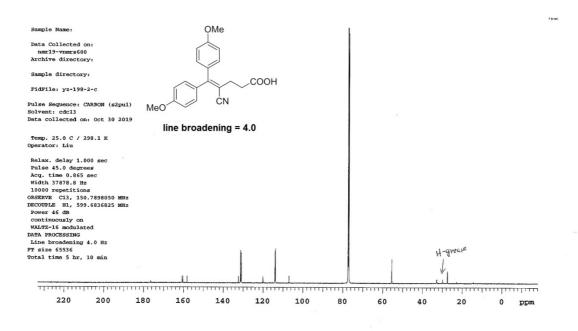
Agilent Technologies Sample Name: Data Collected on: nmr18-vnmrs500 Archive directory: ОМе Sample directory: FidFile: PROTON CHO Pulse Sequence: PROTON (s2pul) Solvent: cdc13 Data collected on: Oct 4 2019 ćΝ MeO Temp. 25.0 C / 298.1 K Operator: Liu Relax. delay 10.000 sec Pulse 45.0 degrees Acq. time 2.045 sec Width 8012.8 Hz 4 repetitions OSSERVE #1, 499.8762807 MHz DATA PROCESSING FT size 32768 Total time 0 min 48 sec UU 1 [---------- T - T Т Т Т 7 10 9 8 6 5 4 3 2 1 ppm 7-66.0 6.03-E











References:

- 1. (a) Xu, S.; Zhang, Y.; Li, B.; Liu, S.-Y. J. Am. Chem. Soc. 2016, 138, 14566-14569. (b) Xu, S.; Haeffner,
- F.; Li, B.; Zakharov, L. N.; Liu, S.-Y. Angew. Chem. Int. Ed. 2014, 53, 6795-6799.
- 2. Huang, S.; Kotzner, L.; De, C. K.; List, B. J. Am. Chem. Soc. 2015, 137, 3446-3449.
- 3. Liu, J.; Ren, Q.; Zhang, X.; Gong, H. Angew. Chem. Int. Ed. 2016, 55, 15544-15548.
- 4. Li, H.; Zhang, Z.; Shangguan, X.; Huang, S.; Chen, J.; Zhang, Y.; Wang, J. *Angew. Chem. Int. Ed.* **2014**, 53, 11921-11925.
- 5. Ren, H.; Krasovskiy, A.; Knochel, P. Org. Lett. 2004, 6, 4215-4217.
- 6. Stille, J. K.; Simpson, J. H. J. Am. Chem. Soc. 1987, 109, 2138-2152.
- 7. Lehr, K.; Schulthoff, S.; Ueda, Y.; Mariz, R.; Leseurre, L.; Gabor, B.; Fürstner, A. *Chem. Eur. J.* **2015**, *21*, 219-227.
- 8. Baladi, T.; Granzhan, A.; Piguel, S. Eur. J. Org. Chem. 2016, 2421-2434.
- 9. Dolby, L. J.; Wilkins, C.; Frey, T. G. J. Org. Chem. 1966, 31, 1110-1116.
- 10. Park, S.; Malcolmson, S. J. ACS Catal. 2018, 8, 8468-8476.
- 11. Kinoshita, H.; Ishikawa, T.; Miura, K. Org. Lett. 2011, 13, 6192-6195.
- 12. Dateer, R. B.; Pati, K.; Liu, R. S. Chem. Commun, 2012, 48, 7200-7202.
- 13. Kraus, G. A.; Dong, P.; Qu, Y.; Evans, A.; Carpenter, S. Tetrahedron Lett. 2016, 57, 5185-5187.
- 14. Mokar, B. D.; Liu, R. S. Chem. Commun. 2014, 50, 8966-8969.
- 15. Rossi, R.; Carpita, A. Tetrahedron Lett. 1986, 27, 2529-2532.
- 16. Andreini, B. P.; Carpita, A.; Rossi, R. Tetrahedron Lett. 1988, 29, 2239-2242.