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Supplemental Information

Protein Stability Buffers the Cost of

Translation Attenuation following

elF2α Phosphorylation

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Supplemental Information

Table S1: Top 10 GO terms of translationally regulated mRNAs as determined by anota2seq, Related to Figure 1

Top 10 GO terms of relative translationally down- or upregulated mRNAs as determined by anota2seq at 2 and 4 hours following Tm injection.

Category	GO terms	count	%	p value	adjusted
					p value
GOTERM_BP_DIRECT	translation	136	8.2	1.90E-52	6.60E-49
GOTERM_BP_DIRECT	transport	232	14	2.50E-15	4.50E-12
GOTERM_BP_DIRECT	mitochondrial translation	21	1.3	1.00E-13	1.20E-10
GOTERM_BP_DIRECT	protein transport	99	6	2.40E-13	2.20E-10
GOTERM_BP_DIRECT	rRNA processing	36	2.2	1.10E-11	8.10E-09
GOTERM_BP_DIRECT	oxidation-reduction process	101	6.1	1.20E-10	7.30E-08
GOTERM_BP_DIRECT	glutathione metabolic process	21	1.3	1.50E-10	7.40E-08
GOTERM_BP_DIRECT	antigen processing and	16	1	9.10E-10	4.00E-07
	presentation of exogenous peptide				
	antigen via MHC class I, TAP-				
	dependent				
GOTERM_BP_DIRECT	proton transport	21	1.3	4.90E-09	1.90E-06
GOTERM_BP_DIRECT	vesicle-mediated transport	43	2.6	1.20E-08	4.20E-06

Translationally downregulated mRNAs at 2 hours

Translationally downregulated mRNAs at 4 hours

Category	GO terms	count	%	p value	adjusted
					p value
GOTERM_BP_DIRECT	translation	105	10.	7.20E-50	1.80E-46
GOTERM_BP_DIRECT	transport	144	14.	4.90E-11	6.10E-08
GOTERM_BP_DIRECT	oxidation-reduction process	69	6.9	8.00E-10	6.60E-07
GOTERM_BP_DIRECT	proton transport	17	1.7	4.20E-09	2.60E-06
GOTERM_BP_DIRECT	ATP synthesis coupled proton	11	1.1	2.50E-08	1.30E-05
GOTERM_BP_DIRECT	ribosomal small subunit assembly	10	1	1.50E-07	6.20E-05
GOTERM_BP_DIRECT	cytoplasmic translation	12	1.2	1.90E-07	6.60E-05
GOTERM_BP_DIRECT	mitochondrial translation	12	1.2	3.60E-07	1.10E-04
GOTERM_BP_DIRECT	rRNA processing	20	2	4.50E-06	1.20E-03
GOTERM_BP_DIRECT	ATP metabolic process	11	1.1	9.20E-06	2.30E-03

Translationally upregulated mRNAs at 2 hours

Category	GO terms	count	%	p value	adjusted
					p value
GOTERM_BP_DIRECT	mRNA processing	95	5.1	3.50E-25	1.60E-21
GOTERM_BP_DIRECT	metabolic process	116	6.2	6.20E-24	1.50E-20
GOTERM_BP_DIRECT	RNA splicing	70	3.8	2.40E-18	3.80E-15
GOTERM_BP_DIRECT	protein transport	110	5.9	1.00E-12	1.20E-09
GOTERM_BP_DIRECT	covalent chromatin modification	61	3.3	4.10E-11	3.80E-08
GOTERM_BP_DIRECT	transcription, DNA-templated	255	13.	6.70E-11	5.30E-08
GOTERM_BP_DIRECT	flavonoid biosynthetic process	15	0.8	1.80E-10	1.20E-07

GOTERM_BP_DIRECT	flavonoid glucuronidation	15	0.8	1.80E-10	1.20E-07
GOTERM_BP_DIRECT	oxidation-reduction process	113	6.1	4.30E-10	2.50E-07
GOTERM_BP_DIRECT	DNA repair	65	3.5	1.40E-09	7.40E-07

Translationally upregulated mRNAs at 4 hours

Category	GO terms	count	%	p value	adjusted
					p value
GOTERM_BP_DIRECT	flavonoid glucuronidation	12	1.8	6.20E-12	1.40E-08
GOTERM_BP_DIRECT	flavonoid biosynthetic process	12	1.8	6.20E-12	1.40E-08
GOTERM_BP_DIRECT	xenobiotic glucuronidation	9	1.4	7.20E-12	8.20E-09
GOTERM_BP_DIRECT	metabolic process	38	5.7	1.50E-07	1.20E-04
GOTERM_BP_DIRECT	transcription, DNA-templated	93	14	6.90E-06	3.90E-03
GOTERM_BP_DIRECT	mRNA processing	25	3.8	7.20E-05	3.20E-02
GOTERM_BP_DIRECT	protein ubiquitination involved in	15	2.3	7.30E-05	2.80E-02
	ubiquitin-dependent protein				
	catabolic process				
GOTERM_BP_DIRECT	regulation of transcription, DNA-	103	15.	7.40E-05	2.40E-02
GOTERM_BP_DIRECT	response to nutrient	10	1.5	4.10E-04	1.10E-01
GOTERM_BP_DIRECT	protein ubiquitination	25	3.8	4.20E-04	1.00E-01

Table S2: Oligonucleotides used in this study

MmPpp1r15a_R: tctcaggtcctccttcctca	Sigma-Aldrich	N/A
MmChop_F: cctagcttggctgacagagg	Sigma-Aldrich	N/A
MmChop_R: ctgctccttctccttcatgc	Sigma-Aldrich	N/A
MmBiP_F: ctttgatcagcgggtcatgg	Sigma-Aldrich	N/A
MmBiP_R: agctcttcaaatttggcccg	Sigma-Aldrich	N/A
MmGrp94_F: agtcgggaagcaacagagaa	Sigma-Aldrich	N/A
MmGrp94_R: tctccatgttgccagaccat	Sigma-Aldrich	N/A
MmNedd4_F: atgtggatgctgggagttga	Sigma-Aldrich	N/A
MmNedd4_R: ctgcacgctggtaaggattc	Sigma-Aldrich	N/A
MmRps6_F: aaggtaagaagcccaggacc	Sigma-Aldrich	N/A
MmRps6_R: ctgttcctggcgcttttctt	Sigma-Aldrich	N/A
<i>MmCul4b</i> _F: cccttgtcttccttgcacac	Sigma-Aldrich	N/A
MmCul4b_R: aggaagcaaagcagagtcct	Sigma-Aldrich	N/A
MmAsns_F: aggaggcatcgagaaaacca	Sigma-Aldrich	N/A
MmAsns_R: aagaagggagtggtgggagtg	Sigma-Aldrich	N/A
HsRpl4_F: tgggacgtttctgcatttgg	Sigma-Aldrich	N/A
HsRpl4_R: cgatggatcttcttgcgtgg	Sigma-Aldrich	N/A
HsMrpI12_F: ggaggcggtggaagaagata	Sigma-Aldrich	N/A
HsMrpI12_R: gactccaccagcttctttgc	Sigma-Aldrich	N/A
HsMrpl28_F: ggacctgtgctccaagtttg	Sigma-Aldrich	N/A
HsMrpl28_R: ttgaacaggggtacagggtc	Sigma-Aldrich	N/A
HsMrps15_F: ggccaacaagaaggagatgc	Sigma-Aldrich	N/A
HsMrps15_R: ttgtgggctttgtcctttcg	Sigma-Aldrich	N/A
HsMrps25_F: cagggaagaggaggaggaga	Sigma-Aldrich	N/A
HsMrps25_R: aacccttgggtgcagatcat	Sigma-Aldrich	N/A
MmMrpI12_F: tcctcctcttccctgag	Sigma-Aldrich	N/A
MmMrpl12_R: cctccaaggctgctttgatc	Sigma-Aldrich	N/A
MmMrpl28_F: ccccggtacactacaaacct	Sigma-Aldrich	N/A
MmMrpl28_R: tccacaccttcttcaccctc	Sigma-Aldrich	N/A
MmMrps15_F: caagcaggacgatgaaccac	Sigma-Aldrich	N/A
MmMrps15_R: attgcggatcctgacagtca	Sigma-Aldrich	N/A
MmMrps25_F: cccttcctgcggttctatct	Sigma-Aldrich	N/A

MmMrps25_R: ggccttctacttcgcacatg	Sigma-Aldrich	N/A
Cyclophilin B_F: ggagatggcacaggaggaa	Sigma-Aldrich	N/A
Cyclophilin B_R: gcccgtagtgcttcagctt	Sigma-Aldrich	N/A
Luciferase_F: aaaggatatcaggtggcccc	Sigma-Aldrich	N/A
Luciferase_R: ccacaaacacaactcctccg	Sigma-Aldrich	N/A
Atp5d_5UTR_F:	Sigma-Aldrich	N/A
gatacaactagtccttcggagaatcctgtgc		
Atp5d_5UTR_R:	Sigma-Aldrich	N/A
gatacaccatggggtggcggaaaagtcagtg		
Atf4_5UTR_F:	Sigma-Aldrich	N/A
gatacaactagtggctaggtgtcccaccc		
Atf4_5UTR_R:	Sigma-Aldrich	N/A
gatacagaattcgttgtggggctttgctggattcg		
Mrps28_5UTR_F:	Sigma-Aldrich	N/A
gatacagaattcaggcaagctttaaactaccggac		
Mrps28_5UTR_R:	Sigma-Aldrich	N/A
gatacaccatgggacgcctgcgcgacctc		
Mrps33_5UTR_F:	Sigma-Aldrich	N/A
gatacaactagttccgggttgtccgccgac		
Mrps33_5UTR_R:	Sigma-Aldrich	N/A
gatacaccatggttcttcaccagtagggaacaggattcc	<u></u>	
Rps15a_5UTR_F:	Sigma-Aldrich	N/A
gatacaactagtataattccgtgcgtttgcccggcccactctc		
Rps15a_5UTR_R:	Sigma-Aldrich	N/A
gatacaccatggggtggcggcggggggaagatggcagaa		
	Signa Aldrich	
Rps18_50TR_F:	Sigma-Aldrich	IN/A
	Sigma Aldrich	ΝΙ/Λ
npsio_501n_n.	Sigma-Alunch	IN/A
Rn/35 5LITR F	Sigma-Aldrich	Ν/Δ
		1 1/7 1
gguologiguuougigguuguuguguuolo	Sigma-Aldrich	N/A
<i>Rpl35_</i> 5UTR_F:		
catgccatggttcggcgttgttcacg		
	Sigma-Aldrich	N/A
Rps24_5UTR_F:		
ggactagtgctggggtccttcctt	<u></u>	
Pro24 FLITP P.	Sigma-Aldrich	N/A
rpsz4_50 m_n.		
	Sigma-Aldrich	N/A
<i>Rps15</i> 5UTR F:		
gatacaactagtgtacttccttttccgagtaaccgccaag		
	Sigma-Aldrich	N/A
<i>Rps15_</i> 5UTR_R:	-	
gatacaccatggcttggcggttactcggaaaaggaagtac		



Figure S1. Gene ontology enrichment map of mRNAs transcriptionally upregulated after Tm, Related to Figure 1

Significant (P<0.001) gene ontology enrichment maps of the 900 mRNAs transcriptionally upregulated (2-fold cut-off) 4 hours after Tm injection.



Figure S2. GO analysis of mRNA translationally affected following Tm, Related to Figure 3

(A) Significant (P<0.001) GO enrichment maps of translationally downregulated mRNAs as determined by anota2seq at 2 and 4 hours following Tm injection.(B) Same as (A) for genes resistant to translational attenuation (relative upregulation).



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E3 ligases upregulated (relative upregulation) at 2 h

Klhl9	Ubr2	Fbxo11	Brca1
Klhl28	Trip12	Usp36	G2E3
Klhdc7A	Bag6	Trim23	Usp34
Cdc73	Ddb1	Ubr3	Fbxo21
Pja2	Dzip3	Rnf111	Rnf123
Klhl22	Usp24	Fbxo10	Usp33
Cand1	Asb3	Cyld	Usp22
Usp4	Nfe2L2	Usp9X	Dcaf8
Shprh	Abtb1	Dcaf17	Ddb2
Usp47	Tnks	Sash1	Usp48
Leo1	Usp19	Pdzrn3	
Ltn1	Hectd3	Kbtbd4	
Uvssa	Pja1	Ctr9	
HItf	Trim33	Cdc20	
Med17	Sqstm1	Ttc3	

E3 ligases upregulated (relative upregulation) at 4 h

Klhl28	Nedd4
Crbn	Usp47
Socs4	Ubr5
Klhl24	Arntl
Vps4B	Fbxo21
Klhl22	Fbxo30
Cul4B	Ube3A
Rnf216	Mib2
Vps37A	Asb3
Klhdc7A	Cbll1
Rnf125	
Usp34	
Ubr7	
Cyld	
Lrrc41	

Figure S3. Relative translational regulation of E3 ligases following Tm, Related to Figure 3

(A) Boxplot showing the median translation efficiency (TA) as determined by anota2seq at 2 and 4 hours following Tm treatment.

(B) List of E3 ligases translationally upregulated (relative upregulation) at 2 and 4 hours following Tm, respectively.



Figure S4. mRNAs abundance of selected transcripts, Related to Figure 3 Log2 FPKM values for indicated genes at 0, 2 and 4 hours following Tm.



Figure S5. Cytosolic and mitochondrial ribosomal proteins are translationally downregulated by Tm, Related to Figure 3

(A) Heat map showing z-scores of anota2seq translation efficiencies of all 79 cytosolic ribosomal proteins (CytoRP) subunit-encoding mRNAs at 0, 2 and 4 hours following Tm injection.

(B) Same as (A) but with all 82 mitochondrial ribosomal proteins (MitoRP) subunits.



Figure S6. Translation of mitochondrial ribosomal proteins is not sensitive to acute mTORC1 inhibition, Related to Figure 4

(A) Representative polysome profile (optical density of the fractions at 254 nm) on 5-50% sucrose gradients of HeLa cells treated with DMSO or 250 nM of Torin1 for 2 hours.

(B) *Rpl4*, *Mrpl12*, *Mrpl28*, *Mrps25* and *Mrps15* mRNA abundance from heavy polysomal fractions (more than 3 ribosomes) from (A) assessed by qPCR. Data are means \pm SEM (n = 5-9). **P<0.01 as determined by the unpaired Student t-test.



Figure S7. Composition of UTRs regulates the translational response to $eIF2\alpha$ phosphorylation, Related to Figure 5

Firefly luciferase activity in NIH3T3 cells transfected with reporter mRNAs containing the 5'UTRs of indicated mRNAs and treated with either vehicle or 2.5 μ g/ml Tunicamycin (Tm) for 2 hours. Firefly luciferase activity was normalized against a transfection control (see Methods) and vehicle-treated cells. Data are means \pm SEM (n=6). Statistical analysis was carried out in Prism 8 using a two-way ANOVA test.



Figure S8. Translationally downregulated mRNAs encode for highly stable proteins, Related to Figure 6

Same as in Figure 6E with protein half-lives analyzed from (Schwanhäusser et al, 2013).

(A) Boxplot of half-lives of proteins encoded by down- or upregulated (Down or Up) mRNAs at 2 hours following Tm injection.

(B) Same as (A) comparing the half-lives of proteins encoded by translationally downregulated mRNAs with all 79 CytoRP subunits 2 hours after Tm.

(C) Same as (B) with all 82 MitoRP subunits. (A-C) ***P<0.001 as determined by the unpaired Wilcoxon test.



Figure S9. eIF2α phosphorylation induced by different stressors downregulates translation of mRNAs encoding for stable proteins, Related to Figure 6

(A) Boxplot of half-lives of proteins encoded by the translationally down or upregulated (Down or Up) mRNAs using data from (Sidrauski et al, 2015) 1 hours following Tm treatment of HEK293T cells.

(B) Same as (A) but excluding the RPs from the category of translationally downregulated mRNAs (Down w/o RP).

(C) Same as (A) but with data from (Andreev et al, 2015) acquired 30 minutes following arsenite treatment of HEK293T cells.

(D) Same as (C) but excluding the RPs from the category of translationally downregulated mRNAs (Down w/o RP).

(A-D) Protein half-lives were analyzed from (Ly et al, 2018)

*P<0.05, ***P<0.001 as determined by the unpaired Wilcoxon test.