# SUPPORTING INFORMATION

# Glucose and maltose surface functionalized

# thermoresponsive poly(N-vinylcaprolactam) nanogels

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Figure S1. <sup>1</sup>H-NMR spectrum of Glc-N<sub>3</sub>



Figure S2.<sup>13</sup>C-NMR spectrum of Glc-N<sub>3</sub>

## <span id="page-2-0"></span>**NMR-spectra of Glc-TEG-N<sup>3</sup>**



Figure S3. <sup>1</sup>H-NMR spectrum of Glc-TEG-N<sub>3</sub>.



Figure S4.<sup>13</sup>C-NMR spectrum of Glc-TEG-N<sub>3</sub>.

## <span id="page-3-0"></span>**NMR-spectra of Mal-N<sup>3</sup>**



Figure S5. <sup>1</sup>H-NMR spectrum of Glc-TEG-N3**.**



Figure S6.<sup>13</sup>C-NMR spectrum of Glc-TEG-N<sub>3</sub>.

# <span id="page-4-0"></span>**NMR-spectra of Glc-TEG-N<sup>3</sup>**



Figure S7. <sup>1</sup>H-NMR spectrum of Glc-TEG-N3**.**



Figure S8.<sup>13</sup>C-NMR spectrum of Glc-TEG-N<sub>3</sub>.

#### <span id="page-5-0"></span>**Kinetics of PNVCL nanogel synthesis**



Figure S9. 1H-NMR spectrum of conversion sample taken after 0h (left) and after 20h (right)



Figure S10. Kinetics of the polymerization according to  ${}^{1}$ H-NMR

# <span id="page-7-0"></span>**<sup>1</sup>H-NMR spectrum of PA**



Figure S11. <sup>1</sup>H-NMR spectrum of PA in  $D_2O$ 

## <span id="page-8-0"></span>**<sup>1</sup>H-NMR spectrum of PNVCL nanogel**



Figure S12. 1H-NMR spectrum of PNVCL nanogel

## <span id="page-9-0"></span>**<sup>1</sup>H-NMR spectrum of PNVCL-TEG-Glc**



Figure S13. 1H-NMR spectrum of PNVCL-TEG-Glc

## <span id="page-10-0"></span>**<sup>1</sup>H-NMR spectrum of PNVCL-Mal**



Figure S14. 1H-NMR spectrum of PNVCL-Mal

## <span id="page-11-0"></span>**<sup>1</sup>H-NMR spectrum of PNVCL-TEG-Mal**



Figure S15. 1H-NMR spectrum of PNVCL-TEG-Mal

#### <span id="page-12-0"></span>**Kinetics of a sugar functionalization of PNVCL-PA nanogel**

To make sure that the reaction time was long enough, a replica of the conjugation reaction to obtain PNVLC-Glc was performed in  $D_2O$  and followed with <sup>1</sup>H-NMR. The intensity of a peak at 5.8 ppm, was used to follow the reaction (1H-spectrum of PNVCL-GIc with peak assignations is presented in main article Figure 6) and results are presented in Figure S12. It is seen that the reaction has happened during  $8 h$ . <sup>1</sup>H-NR of the purified product is presented in Figure S13. The nanogel synthesized in  $D_2O$  differs from the PNVCL-GIc nanogel, as there is a deuterium ion instead of a proton attached to the triazole ring. Akula et al. have studied this phenomenon of CuAAc reactions in  $D_2O$  as "a efficient route for the synthesis of deuterated 1,2,3-triazoles".S1



Figure S16. Kinetic of reaction between Glc-N<sub>3</sub> and PNVL-PA nanogel based on a <sup>1</sup>H-NMR-experiment

S1. Akula HK, Lakshman MK. Synthesis of deuterated 1,2,3-triazoles. *The Journal of organic* 

*chemistry*. 2012;77(20):8896

<span id="page-13-0"></span>**<sup>1</sup>H-NMR spectrum of PNVCL-Glc prepared via CuAAC in D2O** 



Figure S17. 1H-NMR spectrum of PNVCL-Glc prepare via CuAAC in D2O

<span id="page-14-0"></span>**Size distributions (CONTIN) of PNVCL-sugars at 25 oC**



Figure S18. Hydrodynamic size distributions (CONTIN) of selected nanogels; 0.25 mg/ml in 10 mM HEPES (pH=7.4) at 25 °C

#### <span id="page-15-0"></span>**Hydrodynamic size of PNVCL-sugars as a function of temperature**



Figure S19. Hydrodynamic mean size of selected nanogels as functions of temperature; 0.25 mg/ml in 10 mM HEPES (pH=7.4) at 25 °C

#### <span id="page-16-0"></span>**Salt induced aggregation at 50 <sup>o</sup>C**

Selected salt induced aggregation tests are shown Figure S16. Two points were obtained from each curve  $C_1$ and  $C_2$ . Aggregation concentration,  $C_1$ , was taken from the point where transmittance as a function of NaCl concentration broke the linear trend. Precipitation concentration  $(C_2)$  was determined a point, where the transmittance had increased compared to previous concentration. The increase of transmittance was due to visible aggregates sedimenting out of the solution in the given 15 min stabilization time. Both aggregation  $(C_1)$ and precipitation  $(C_2)$  concentrations are listed in table S1. In addition, size distribution of PNVCL-PA in selected NaCl concentrations was measured with light scattering, see Figure S17. There was no change in particle size before and only a moderate change above the  $C_1$  concentration. This evidenced the gradual process of particle aggregation. First nanogels start to associate moderately at C1 and then upon further increasing the NaCl the aggregates become larger and finally the aggregates become too large to stay in solution and precipitate out of solution.



Figure S20. Salt induced aggregation of PNVCL-Glc and PNVCL-PA at 50 °C

<b>Product</b>	$C_{\text{A}}(mM)$ of NaCl	$C_{\text{g}}(mM)$ of NaCl
PNVCL-PA nanogel		16
PNVCL-Glc	21	31
PNVCL-TEG-Glc	21	34
PNVCL-Mal	20	29
PNVCL-TEG-Mal	21	31

Table. S1 Aggregation and precipitation concentrations of NaCl, see Figure S16



Figure S21. Size distribution of PNVLC-PA at various NaCl concetrations at 50 °C