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Supporting Information

A macromolecule reversing antibiotic resistance phenotype and repurposing drugs as potent antibiotics

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Scheme S1. Chemical structures of polymers used in this study.



Figure S1. The polymer pEt_20 potentiates rifampicin and auranofin as potent antibiotics against drug-susceptible *A. baumannii* (BAA-1709). **a** MICs and **b** MBCs of rifampicin and auranofin with and without pEt_20 in comparison with colistin sulfate; **c** MIC fold change and **d** MBC fold change of rifampicin and auranofin in the presence of pEt_20 or colistin sulfate; **d** Killing efficiency of rifampicin and **e** killing efficiency of auranofin in the presence of pEt_20 in comparison with colistin sulfate. The polymer pEt_20 potentiated rifampicin and auranofin more effectively than colistin sulfate, leading to much greater MIC and MBC reduction (2048- *vs.* 32-fold reduction in MIC and 256- *vs.* 2-fold reduction in MBC for rifampicin; 128- *vs.* 8-fold reduction in MIC and 64- *vs.* 2-fold reduction in MBC for auranofin). The pEt_20 combinations showed a much stronger bactericidal effect (\geq 99.9% killing efficiency) than the colistin sulfate combinations. MIC and MBC data are representatives of three biological replicates, and killing efficiency is presented as Mean \pm S.D. (n=3).



Figure S2. The polymer pEt_20 enhances antimicrobial activity of azithromycin against *A*. *baumannii* in comparison with colistin sulfate. **a** MIC and MBC of azithromycin against drug-susceptible *A. baumannii* (BAA-1709) and **b** MIC and MBC of azithromycin against MDR *A. baumannii* (BAA-1789) with or without pEt_20 in comparison with colistin sulfate; **c** Killing efficiency of azithromycin against *A. baumannii* (BAA-1709) and **b** MIC and without pEt_20 in comparison with colistin sulfate; **c** Killing efficiency of azithromycin against *A. baumannii* (BAA-1709) and **d** Killing efficiency of azithromycin against *A. baumannii* (BAA-1789) with and without pEt_20 in comparison with colistin sulfate. The polymer pEt_20 reduced azithromycin MIC and MBC more effectively than colistin sulfate, and pEt_20/azithromycin combination showed a stronger bactericidal effect (\geq 99.9% killing efficiency) than colistin sulfate/azithromycin combination. MIC and MBC data are representatives of three biological replicates, and killing efficiency is presented as Mean ± S.D. (n=3).



Figure S3. Killing kinetics of pEt_20 ($\frac{1}{2} \times MIC$, i.e. 7.8 µg/mL), rifampicin (0.015 µg/mL), pEt_20/rifampicin combination, colistin sulfate ($\frac{1}{2} \times MIC$, i.e. 0.50 µg/mL), and colistin sulfate /rifampicin combination against *A. baumannii* BAA-1709 (~10⁵ CFU/mL). pEt_20/rifampicin combination killed the bacteria effectively within 1 h (3 log reduction in bacterial counts, i.e. >99.9% killing), while colistin sulfate /rifampicin combination did not show bactericidal activity at 18 h (~0% killing efficiency as compared to CFU at 0 h). Limit of detection: 50 CFU/mL. The data are representative of three biological replicates.



Figure S4. pEt_20 translocated bacterial membrane and bound with cytosolic DNA. **a** Confocal microscopic images of *A. baumannii* BAA-1709 after 30 min treatment with pEt_20 at $0.5 \times$ MIC (7.8 µg/mL) (Red regions: MM 4-64 stained membrane; Blue regions: Hoechst stained nucleic acids; Green regions: Alexa Fluor 488-labeled pEt_20); **b** Electrophoretic mobility of cytosolic DNA of *A. baumannii* BAA-1709 in pEt_20/DNA complexes at various weight ratios of polymer to DNA as specified. pEt_20 effectively bound DNA and completely inhibited the mobility of DNA at the polymer to DNA mass ratio of 3 and above.



Figure S5. Increase in intracellular ROS of *A. baumannii* BAA-1709 after treatment with pEt_20 or pEt_20/rifampicin combination. Confocal microscopic analyses [Red dye (MM 4-64): membrane staining; Green dye (CellRox Green): ROS probe] of the bacteria before and after 10-min treatment with rifampicin (0.50 μ g/mL), pEt_20 (7.8 μ g/mL) and their combination (pEt_20: 7.8 μ g/mL; rifampicin: 0.50 μ g/mL). (Scale bar: 10 μ m)



Figure S6. Increase in intracellular ROS of rifampicin-resistant *A. baumannii* BAA-1709 mutant. Fluorescence intensity analysis of intracellular ROS probe CellRox Green in rifampicin-resistant *A. baumannii* BAA-1709 mutant (~10⁷ CFU/mL) after treatment with rifampicin (0.50 μ g/mL), pEt_20 (7.8 μ g/mL) and their combination (pEt_20: 7.8 μ g/mL; rifampicin: 0.50 μ g/mL) over various periods of time. The results showed that the combination significantly enhanced intracellular ROS generation. pEt_20 translocated bacterial membrane followed by binding of cytosolic proteins or genes, facilitating ROS generation and thus killing the bacteria.



Figure S7. (for Figure 5f) The combination of pEt_20 and rifampicin significantly enhances intracellular ROS generation as indicated by flow cytometry results. **a-d** Gating strategy used for flow cytometry analysis of the intracellular probe CellRox Green-treated *A. baumannii* BAA-1709 cells. The P1 gating was to exclude the dead cells, which is located in the bottom left corner in FCS/SSC plots; The P2 gating was to choose the single cells in FSC-A/FSC-H plots; 10,000 cells were counted for each sample; **e** Individual flow cytometry histograms of CellRox Green-treated *A. baumannii* BAA-1709 cells after treatment with rifampicin (0.50 μ g/mL), pEt_20 (7.8 μ g/mL) and their combination (pEt_20: 7.8 μ g/mL; rifampicin: 0.50 μ g/mL)



Polymer-Rifampicin Combination



Figure S8. RNA-seq based comparative transcriptomics and functional annotation across different treatment conditions. **a** A venn diagram showing the number of differentially regulated genes upon the treatment of pEt_20, rifampicin or pEt_20/rifampicin combination, relative to untreated controls; Pink colored circle highlights the pEt_20/rifampicin combination treatment condition; **b** Heatmap depicts differentially expressed genes that are significant in "Polymer-Rifampicin" treatment condition in contrast to "Polymer only" and "Rifampicin only" treatments wherein the same genes from the latter might have not been

а

b

statistically significant. Gradient of green to red was used to depict intensity of down and up regulation in terms of log2 fold change of treatment conditions relative to control. Intermittent white lines in the heatmap denote genes that fell below coverage cut off for RNA-seq reads, therefore differential expression is undetermined. Colored bar panel on the left of heatmap refers to manually assigned putative functional categories (colored boxes and the legend text) of the corresponding genes in the heatmap.



Figure S9. Combination with pEt_20 does not increase cytotoxicity of rifampicin to mammalian cells. **a** Hemolysis level of rifampicin, pEt_20 and their combination at different concentrations; All treatments did not cause hemolysis up to 500 μ g/mL; **b** Viability of human embryonic kidney cells (HEK293T) after 24 h-treatment with rifampicin or pEt_20/rifampicin combination. pEt_20 did not affect cytotoxicity profile of rifampicin. At 31.3 μ g/mL or below, there was no cytotoxicity of rifampicin observed; **c** The confocal microscopic results indicated that the polymer (pEt_20, 7.8 μ g/mL) was localized in cytoplasma instead of nucleus. Treatment time: 1 h.



Figure S10. *In vivo* biocompatibility of pEt_20/rifampicin combination and antibacterial efficacy of pEt_20/rifampicin combination at different rifampicin doses. **a** Serum biochemistry analysis; Compared to PBS, rifampicin and pEt_20/rifampicin combination did not alter liver and kidney functions nor affect sodium ion and potassium ion concentrations (p > 0.05), indicating that rifampicin or the combination did not induce *in vivo* toxicity. Microscopic images of hematoxylin and eosin (H&E) - stained liver (**b**) and kidney (**c**) at 14 days post treatment with rifampicin (5.0 mg/kg) or the combination of rifampicin (5.0 mg/kg) and pEt_20 (2.0 mg/kg); The insignificant difference of tissue structures between pEt_20/rifampicin combination-treated mice and untreated or rifampicin-treated mice indicated that the combination did not induce hepatotoxicity and nephrotoxicity; **d** An increased rifampicin dose led to higher survival rate (n=10).

Table S1. The polymer pEt_20 sensitized MCR-1 positive colistin-resistant *E. coli* strains to colistimethate sodium treatment. Polymer concentration: $0.5 \times MIC$ (7.8 µg/mL). The polymer reversed colistin resistance phenotype, reducing its MIC from 15.6-31.3 µg/mL to 0.49 µg/mL. It also reduced MBC of colistimethate sodium by 8-fold from 15.6 and 31.3 µg/mL to 1.95 and 3.91 µg/mL, respectively. MIC and MBC data is representative of three biological replicates.

| Bacteria . | MIC of coli | stin (µg/mL) | MBC of colistin (µg/mL) | | |
|--------------------|-------------|---------------|-------------------------|---------------|--|
| | w/o polymer | With polymer* | w/o polymer | With polymer* | |
| <i>E. coli</i> - 1 | 31.3 | 0.49 | 31.3 | 3.9 | |
| <i>E. coli</i> - 2 | 15.6 | 0.49 | 15.6 | 1.95 | |

Table S2. The polymer pEt_20 repurposed rifampicin derivatives (rifaximin and rifabutin) as potent antibiotics against *A. baumannii* (BAA-1709). The polymer at $0.5 \times MIC$ (i.e. 7.8 µg/mL) reduced MIC of rifampicin derivatives (rifaximin and rifabutin) by 512-fold. MIC data is representative of three biological replicates.

| Antibiotic | MIC (µg/mL) | | | |
|------------|-------------|-------------------------------|--|--|
| | w/o polymer | With polymer (fold reduction) | | |
| Rifaximin | 3.91 | 0.0076 (512) | | |
| Rifabutin | 7.81 | 0.015 (512) | | |

Table S3. Effect of various compounds including cationic polycarbonate with quaternary ammonium (Qua_20), polyarginine with 10 amino acids (R_{10}), colistin sulfate, pEt_10, pEt_20 and pEt_40 on MIC of rifampicin against *A. baumannii* (BAA-1709). Use of guanidinium-functionalized polycarbonates (pEt_10, pEt_20 and pEt_40) potentiated rifampicin more effectively than the rest compounds, reducing rifampicin MIC by 2048 to 4096-fold.

| Compounds | Rifampicin MIC (fold reduction) in the presence of different compounds at 0.5×MIC (µg/mL) | | | |
|---------------------------|--|--|--|--|
| Qua_20 (MIC: 15.6 µg/mL) | 0.98 (4) | | | |
| R10 (MIC: 31.3 µg/mL) | 0.98 (4) | | | |
| Colistin (MIC: 1.0 µg/mL) | 0.244 (16) | | | |
| pEt_10 (MIC: 15.6 µg/mL) | 0.0019 (2048) | | | |
| pEt_20 (MIC: 15.6 µg/mL) | 0.0019 (2048) | | | |
| pEt_40 (MIC: 15.6 µg/mL) | 0.00095 (4096) | | | |

Table S4. RNA-seq on *A. baumannii* BAA-1709 cells treated with rifampin for 5 min, relative to untreated control cells. (In the word file, please double click the table below to see the full list).

| Locus-tag | Gene name | Product description | logFC | logCPM | PValue |
|-----------|-----------|---------------------------------|------------|----------|-------------|
| ABSDF3397 | NA | pseudo | 2.75038382 | 2.639199 | 5.54E-05 |
| ABSDF1359 | NA | IS982 family transposase | 1.49153608 | 4.452744 | 8.42E-05 |
| ABSDF3602 | NA | IS982 family transposase | 1.32775427 | 3.555293 | 0.016632479 |
| ABSDF3045 | NA | pseudo | 1.08966404 | 3.493362 | 0.001096222 |
| ABSDF0431 | rpmC | 50S ribosomal protein L29 | 1.01951977 | 11.84988 | 1.02E-16 |
| ABSDF0254 | NA | pseudo | 1.00522482 | 4.527912 | 0.001514906 |
| ABSDF3597 | hpd | 4-hydroxyphenylpyruvate dioxyge | -1.007257 | 10.66986 | 8.01E-28 |
| ABSDF1992 | NA | hypothetical protein | -1.0134616 | 7.82507 | 5.98E-30 |
| ABSDF2122 | NA | hypothetical protein | -1.0149208 | 7.965829 | 1.91E-28 |
| ABSDF3013 | NA | hydrolase | -1.0285445 | 9.276849 | 7.34E-26 |
| ABSDF0522 | NA | pseudo | -1.0382909 | 7.425346 | 1.97E-30 |
| ABSDF3489 | NA | hypothetical protein | -1.0396111 | 10.95793 | 4.61E-27 |
| ABSDF2566 | NA | hypothetical protein | -1.0400809 | 8.222832 | 1.36E-24 |
| ABSDF1698 | NA | signal peptide | -1.0616396 | 3.469103 | 3.43E-07 |
| ABSDF1830 | NA | hypothetical protein | -1.0628058 | 5.313994 | 8.45E-13 |
| ABSDF0317 | bfrB | bacterioferritin | -1.0873115 | 8.604489 | 1.39E-32 |
| ABSDF2565 | NA | hypothetical protein | -1.100315 | 10.79073 | 3.80E-35 |
| ABSDF2228 | NA | hypothetical protein | -1.1372366 | 7.201906 | 8.09E-33 |
| ABSDF2429 | NA | pseudo | -1.1417021 | 2.668859 | 0.00030075 |
| ABSDF2655 | otsB | trehalose-6-phosphate phophatas | -1.148711 | 4.582244 | 4.56E-08 |
| ABSDF3026 | NA | hypothetical protein | -1.1621485 | 5.919265 | 1.55E-22 |
| ABSDF1910 | NA | methyltransferase | -1.1764917 | 9.557042 | 9.10E-16 |
| ABSDF2192 | NA | IS5 family transposase | -1.1984725 | 2.945334 | 1.75E-07 |
| ABSDF1165 | NA | IS982 family transposase | -1.2000702 | 1.257168 | 6.25E-05 |
| ABSDF2834 | NA | pilus assembly protein FilB | -1.2092649 | 7.225845 | 1.32E-22 |
| ABSDF2046 | aro | phospho-2-dehydro-3-deoxyhept | -1.2138201 | 8.438065 | 4.98E-36 |

Table S5. RNA-seq on *A. baumannii* BAA-1709 cells treated with pEt_20 for 5 min, relative to untreated control cells. (In the word file, please double click the table below to see the full list).

| Locus-tag | Gene name | Product description | logFC | logCPM | PValue |
|-----------|-----------|---------------------------------|----------|------------|-------------|
| ABSDF3350 | NA | signal peptide | 3.24886 | 7.82486274 | 1.53E-193 |
| ABSDF1359 | NA | IS982 family transposase | 3.061063 | 5.97541646 | 1.92E-07 |
| ABSDF2215 | NA | hypothetical protein | 2.555296 | 4.97783899 | 4.95E-55 |
| ABSDF3326 | NA | thymidylate synthase | 2.550949 | 6.87162395 | 3.29E-145 |
| ABSDF3325 | NA | hypothetical protein | 2.451616 | 3.22231838 | 3.17E-21 |
| ABSDF1200 | NA | hypothetical protein | 2.446091 | 8.75424176 | 3.35E-213 |
| ABSDF2041 | fadH | pseudo | 2.358722 | 4.82445168 | 1.92E-55 |
| ABSDF0533 | NA | hypothetical protein | 2.353601 | 3.95914602 | 1.32E-38 |
| ABSDF2313 | NA | hypothetical protein | 2.34164 | 5.32799649 | 1.18E-73 |
| ABSDF3324 | NA | hypothetical protein | 2.296721 | 6.82141566 | 4.40E-91 |
| ABSDF2042 | fadH | pseudo | 2.280224 | 6.05410291 | 5.05E-95 |
| ABSDF3602 | NA | IS982 family transposase | 2.272546 | 4.53020397 | 0.000131123 |
| ABSDF3329 | NA | hypothetical protein | 2.263146 | 2.17115015 | 2.38E-19 |
| ABSDF3353 | NA | hypothetical protein | 2.142744 | 3.56947447 | 1.83E-26 |
| ABSDF1207 | NA | membrane protein branched-cl | 2.116231 | 3.40345873 | 3.12E-24 |
| ABSDF1206 | NA | pseudo | 2.104189 | 2.46675289 | 7.93E-21 |
| ABSDF0637 | NA | pseudo | 2.09452 | 2.78157203 | 6.01E-20 |
| ABSDF2040 | fadH | pseudo | 2.089076 | 4.57773832 | 1.95E-32 |
| ABSDF3330 | NA | ATP-binding | 2.086455 | 4.44557645 | 5.55E-40 |
| ABSDF1467 | NA | IS982 family transposase | 2.083449 | 3.92440752 | 0.002317 |
| ABSDF0727 | NA | SAM-dependent methyltransfera | 2.074423 | 2.66099594 | 1.21E-19 |
| ABSDF3327 | NA | hypothetical protein | 2.016346 | 5.92970748 | 3.13E-69 |
| ABSDF0049 | NA | oxidoreductase | 2.001574 | 6.84986521 | 1.06E-93 |
| ABSDF0862 | NA | pseudo | 1.955488 | 3.18702908 | 3.02E-19 |
| ABSDF0075 | NA | polysaccharide biosynthesis pro | 1.943847 | 3.13174817 | 1.00E-18 |
| ABSDF1269 | NA | hypothetical protein | 1.937373 | 7.01237651 | 5.31E-107 |
| ABSDF2043 | fadH | pseudo | 1.935158 | 6.19161655 | 6.69E-71 |

Table S6. RNA-seq on *A. baumannii* BAA-1709 cells treated with pEt_20/rifampicin combination for 5 min, relative to untreated control cells. (In the word file, please double click the table below to see the full list).

| Locus-tag | Gene name | Product description | logFC | logCPM | PValue |
|-----------|-----------|---------------------------------------|----------|------------|-----------|
| ABSDF007 | NA | polysaccharide biosynthesis protein | 2.786488 | 3.63568484 | 2.66E-27 |
| ABSDF335 | NA | signal peptide | 1.975524 | 6.57871484 | 8.11E-64 |
| ABSDF135 | NA | IS982 family transposase | 1.779427 | 4.72558742 | 0.0207515 |
| ABSDF186 | NA | acyl-CoA dehydrogenase | 1.692394 | 5.49847857 | 3.65E-47 |
| ABSDF090 | pal | peptidoglycan-associated lipoprotein | 1.687457 | 9.36354221 | 4.31E-92 |
| ABSDF193 | NA | hypothetical protein | 1.686021 | 4.48551059 | 1.99E-27 |
| ABSDF077 | sucD | succinyl-CoA synthetase subunit alpha | 1.685146 | 12.0426669 | 6.03E-93 |
| ABSDF126 | NA | hypothetical protein | 1.625216 | 6.61439267 | 1.39E-75 |
| ABSDF193 | NA | hypothetical protein | 1.618615 | 7.48865192 | 5.37E-85 |
| ABSDF194 | NA | homoserine dehydrogenase HDH | 1.568337 | 3.8455438 | 1.27E-18 |
| ABSDF332 | NA | hypothetical protein | 1.537554 | 5.39889345 | 3.95E-38 |
| ABSDF039 | NA | toluene-tolerance protein Ttg2E | 1.526315 | 8.01998117 | 2.94E-84 |
| ABSDF077 | sucC | succinyl-CoA synthetase subunit beta | 1.490374 | 12.1985951 | 2.56E-84 |
| ABSDF204 | fadH | pseudo | 1.489227 | 3.9438923 | 1.81E-15 |
| ABSDF175 | NA | phage-like protein | 1.489131 | 2.46309211 | 1.78E-10 |
| ABSDF177 | NA | hypothetical protein | 1.472403 | 1.91186459 | 1.90E-09 |
| ABSDF036 | NA | signal peptide | 1.450386 | 5.83995619 | 3.73E-48 |
| ABSDF008 | galE | UDP-glucose 4-epimerase | 1.43173 | 8.57521825 | 2.58E-67 |
| ABSDF332 | NA | hypothetical protein | 1.397009 | 2.20525393 | 1.71E-08 |
| ABSDF077 | lpd | dihydrolipoamide dehydrogenase | 1.391102 | 12.3110094 | 1.54E-63 |
| ABSDF099 | NA | phage-like protein | 1.347177 | 2.04083423 | 4.94E-07 |
| ABSDF054 | NA | hypothetical protein | 1.340812 | 5.19923806 | 5.95E-27 |
| ABSDF112 | purB | adenylosuccinate lyase | 1.338083 | 9.35957961 | 8.30E-74 |
| ABSDF088 | lolD | outer membrane lipoproteins ABC trans | 1.326679 | 7.13810412 | 9.56E-76 |
| ABSDF053 | NA | hypothetical protein | 1.323273 | 2.98011811 | 6.44E-11 |
| ABSDF101 | NA | hypothetical protein | 1.317878 | 2.92379812 | 1.57E-07 |
| ABSDF203 | eno | phosphopyruvate hydratase | 1.315891 | 10.1433587 | 3.71E-44 |

Table S7. Differentially regulated genes upon the treatment with pEt_20/rifampicin combination but not with rifampicin (Rif) treatment or polymer treatment alone. (In the word file, please double click the table below to see the full list).

| Locus_tag | Protein name | pEt_20 | Rifvs.Co | pEt_20+Rif v | Putative functional annotation |
|------------|--------------------------|--------|----------|--------------|--------------------------------|
| ABSDF0011 | RND type efflux pump in | 0.243 | -0.8579 | -1.138104 | Antibiotic resistance |
| ABSDF0008 | RND type efflux pump in | -0.1 | -0.7188 | -1.297772 | Antibiotic resistance |
| ABSDF0010 | RND type efflux pump in | -0.06 | -0.704 | -1.3694 | Antibiotic resistance |
| ABSDE2476 | bacteriophage protein | 0.042 | 0 77761 | 1 0105386 | Bacteriophage associated |
| ABSDE2471 | bacteriophage protein | 0.332 | 0 17653 | 1 0023388 | Bacteriophage associated |
| ABSDE2473 | bacteriophage protein | 0.044 | 0 44359 | 1 0780/38 | Bacteriophage associated |
| | bacteriophage protein | 0.044 | 0.14406 | 1.0062025 | Pacteriophage associated |
| | | 0.221 | 0.14400 | 1.0903923 | Dacteriophage associated |
| A DODE4000 | | 0.004 | 0.25672 | 1.1152003 | Bacteriophage associated |
| ABSDF1029 | bacteriopnage protein p | 0.246 | 0.32149 | 1.1491374 | Bacteriophage associated |
| ABSDF2502 | bacteriophage regulatory | 0.706 | 0.30696 | 1.1470215 | Bacteriophage associated |
| ABSDF2608 | phage-like protein | -0.57 | NA | 1.2641299 | Bacteriophage associated |
| ABSDF1754 | phage-like protein | 0.316 | 0.50618 | 1.489131 | Bacteriophage associated |
| ABSDF1785 | phage-like protein | -0.2 | 0.26123 | 1.0473229 | Bacteriophage associated |
| ABSDF1004 | phage-like protein | 0.224 | 0.14779 | 1.2692441 | Bacteriophage associated |
| ABSDF0997 | phage-like protein | 0.069 | 0.52728 | 1.3471773 | Bacteriophage associated |