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Supporting Information

A macromolecule reversing antibiotic resistance phenotype and repurposing drugs as potent antibiotics

Xin Ding, Chuan Yang, Wilfried Moreira, Peiyan Yuan, Balamurugan Periaswamy, Paola Florez de Sessions, Huimin Zhao, Jeremy Tan, Ashlynn Lee, Kai Xun Ong, Nathaniel Park, Zhen Chang Liang, James L. Hedrick and Yi Yan Yang**

Scheme S1. Chemical structures of polymers used in this study.

Figure S1. The polymer pEt 20 potentiates rifampicin and auranofin as potent antibiotics against drug-susceptible *A. baumannii* (BAA-1709). **a** MICs and **b** MBCs of rifampicin and auranofin with and without pEt_20 in comparison with colistin sulfate; **c** MIC fold change and **d** MBC fold change of rifampicin and auranofin in the presence of pEt_20 or colistin sulfate; **d** Killing efficiency of rifampicin and **e** killing efficiency of auranofin in the presence of pEt_20 in comparison with colistin sulfate. The polymer pEt_20 potentiated rifampicin and auranofin more effectively than colistin sulfate, leading to much greater MIC and MBC reduction (2048- *vs.* 32-fold reduction in MIC and 256- *vs.* 2-fold reduction in MBC for rifampicin; 128- *vs.* 8-fold reduction in MIC and 64- *vs.* 2-fold reduction in MBC for auranofin). The pEt 20 combinations showed a much stronger bactericidal effect $(\geq 99.9\%)$ killing efficiency) than the colistin sulfate combinations. MIC and MBC data are representatives of three biological replicates, and killing efficiency is presented as Mean \pm S.D. (n=3).

Figure S2. The polymer pEt 20 enhances antimicrobial activity of azithromycin against *A*. *baumannii* in comparison with colistin sulfate. **a** MIC and MBC of azithromycin against drugsusceptible *A. baumannii* (BAA-1709) and **b** MIC and MBC of azithromycin against MDR *A. baumannii* (BAA-1789) with or without pEt_20 in comparison with colistin sulfate; **c** Killing efficiency of azithromycin against *A. baumannii* (BAA-1709) and **d** Killing efficiency of azithromycin against *A. baumannii* (BAA-1789) with and without pEt_20 in comparison with colistin sulfate. The polymer pEt_20 reduced azithromycin MIC and MBC more effectively than colistin sulfate, and pEt_20/azithromycin combination showed a stronger bactericidal effect (≥99.9% killing efficiency) than colistin sulfate/azithromycin combination. MIC and MBC data are representatives of three biological replicates, and killing efficiency is presented as Mean \pm S.D. (n=3).

Figure S3. Killing kinetics of pEt_20 ($\frac{1}{2} \times$ MIC, i.e. 7.8 µg/mL), rifampicin (0.015 µg/mL), pEt_20/rifampicin combination, colistin sulfate ($\frac{1}{2} \times$ MIC, i.e. 0.50 µg/mL), and colistin sulfate /rifampicin combination against *A. baumannii* BAA-1709 (~10⁵ CFU/mL). pEt_20/rifampicin combination killed the bacteria effectively within 1 h (3 log reduction in bacterial counts, i.e. >99.9% killing), while colistin sulfate /rifampicin combination did not show bactericidal activity at 18 h (~0% killing efficiency as compared to CFU at 0 h). Limit of detection: 50 CFU/mL. The data are representative of three biological replicates.

Figure S4. pEt_20 translocated bacterial membrane and bound with cytosolic DNA. **a** Confocal microscopic images of *A. baumannii* BAA-1709 after 30 min treatment with pEt_20 at 0.5×MIC (7.8 µg/mL) (Red regions: MM 4-64 stained membrane; Blue regions: Hoechst stained nucleic acids; Green regions: Alexa Fluor 488-labeled pEt_20); **b** Electrophoretic mobility of cytosolic DNA of *A. baumannii* BAA-1709 in pEt_20/DNA complexes at various weight ratios of polymer to DNA as specified. pEt 20 effectively bound DNA and completely inhibited the mobility of DNA at the polymer to DNA mass ratio of 3 and above.

Figure S5. Increase in intracellular ROS of *A. baumannii* BAA-1709 after treatment with pEt 20 or pEt 20/rifampicin combination. Confocal microscopic analyses [Red dye (MM 4-64): membrane staining; Green dye (CellRox Green): ROS probe] of the bacteria before and after 10-min treatment with rifampicin (0.50 μg/mL), pEt_20 (7.8 μg/mL) and their combination (pEt 20: 7.8 μg/mL; rifampicin: 0.50 μg/mL). (Scale bar: 10 μm)

Figure S6. Increase in intracellular ROS of rifampicin-resistant *A. baumannii* BAA-1709 mutant. Fluorescence intensity analysis of intracellular ROS probe CellRox Green in rifampicin-resistant *A. baumannii* BAA-1709 mutant (~10⁷ CFU/mL) after treatment with rifampicin (0.50 μg/mL), pEt 20 (7.8 μg/mL) and their combination (pEt 20: 7.8 μg/mL; rifampicin: 0.50 μg/mL) over various periods of time. The results showed that the combination significantly enhanced intracellular ROS generation. pEt_20 translocated bacterial membrane followed by binding of cytosolic proteins or genes, facilitating ROS generation and thus killing the bacteria.

Figure S7. (for Figure 5f) The combination of pEt_20 and rifampicin significantly enhances intracellular ROS generation as indicated by flow cytometry results. **a-d** Gating strategy used for flow cytometry analysis of the intracellular probe CellRox Green-treated *A. baumannii* BAA-1709 cells. The P1 gating was to exclude the dead cells, which is located in the bottom left corner in FCS/SSC plots; The P2 gating was to choose the single cells in FSC-A/FSC-H plots; 10,000 cells were counted for each sample; **e** Individual flow cytometry histograms of CellRox Green-treated *A. baumannii* BAA-1709 cells after treatment with rifampicin (0.50 μg/mL), pEt 20 (7.8 μg/mL) and their combination (pEt 20: 7.8 μg/mL; rifampicin: 0.50 μ g/mL) for 5 min.

Figure S8. RNA-seq based comparative transcriptomics and functional annotation across different treatment conditions. **a** A venn diagram showing the number of differentially regulated genes upon the treatment of pEt_20, rifampicin or pEt_20/rifampicin combination, relative to untreated controls; Pink colored circle highlights the pEt_20/rifampicin combination treatment condition; **b** Heatmap depicts differentially expressed genes that are significant in "Polymer-Rifampicin" treatment condition in contrast to "Polymer only" and "Rifampicin only" treatments wherein the same genes from the latter might have not been

a

b

statistically significant. Gradient of green to red was used to depict intensity of down and up regulation in terms of log2 fold change of treatment conditions relative to control. Intermittent white lines in the heatmap denote genes that fell below coverage cut off for RNA-seq reads, therefore differential expression is undetermined. Colored bar panel on the left of heatmap refers to manually assigned putative functional categories (colored boxes and the legend text) of the corresponding genes in the heatmap.

Figure S9. Combination with pEt_20 does not increase cytotoxicity of rifampicin to mammalian cells. **a** Hemolysis level of rifampicin, pEt_20 and their combination at different concentrations; All treatments did not cause hemolysis up to 500 µg/mL; **b** Viability of human embryonic kidney cells (HEK293T) after 24 h-treatment with rifampicin or pEt_20/rifampicin combination. pEt_20 did not affect cytotoxicity profile of rifampicin. At 31.3 µg/mL or below, there was no cytotoxicity of rifampicin observed; **c** The confocal microscopic results indicated that the polymer (pEt_20 , 7.8 µg/mL) was localized in cytoplasma instead of nucleus. Treatment time: 1 h.

Figure S10. *In vivo* biocompatibility of pEt 20/rifampicin combination and antibacterial efficacy of pEt_20/rifampicin combination at different rifampicin doses. **a** Serum biochemistry analysis; Compared to PBS, rifampicin and pEt_20/rifampicin combination did not alter liver and kidney functions nor affect sodium ion and potassium ion concentrations (p > 0.05), indicating that rifampicin or the combination did not induce *in vivo* toxicity. Microscopic images of hematoxylin and eosin (H&E) - stained liver (**b**) and kidney (**c**) at 14 days post treatment with rifampicin (5.0 mg/kg) or the combination of rifampicin (5.0 mg/kg) and pEt_20 (2.0 mg/kg); The insignificant difference of tissue structures between pEt_20/rifampicin combination-treated mice and untreated or rifampicin-treated mice indicated that the combination did not induce hepatotoxicity and nephrotoxicity; **d** An increased rifampicin dose led to higher survival rate $(n=10)$.

Table S1. The polymer pEt 20 sensitized MCR-1 positive colistin-resistant *E. coli* strains to colistimethate sodium treatment. Polymer concentration: 0.5×MIC (7.8 µg/mL). The polymer reversed colistin resistance phenotype, reducing its MIC from 15.6-31.3 μg/mL to 0.49 μg/mL. It also reduced MBC of colistimethate sodium by 8-fold from 15.6 and 31.3 μg/mL to 1.95 and 3.91 µg/mL, respectively. MIC and MBC data is representative of three biological replicates.

Table S2. The polymer pEt_20 repurposed rifampicin derivatives (rifaximin and rifabutin) as potent antibiotics against \overline{A} . *baumannii* (BAA-1709). The polymer at 0.5×MIC (i.e. 7.8 µg/mL) reduced MIC of rifampicin derivatives (rifaximin and rifabutin) by 512-fold. MIC data is representative of three biological replicates.

Table S3. Effect of various compounds including cationic polycarbonate with quaternary ammonium (Qua 20), polyarginine with 10 amino acids (R_{10}) , colistin sulfate, pEt 10, pEt_20 and pEt_40 on MIC of rifampicin against *A. baumannii* (BAA*-*1709). Use of guanidinium-functionalized polycarbonates (pEt_10, pEt_20 and pEt_40) potentiated rifampicin more effectively than the rest compounds, reducing rifampicin MIC by 2048 to 4096-fold.

Table S4. RNA-seq on *A. baumannii* BAA-1709 cells treated with rifampin for 5 min, relative to untreated control cells. (In the word file, please double click the table below to see the full list).

Table S5. RNA-seq on *A. baumannii* BAA-1709 cells treated with pEt_20 for 5 min, relative to untreated control cells. (In the word file, please double click the table below to see the full list).

Table S6. RNA-seq on *A. baumannii* BAA-1709 cells treated with pEt_20/rifampicin combination for 5 min, relative to untreated control cells. (In the word file, please double click the table below to see the full list).

Table S7. Differentially regulated genes upon the treatment with pEt_20/rifampicin combination but not with rifampicin (Rif) treatment or polymer treatment alone. (In the word file, please double click the table below to see the full list).

Locus_tag	Protein name				pEt_20 Rif vs. CopEt_20+Rif v Putative functional annotation
ABSDF0011	RND type efflux pump in 0.243		-0.8579	-1.138104	Antibiotic resistance
ABSDF0008	RND type efflux pump in -0.1		-0.7188	-1.297772	Antibiotic resistance
ABSDF0010	RND type efflux pump in -0.06		-0.704	-1.3694	Antibiotic resistance
ABSDF2476	bacteriophage protein	0.042	0.77761	1.0105386	Bacteriophage associated
ABSDF2471	bacteriophage protein	0.332	0.17653	1.0023388	Bacteriophage associated
ABSDF2473	bacteriophage protein	0.044	0.44359	1.0789438	Bacteriophage associated
ABSDF2497	bacteriophage protein	0.221	0.14406	1.0963925	Bacteriophage associated
ABSDF2469	bacteriophage protein	0.004	0.25672	1.1152003	Bacteriophage associated
ABSDF1029	bacteriophage protein p 0.246		0.32149	1.1491374	Bacteriophage associated
ABSDF2502	bacteriophage regulator 0.706		0.30696	1.1470215	Bacteriophage associated
ABSDF2608	phage-like protein	-0.57	NA	1.2641299	Bacteriophage associated
ABSDF1754	phage-like protein	0.316	0.50618	1.489131	Bacteriophage associated
ABSDF1785	phage-like protein	-0.2	0.26123	1.0473229	Bacteriophage associated
ABSDF1004	phage-like protein	0.224	0.14779	1.2692441	Bacteriophage associated
ABSDF0997	phage-like protein	0.069	0.52728	1.3471773	Bacteriophage associated