

## SUPPORTING INFORMATION

# Synthesis, In Silico and In Vitro Evaluation for Acetylcholinesterase and BACE-1 Inhibitory Activity of Some *N*-Substituted-4-Phenothiazine-Chalcones

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**Table S1.** Results of re-docking (RMSD in Å)

**Table S2:** Docking results and ligand interaction (Co-crystallized 1DX6)

**Table S3.** Docking results and ligand interaction (Co-crystallized 5HU1 chain A and B)

**Table S4.** Dataset of 72 compounds used in the building of 2D-QSAR model for AChE inhibitors

**Table S5.** Dataset of 215 compounds used in the building of 2D-QSAR model for BACE-1 inhibitors

**Table S6.** Equations for calculation of 2D-QSAR validation metrics

**Table S7.** Selected descriptors used for building 2D-QSAR models

**Table S8.** Values of selected descriptors used in prediction of pIC<sub>50</sub> of the synthesized chalcone derivatives (AChE)

**Table S9.** Values of selected descriptors used in prediction of pIC<sub>50</sub> of the synthesized chalcone derivatives (BACE-1)

**Table S10.** Spectra of synthesized chalcone derivatives

**Figure S1.** Interactions of co-crystallized ligand in the protein complex 1DX6 (2D)

**Figure S2.** Interactions of co-crystallized ligand in the protein complex 1DX6 (3D)

**Figure S3.** Alignment of re-dock ligands with the native one in the binding pocket of of 1DX6

**Figure S4.** Interactions of co-crystallized ligand in the protein complex 5HU1-chain A (2D)

**Figure S5.** Interactions of co-crystallized ligand in the protein complex 5HU1-chain A (3D)

**Figure S6.** Alignment of re-dock ligands with the native one in the binding pocket of of 5HU1-chain A

**Figure S7.** Interactions of co-crystallized ligand in the protein complex 5HU1-chain B (2D)

**Figure S8.** Interactions of co-crystallized ligand in the protein complex 5HU1-chain B (3D)

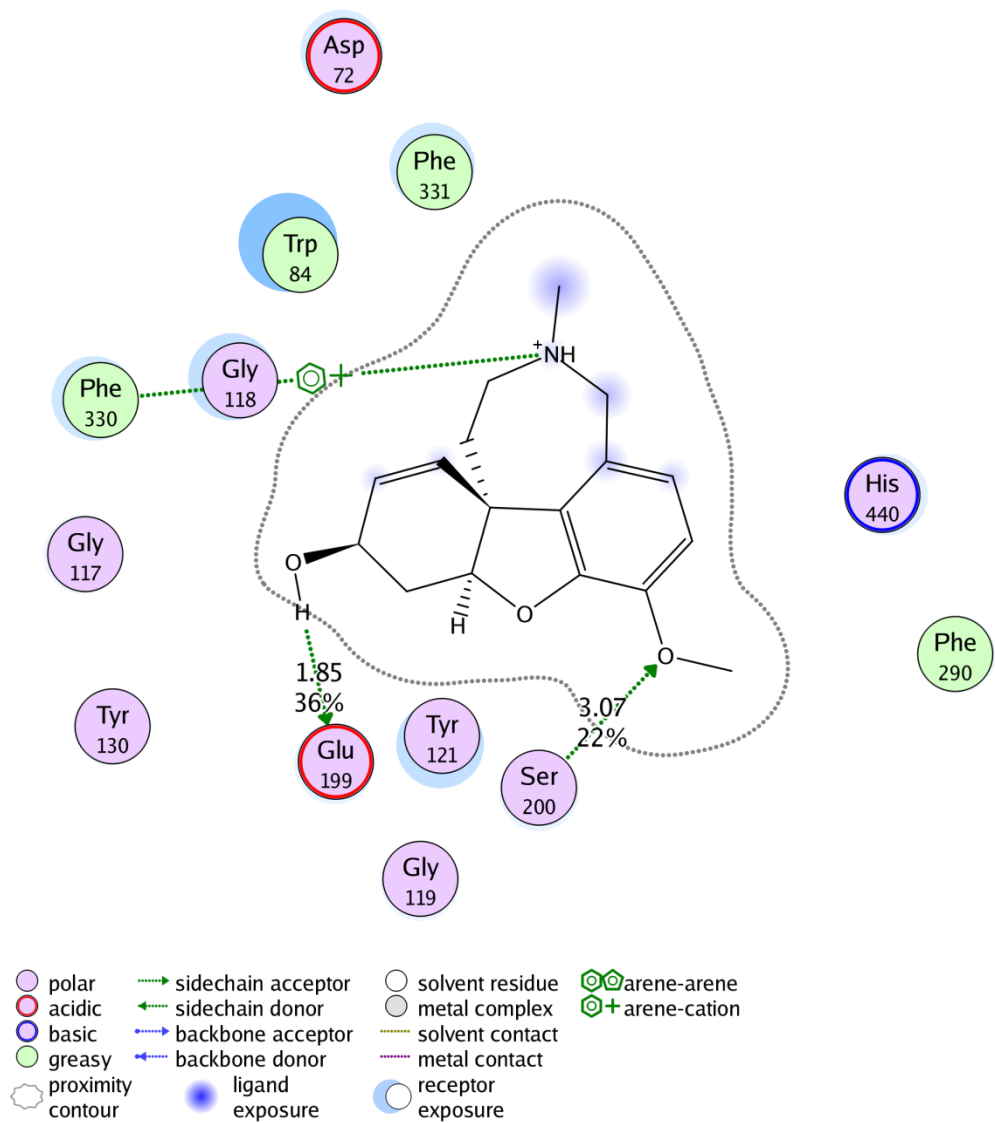
**Figure S9.** Alignment of re-dock ligands with the native one in the binding pocket of of 5HU1-chain B

**Figure S10.** The linear regression between docking score and  $pIC_{50}$  on AChE of synthesized chalcone derivatives (A. observed values from **AC1-3** and **AC5-10**, B. predicted values from **AC1, AC4, AC6, AC8, AC10, AC13**)

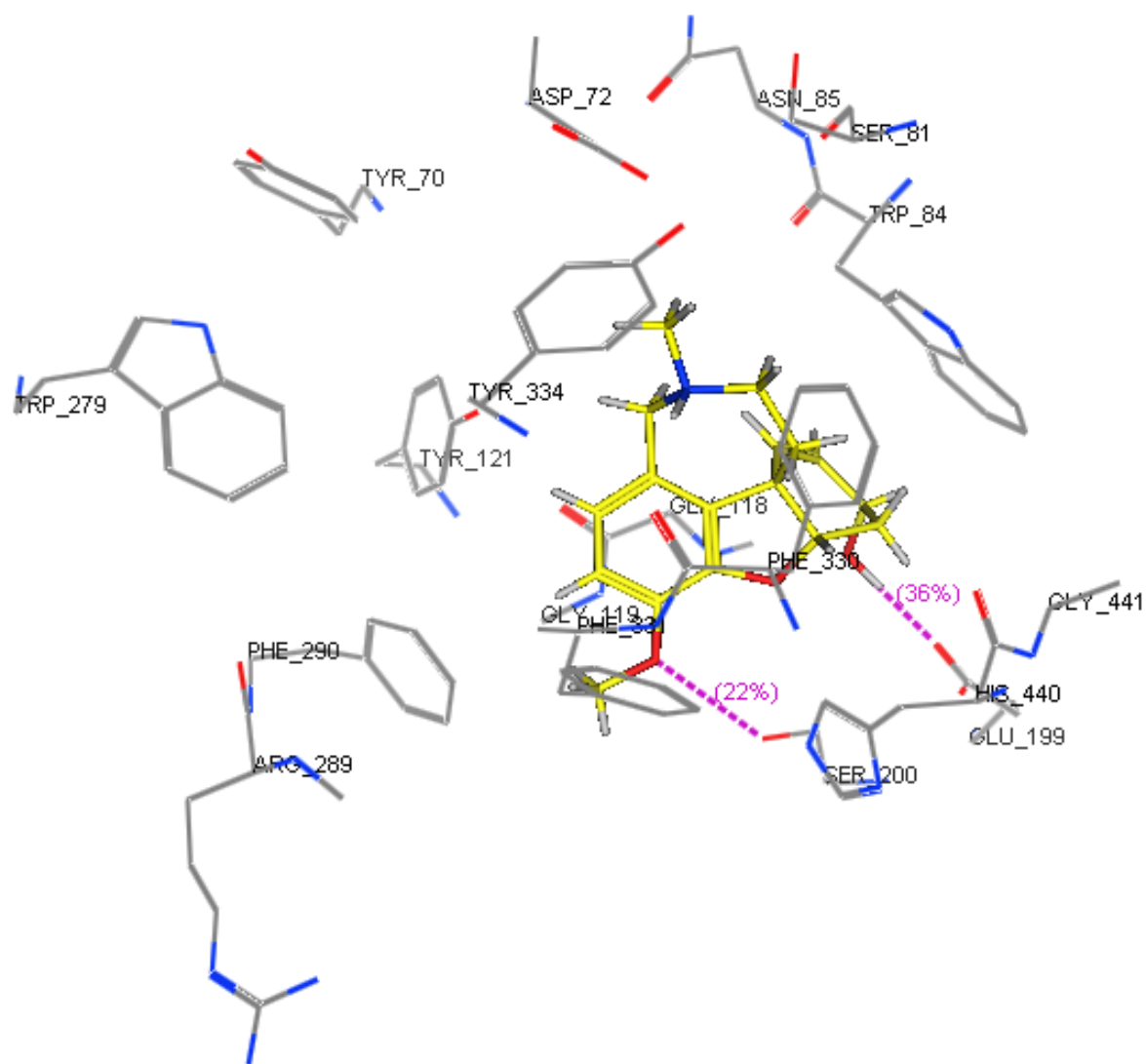
**Table S1.** Results of re-docking (RMSD in Å)

Ligand	AChE	BACE-1	
	1DX6	5HU1 (chain A)	5HU1 (chain B)
1	0.49	0.71	0.53
2	0.50	0.52	0.87
3	0.89	0.89	0.96

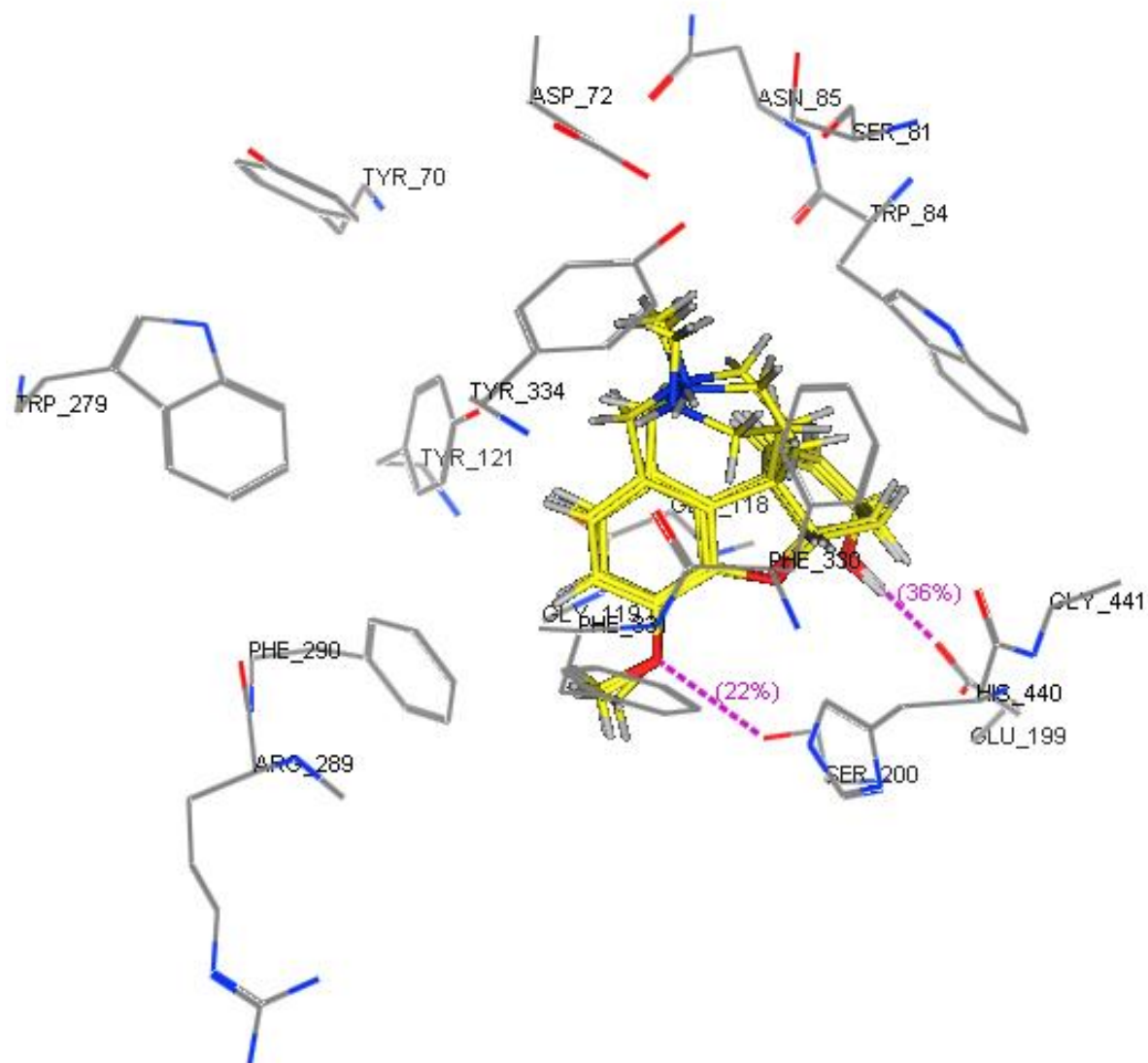
- *Ligand 1: separated from the complex (native form, not prepared).*
- *Ligand 2: separated from the complex and re-prepared using appropriate procedure indicated in the section of Materials and Methods*
- *Ligand 3: built and prepared from the beginning.*



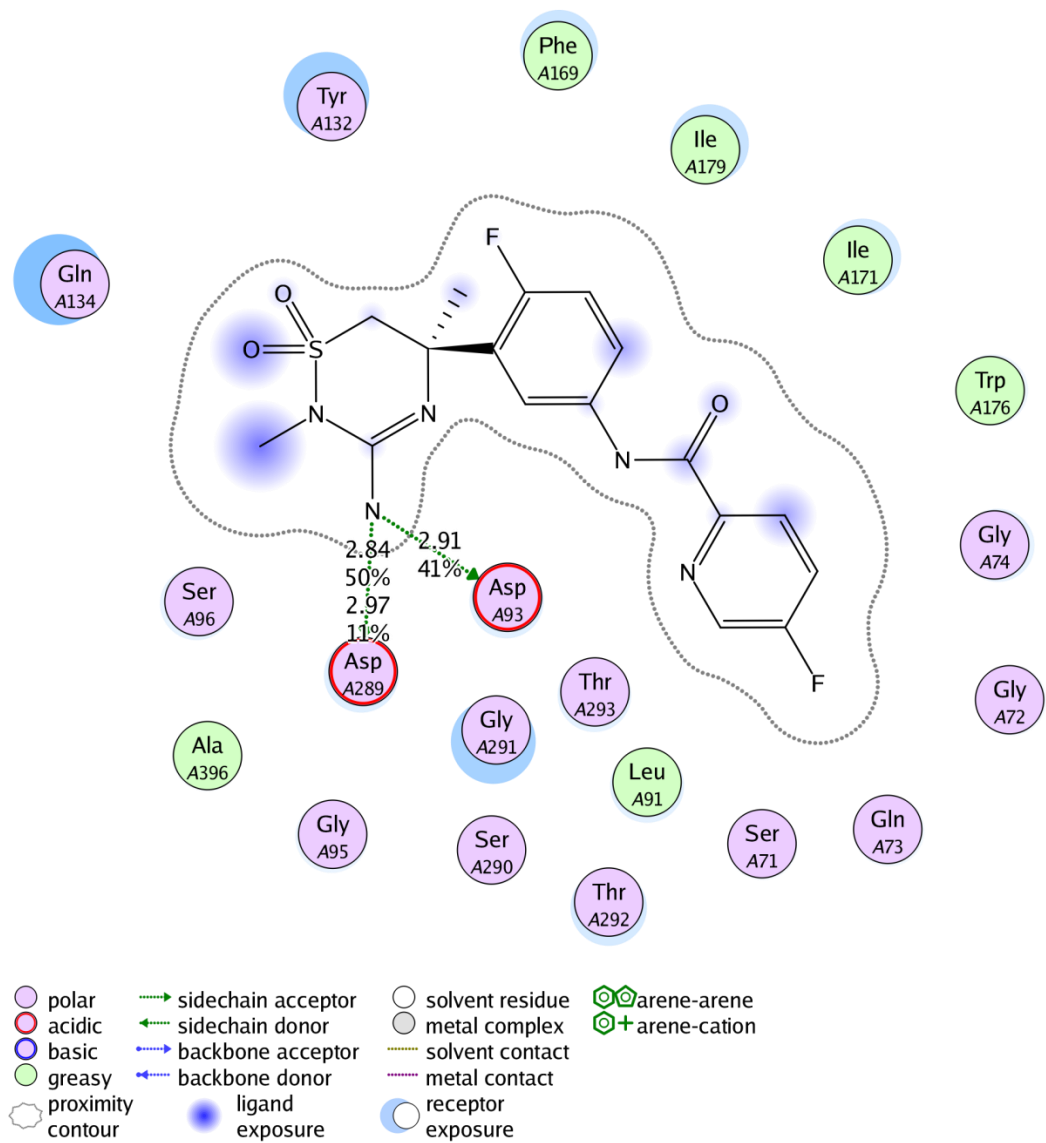
**Figure S1.** Interactions of co-crystallized ligand in the protein complex 1DX6 (2D)



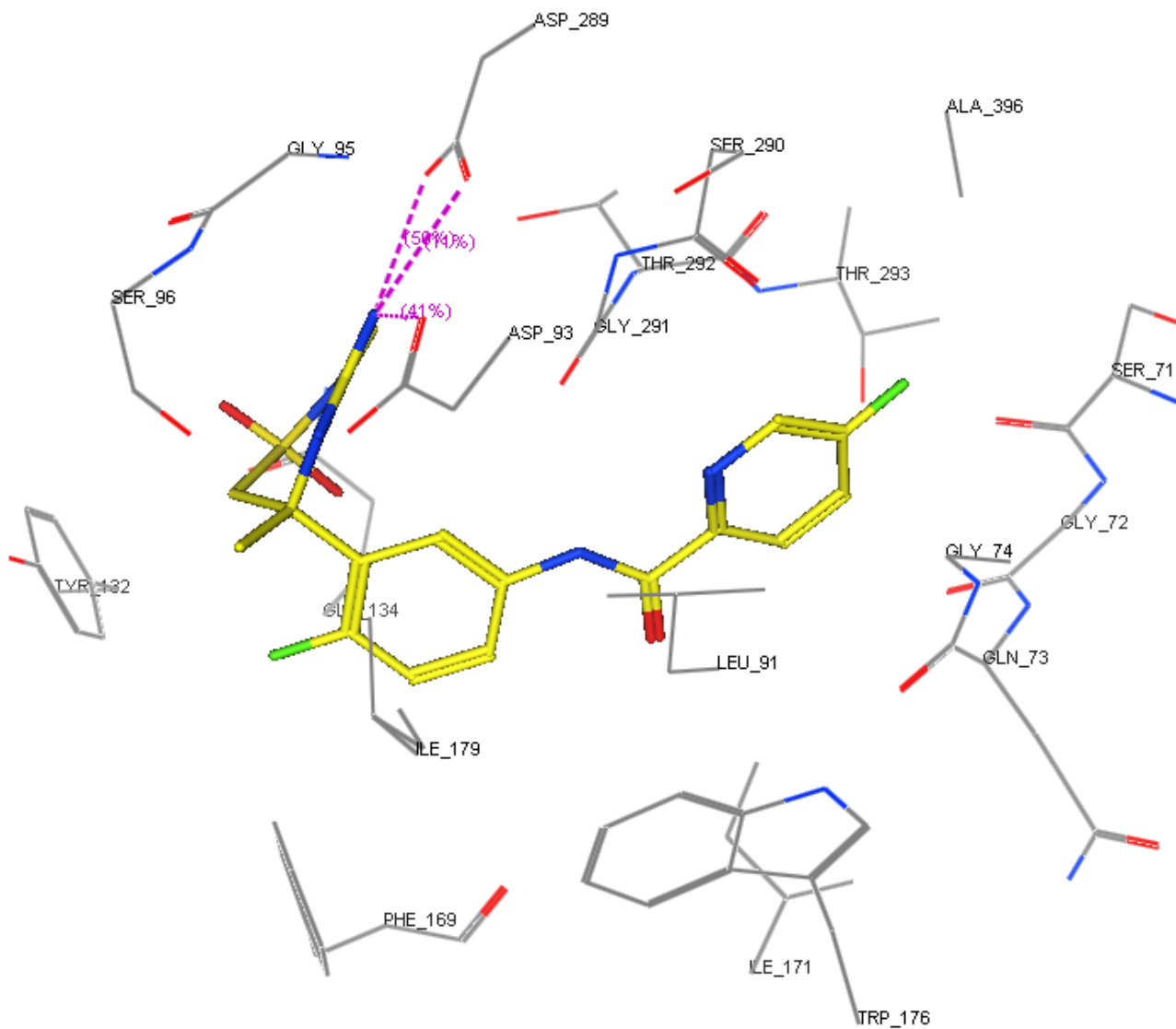
**Figure S2.** Interactions of co-crystallized ligand in the protein complex 1DX6 (3D)



**Figure S3.** Alignment of re-dock ligands with the native one in the binding pocket of 1DX6

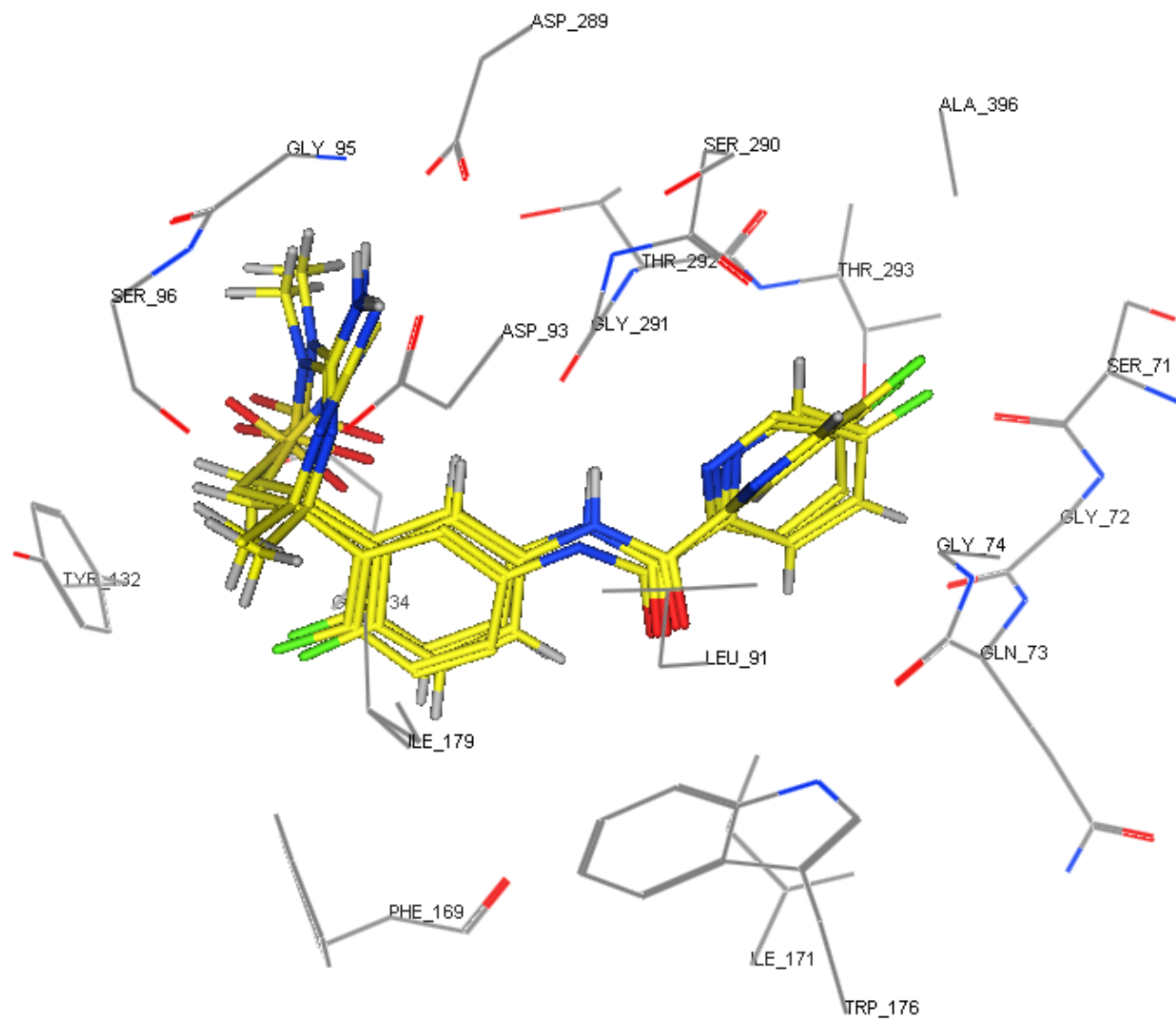


**Figure S4.** Interactions of co-crystallized ligand in the protein complex 5HU1-chain A (2D)

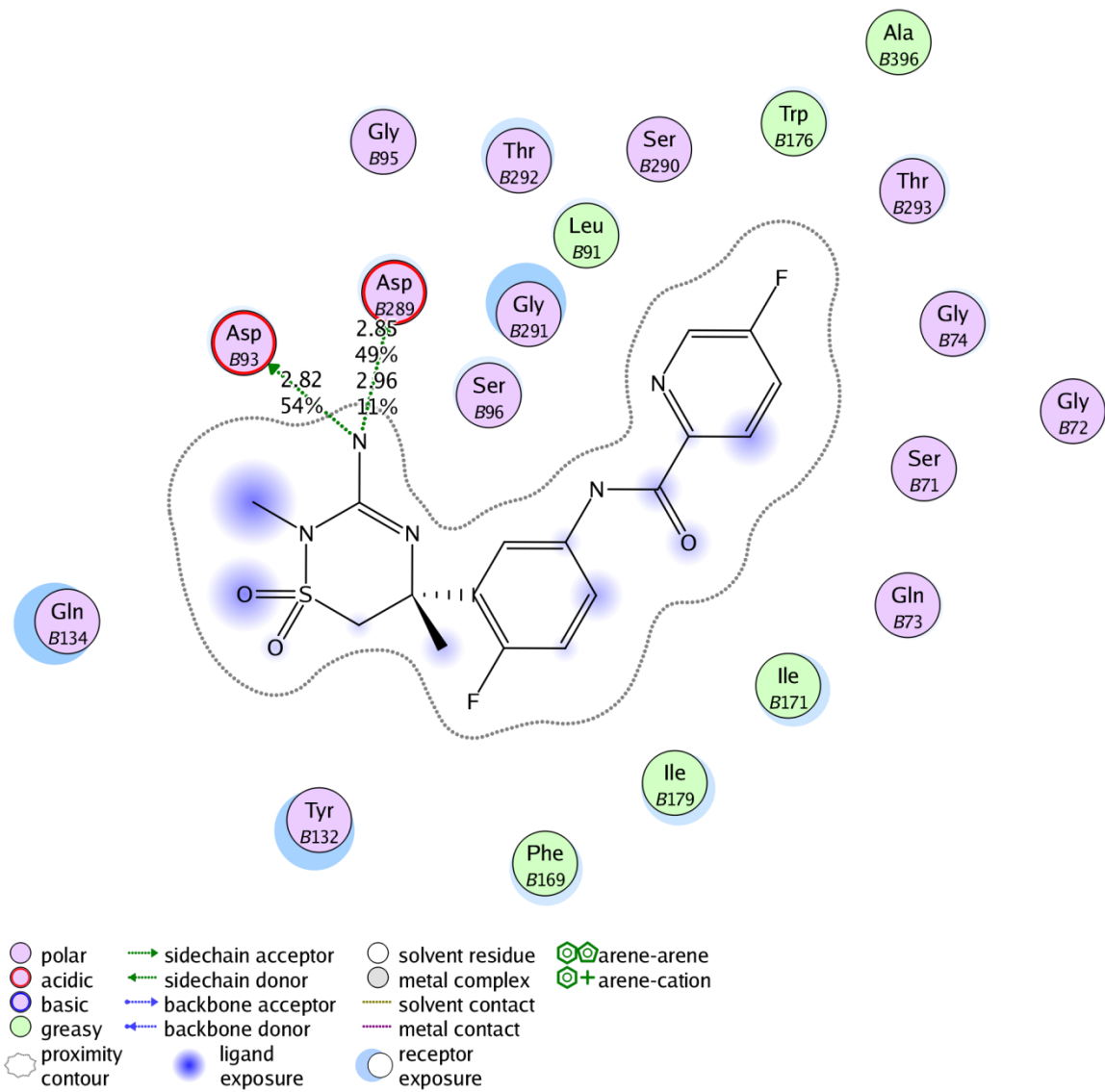


**Figure S5.** Interactions of co-crystallized ligand in the protein complex 5HU1-chain A (3D)

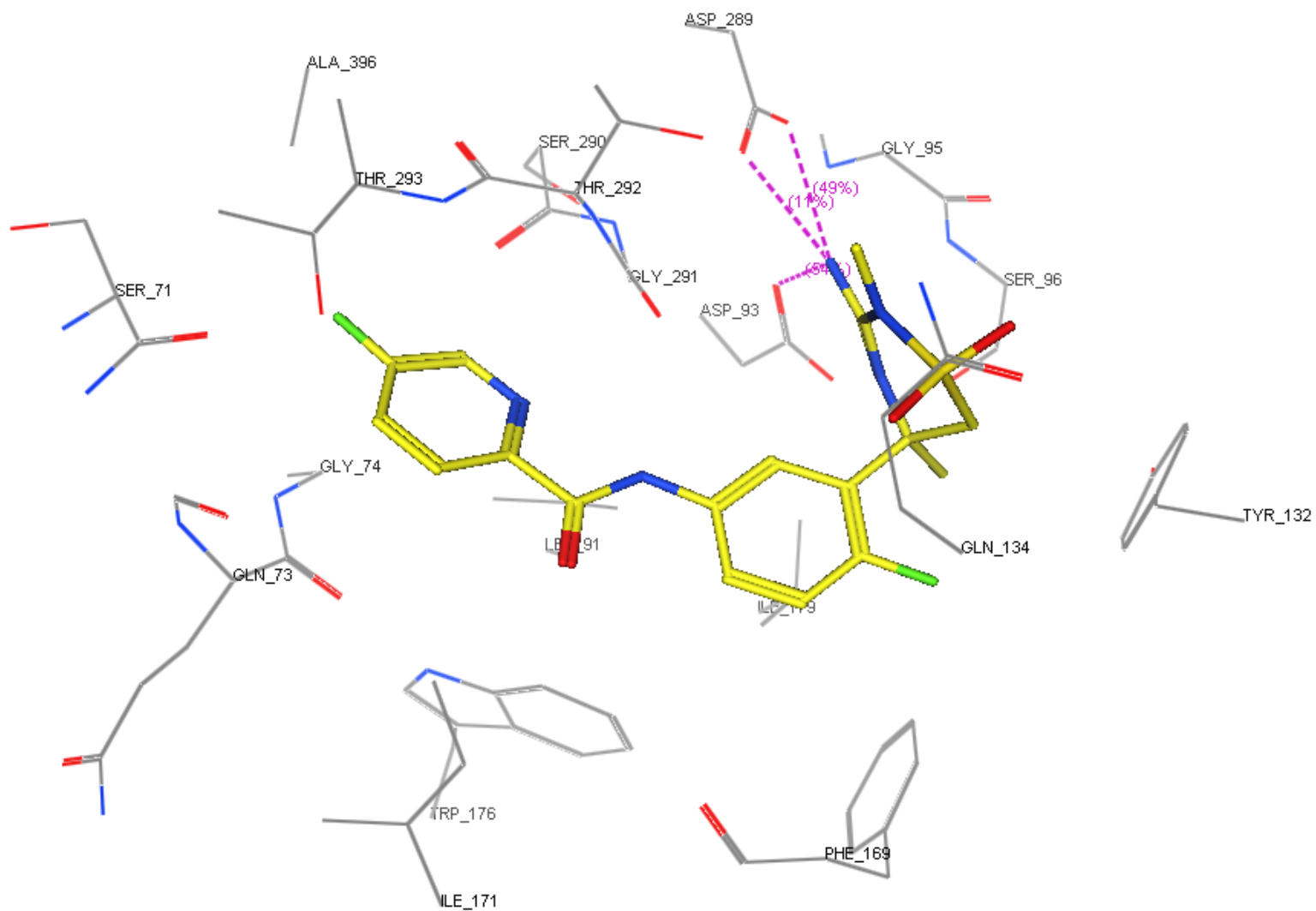




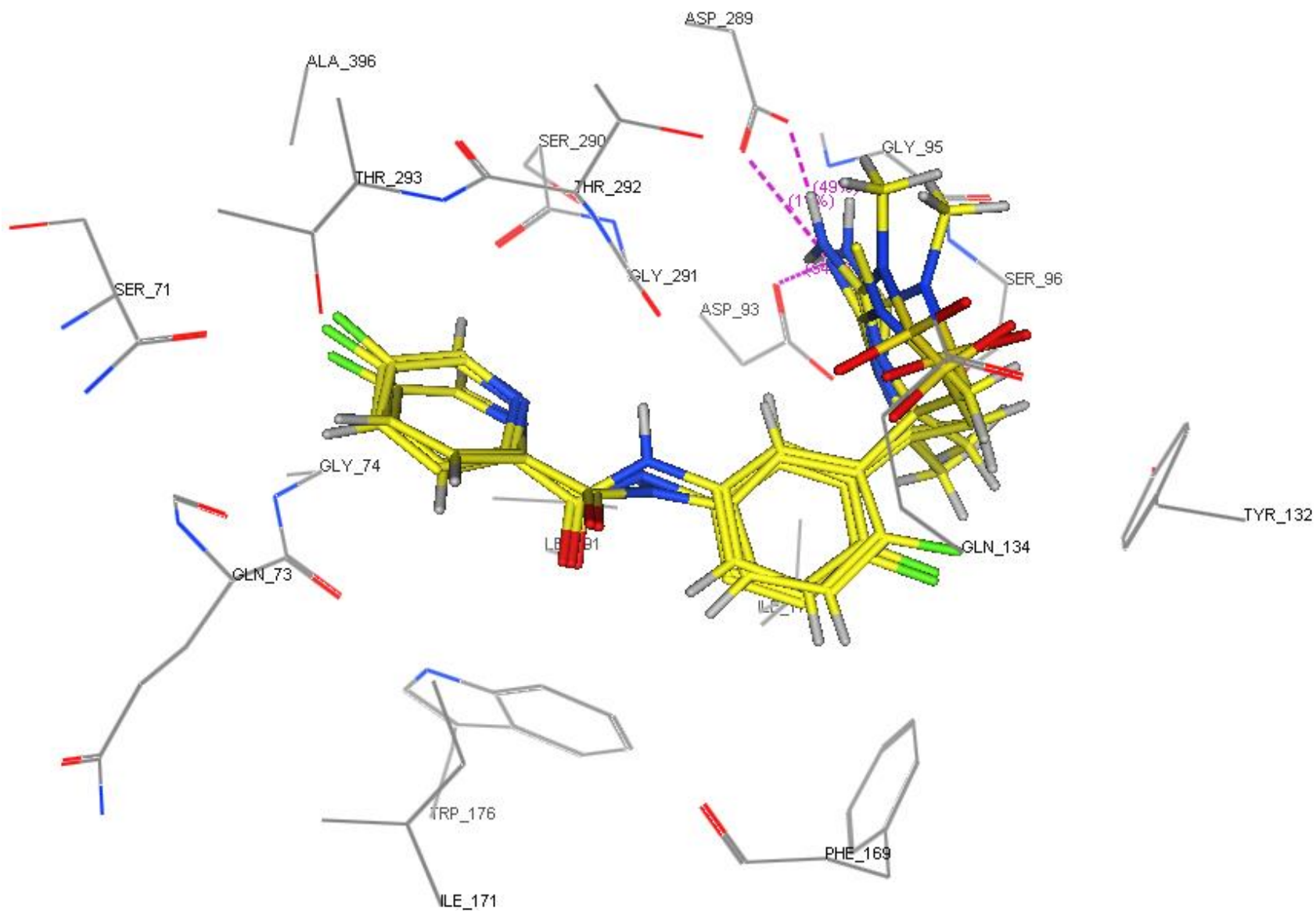
**Figure S6.** Alignment of re-dock ligands with the native one in the binding pocket of of 5HU1-chain A



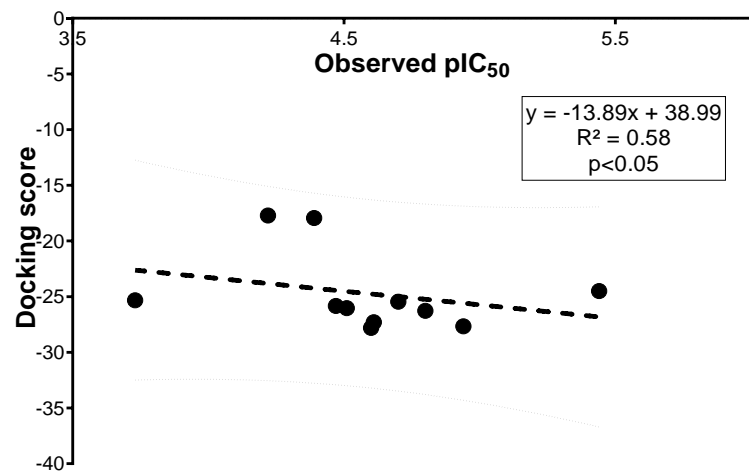
**Figure S7.** Interactions of co-crystallized ligand in the protein complex 5HU1-chain B (2D)



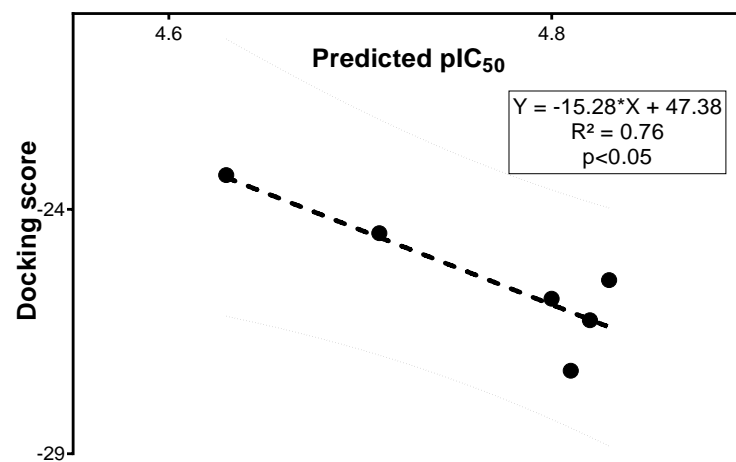
**Figure S8.** Interactions of co-crystallized ligand in the protein complex 5HU1-chain B (3D)



**Figure S9.** Alignment of re-dock ligands with the native one in the binding pocket of of 5HU1-chain B



(A)

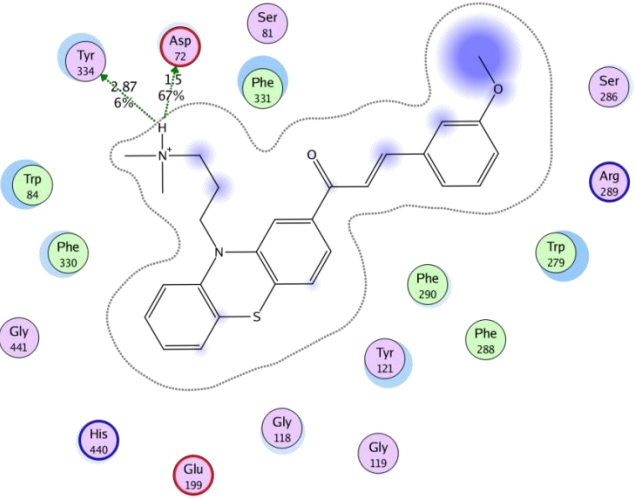
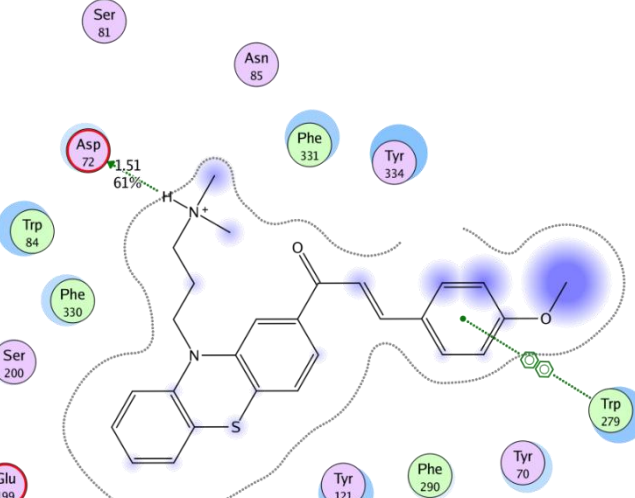


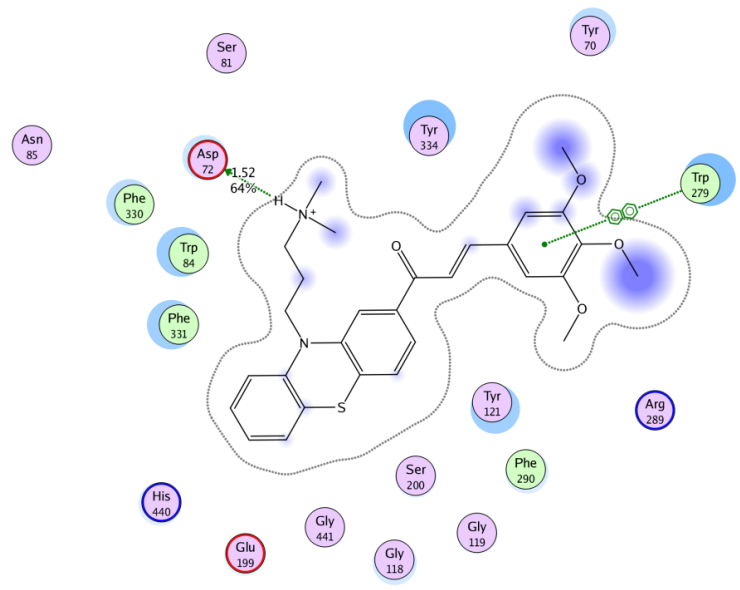
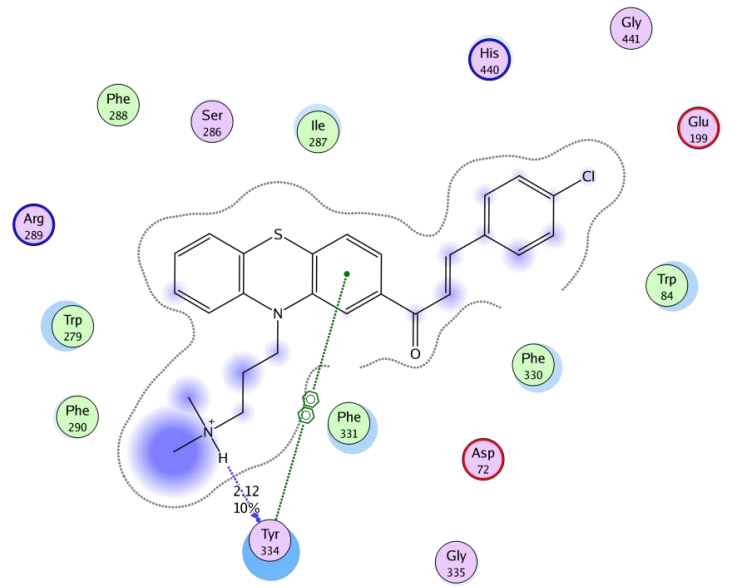
(B)

**Figure S10.** The linear regression between docking score and pIC<sub>50</sub> on AChE of synthesized chalcone derivatives (A. observed values from AC1-3 and AC5-10, B. predicted values from AC1, AC4, AC6, AC8, AC10, AC13)

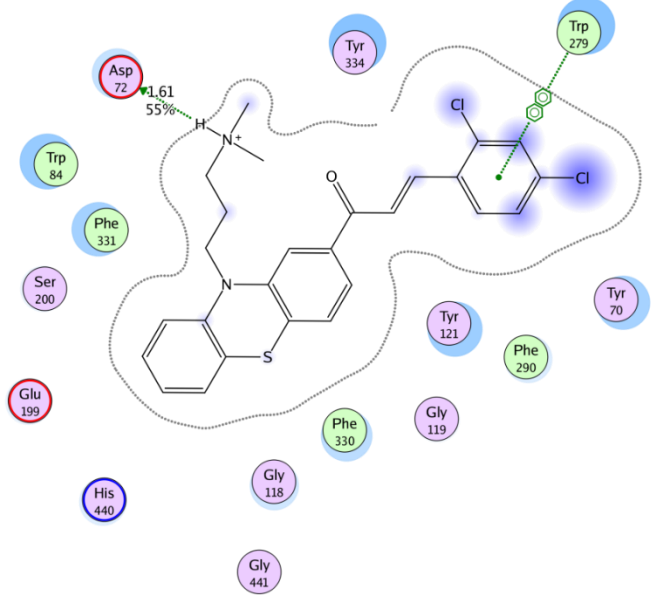
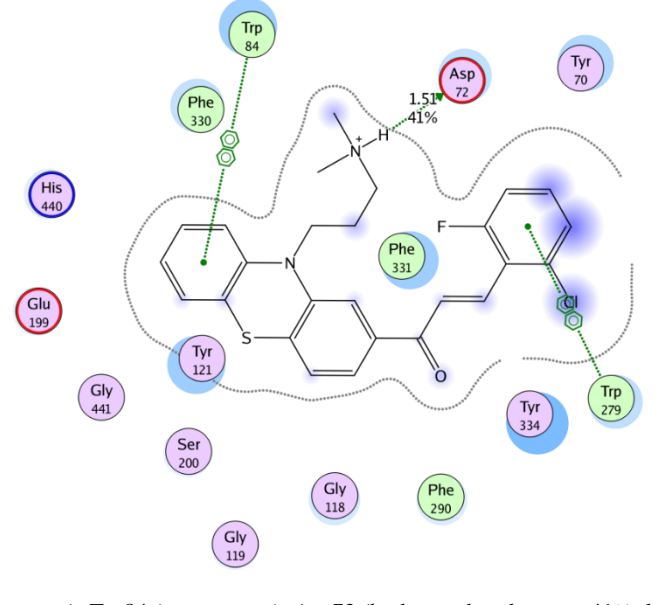
**Table S2: Docking results and ligand interaction (Co-crystallized 1DX6)**

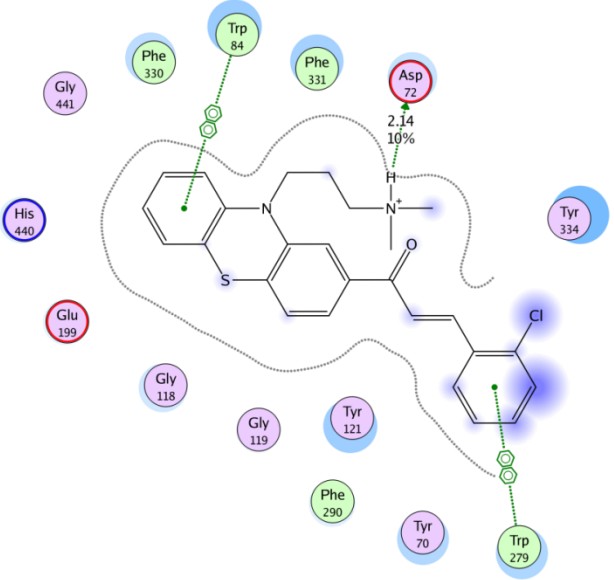
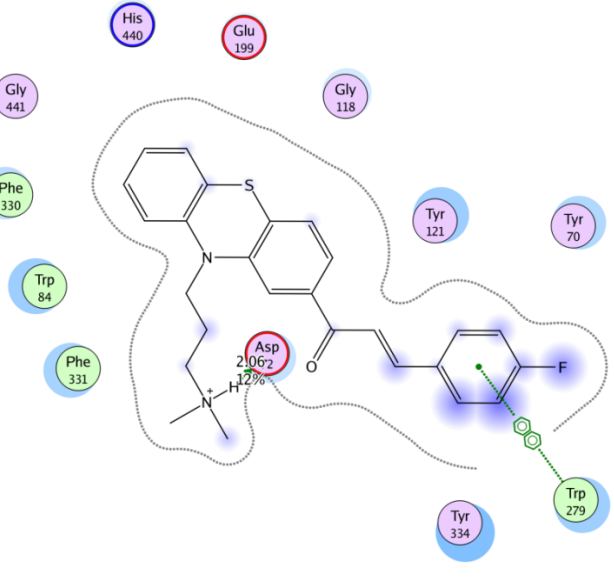
Compound	pIC <sub>50</sub>		Docking score (kJ.mol <sup>-1</sup> )	Ligand interaction
	Observed	Predicted		
AC1	4.47	4.80	-25.83	<p>Trp84 (arene-arene), Asp72 (hydrogen bond, score: 9%, length: 2.14 Å )</p>

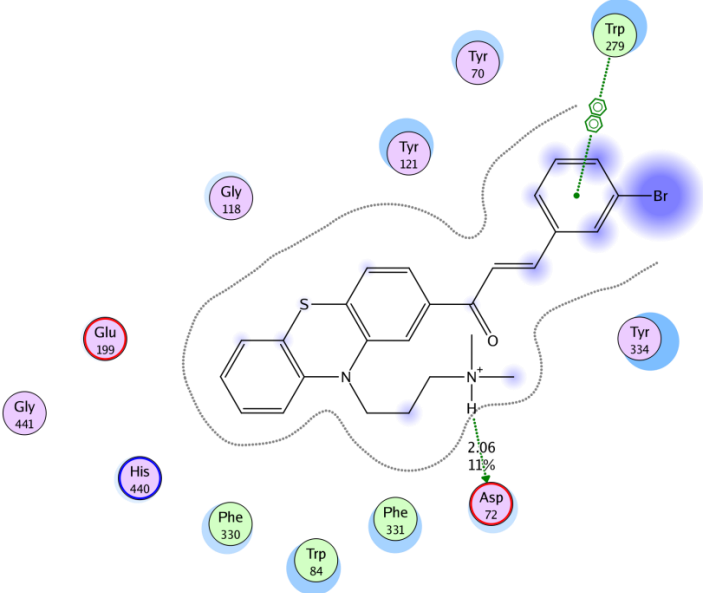
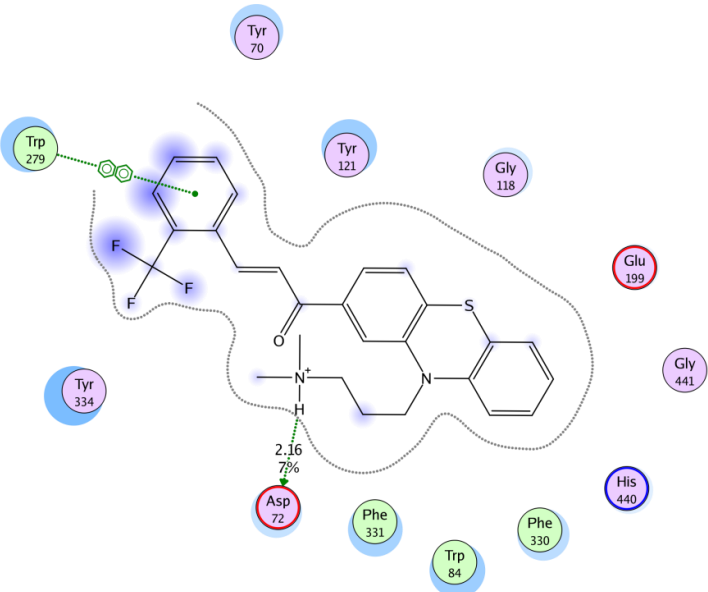
AC2	4.51	4.74	-26.03	 <p>Tyr334 (hydrogen bond, score: 6%, length: 2.87 Å ), Asp72 (hydrogen bond, score: 67%, length: 1.5 Å )</p>
AC3	4.94	4.74	-27.67	 <p>Trp279 (arene-arene), Asp72 (hydrogen bond, score: 61%, length: 1.51 Å )</p>

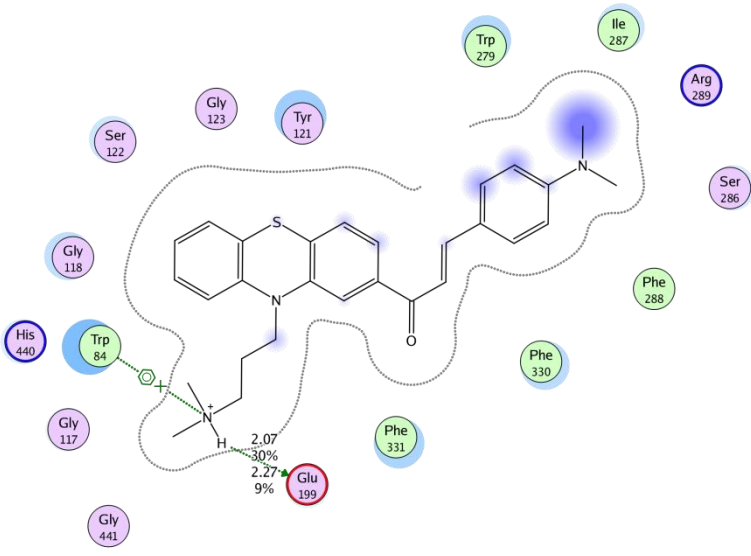
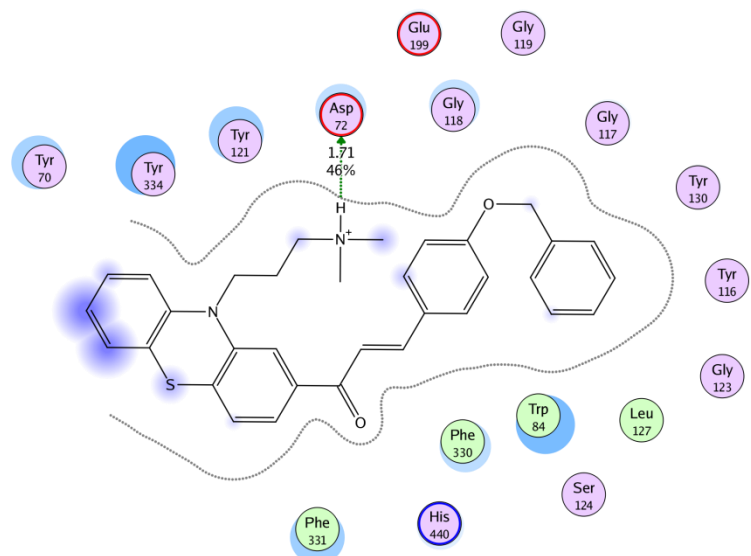
AC4	5.44	4.71	-24.49	 <p>Trp279 (arene-arene), Asp72 (hydrogen bond, score: 64%, length: 1.52 Å)</p>
AC5	4.22	4.82	-17.71	 <p>Tyr334 (arene-arene; hydrogen bond, score 10%, length: 2.12 Å)</p>



AC6	4.70	4.83	-25.45	 <p>Trp279 (arene-arene), Asp72 (hydrogen bond, score: 55%, length: 1.61 Å )</p>
AC7	4.39	4.76	-17.94	 <p>Trp279 (arene-arene), Trp84 (arene-arene), Asp72 (hydrogen bond, score: 41%, length: 1.51 Å )</p>

AC8	4.80	4.82	-26.27	 <p>Trp279 (arene-arene), Trp84 (arene-arene), Asp72 (hydrogen bond, score: 10%, length: 2.14 Å )</p>
AC9	4.60	4.76	-27.80	 <p>Trp279 (arene-arene), Asp72 (hydrogen bond, score: 12%, length: 2.06 Å )</p>

AC10	4.61	4.81	-27.30	 <p>Trp279 (arene-arene), Asp72 (hydrogen bond, score: 11%, length: 2.06 Å )</p>
AC11	3.73	4.59	-25.33	 <p>Trp279 (arene-arene), Asp72 (hydrogen bond, score: 7%, length: 2.16 Å )</p>

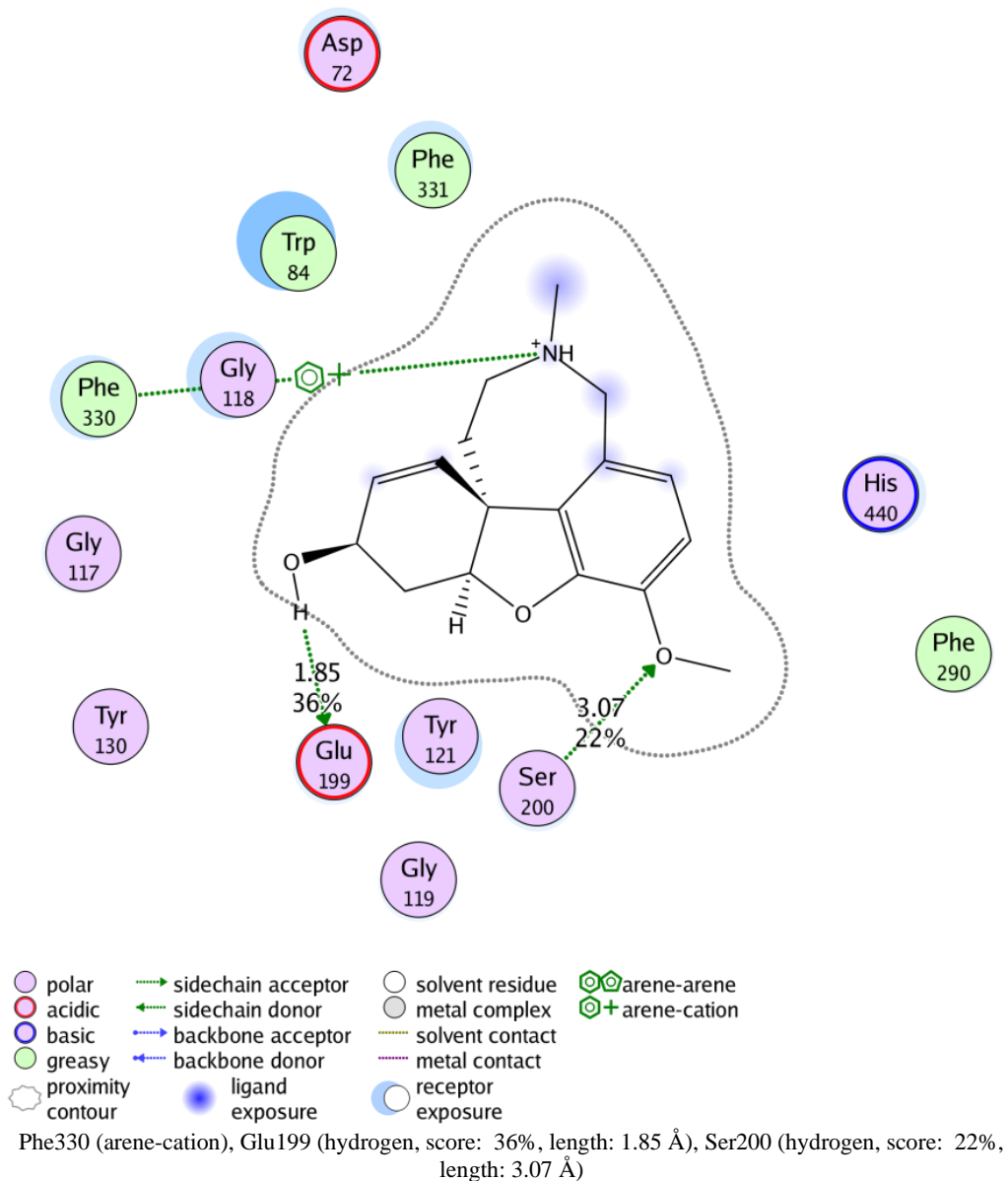
AC12	5.96	4.81	-22.15	 <p>Trp84 (arene-cation), Glu199 (hydrogens, score: 9%; 30%; length: 2.27; 2.07 Å)</p>
AC13	4.93	4.63	-23.30	 <p>Asp72 (hydrogen bond, score: 46%, length: 1.71 Å)</p>

Galanthamine

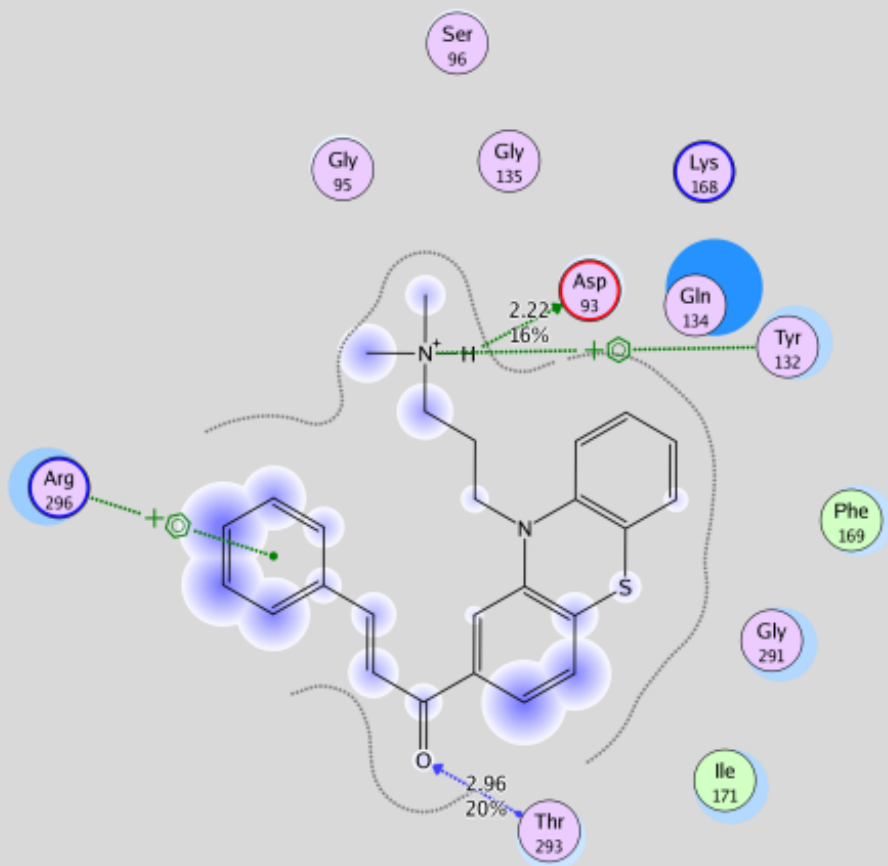
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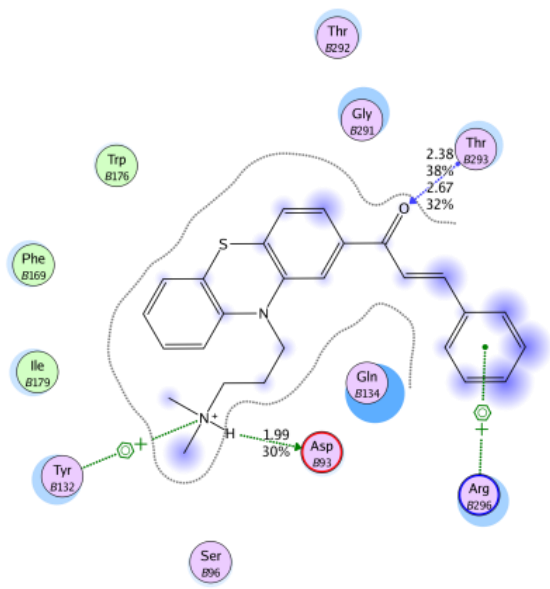
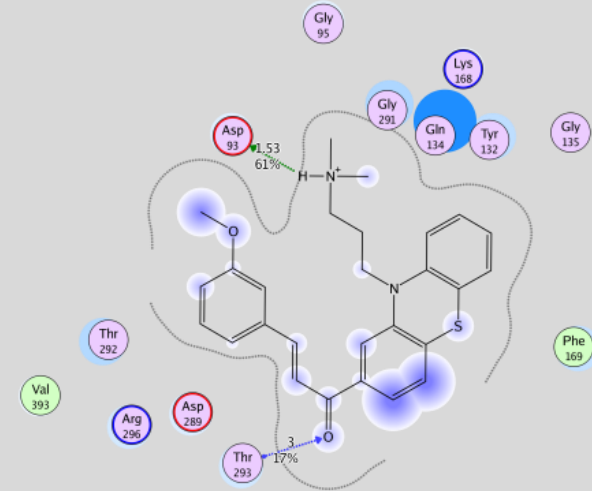
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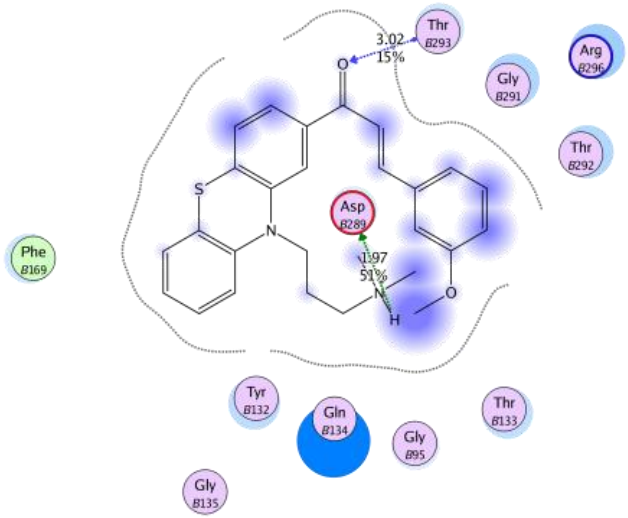
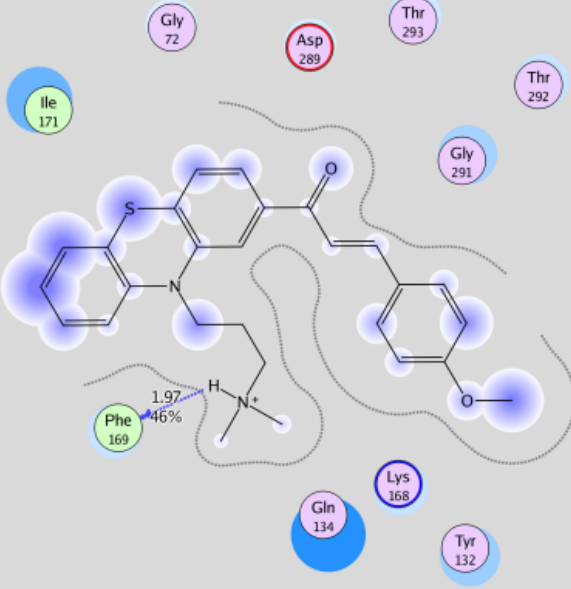
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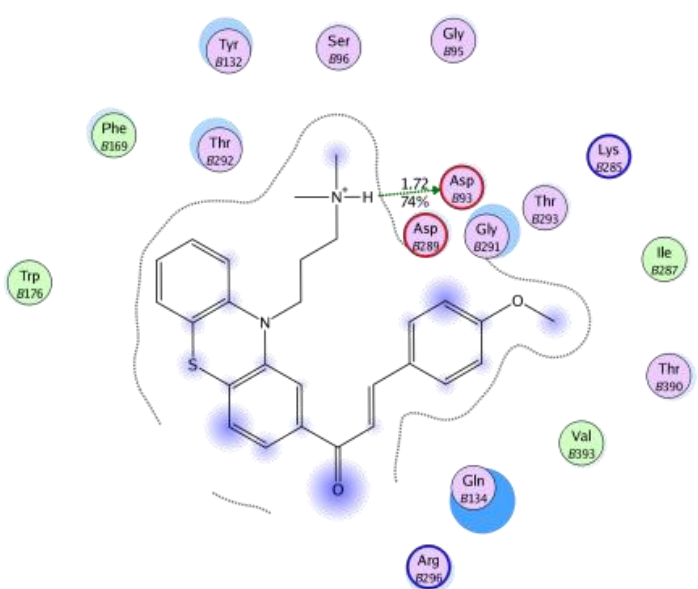
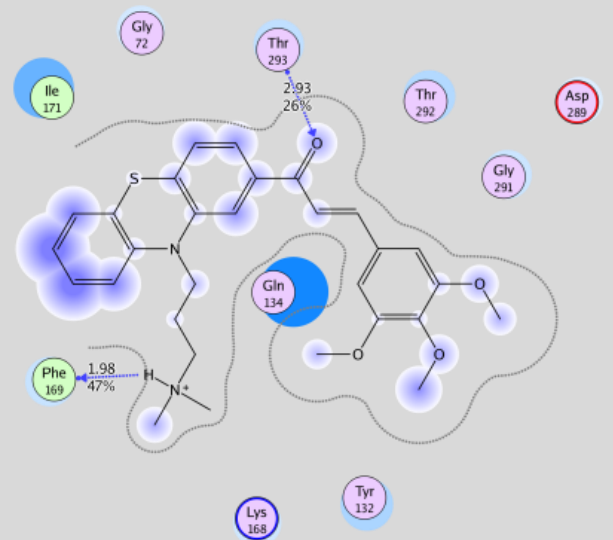
**Table S3.** Docking results and ligand interaction (Co-crystallized 5HU1 chain A and B)

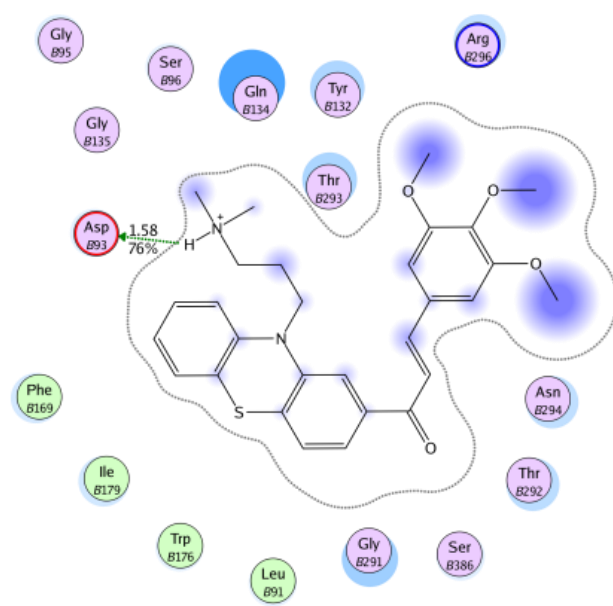
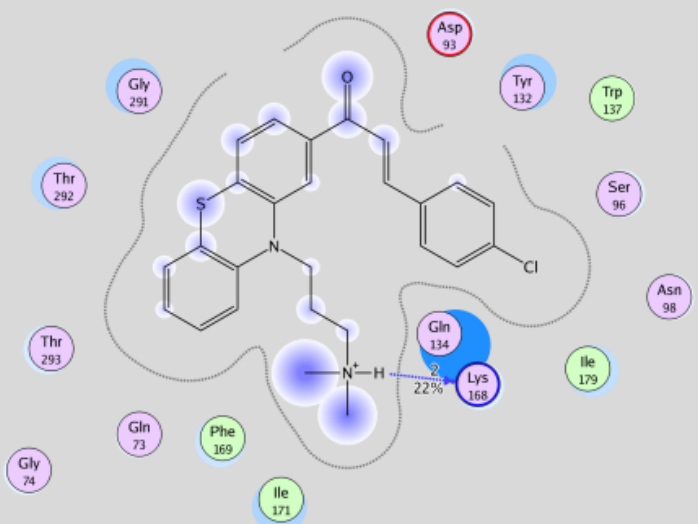
Compound	pIC <sub>50</sub>		Chain	Docking score (kJ.mol <sup>-1</sup> )	Interactions
	Observed	Predicted			
AC1	5.20	6.52	A	-17.82	 <p>TyrA132 (arene-cation), ArgA296 (arene-cation), AspA93 (hydrogen bond, score: 16%, length 2.22 Å), ThrA293 (hydrogen bond, score: 20%, length 2.96 Å)</p>

			B	-17.69	 <p>TyrB132 (arene-cation, ArgB296 (arene-cation), AspB93 (hydrogen bond, score: 30%, length 1.99 Å), ThrB293 (hydrogen bonds, score: 32%; 38%, length: 2.67 Å; 2.38 Å)</p>
AC2	5.52	7.38	A	-17.22	 <p>AspA93 (hydrogen bond, score: 61%, length 1.53 Å), ThrA293 (hydrogen bond, score: 17%, length 3 Å)</p>

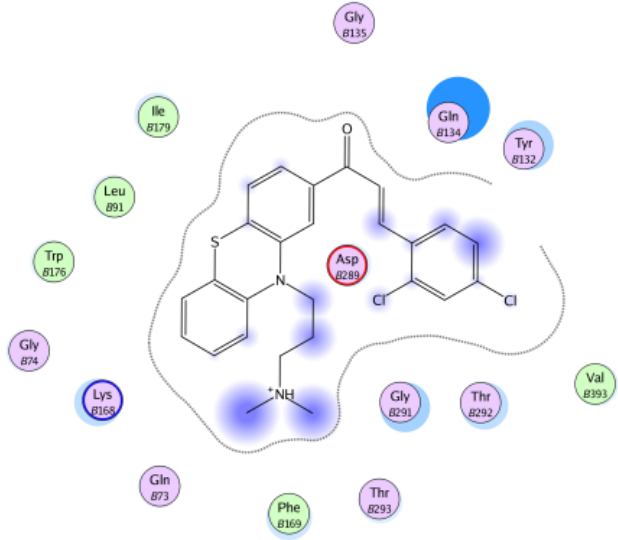
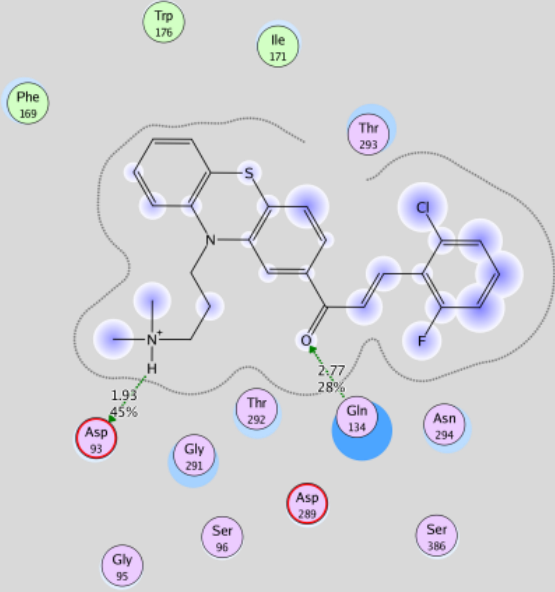
			B	-17.81	 <p>ThrB293 (hydrogen bond, score: 15%, length 3.02 Å), AspB289 (hydrogen bond, score: 51%, length 1.97 Å)</p>
AC3	5.35	7.37	A	-16.28	 <p>PheA169 (hydrogen bond, score: 46%, length 1.97 Å)</p>

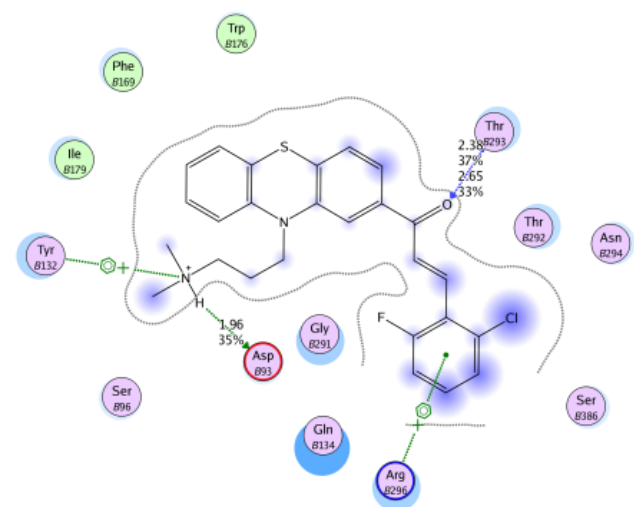
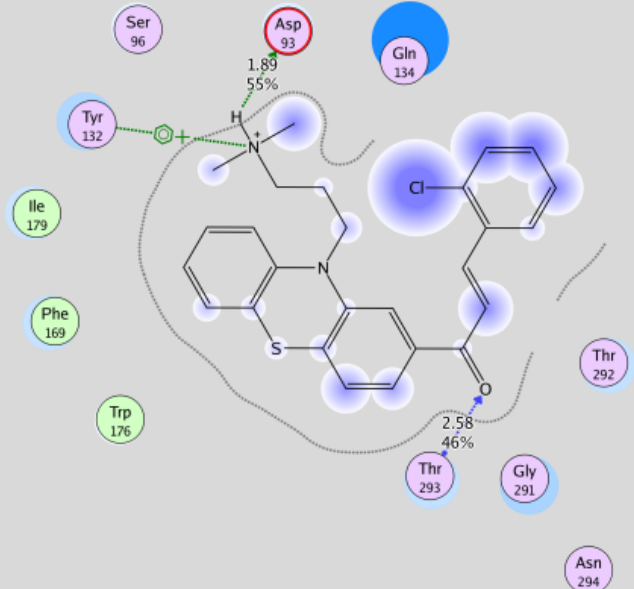


			B	-15.37	 <p>AspB93 (hydrogen bond, score: 74%, length 1.72 Å)</p>
AC4	6.81	8.49	A	-16.71	 <p>ThrA293 (hydrogen bond, score: 26%, length 2.93 Å), PheA169 (hydrogen bond, score: 47%, length 1.98 Å)</p>

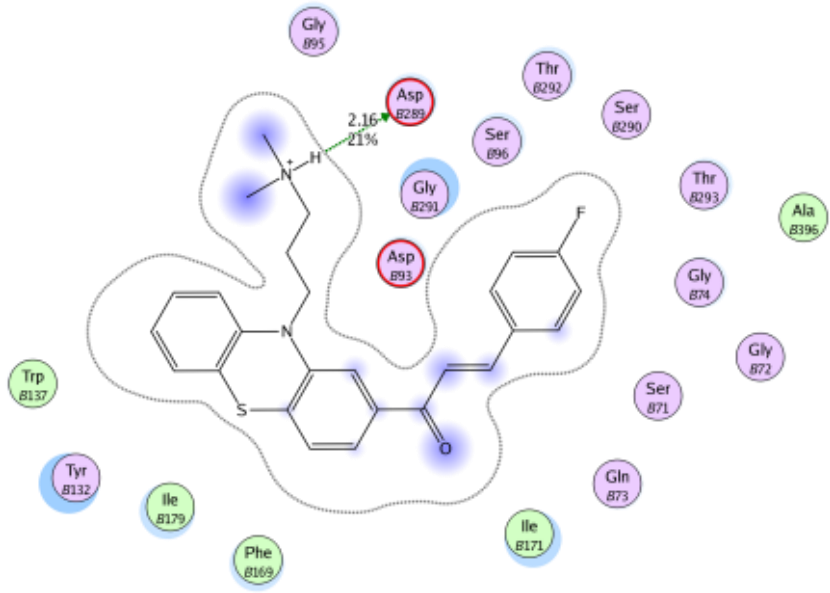
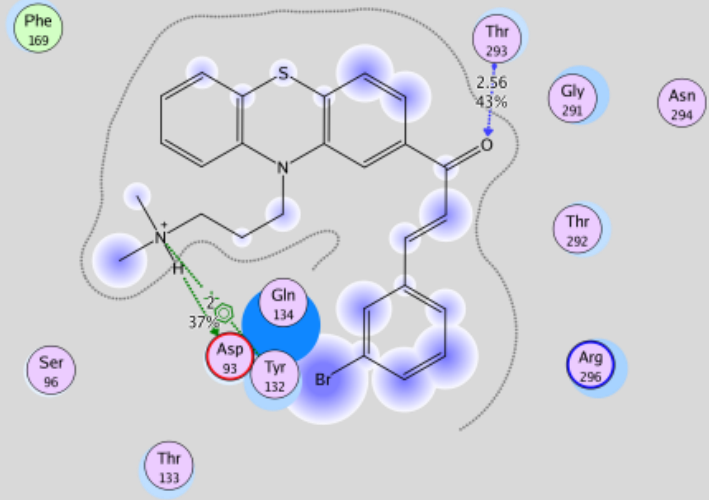
			B	-13.92	 <p>AspB93 (hydrogen bond, score: 76%, length 1.58 Å)</p>
AC5	5.91	7.25	A	-20.77	 <p>LysA168 (hydrogen bond, score: 22%, length 2 Å)</p>

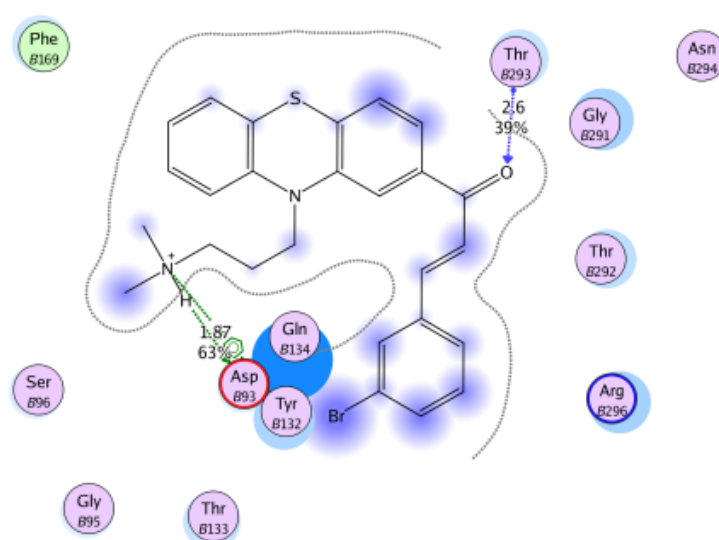
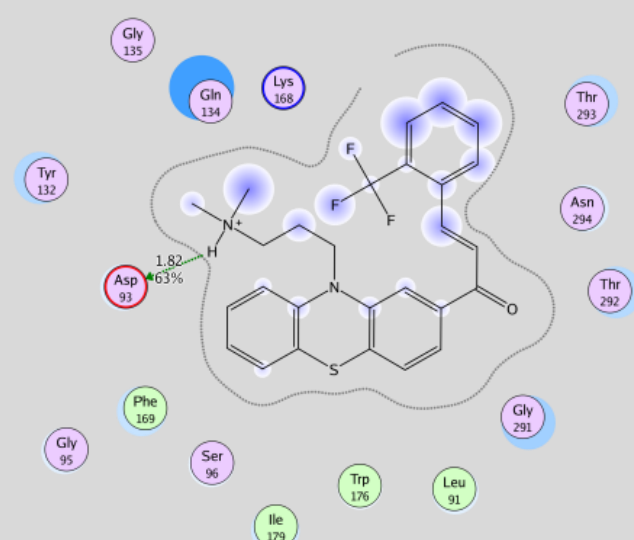
			B	-16.87	<p>AspB93 (hydrogen bond, score: 44%, length 1.9 Å), TyrB132 (arene-cation)</p>
AC6	6.30	7.68	A	-18.79	<p>AspA93 (hydrogen bond, score: 43%, length 1.91 Å), ThrA293 (hydrogen bond, score: 34%, length 2.86 Å) ArgA296 (arene-cation)</p>

			B	-16.73	 <p>No strong interaction</p>
AC7	6.35	7.63	A	-19.51	 <p>AspA93 (hydrogen bond, score: 45%, length 1.93 Å), GlnA134 (hydrogen bond, score: 28%, length 2.77 Å)</p>

			B	-16.85	 <p>AspB93 (hydrogen bond, score: 35%, length 1.96 Å), ThrB293 (hydrogen bonds, score: 33%; 37%, length: 2.65 Å; 2.38 Å), TyrB132 (arene-cation), ArgB296 (arene-cation)</p>
AC8	5.53	7.15	A	-20.36	 <p>AspA93 (hydrogen bond, score: 55%, length 1.89 Å), ThrA293 (hydrogen bond, score: 46%, length 2.58 Å), TyrA132 (arene-cation)</p>

			B	-16.81	<p>AspB93 (hydrogen bond, score: 49%, length 1.82 Å), TyrB132 (arene-cation)</p>
AC9	5.43	7.12	A	-20.97	<p>AspA93 (hydrogen bond, score: 66%, length 1.78 Å), ThrA293 (hydrogen bond, score: 11%, length 3 Å)</p>

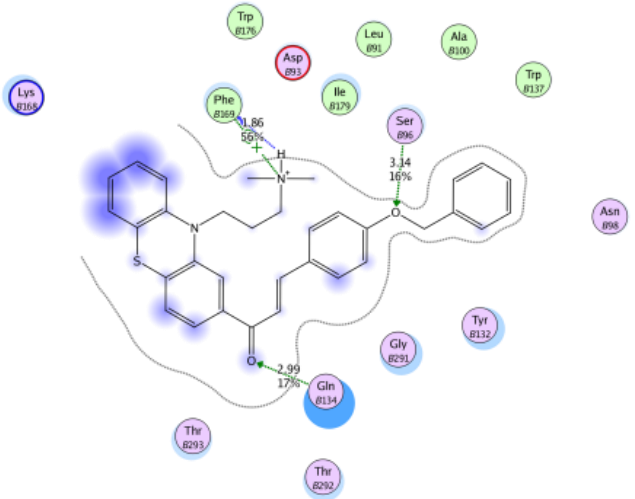
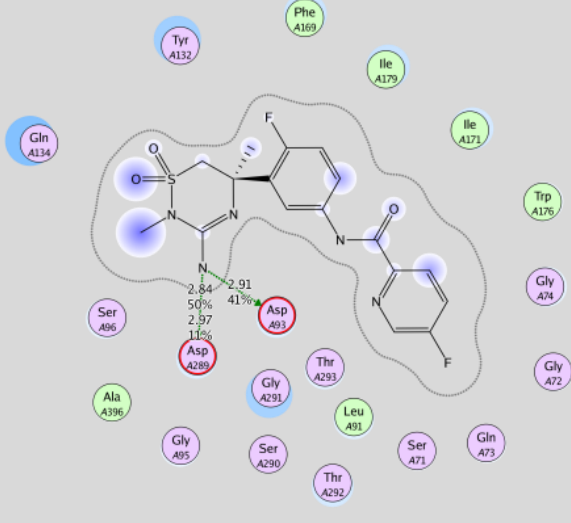
			B	-18.06	 <p>AspB289 (hydrogen bond, score: 21%, length 2.16 Å)</p>
AC10	5.70	7.28	A	-22.51	 <p>AspA93 (hydrogen bond, score: 37%, length 2 Å), ThrA293 (hydrogen bond, score: 43%, length 2.56 Å), TyrA132 (arene-cation)</p>

			B	-20.81	 <p>AspB93 (hydrogen bond, score: 63%, length 1.87 Å), ThrB293 (hydrogen bond, score: 39%, length 2.6 Å), TyrB132 (arene-cation)</p>
AC11	6.40	7.99	A	-19.35	 <p>AspA93 (hydrogen bond, score: 63%, length 1.82 Å)</p>



			B	-16.41	<p>AspB93 (hydrogen bond, score: 25%, length 2.02 Å), ThrB293 (hydrogen bond, score: 13%, length 2.93 Å), TyrB132 (arene-cation)</p>
AC12	6.46	8.08	A	-18.25	<p>AspA93 (hydrogen bond, score: 57%, length 1.85 Å)</p>

			B	-16.18	<p>PheB169 (hydrogen bond, score: 41%, length 2 Å), ThrB293 (hydrogen bond, score: 32%, length 2.9 Å)</p>
AC13	5.52	7.20	A	-11.50	<p>ThrA293 (hydrogen bond, score: 41%, length 2.81 Å), PheA169 (hydrogen bond, score: 29%, length 2.02 Å)</p>

			B	-14.09	 <p>SerB96 (hydrogen bond, score: 16%, length 3.14 Å), PheB169 (hydrogen bond, score: 56%, length 1.86 Å), (arene-cation), GlnB134 (hydrogen bond, score: 17%, length 2.99 Å)</p>
Verubecestat	-	7.66	A		 <p>AspA93 (hydrogen bond, score: 41%, length 2.91Å), AspA289 (hydrogen bonds, score: 50%; 11%, length 2.84 Å, 2.97 Å)</p>

			B		<p>       polar → sidechain acceptor    solvent residue    arene-arene        acidic ← sidechain donor    metal complex    arene-cation        basic → backbone acceptor    solvent contact    metal contact        greasy ← backbone donor    ligand exposure    receptor exposure        proximity contour    ligand exposure    receptor exposure     </p> <p>AspB93 (hydrogen bond, score: 54%, length 2.82 Å), AspB289 (hydrogen bonds, score: 49%; 11%, length 2.85 Å, 2.96 Å)</p>
Quercetin	5.02	5.24	A	-22.23	<p>GlnA134 (hydrogen bonds, score: 22%, length: 2.68 Å; score: 11%, length 3.03 Å), GlyA291 (hydrogen bond, score: 23%, length: 1.9 Å)</p>

			B	-23.95	<p>GlnB134 (hydrogen bonds, score: 12%, length: 2.98 Å; score), GlyB291 (hydrogen bond, score: 31%, length: 1.71 Å), GlnB73 (hydrogen bond, score: 29%, length: 1.62 Å)</p>
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**Table S4.** Dataset of 72 compounds used in the building of 2D-QSAR model for AChE inhibitors

<b>No.</b>	<b>Name</b>	<b>pIC50</b>	<b>predicted pIC50</b>
1	CHEMBL3133246	5.45	5.21
2	CHEMBL2296124	4.33	4.26
3	CHEMBL237223	4.59	4.72
4	CHEMBL2334727	4.72	4.60
5	CHEMBL3617370	4.51	4.42
6	CHEMBL3617380	4.49	4.46
7	CHEMBL2334728	4.24	4.28
8	CHEMBL416526	4.94	4.73
9	CHEMBL2393074	4.75	4.90
10	CHEMBL2385772	4.48	4.73
11	CHEMBL253386	5.49	5.22
12	CHEMBL3617382	4.63	4.52
13	CHEMBL3125443	4.90	4.67
14	CHEMBL3617375	4.28	4.49
15	CHEMBL3127216	4.84	4.70
16	CHEMBL3617367	4.38	4.27
17	CHEMBL457684	4.78	4.48
18	CHEMBL340807	4.80	4.66
19	CHEMBL2334726	4.49	4.55
20	CHEMBL659	5.62	5.42
21	CHEMBL2334730	4.64	4.78
22	CHEMBL3133247	4.81	5.07
23	CHEMBL517342	4.19	4.38
24	CHEMBL2296125	4.46	4.37
25	CHEMBL3617393	4.47	4.24
26	CHEMBL3617388	4.45	4.47
27	CHEMBL2385773	4.83	4.61
28	CHEMBL2385777	4.34	4.58
29	CHEMBL3127219	4.92	4.75
30	CHEMBL3127043	4.98	4.97
31	CHEMBL1269160	4.57	4.42
32	CHEMBL2334725	4.70	4.76
33	CHEMBL1270150	4.73	4.81
34	CHEMBL500712	4.40	4.21
35	CHEMBL2334736	4.72	4.83
36	CHEMBL2018160	4.63	4.48
37	CHEMBL2334737	4.71	4.61
38	CHEMBL269538	4.19	4.49
39	CHEMBL3617397	4.52	4.32
40	CHEMBL2334745	4.49	4.70
41	CHEMBL2296112	4.25	4.44

42	CHEMBL3617381	4.49	4.53
43	CHEMBL3133250	4.29	4.63
44	CHEMBL12014	4.23	4.50
45	CHEMBL3617372	4.26	4.44
46	CHEMBL3133243	4.97	5.10
47	CHEMBL510090	4.72	4.82
48	CHEMBL2385771	4.30	4.56
49	CHEMBL2296117	4.18	4.13
50	CHEMBL3127217	4.99	4.86
51	CHEMBL2334738	4.69	4.66
52	CHEMBL2296113	4.21	4.40
53	CHEMBL2381433	4.58	4.53
54	CHEMBL3132866	4.98	5.01
55	CHEMBL2385768	4.75	4.66
56	CHEMBL2385780	4.88	4.68
57	CHEMBL2334739	4.52	4.35
58	CHEMBL3133245	5.36	5.17
59	CHEMBL1270255	5.01	5.24
60	CHEMBL3133248	4.66	4.90
61	CHEMBL3617362	4.22	4.42
62	CHEMBL2381434	4.67	4.49
63	CHEMBL1535235	4.61	4.53
64	CHEMBL129177	4.47	4.70
65	CHEMBL2385784	4.53	4.76
66	CHEMBL2385779	4.42	4.23
67	CHEMBL3617396	4.39	4.26
68	CHEMBL1085869	5.67	5.51
69	CHEMBL2334747	4.51	4.56
70	CHEMBL2385766	4.35	4.57
71	CHEMBL463856	4.33	4.26
72	CHEMBL2334732	4.86	4.81

**Table S5.** Dataset of 215 compounds used in the building of 2D-QSAR model for BACE-1 inhibitors

No.	Name	pIC50	Predicted pIC50
1	BMC-2009-3671-17	6.92	7.02
2	BMC-2004-247-8	6.46	6.57
3	BMC-2007-1023-8	8.30	7.97
4	JMC-2012-27-30	5.49	5.65
5	BMC-2004-248-21	7.17	6.96
6	BMC-2009-3675-11	7.96	8.19
7	BMC-2009-3671-20	6.82	7.33
8	BMC-2009-3677-26	7.49	7.93
9	BMC-2008-1019-14	6.26	6.86
10	BMC-2014-2038-39	7.74	7.51
11	BMC-2013-4677-30	6.83	6.54
12	BMC-2004-248-19	7.37	6.74
13	BMC-2013-4677-37	5.88	6.33
14	BMC-2013-4241-10	6.46	6.45
15	BMC-2014-2036-24	7.49	8.02
16	BMC-2013-4675-7	7.35	6.70
17	BMC-2009-3667-16	7.72	7.63
18	BMC-2014-2039-43	7.28	7.39
19	EODD-2013-723-102	6.62	6.99
20	JMC-2012-26-15	5.00	5.15
21	JMC-2012-27-37	5.70	5.84
22	EODD-2013-718-42c	8.00	8.56
23	JMC-2012-9013-24	7.11	7.90
24	BMC-2014-2041-65	6.24	6.08
25	BMC-2008-1021-30	6.48	7.12
26	BMC-2009-3672-21	8.10	7.85
27	JMC-2012-9013-26	7.58	7.52
28	EODD-2013-720-53	6.82	7.20
29	BMC-2008-1021-31	6.64	6.66
30	BMC-2008-1021-28	7.08	6.58
31	JMC-2012-27-41	6.20	6.58
32	BMC-2009-3675-7	8.40	8.17
33	BMC-2009-3673-39	7.70	7.35
34	BMC-2014-2042-71	6.62	6.31
35	EODD-2013-713-11	7.55	7.25
36	BMC-2009-3672-27	7.04	7.46
37	BMC-2014-2042-90	8.07	7.85
38	EODD-2013-719-48	8.48	8.21
39	JMC-2012-27-47	6.49	6.45
40	BMC_2014-2042-81	7.60	8.12
41	BMC-2009-3665-4	8.04	7.43
42	BMC-2009-3673-36	7.64	7.69



43	BMC-2013-4676-17	6.93	7.33
44	BMC-2014-2038-41	7.70	7.60
45	BMC-2013-4677-38	6.07	6.44
46	BMC-2009-3667-20	7.47	7.71
47	EODD-2013-720-58	7.70	7.39
48	JMC-2012-26-24b	6.20	5.74
49	BMC-2014-2036-23	7.70	7.56
50	BMC-2013-4677-53	7.05	7.14
51	BMC-2007-1023-2	6.60	7.23
52	BMC-2009-3667-17	7.64	7.27
53	BMC-2008-1020-23	6.22	6.37
54	BMC-2014-2041-66	6.15	6.14
55	BMC-2013-4241-4	5.99	6.39
56	BMC-2009-3673-34	7.85	7.19
57	BMC-2014-2036-17	7.32	7.35
58	BMC-2008-1020-18	6.12	5.95
59	BMC-2014-2038-37	4.66	5.00
60	BMC-2009-3672-24	8.30	7.96
61	BMC-2007-1024-18	7.40	7.93
62	BMC-2008-1020-24	6.80	6.66
63	JMC-2012-28-64	6.10	6.08
64	BMC-2014-2035-13	8.00	7.51
65	BMC-2014-2040-53	8.15	8.02
66	BMC-2009-3667-18a	7.58	7.41
67	BMC-2009-3666-10	7.64	8.03
68	JMC-2012-9010-17	8.10	7.96
69	BMC-2014-2036-20	7.11	7.79
70	EODD-2013-721-64	6.33	6.65
71	BMC-2013-4677-35	6.38	6.60
72	BMC-2009-3671-12	7.04	7.62
73	EODD-2013-718-44	8.70	8.08
74	EODD-2013-721-65	7.20	6.65
75	BMC-2013-4242-23	7.34	7.00
76	BMC-2013-4241-9	7.92	7.22
77	BMC-2014-2040-62	7.69	7.61
78	BMC-2007-1023-5	7.30	7.87
79	BMC-2004-247-9	6.26	6.30
80	EODD-2013-718-42b	8.04	8.33
81	BMC-2009-3676-18	8.10	7.76
82	BMC-2007-1023-6	8.22	8.01
83	BMC-2009-3668-25	8.22	7.78
84	EODD-2013-718-45	7.68	7.34
85	BMC-2004-246-4	6.72	6.46
86	BMC-2013-4241-14	7.51	7.15
87	BMC-2009-3667-19	7.55	7.59

88	BMC-2007-1025-22	8.22	7.68
89	EODD-2013-716-31	8.70	8.27
90	BMC-2014-2042-85	6.53	7.06
91	BMC-2007-1023-11	8.30	8.05
92	BMC-2009-3671-16	7.58	8.09
93	BMC-2014-2039-42	7.08	7.22
94	BMC-2009-3666-11	7.74	8.32
95	BMC-2014-2038-40	7.70	7.27
96	BMC-2004-248-13	6.68	7.04
97	BMC-2014-2039-47	7.57	7.82
98	BMC-2009-3672-25	7.22	7.43
99	BMC-2009-3677-23	7.30	7.25
100	BMC-2009-3666-7	7.20	7.33
101	JMC-2012-28-83	5.10	5.52
102	BMC-2014-2035-14	7.70	8.20
103	BMC-2007-1023-10	8.30	8.19
104	BMC-2014-2035-15	8.00	8.16
105	BMC-2014-2036-22	7.04	6.54
106	EODD-2013-718-42a	7.89	8.20
107	BMC-2014-2042-72	7.58	7.34
108	BMC-2013-4242-24	6.43	6.98
109	BMC-2009-3672-26	8.04	7.58
110	BMC-2009-3676-21	6.60	6.71
111	BMC-2009-3677-24	7.40	7.21
112	BMC-2013-4242-20	6.92	6.62
113	BMC-2009-3675-9	8.00	7.47
114	BMC-2013-4241-5	6.26	6.22
115	JMC-2012-26-19b	5.20	5.44
116	BMC-2014-2042-87	7.31	7.24
117	BMC-2014-2042-84	6.49	7.03
118	BMC-2013-4676-19	7.38	7.76
119	BMC-2013-4241-3	5.05	5.61
120	BMC-2009-3668-27	7.40	7.26
121	BMC-2009-3677-25	7.70	7.26
122	BMC-2009-3675-10	7.66	7.50
123	BMC-2007-1025-21	8.40	7.77
124	BMC-2007-1024-17	8.30	8.17
125	BMC-2004-246-5	6.89	6.25
126	BMC-2009-3676-19	7.06	7.16
127	BMC-2013-4675-4	6.24	6.39
128	BMC-2014-2041-64	6.92	6.23
129	BMC-2004-246-6	5.95	6.41
130	BMC-2013-4242-22	7.64	7.14
131	BMC-2007-1024-15	7.92	7.44
132	BMC-2013-4241-12	6.86	6.99

133	BMC-2009-3673-35	8.30	7.86
134	BMC-2008-1019-13	6.52	6.58
135	BMC-2013-4241-16	6.12	6.63
136	BMC-2014-2042-73	7.52	7.45
137	JMC-2012-27-34	5.70	5.91
138	BMC-2013-4677-36	6.65	6.43
139	BMC-2004-248-20	6.75	6.64
140	BMC-2009-3672-23	7.64	7.81
141	JMC-2012-9010-18	7.64	7.67
142	BMC-2009-3675-13	7.64	7.55
143	JMC-2012-28-65	6.30	6.03
144	BMC-2013-4241-15	7.35	7.51
145	BMC-2004-246-7	5.76	6.23
146	BMC-2013-4677-48	8.04	7.86
147	BMC-2013-4241-13	7.41	7.34
148	BMC-2013-4675-6	6.42	6.42
149	BMC-2009-3671-9	8.70	8.29
150	BMC-2009-3671-11	7.72	8.26
151	BMC-2008-1019-12	7.48	6.96
152	BMC-2008-1021-27	7.70	7.30
153	BMC-2014-2039-46	7.27	7.16
154	BMC-2014-2035-9	7.10	6.33
155	JMC-2012-9010-15	7.74	7.98
156	BMC-2007-1024-13	7.47	7.41
157	BMC-2014-2042-79	7.40	7.59
158	EODD-2013-716-32	8.40	8.23
159	BMC-2008-1021-29	7.00	7.42
160	JMC-2012-9013-25	7.48	7.95
161	BMC-2004-246-3	7.09	7.26
162	BMC-2014-2042-80	7.47	7.90
163	BMC-2004-247-10	6.46	6.20
164	JMC-2012-9013-23	7.54	8.48
165	BMC-2008-1020-26	7.85	7.24
166	BMC-2009-3667-18b	7.40	7.51
167	EODD-2013-713-12a	8.41	8.84
168	BMC-2009-3677-27	7.10	7.58
169	BMC-2004-248-14	6.68	6.49
170	BMC-2007-1024-12	6.92	6.75
171	EODD-2013-718-41	7.30	7.95
172	BMC-2009-3675-14	7.32	7.60
173	BMC-2008-1019-8	5.61	6.20
174	BMC-2008-1020-22	5.77	6.13
175	BMC-2008-1019-6	6.74	6.26
176	BMC-2009-3671-15	8.15	8.22
177	BMC-2013-4242-19	7.21	6.91

178	BMC-2014-2042-82	7.55	7.36
179	EODD-2013-721-61	6.96	7.39
180	BMC-2008-1020-16	7.40	7.28
181	BMC-2014-2042-76	8.70	8.94
182	BMC-2009-3675-5	8.30	7.93
183	BMC-2009-3675-16	7.70	7.51
184	BMC-2007-1023-9	7.96	8.00
185	BMC-2014-2042-78	7.70	7.43
186	BMC-2008-1019-7	5.95	6.35
187	BMC-2013-4675-3	6.27	6.20
188	JMC-2012-26-8b	4.60	4.57
189	EODD-2013-713-10	7.06	7.07
190	JMC-2012-26-19a	5.00	5.44
191	BMC-2014-2035-12	5.47	5.08
192	BMC-2013-4676-18	6.90	7.35
193	JMC-2012-9013-19	8.27	8.63
194	BMC-2004-248-17	7.22	6.58
195	BMC-2014-2040-54	8.22	8.06
196	BMC-2009-3671-13	7.82	7.98
197	JMC-2012-26-24a	6.00	6.04
198	BMC-2008-1020-19	7.77	7.61
199	EODD-2013-719-50	7.77	8.60
200	JMC-2012-27-54a	5.40	5.90
201	EODD-2013-717-39a	9.40	9.53
202	BMC-2007-1024-14	7.74	8.09
203	BMC-2013-4677-31	6.73	6.68
204	BMC-2009-3672-22	8.70	8.02
205	BMC-2009-3667-23	6.38	6.32
206	BMC-2013-4675-5	7.03	6.31
207	BMC-2013-4677-32	7.22	7.24
208	BMC-2014-2042-88	7.44	7.03
209	BMC-2007-1023-7	8.52	7.97
210	JMC-2012-26-8a	4.79	4.94
211	JMC-2012-9013-22	7.69	8.31
212	BMC-2013-4241-18	7.02	6.58
213	BMC-2014-2042-75	8.00	8.54
214	JMC-2012-9010-16	7.45	8.05
215	BMC-2007-1024-16	8.10	7.53

**Table S6. Equations for calculation of 2D-QSAR validation metrics**

These metrics were calculated according to the equations (Eq.) (1-10).

$$\text{RMSE} = \sqrt{\frac{\sum_{i=1}^{n_{TR}} (\hat{y}_i - y_i)^2}{n}} \quad (1)$$

$$R^2 = 1 - \frac{\sum_{i=1}^{n_{TR}} (\hat{y}_i - y_i)^2}{\sum_{i=1}^{n_{TR}} (y_i - \bar{y})^2} \quad (2)$$

$$\text{RMSE}_{\text{LOO}} = \sqrt{\frac{\sum_{i=1}^{n_{TR}} (\hat{y}'_i - y_i)^2}{n}} \quad (3)$$

$$R^2_{\text{LOO}} = 1 - \frac{\sum_{i=1}^{n_{TR}} (\hat{y}'_i - y_i)^2}{\sum_{i=1}^{n_{TR}} (y_i - \bar{y})^2} \quad (4)$$

$$R^2_{\text{PRED}} = 1 - \frac{\sum_{l=1}^{n_{VAL}} (y_{\text{pred}(\text{validation})} - y_{\text{validation}})^2}{\sum_{l=1}^{n_{VAL}} (y_{\text{validation}} - \bar{y}_{\text{training}})^2} \quad (5)$$

$$r_m^2 = r^2 (1 - \sqrt{r^2 - r_0^2}) \quad (6)$$

$$r'^2_m = r'^2 (1 - \sqrt{r'^2 - r'^2_0}) \quad (7)$$

$$\overline{r_m^2} = \frac{r_m^2 + r'^2_m}{2} \quad (8)$$

$$\Delta r_m^2 = |r_m^2 - r'^2_m| \quad (9)$$

$$\text{CCC} = \frac{2 \sum_{i=1}^{n_{VAL}} (y_i - \bar{y})(\hat{y}_i - \bar{\hat{y}})}{\sum_{i=1}^{n_{VAL}} (y_i - \bar{y})^2 + \sum_{i=1}^{n_{VAL}} (\hat{y}_i - \bar{\hat{y}})^2 + n_{VAL} (\bar{y} - \bar{\hat{y}})^2} \quad (10)$$

In Eq. (1), (2), and (10),  $y_i$  and  $\hat{y}_i$  are, respectively, the observed and predicted activity values, while  $\bar{y}$  and  $\bar{\hat{y}}$  are, respectively, the mean values of  $y_i$  and  $\hat{y}_i$ . In Eq.(3) and (4),  $y_i$  and  $\hat{y}'_i$  are, respectively, the observed and predicted activity values in LOO cross-validation. Eq. (6) and (7) were utilized to calculate correlation coefficients between observed and predicted activity values of the compounds of training set with ( $r^2$ ) or without intercept ( $r_0^2$ ) in case of using predicted data on the y-axis and experimental data on the x-axis, while ( $r'^2$ ) and ( $r'^2_0$ ) are, respectively, the same coefficients in the opposite case. The most stringent validation criteria thresholds including  $r_m^2 \geq 0.65$ ;  $\text{CCC} \geq 0.85$ ;  $\overline{r_m^2} \geq 0.5$ ; and  $\Delta r_m^2 \leq 0.2$  were applied to verify the external predictivity of good models [1,2]

**Table S7.** Selected descriptors used for building 2D-QSAR models

Code	Category	Description
BCUT_SlogP_3	Adjacency and distance matrix	A Burden's parameter using atomic contribution to logP (using the Wildman and Crippen SlogP method [3]) instead of partial charge.
BCUT_PEOE_1	Adjacency and distance matrix	A descriptor relating topological shape and partial charges
petitjean	Adjacency and distance matrix	Value of (diameter - radius) / diameter.
reactive	Physical property	An indicator of the presence of reactive groups. A non-zero value indicates that the molecule contains a reactive group. The table of reactive groups is based on the Oprea set [4] and includes metals, phospho-, N/O/S-N/O/S single bonds, thiols, acyl halides, Michael Acceptors, azides, esters, etc.
logS	Physical property	The log of the aqueous solubility (mol/L).
PEOE_VSA-0, PEOE_VSA+1, PEOE_VSA-3, PEOE_VSA-6	Partial charge	Sum of the proximate accessible <i>van der Waals</i> surface area ( $\text{\AA}^2$ ), $v_i$ , calculation for each atom over all the atoms $i$ , such that partial charge of atom $i$ is in a specified range.
SlogP_VSA2, SlogP_VSA3, SlogP_VSA5	Subdivided surface areas	Sum of the proximate accessible <i>van der Waals</i> surface area ( $\text{\AA}^2$ ), $v_i$ , calculated for each atom over all the atoms, such that partition coefficient for atom $i$ is in a specified range
SMR_VSA2	Subdivided surface areas	Sum of the proximate accessible <i>van der Waals</i> surface area ( $\text{\AA}^2$ ), $v_i$ , calculation for each atom over all the atoms $i$ , such that molar refractivity for atom $i$ is in a specified range.
a_ICM	Atom counts and bond counts	The entropy of the element distribution in the molecule (including implicit hydrogens but not lone pair pseudo-atoms)
chiral_u	Atom counts and bond counts	The number of unconstrained chiral centers.
rings	Atom counts and bond counts	The number of rings
a_Nn	Atom counts and bond counts	The number of nitrogen atoms.

**Table S8.** Values of selected descriptors used in prediction of pIC<sub>50</sub> of the synthesized chalcone derivatives (AChE)

Compound	BCUT_SLOGP_3	reactive	PEOE_VSA+1	PEOE_VSA-3	SlogP_VSA2	SMR_VSA2	Predicted pIC <sub>50</sub>
AC1	2.61	1	59.72	0.00	0.00	6.23	<b>4.80</b>
AC2	2.61	1	76.70	0.00	0.00	6.23	<b>4.74</b>
AC3	2.61	1	76.70	0.00	0.00	6.23	<b>4.74</b>
AC4	2.61	1	85.31	0.00	0.00	6.23	<b>4.71</b>
AC5	2.61	1	55.45	0.00	0.00	6.23	<b>4.82</b>
AC6	2.61	1	51.18	0.00	0.00	6.23	<b>4.83</b>
AC7	2.61	1	55.70	0.00	9.12	15.35	<b>4.76</b>
AC8	2.61	1	55.45	0.00	0.00	6.23	<b>4.82</b>
AC9	2.61	1	55.45	0.00	9.12	15.35	<b>4.76</b>
AC10	2.61	1	55.45	0.00	0.00	9.28	<b>4.81</b>
AC11	2.61	1	57.66	35.73	0.00	13.99	<b>4.59</b>
AC12	2.61	1	55.45	0.00	0.00	9.35	<b>4.81</b>
AC13	2.61	1	108.72	0.00	0.00	6.23	<b>4.63</b>
Galantamine	2.77	0	114.49	0.00	12.94	0.00	<b>5.14</b>

**Table S9.** Values of selected descriptors used in prediction of pIC<sub>50</sub> of the synthesized chalcone derivatives (BACE-1)

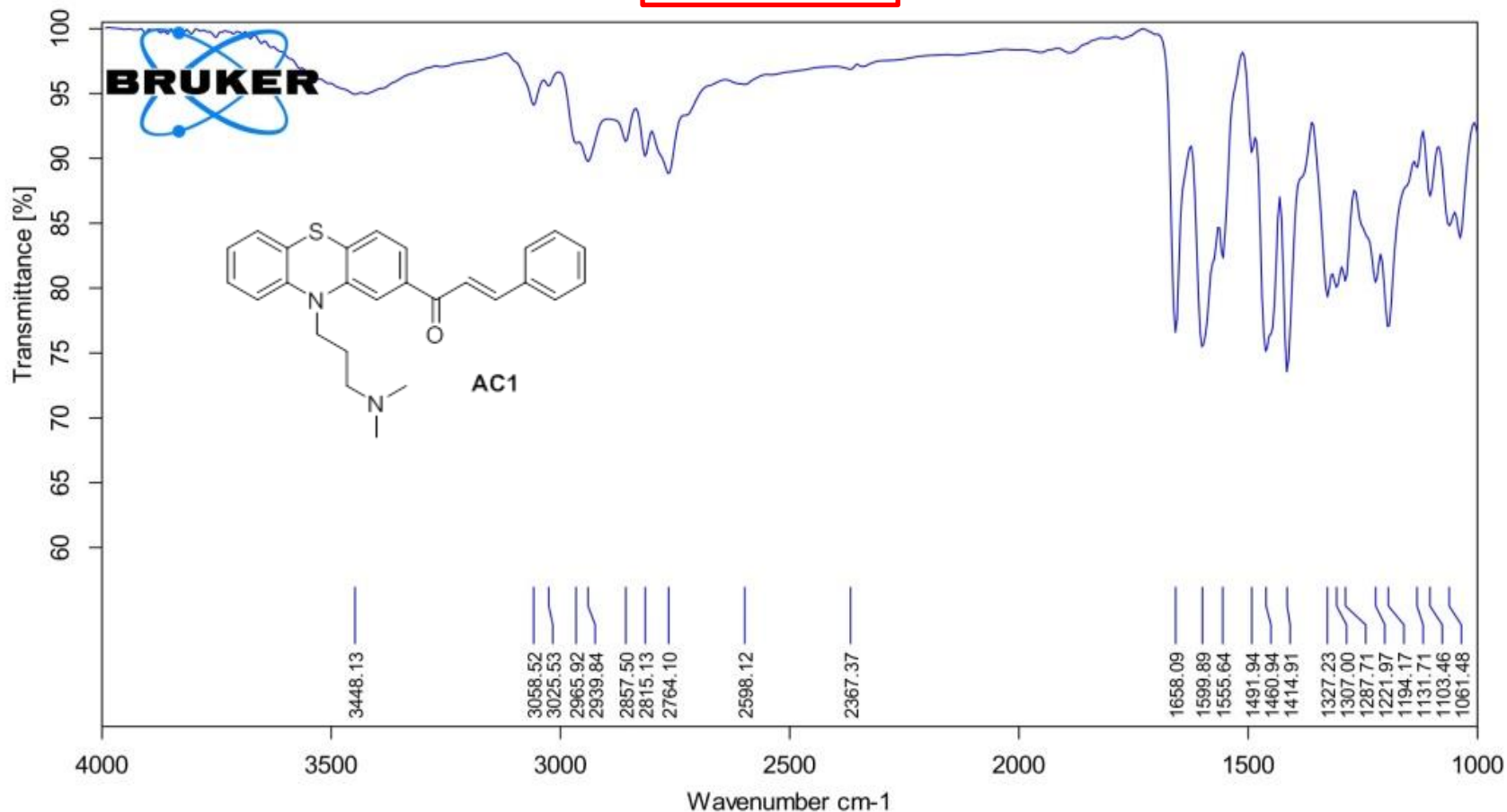
Compound	petitjean	BCUT_PEOE_1	a_ICM	chiral_u	rings	a_nN	PEOE_VSA-0	PEOE_VSA-6	logS	SlogP_VSA3	SlogP_VSA5	Predicted pIC <sub>50</sub>
AC1	0.47	-0.63	1.41	0	4	2	144.11	0.00	-6.42	36.88	68.90	<b>6.52</b>
AC2	0.50	-0.61	1.46	0	4	2	168.62	2.50	-6.47	36.88	104.28	<b>7.38</b>
AC3	0.47	-0.61	1.46	0	4	2	168.62	2.50	-6.47	36.88	104.28	<b>7.37</b>
AC4	0.47	-0.59	1.52	0	4	2	168.62	7.51	-6.57	36.88	175.05	<b>8.49</b>
AC5	0.50	-0.63	1.52	0	4	2	168.62	0.00	-7.15	36.88	68.90	<b>7.25</b>
AC6	0.50	-0.64	1.59	0	4	2	180.87	0.00	-7.89	36.88	68.90	<b>7.68</b>
AC7	0.47	-0.64	1.62	0	4	2	178.67	0.00	-7.45	36.88	68.90	<b>7.63</b>
AC8	0.47	-0.64	1.52	0	4	2	168.62	0.00	-7.15	36.88	68.90	<b>7.15</b>
AC9	0.50	-0.63	1.52	0	4	2	168.62	0.00	-6.71	36.88	68.90	<b>7.12</b>
AC10	0.47	-0.63	1.52	0	4	2	168.62	0.00	-7.51	36.88	71.95	<b>7.28</b>
AC11	0.47	-0.56	1.62	0	4	2	156.36	0.00	-7.48	36.88	68.90	<b>7.99</b>
AC12	0.47	-0.58	1.42	0	4	3	215.10	0.00	-6.35	36.88	134.69	<b>8.08</b>
AC13	0.48	-0.64	1.41	0	5	2	170.83	2.50	-8.24	36.88	68.90	<b>7.20</b>
Verubecestat	0.50	-0.58	2.00	1	3	5	112.04	12.47	-3.62	17.23	49.68	<b>7.66</b>
Quercetin	0.50	-0.63	-1.52	0	3	0	61.27	41.34	-2.77	25.39	5.24	<b>5.24</b>

Table S10. Spectra of synthesized chalcone derivatives

VIEN CONG NGHE HOA HOC  
01 Mac Dinh Chi - Q.1 - Tp HCM

IR

Tel: 08.38296127  
Fax: 08.38293889



E:\OPUS 7\2017\THANG 6\20170606\C12.0

C12

SENSOR 27 - BRUKER - GERMANY

6/8/2017



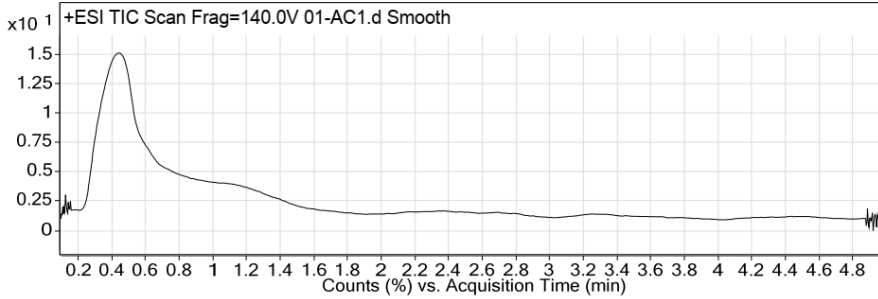
# MS

## Qualitative Analysis Report

<b>Data Filename</b>	01-AC1.d	<b>Sample Name</b>	01-AC1
<b>Sample Type</b>	Sample	<b>Position</b>	P2-A9
<b>Instrument Name</b>	Instrument 1	<b>User Name</b>	
<b>Acq Method</b>	Cot ngan - MSMS_Pos.m	<b>Acquired Time</b>	22/08/2020 10:06:30 AM
<b>IRM Calibration Status</b>	Success	<b>DA Method</b>	COTNGAN.M.m
<b>Comment</b>			
<b>Sample Group</b>		<b>Info.</b>	
<b>Stream Name</b>	LC 1	<b>Acquisition SW</b>	6200 series TOF/6500 series
		<b>Version</b>	Q-TOF B.06.01 (B6172 SP1)

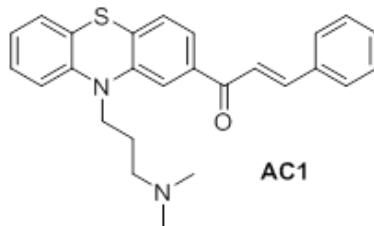
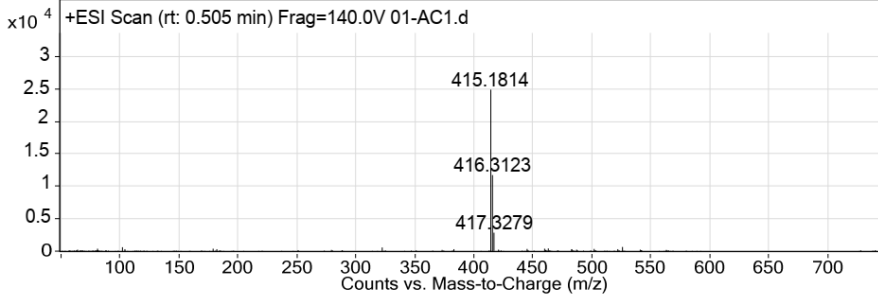
### User Chromatograms

Fragmentor Voltage 140 Collision Energy 0 Ionization Mode ESI



### User Spectra

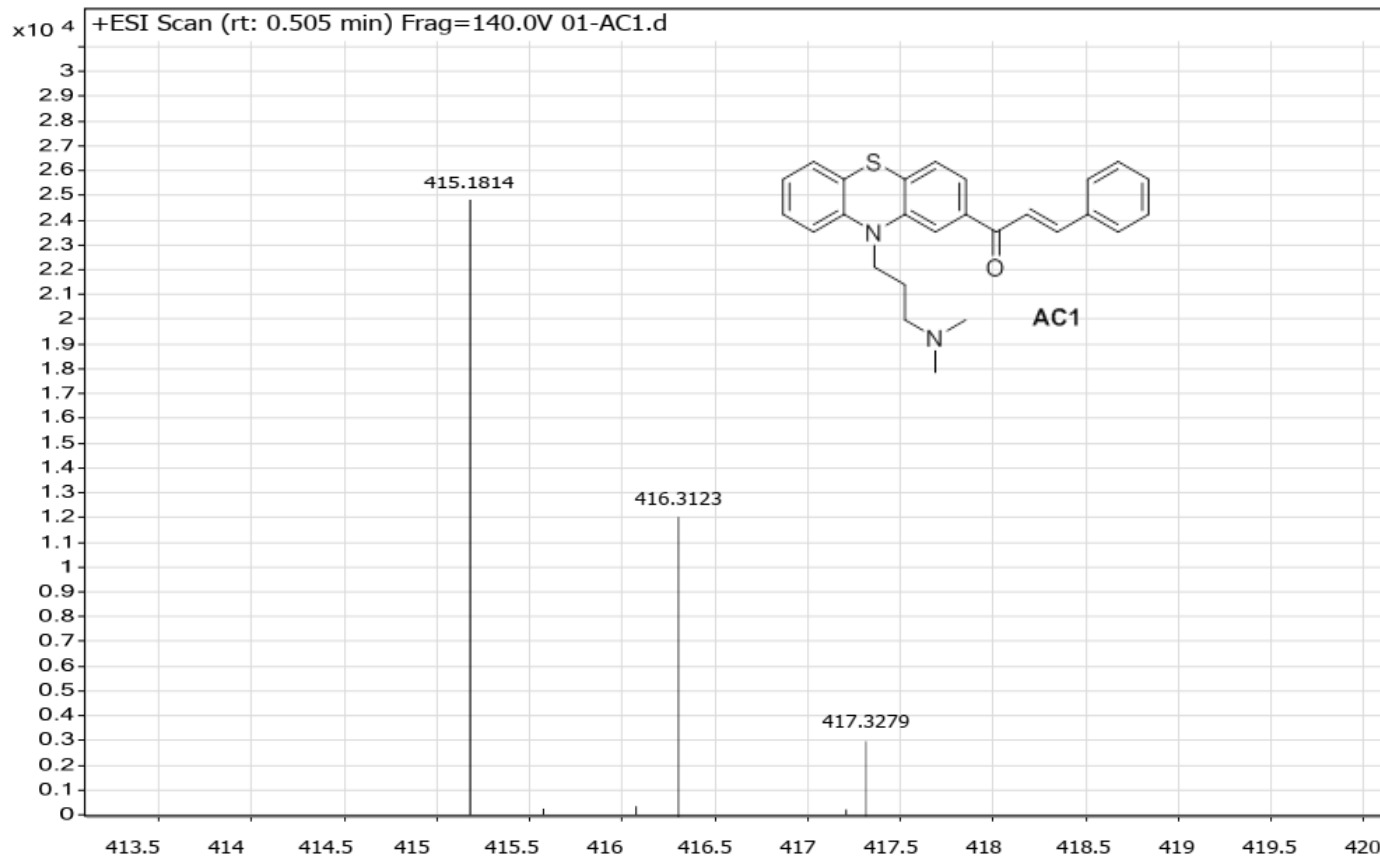
Fragmentor Voltage 140 Collision Energy 0 Ionization Mode ESI



--- End Of Report ---

# MS

Sample Name	01-AC1	Position	P2-A9	Instrument Name	Instrument 1	User Name	
Inj Vol	2	InjPosition		SampleType	Sample	IRM Calibration Status	Success
Data Filename	01-AC1.d	ACQ Method	Cot ngan - MSMS_Pos.	Comment		Acquired Time	22/08/2020 10:06:30 AM

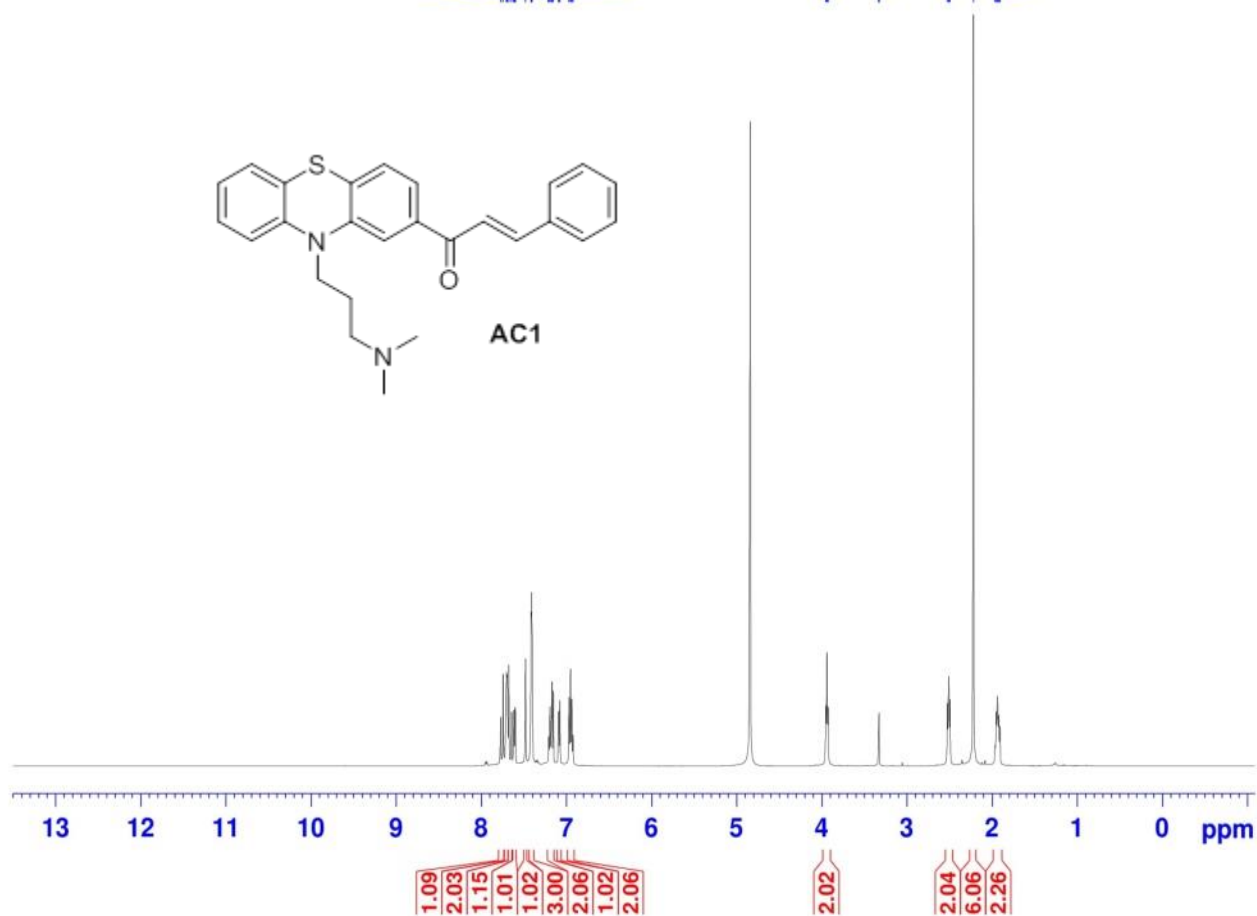
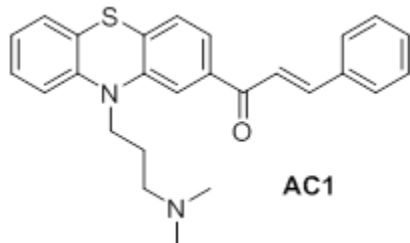


# <sup>1</sup>H-NMR

C12-MeOD-1H



7.776  
7.744  
7.705  
7.699  
7.678  
7.646  
7.620  
7.618  
7.604  
7.602  
7.483  
7.417  
7.410  
7.405  
7.212  
7.210  
7.195  
7.195  
7.172  
7.156  
7.095  
7.093  
7.080  
6.971  
6.954  
6.939  
6.924  
3.954  
3.941  
3.927  
3.333  
3.330  
3.327  
2.526  
2.511  
2.496  
2.221  
1.966  
1.952  
1.938  
1.923  
1.909



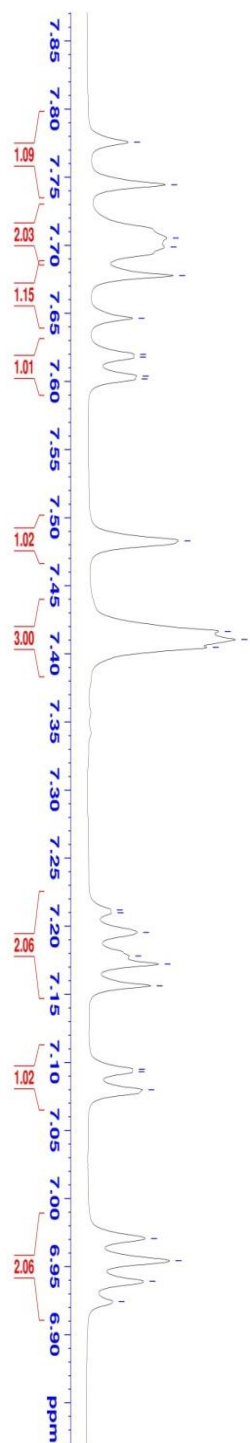
Current Data Parameters  
NAME 114DAO\_C12  
EXPNO 1  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20170704  
Time 11.31  
INSTRUM spect  
PROBHD 5 mm PABBO BB/  
PULPROG zg30  
TD 65536  
SOLVENT MeOD  
NS 16  
DS 2  
SWH 10000.000 Hz  
FIDRES 0.152588 Hz  
AQ 3.2767999 sec  
RG 50.73  
DW 50.000 usec  
DE 6.50 usec  
TE 303.0 K  
D1 1.00000000 sec  
TD0 1

===== CHANNEL f1 =====  
SFO1 500.2030889 MHz  
NUC1 1H  
P1 10.00 usec  
PLW1 22.00000000 W

F2 - Processing parameters  
SI 65536  
SF 500.2000017 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

# <sup>1</sup>H-NMR



7.776  
7.744  
7.705  
7.699  
7.678  
7.646  
7.620  
7.618  
7.604  
7.602

C12-MeOD-1H

7.483  
7.417  
7.410  
7.405

7.212  
7.210  
7.195  
7.178  
7.172  
7.156

7.095  
7.093  
7.080

6.971  
6.954  
6.939  
6.924

BRUKER



3.954  
3.941  
3.927

3.333  
3.330  
3.327

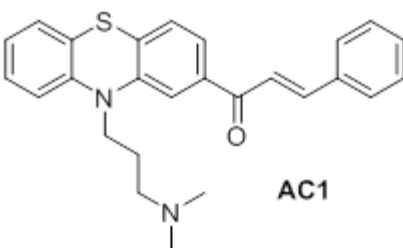
2.526  
2.511  
2.496

2.221

1.966  
1.952  
1.938  
1.923  
1.909

C12-MeOD-1H

BRUKER



# <sup>13</sup>C-NMR

C12-MeOD-C13CPD



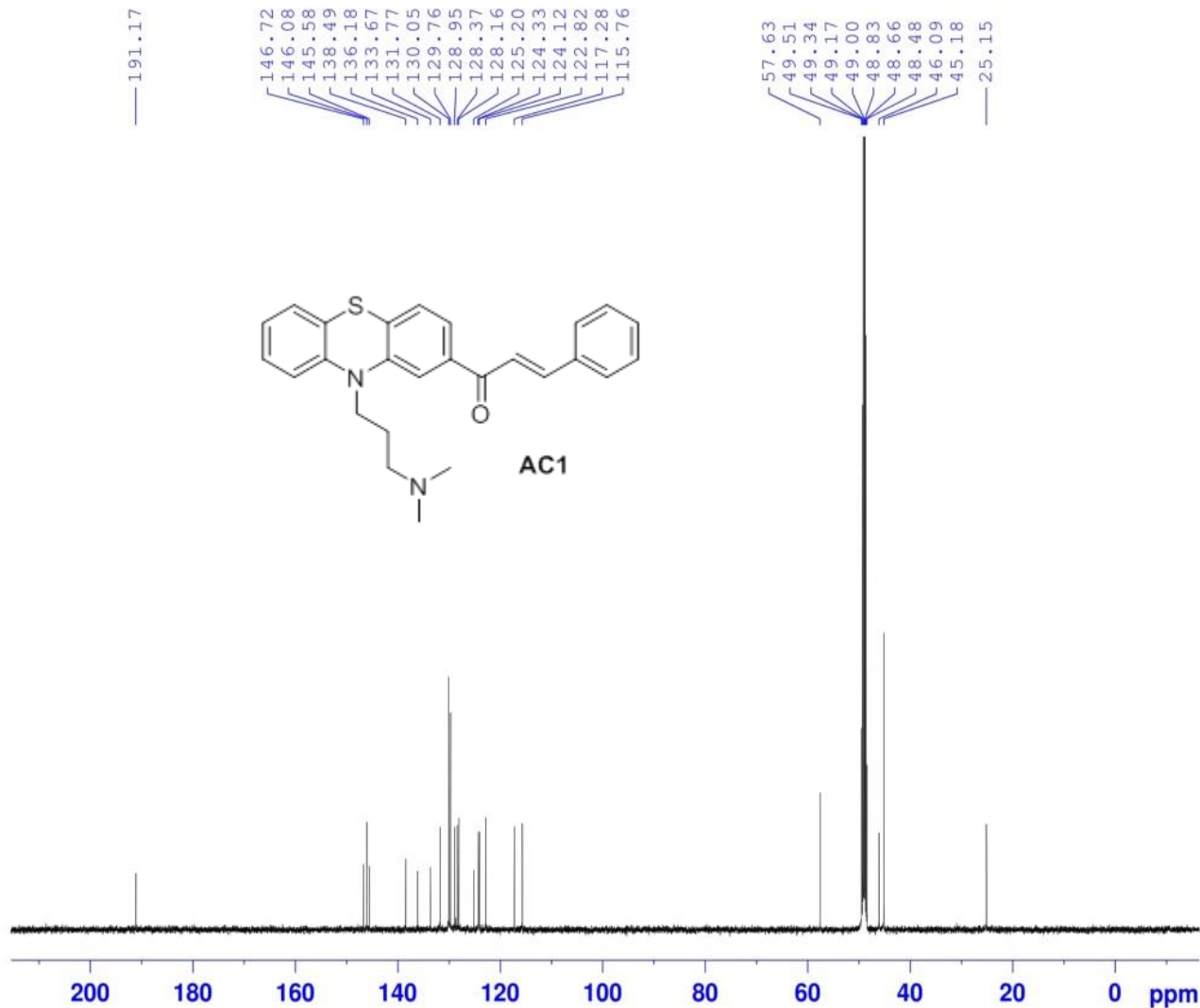
Current Data Parameters  
NAME 114DAO\_C12  
EXPNO 2  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20170705  
Time 14.51  
INSTRUM spect  
PROBHD 5 mm PABBO BB/  
PULPROG zgpg30  
TD 65536  
SOLVENT MeOD  
NS 128  
DS 4  
SWH 29761.904 Hz  
FIDRES 0.454131 Hz  
AQ 1.1010048 sec  
RG 198.57  
DW 16.800 usec  
DE 6.50 usec  
TE 303.0 K  
D1 2.00000000 sec  
D11 0.03000000 sec  
TD0 1

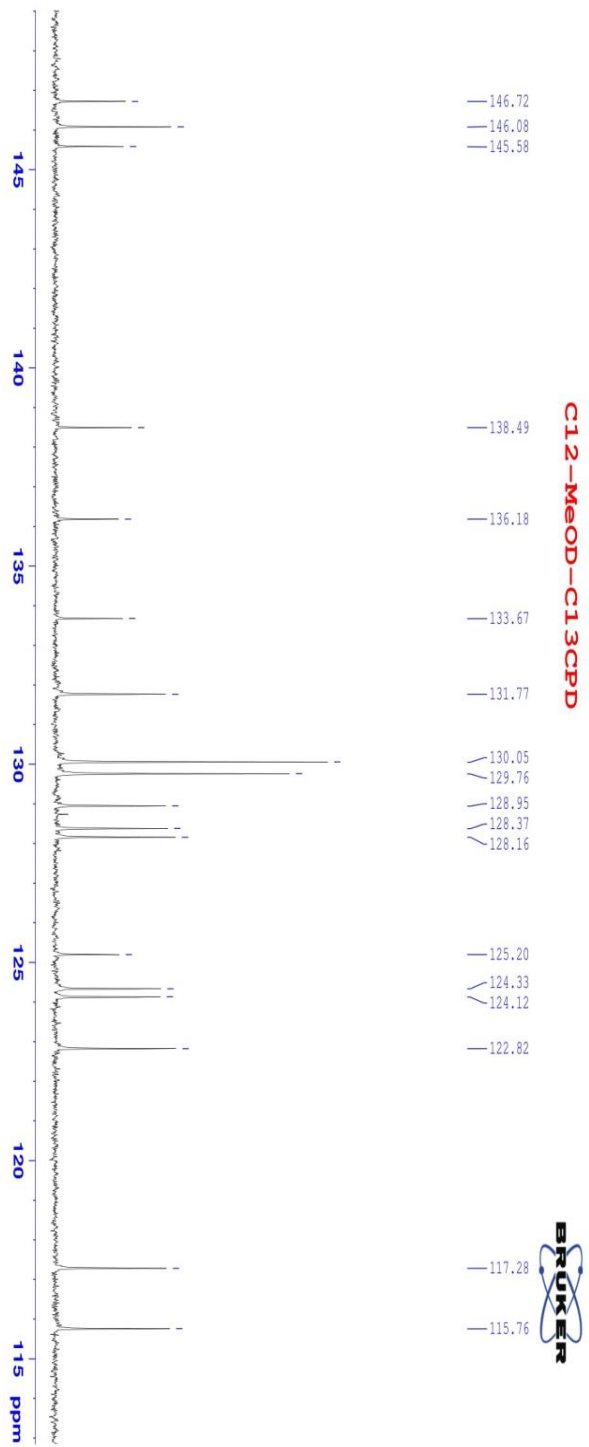
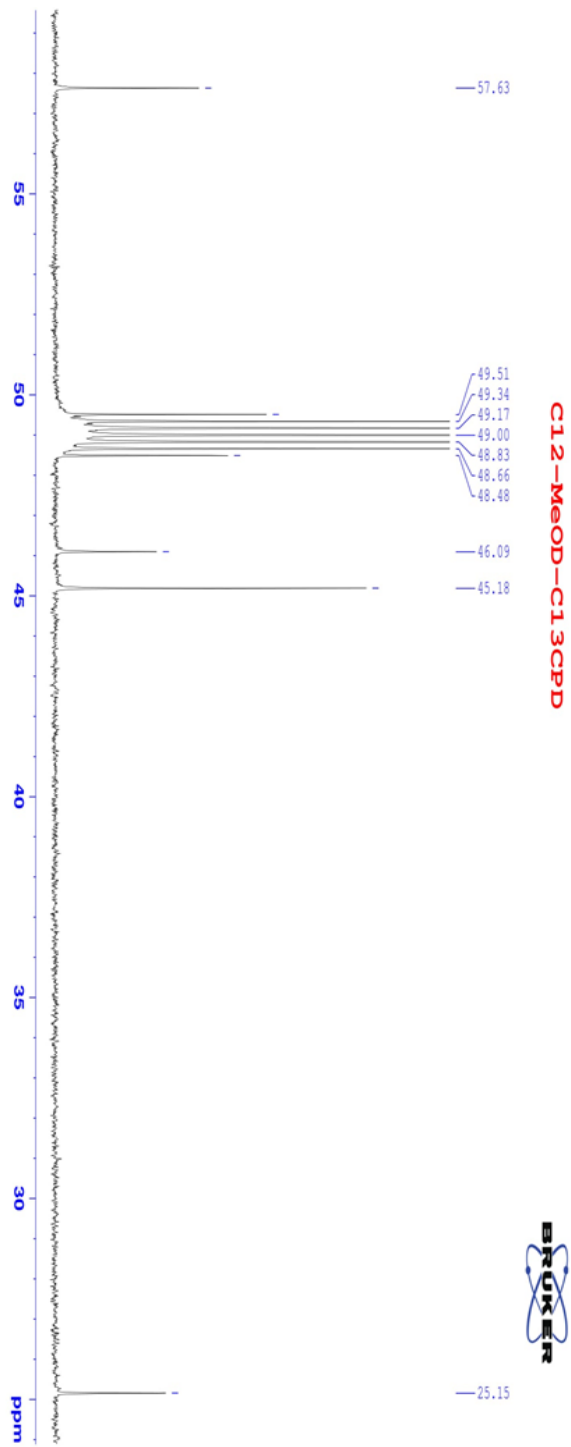
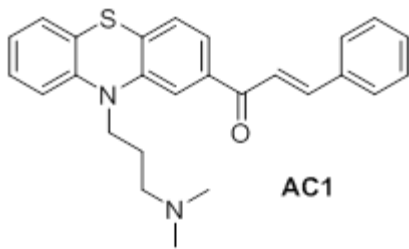
==== CHANNEL f1 =====  
SFO1 125.7879670 MHz  
NUC1 13C  
P1 10.00 usec  
PLW1 88.00000000 W

==== CHANNEL f2 =====  
SFO2 500.2020008 MHz  
NUC2 1H  
CPDPRG[2] waltz16  
PCPD2 80.00 usec  
PLW2 22.00000000 W  
PLW12 0.34375000 W  
PLW13 0.22000000 W

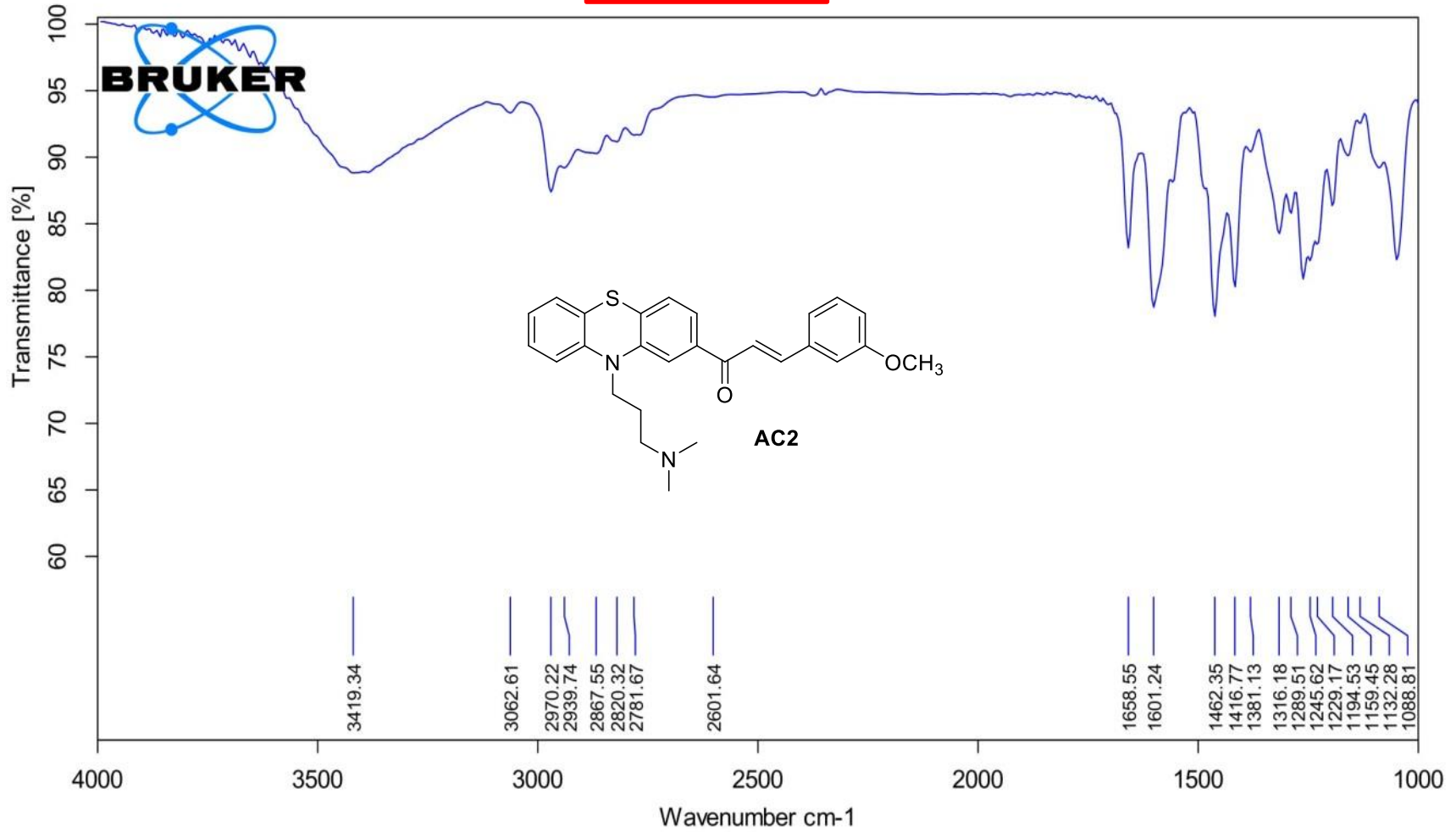
F2 - Processing parameters  
SI 32768  
SF 125.7752211 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40



# $^{13}\text{C}$ -NMR



IR



# MS

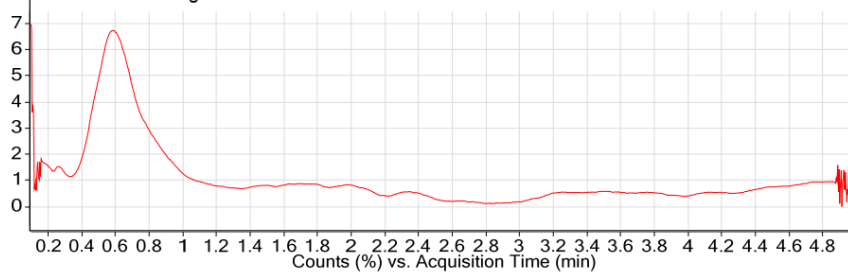
## Qualitative Analysis Report

<b>Data Filename</b>	02-AC2.d	<b>Sample Name</b>	02-AC2
<b>Sample Type</b>	Sample	<b>Position</b>	P2-C6
<b>Instrument Name</b>	Instrument 1	<b>User Name</b>	
<b>Acq Method</b>	Cot ngan - MSMS_Pos.m	<b>Acquired Time</b>	22/08/2020 10:13:59 AM
<b>IRM Calibration Status</b>	Success	<b>DA Method</b>	COTNGAN.M.m
<b>Comment</b>			
<b>Sample Group</b>		<b>Info.</b>	
<b>Stream Name</b>	LC 1	<b>Acquisition SW</b>	6200 series TOF/6500 series
		<b>Version</b>	Q-TOF B.06.01 (B6172 SP1)

### User Chromatograms

Fragmentor Voltage 140 Collision Energy 0 Ionization Mode ESI

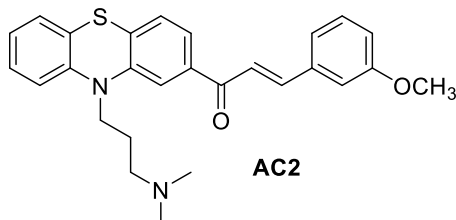
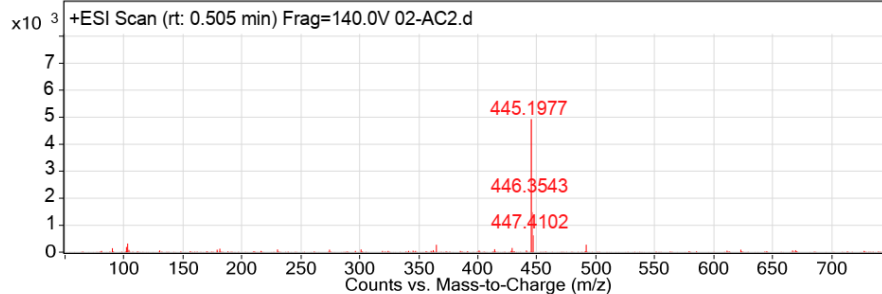
+ESI TIC Scan Frag=140.0V 02-AC2.d Smooth



### User Spectra

Fragmentor Voltage 140 Collision Energy 0 Ionization Mode ESI

+ESI Scan (rt: 0.505 min) Frag=140.0V 02-AC2.d

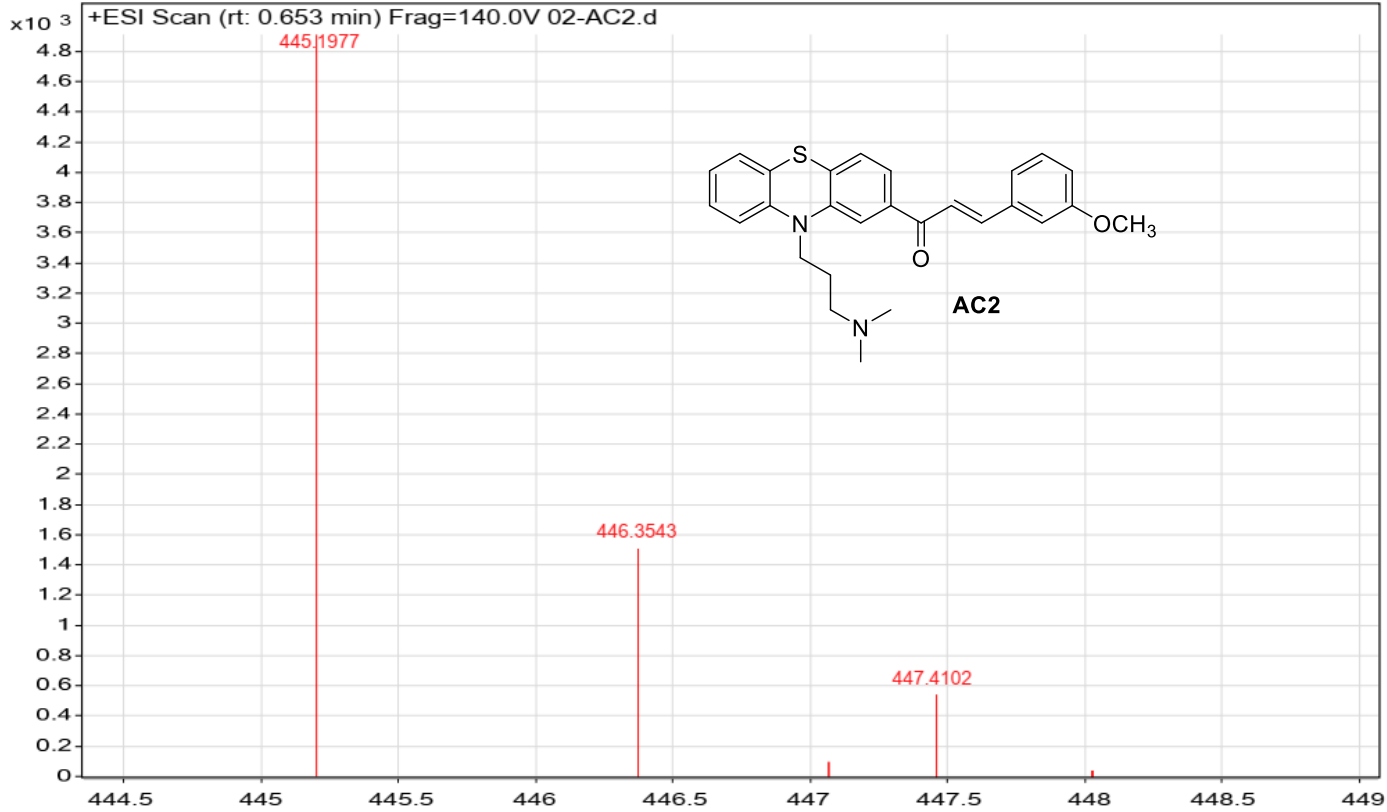


--- End Of Report ---



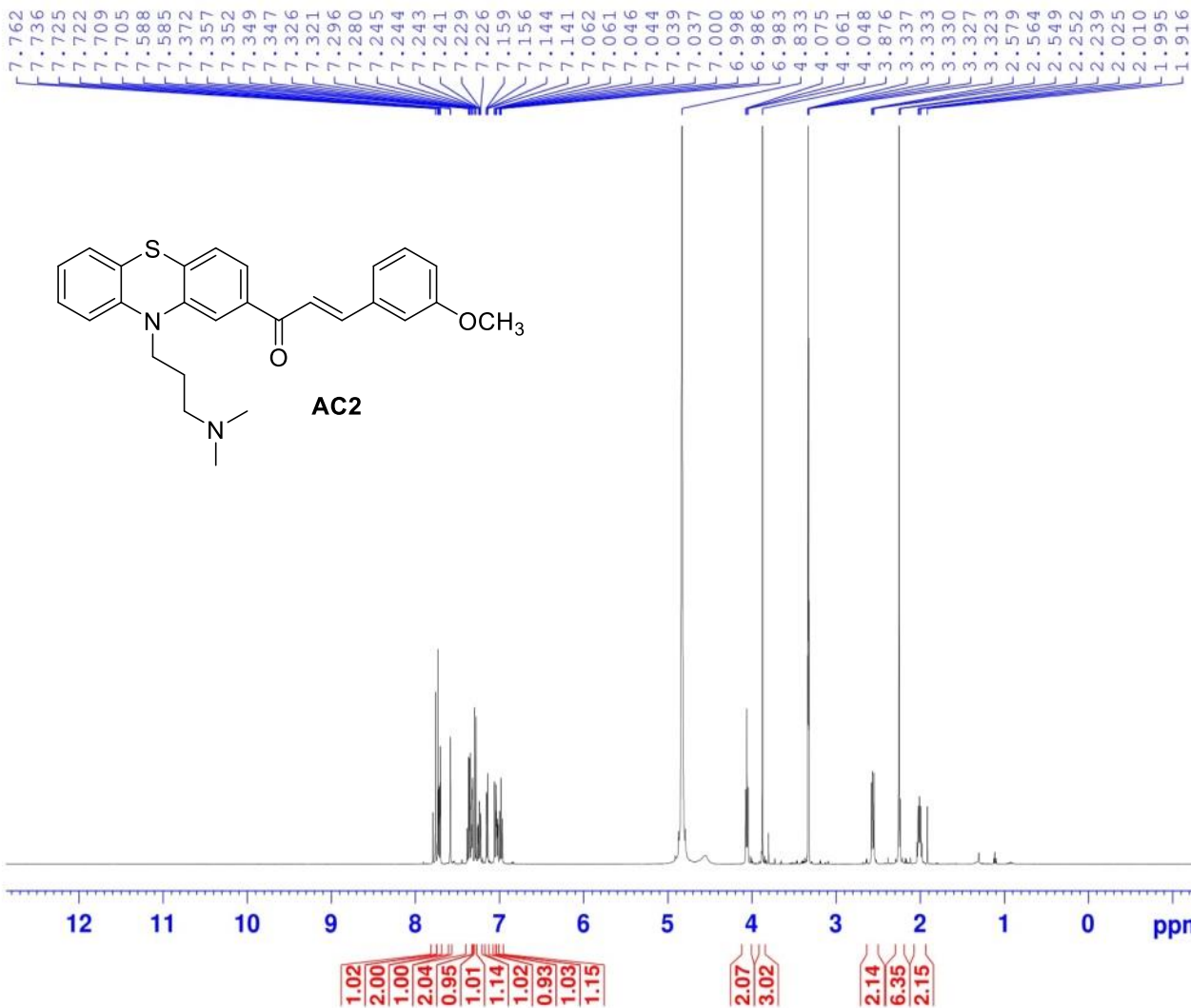
# MS

Sample Name	02-AC2	Position	P2-C6	Instrument Name	Instrument 1	User Name	
Inj Vol	2	InjPosition		SampleType	Sample	IRM Calibration Status	Success
Data Filename	02-AC2.d	ACQ Method	Cot ngan - MSMS_Pos.	Comment		Acquired Time	22/08/2020 10:13:59 AM



# <sup>1</sup>H-NMR

C3-MeOD-1H



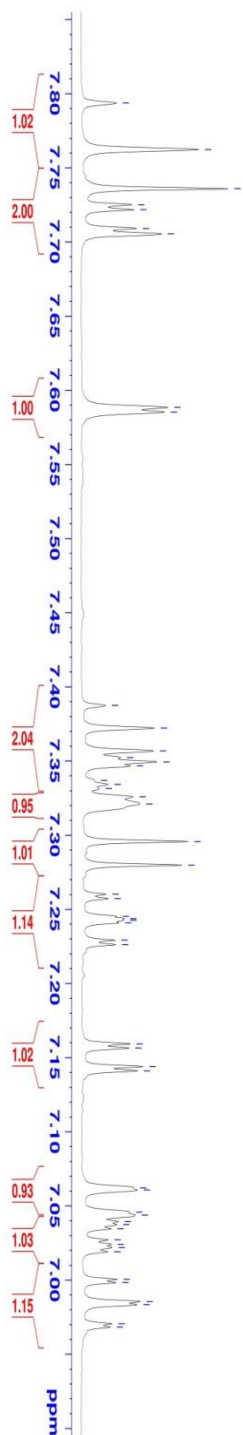
Current Data Parameters  
NAME 113D\_C3  
EXPNO 1  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20170606  
Time 16.02  
INSTRUM spect  
PROBHD 5 mm PABBO BB/  
PULPROG zg30  
TD 65536  
SOLVENT MeOD  
NS 16  
DS 2  
SWH 10000.000 Hz  
FIDRES 0.152588 Hz  
AQ 3.2767999 sec  
RG 127.68  
DW 50.000 usec  
DE 6.50 usec  
TE 303.0 K  
D1 1.00000000 sec  
TD0 1

===== CHANNEL f1 =====  
SFO1 500.2030889 MHz  
NUC1 1H  
P1 10.00 usec  
PLW1 22.00000000 W

F2 - Processing parameters  
SI 65536  
SF 500.2000000 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

# <sup>1</sup>H-NMR

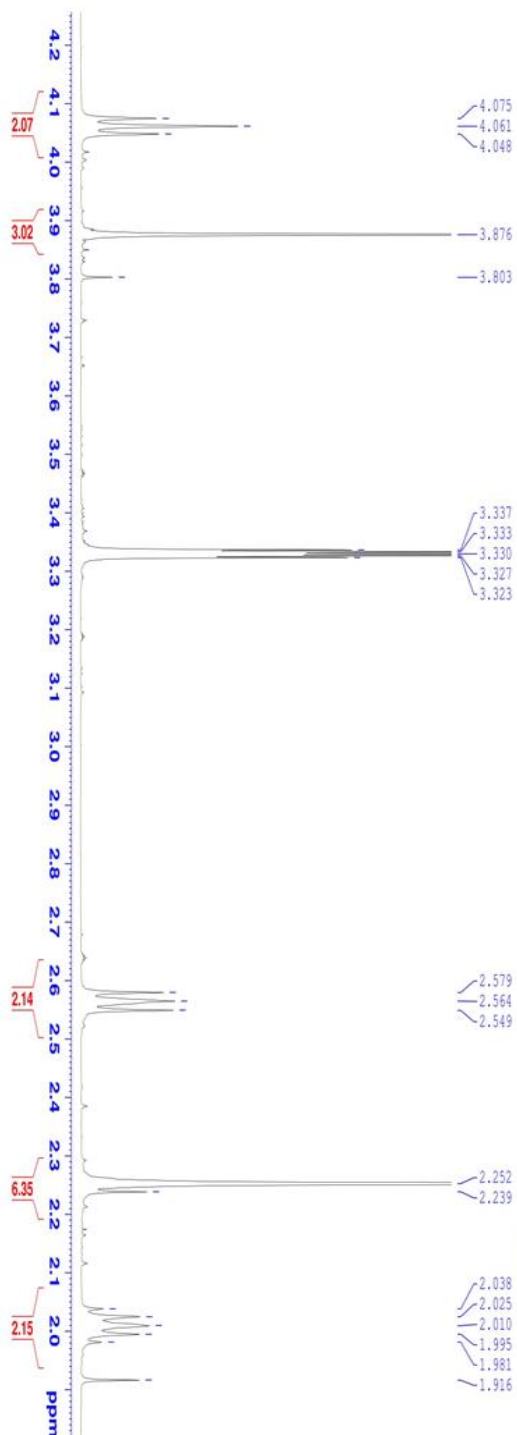


7.794  
7.762  
7.736  
7.725  
7.722  
7.709  
7.705

7.588  
7.585

7.387  
7.372  
7.357  
7.352  
7.349  
7.347  
7.337  
7.334  
7.331  
7.326  
7.321  
7.296  
7.280  
7.260  
7.257  
7.245  
7.244  
7.243  
7.241  
7.229  
7.226  
7.159  
7.156  
7.144  
7.141  
7.062  
7.061  
7.046  
7.044  
7.039  
7.037  
7.035  
7.027  
7.024  
7.022  
7.019  
7.000  
6.998  
6.986  
6.983

**C3-MeOD-1H**



4.075  
4.061  
4.048

3.876  
3.803

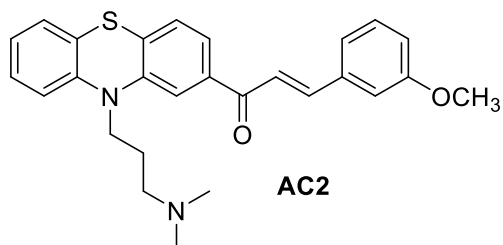
3.337  
3.333  
3.330  
3.327  
3.323

2.579  
2.564  
2.549

2.252  
2.239

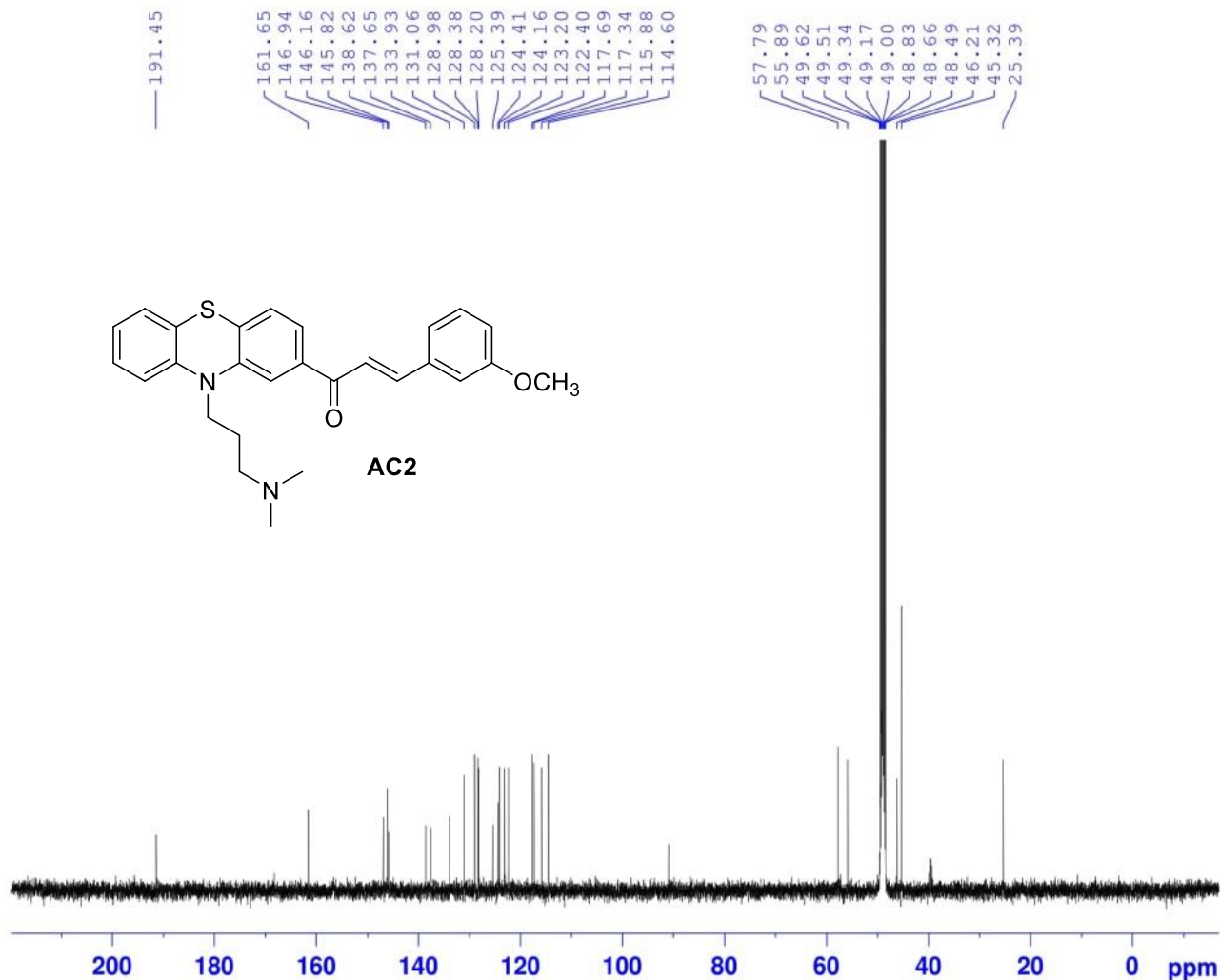
2.038  
2.025  
2.010  
1.995  
1.981  
1.916

**C3-MeOD-1H**



# <sup>13</sup>C-NMR

C3-MeOD-C13CPD



Current Data Parameters  
NAME 113D\_C3  
EXPNO 2  
PROCNO 1

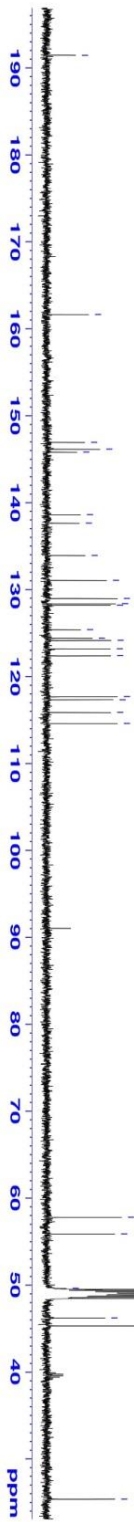
F2 - Acquisition Parameters  
Date\_ 20170606  
Time\_ 17.57  
INSTRUM spect  
PROBHD 5 mm PABBO BB/  
PULPROG zgpg30  
TD 65536  
SOLVENT MeOD  
NS 2048  
DS 4  
SWH 29761.904 Hz  
FIDRES 0.454131 Hz  
AQ 1.1010048 sec  
RG 198.57  
DW 16.800 usec  
DE 6.50 usec  
TE 303.0 K  
D1 2.00000000 sec  
D11 0.03000000 sec  
TD0 1

----- CHANNEL f1 -----  
SFO1 125.7879670 MHz  
NUC1 13C  
P1 10.00 usec  
PLW1 88.00000000 W

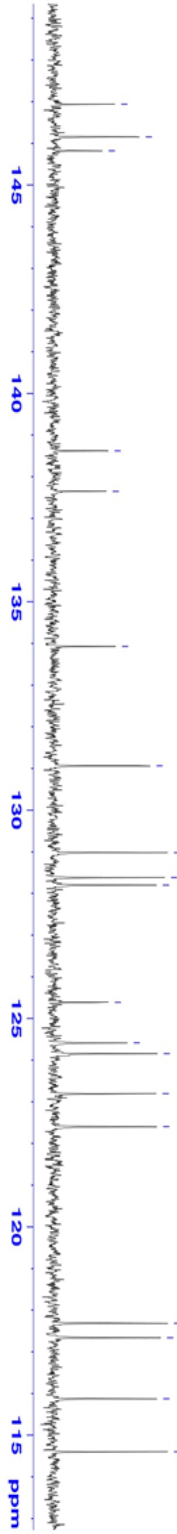
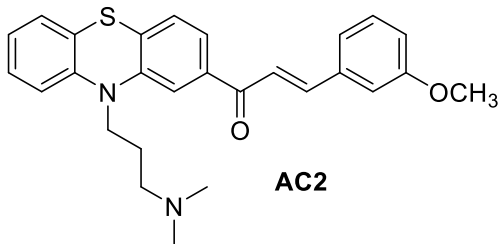
----- CHANNEL f2 -----  
SFO2 500.2020008 MHz  
NUC2 1H  
CPDPRG[2] waltz16  
PCPD2 80.00 usec  
PLW2 22.00000000 W  
PLW12 0.34375000 W  
PLW13 0.22000000 W

F2 - Processing parameters  
SI 32768  
SF 125.7753900 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40

# $^{13}\text{C}$ -NMR



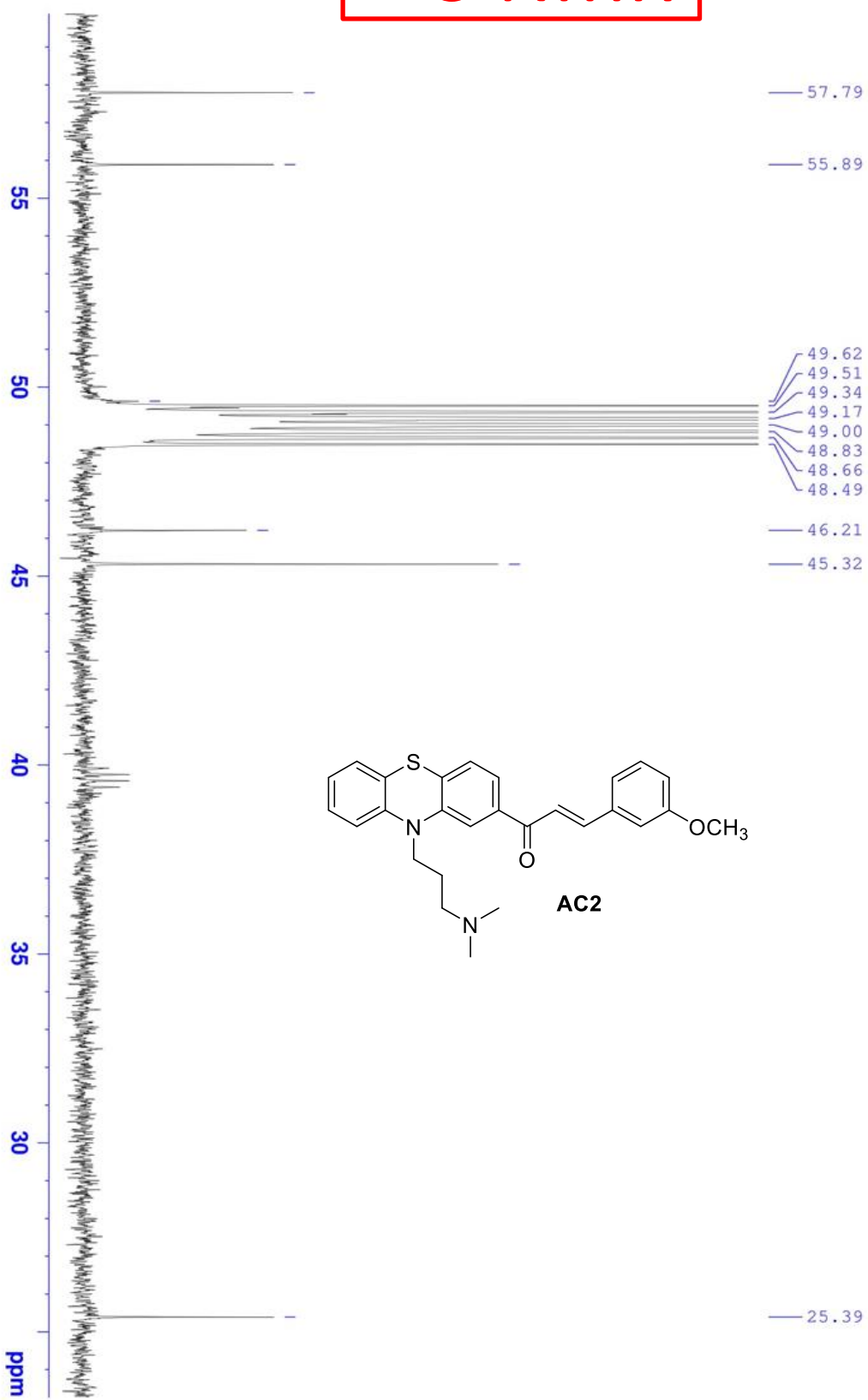
C3-MeOD-C13CPD



C3-MeOD-C13CPD

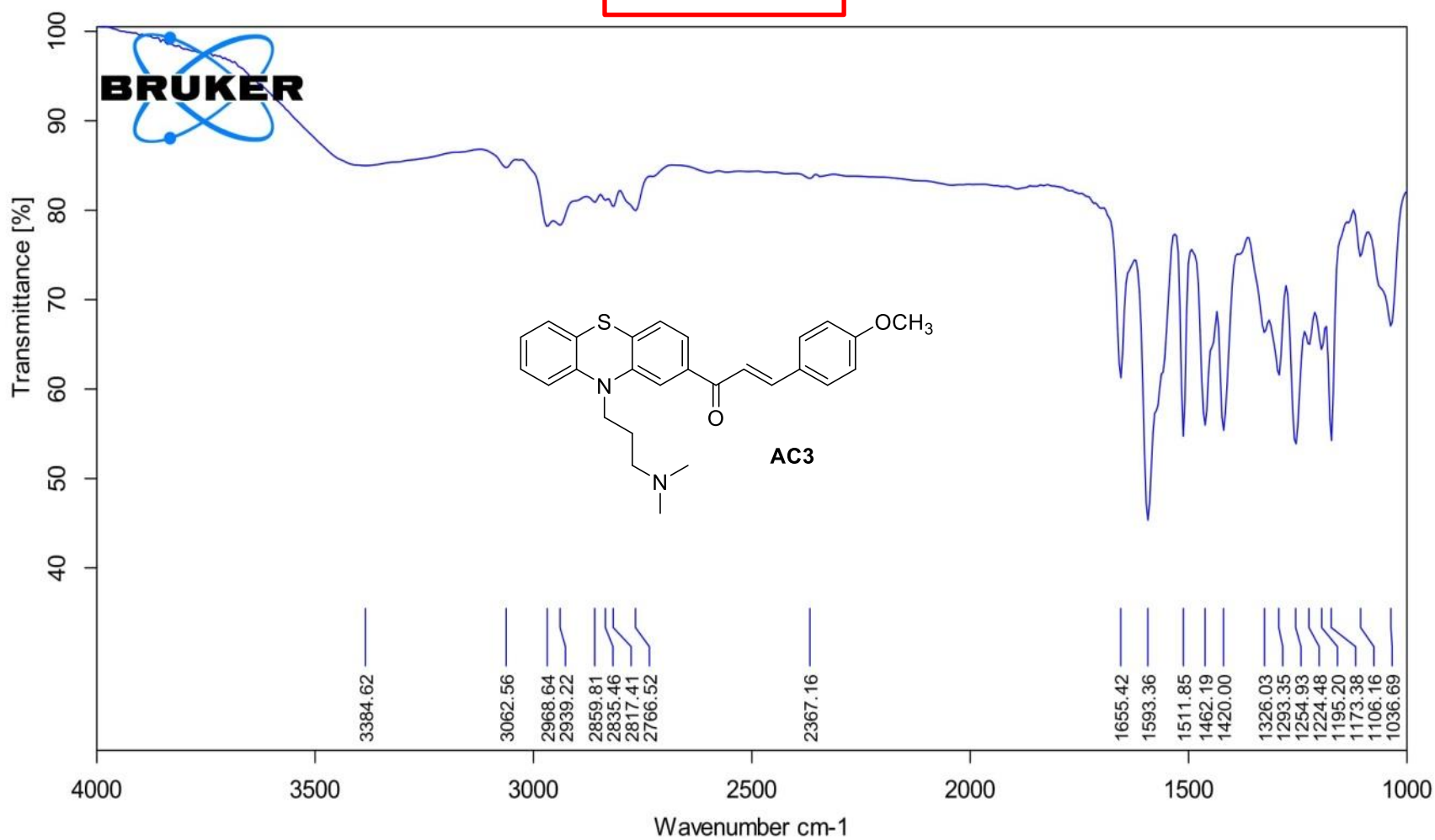


# $^{13}\text{C}$ -NMR



C3-MeOD-C13CPD





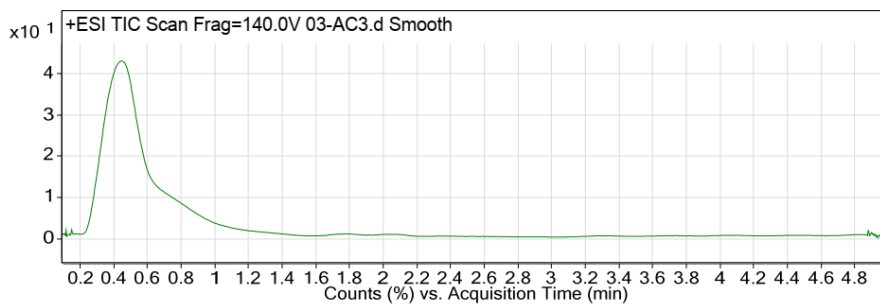
# MS

## Qualitative Analysis Report

<b>Data Filename</b>	03-AC3.d	<b>Sample Name</b>	03-AC3
<b>Sample Type</b>	Sample	<b>Position</b>	P2-C4
<b>Instrument Name</b>	Instrument 1	<b>User Name</b>	
<b>Acq Method</b>	Cot ngan - MSMS_Pos.m	<b>Acquired Time</b>	22/08/2020 10:21:51 AM
<b>IRM Calibration Status</b>	Success	<b>DA Method</b>	COTNGAN.M.m
<b>Comment</b>			
<b>Sample Group</b>		<b>Info.</b>	
<b>Stream Name</b>	LC 1	<b>Acquisition SW</b>	6200 series TOF/6500 series
		<b>Version</b>	Q-TOF B.06.01 (B6172 SP1)

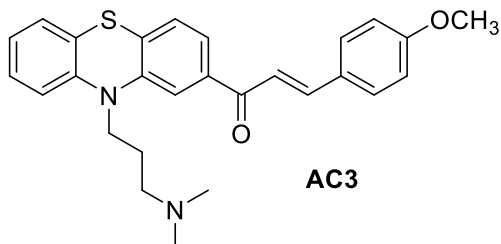
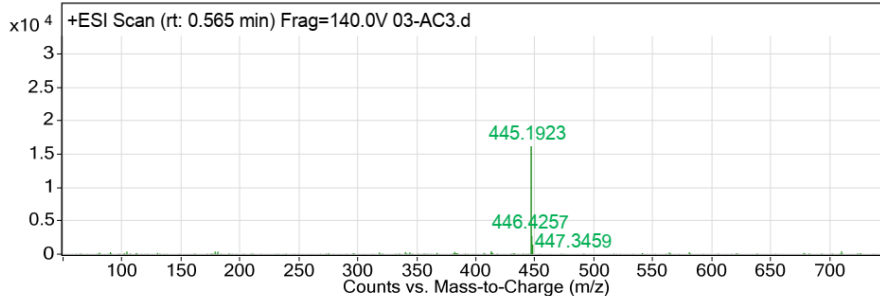
### User Chromatograms

Fragmentor Voltage 140 Collision Energy 0 Ionization Mode ESI



### User Spectra

Fragmentor Voltage 140 Collision Energy 0 Ionization Mode ESI

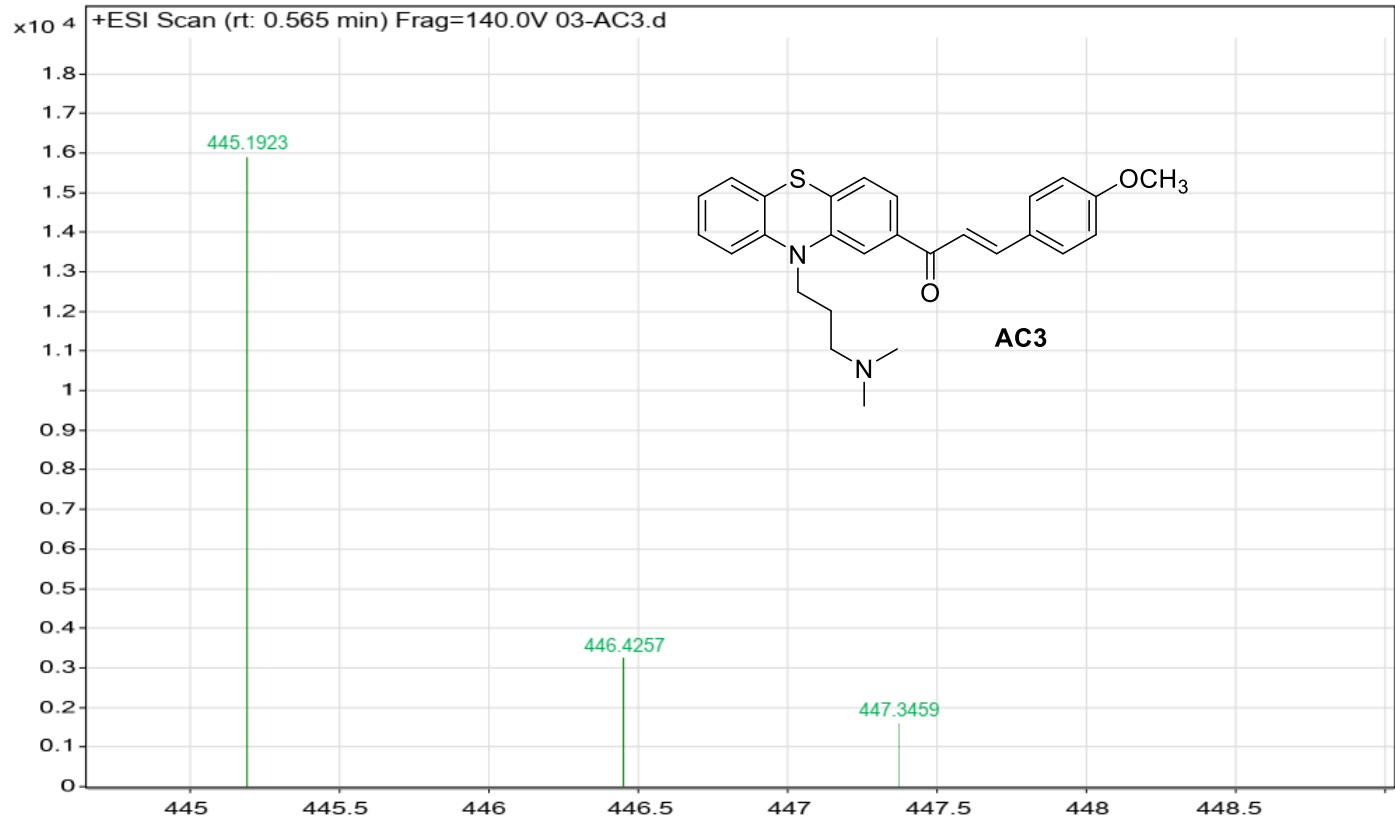


--- End Of Report ---



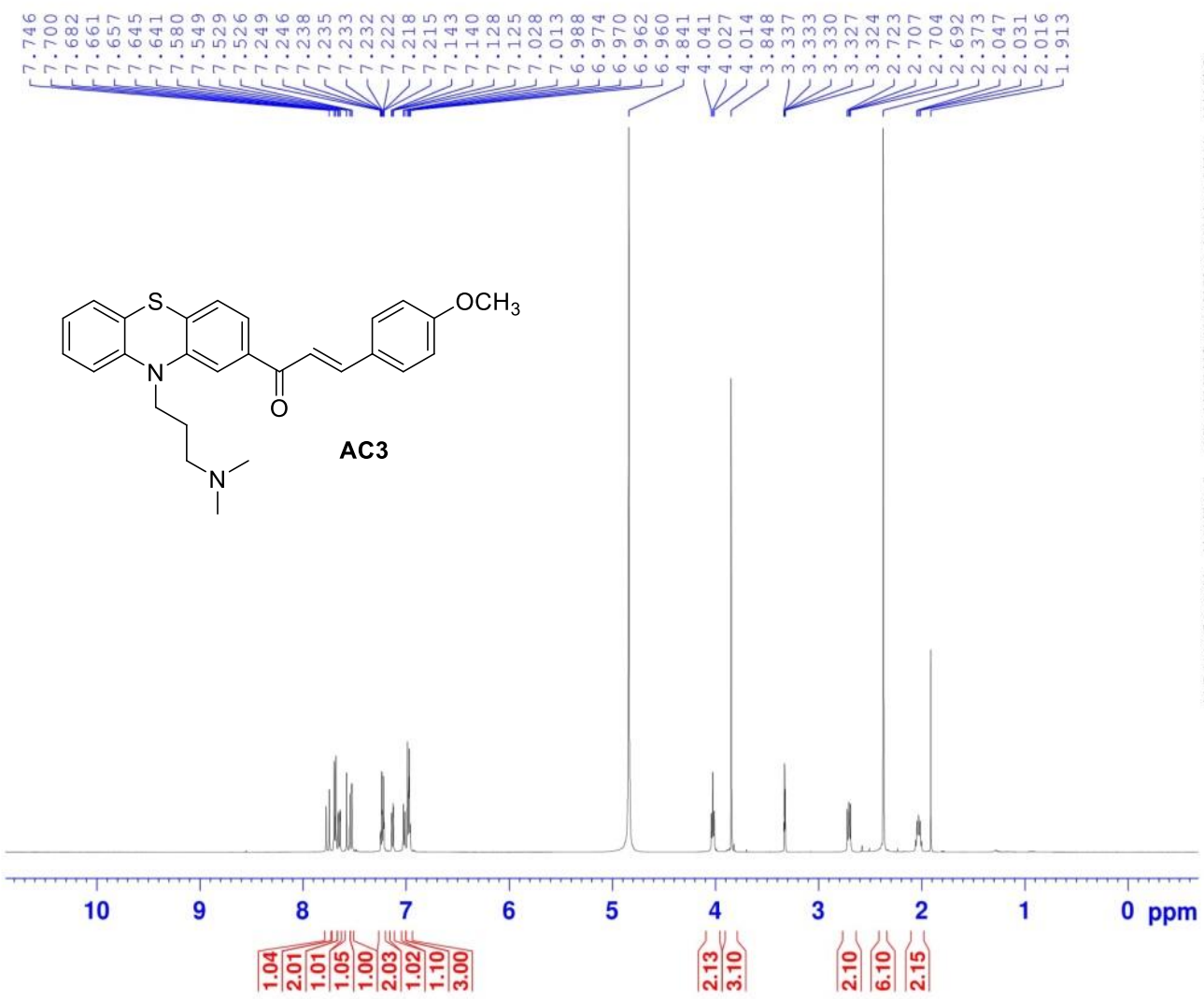
# MS

Sample Name	03-AC3	Position	P2-C4	Instrument Name	Instrument 1	User Name	
Inj Vol	2	InjPosition		SampleType	Sample	IRM Calibration Status	Success
Data Filename	03-AC3.d	ACQ Method	Cot ngan - MSMS_Pos.	Comment		Acquired Time	22/08/2020 10:21:51 PM



# <sup>1</sup>H-NMR

C2-MeOD-1H



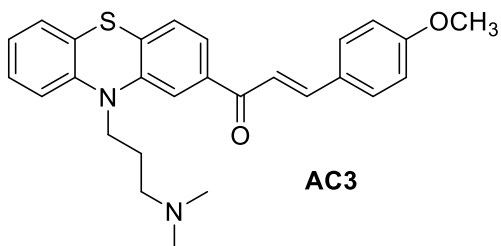
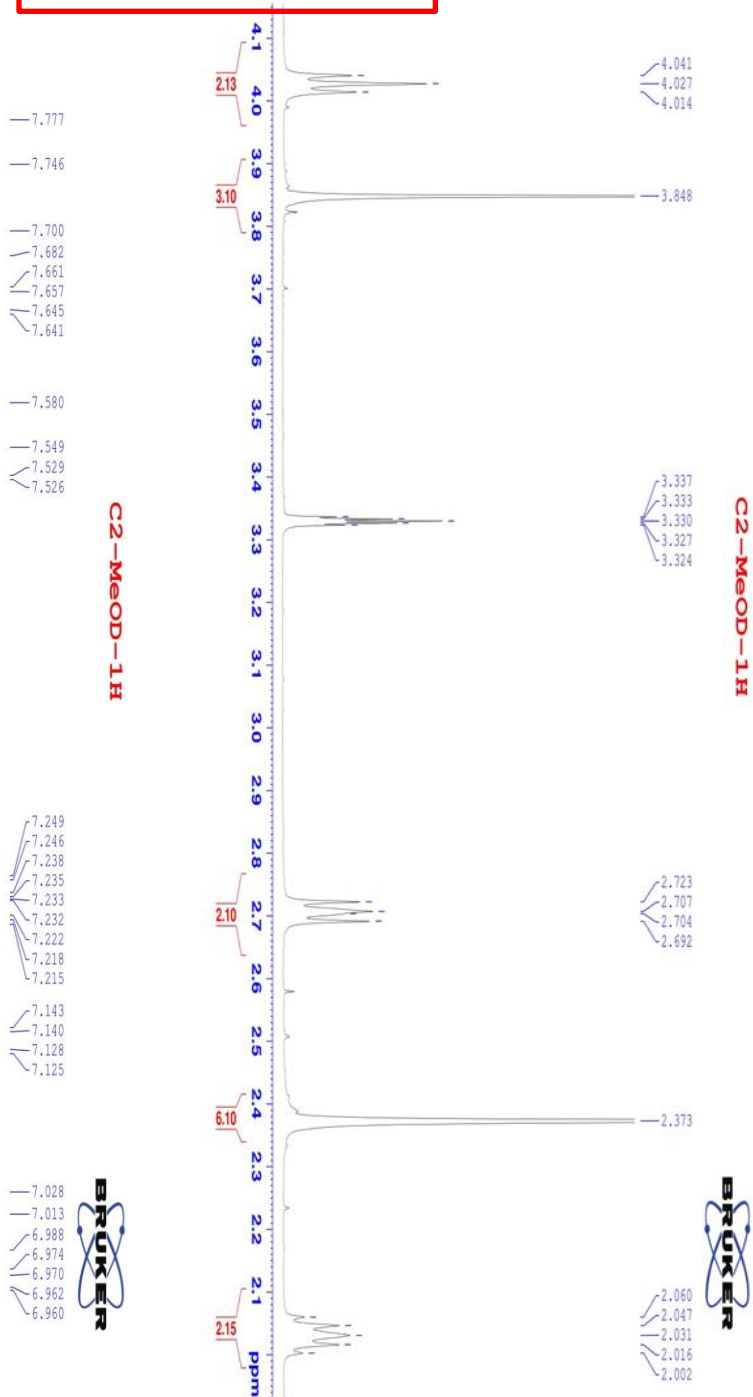
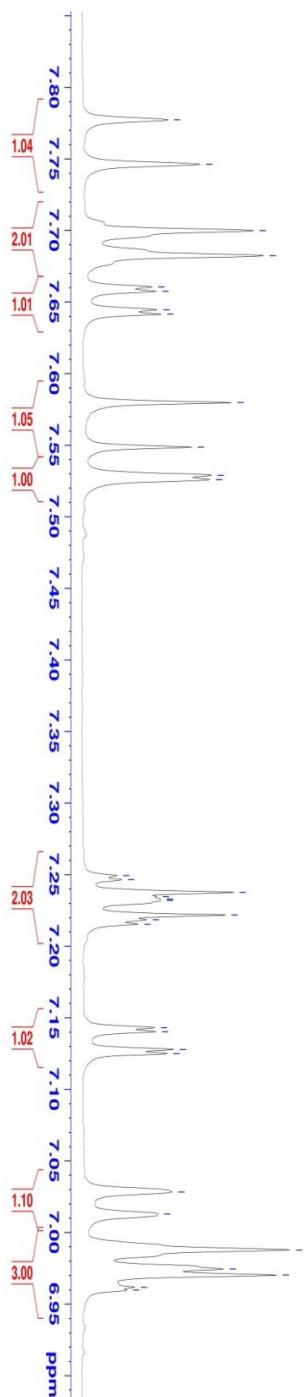
```
Current Data Parameters
NAME          113D_C2
EXPNO         1
PROCNO        1

F2 - Acquisition Parameters
Date_         20170606
Time          15.13
INSTRUM       spect
PROBHD        5 mm PABBO BB/
PULPROG       zg30
TD            65536
SOLVENT       MeOD
NS            16
DS            2
SWH           10000.000 Hz
FIDRES        0.152588 Hz
AQ            3.2767999 sec
RG            79.36
DW            50.000 usec
DE            6.50 usec
TE            303.0 K
D1            1.00000000 sec
TD0           1

===== CHANNEL f1 =====
SFO1          500.2030889 MHz
NUC1           1H
P1            10.00 usec
PLW1          22.00000000 W

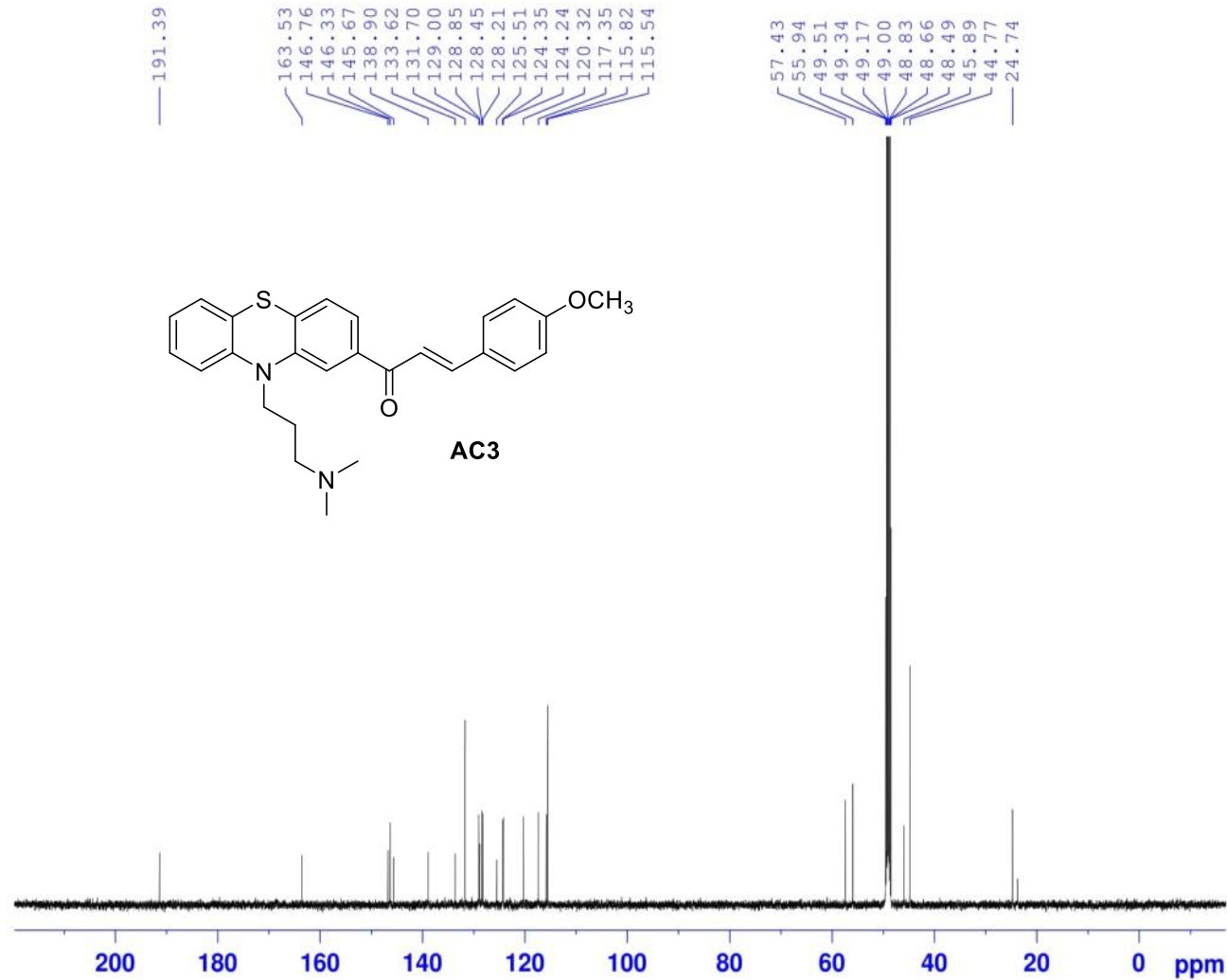
F2 - Processing parameters
SI            65536
SF            500.2000002 MHz
WDW           EM
SSB           0
LB            0.30 Hz
GB            0
PC            1.00
```

# <sup>1</sup>H-NMR



# $^{13}\text{C}$ -NMR

C2-MeOD-C13CPD



Current Data Parameters  
NAME 113D\_C2  
EXPNO 2  
PROCNO 1

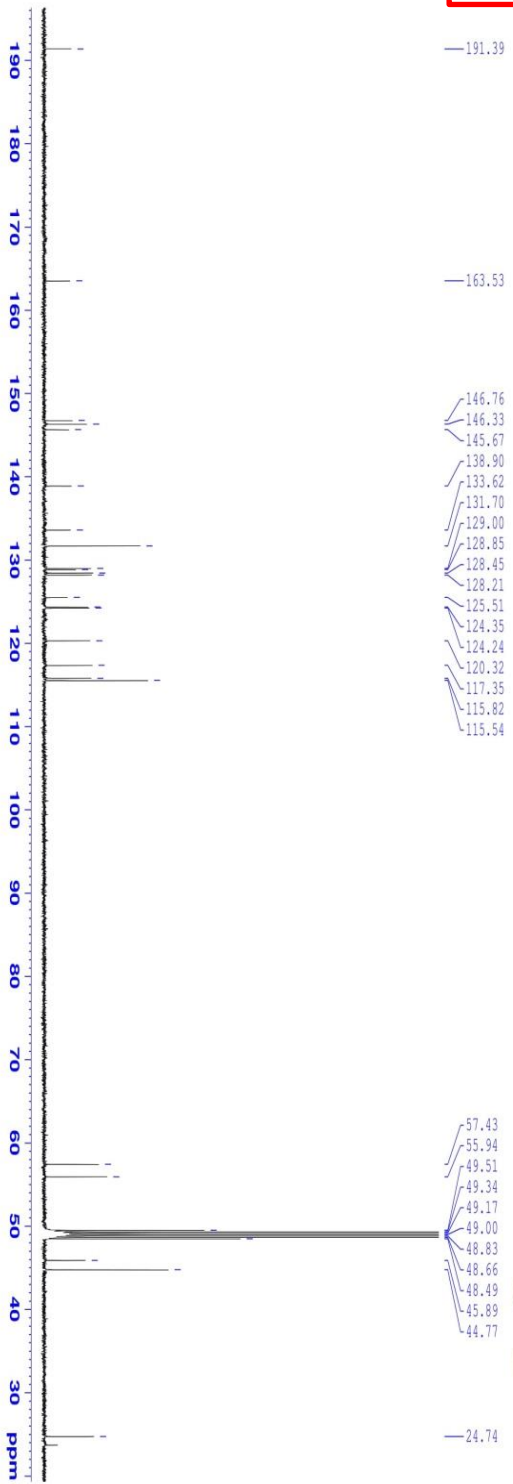
F2 - Acquisition Parameters  
Date\_ 20170606  
Time 15.57  
INSTRUM spect  
PROBHD 5 mm PABBO BB/  
PULPROG zgpg30  
TD 65536  
SOLVENT MeOD  
NS 256  
DS 4  
SWH 29761.904 Hz  
FIDRES 0.454131 Hz  
AQ 1.1010048 sec  
RG 198.57  
DW 16.800 usec  
DE 6.50 usec  
TE 303.0 K  
D1 2.00000000 sec  
D11 0.03000000 sec  
TD0 1

===== CHANNEL f1 =====  
SFO1 125.7879670 MHz  
NUC1 13C  
P1 10.00 usec  
PLW1 88.00000000 W

===== CHANNEL f2 =====  
SFO2 500.2020008 MHz  
NUC2 1H  
CPDPRG[2] waltz16  
PCPD2 80.00 usec  
PLW2 22.00000000 W  
PLW12 0.34375000 W  
PLW13 0.22000000 W

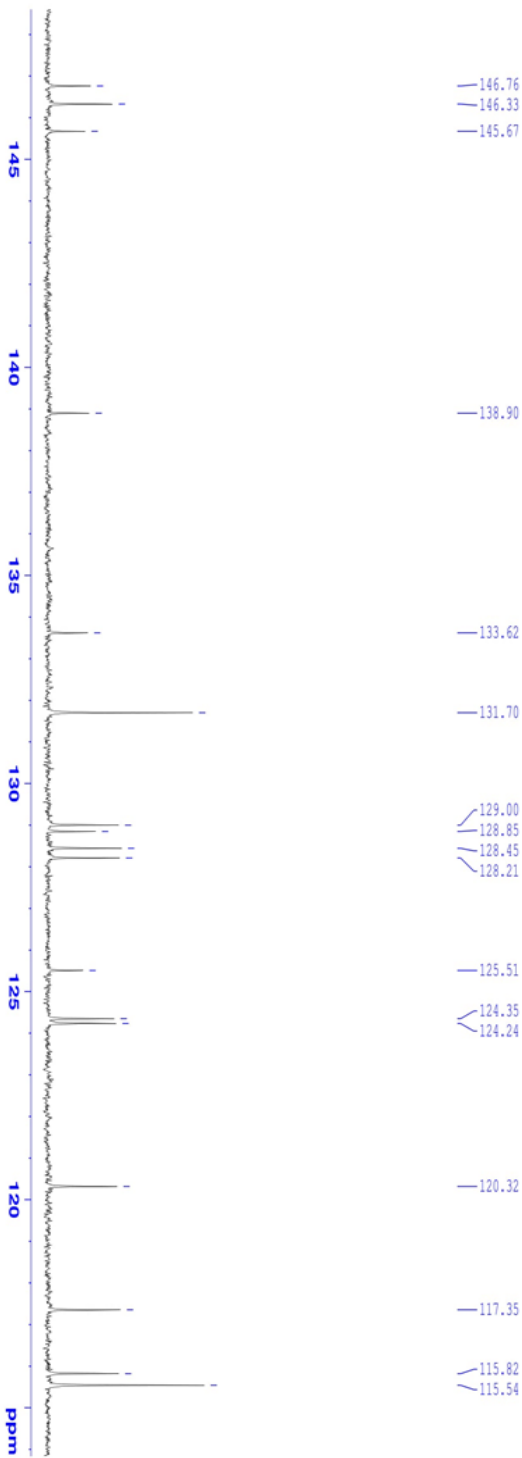
F2 - Processing parameters  
SI 32768  
SF 125.7753900 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40

# $^{13}\text{C}$ -NMR



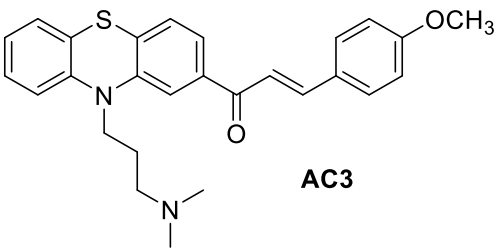
C2-MeOD-C13CPD

BRUKER

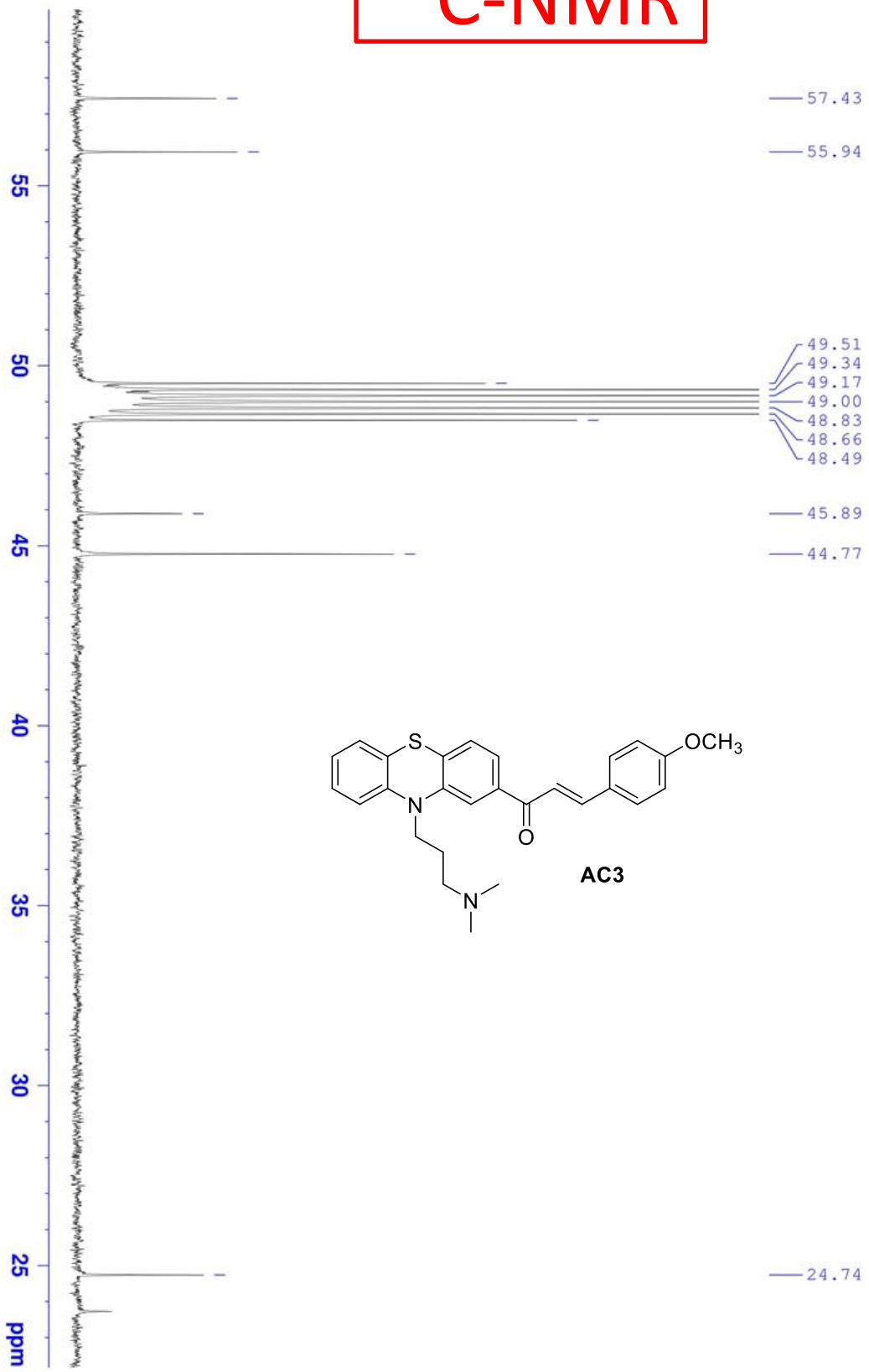


C2-MeOD-C13CPD

BRUKER



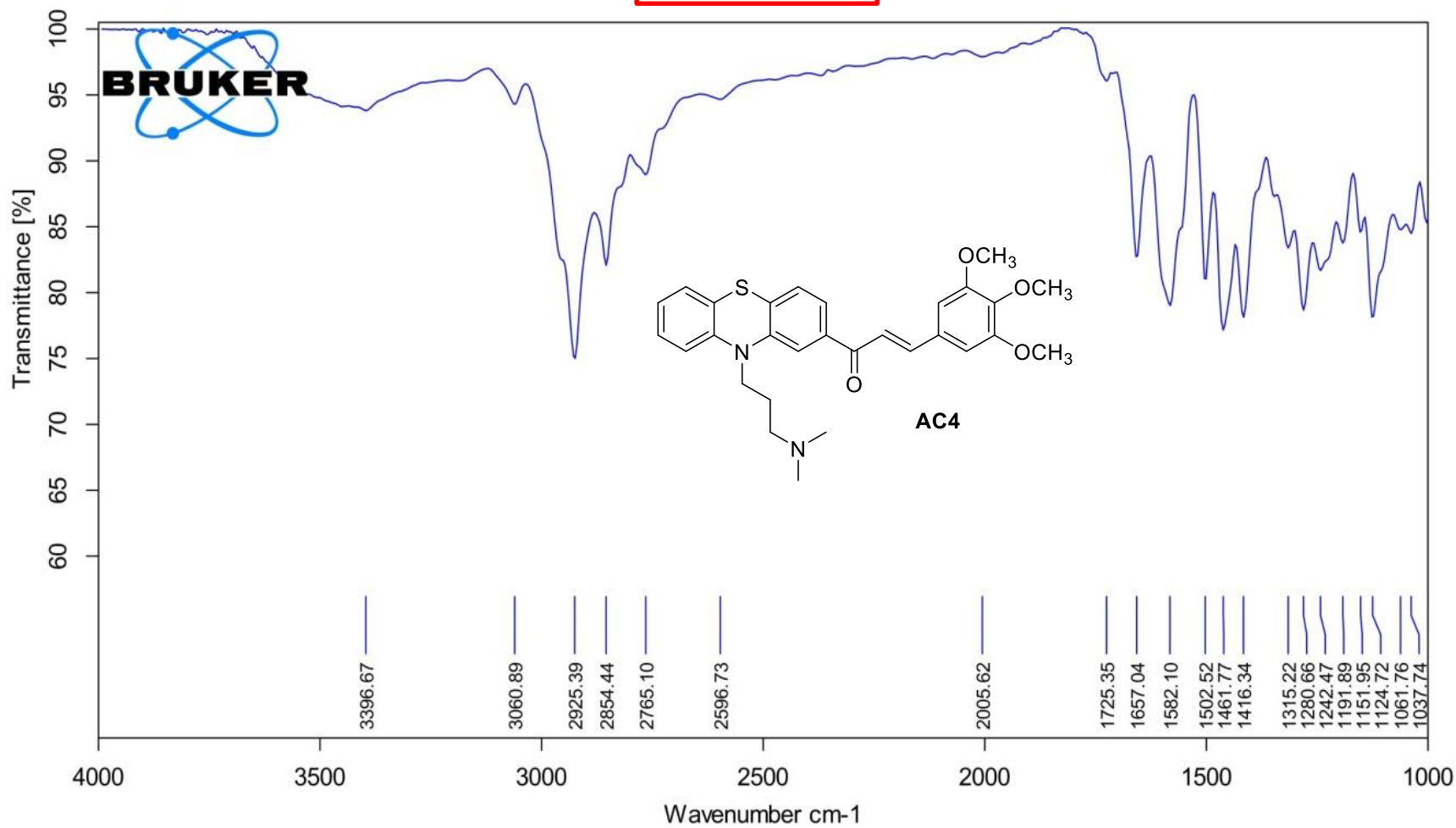
# $^{13}\text{C-NMR}$



**CD-MeOD-Cl3CPD**



IR



E:\OPUS 7\2017\THANG 6\20170606\C4.0	C4	SENSOR 27 - BRUKER - GERMANY	6/6/2017
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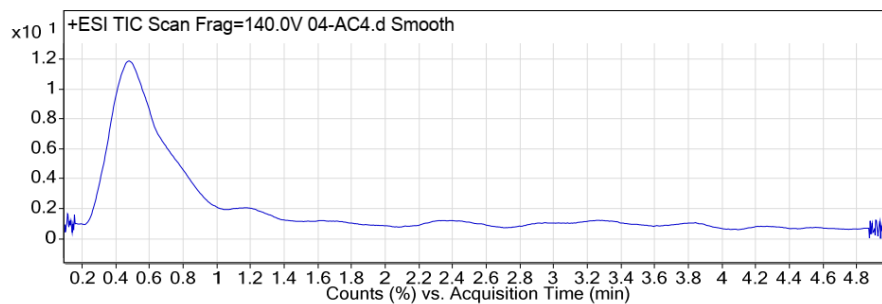
# MS

## Qualitative Analysis Report

<b>Data Filename</b>	04-AC4.d	<b>Sample Name</b>	04-AC4
<b>Sample Type</b>	Sample	<b>Position</b>	P2-B5
<b>Instrument Name</b>	Instrument 1	<b>User Name</b>	
<b>Acq Method</b>	Cot ngan - MSMS_Pos.m	<b>Acquired Time</b>	22/08/2020 10:30:20 AM
<b>IRM Calibration Status</b>	Success	<b>DA Method</b>	COTNGAN.M.m
<b>Comment</b>			
<b>Sample Group</b>		<b>Info.</b>	
<b>Stream Name</b>	LC 1	<b>Acquisition SW</b>	6200 series TOF/6500 series
		<b>Version</b>	Q-TOF B.06.01 (B6172 SP1)

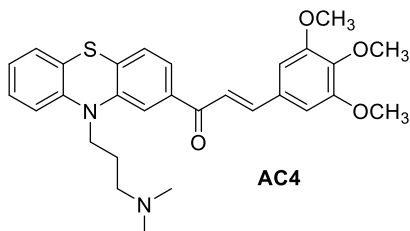
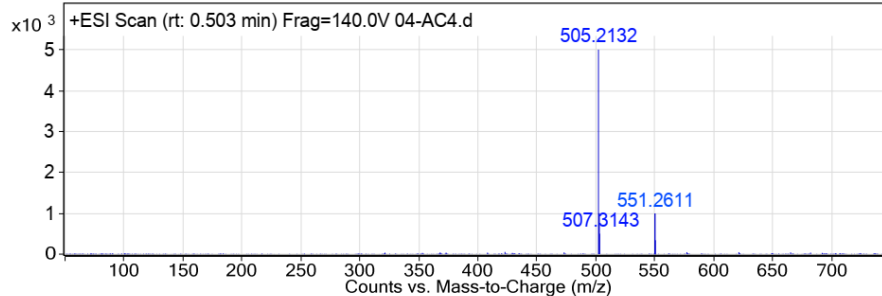
### User Chromatograms

Fragmentor Voltage 140 Collision Energy 0 Ionization Mode ESI



### User Spectra

Fragmentor Voltage 140 Collision Energy 0 Ionization Mode ESI

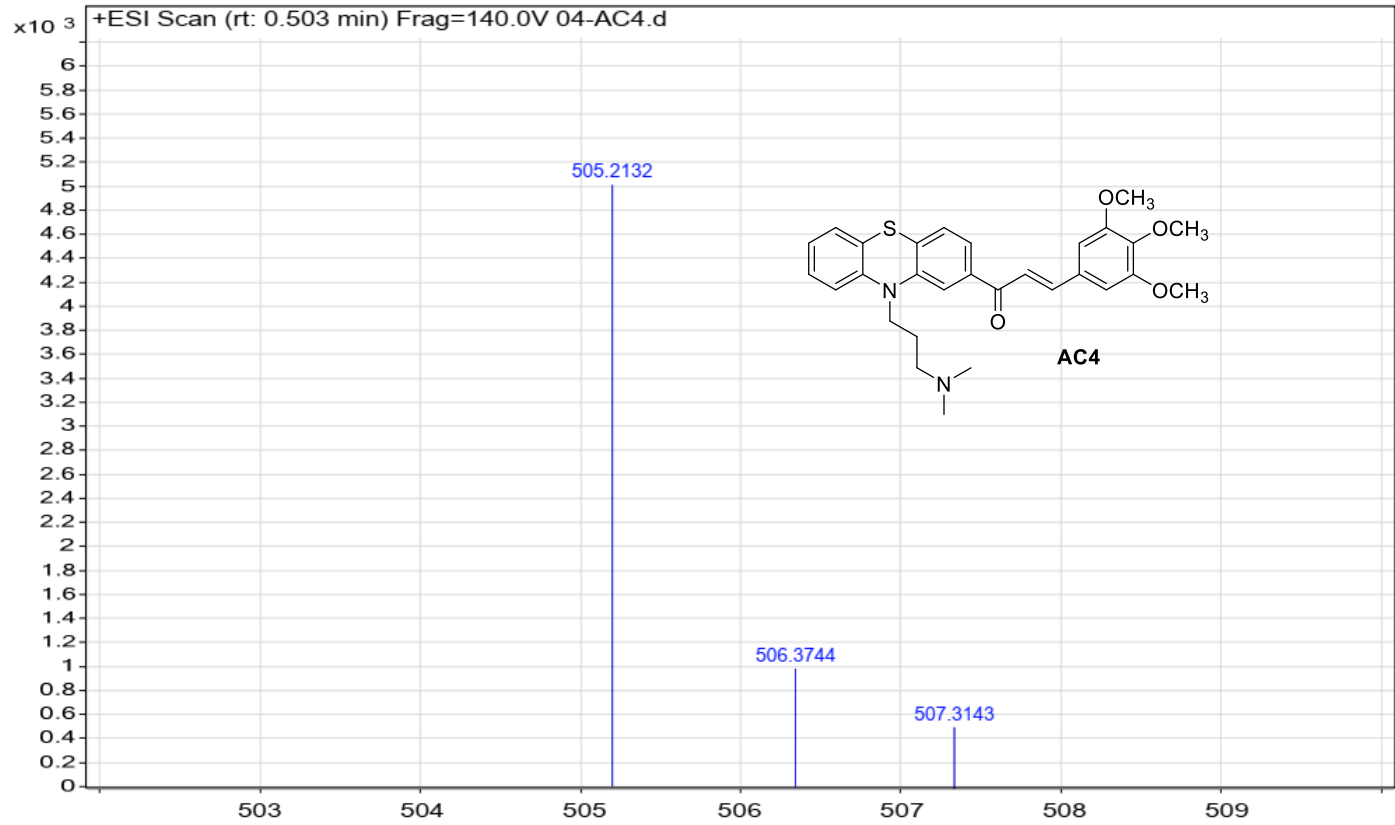


--- End Of Report ---



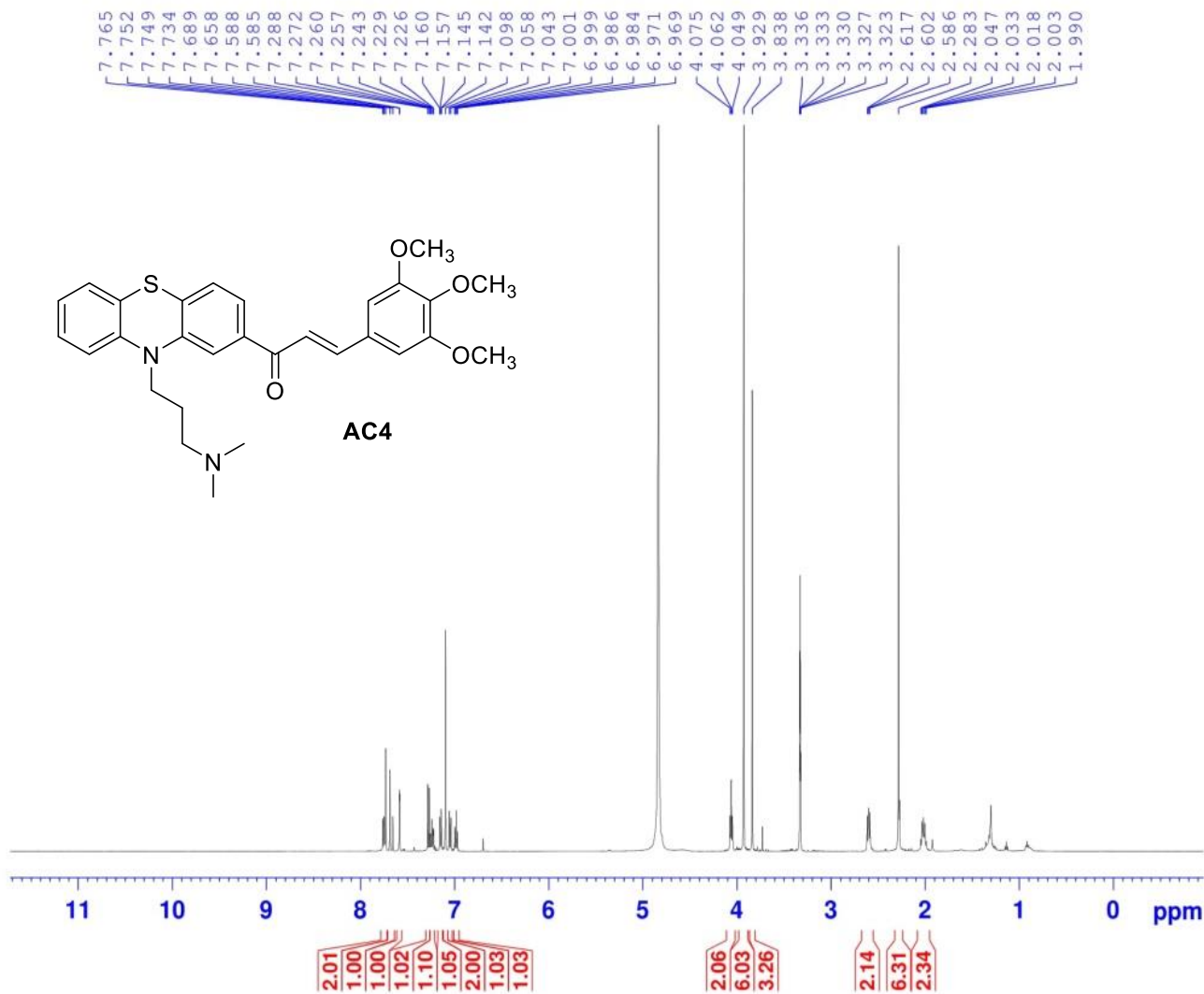
# MS

Sample Name	04-AC4	Position	P2-B5	Instrument Name	Instrument 1	User Name	
Inj Vol	2	InjPosition		SampleType	Sample	IRM Calibration Status	Success
Data Filename	04-AC4.d	ACQ Method	Cot ngan - MSMS_Pos.	Comment		Acquired Time	22/08/2020 10:30:20 AM



# <sup>1</sup>H-NMR

C4-MeOD-1H



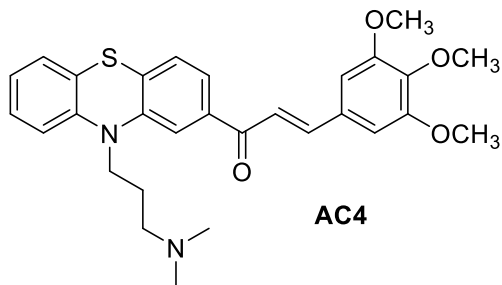
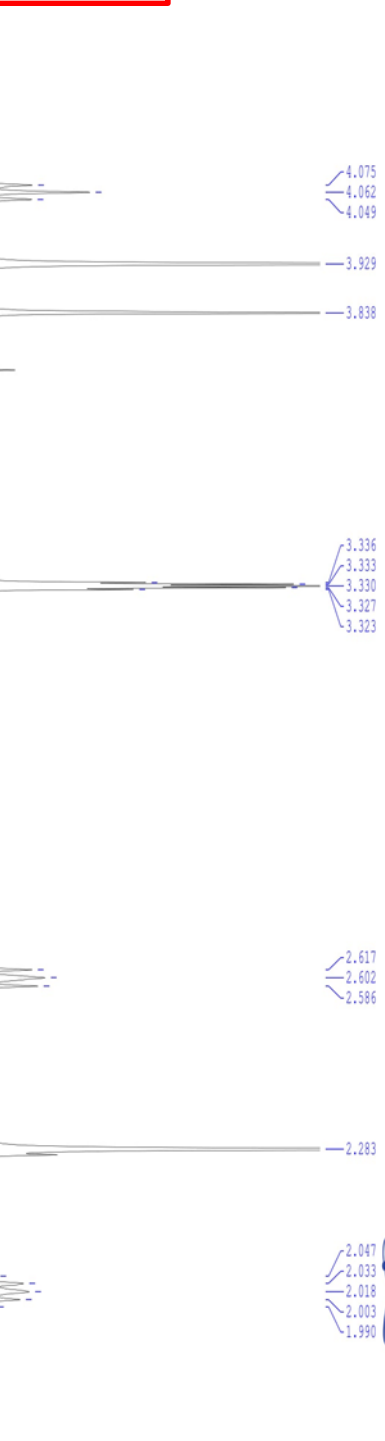
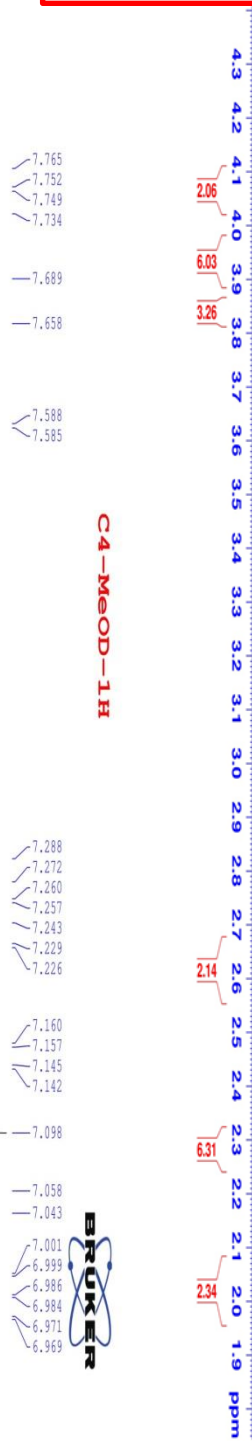
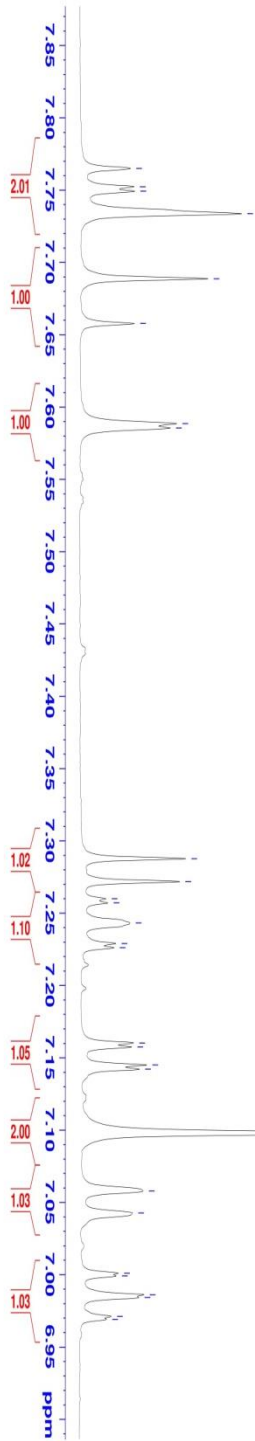
Current Data Parameters  
NAME 113D\_C4  
EXPNO 1  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20170606  
Time 16.07  
INSTRUM spect  
PROBHD 5 mm PABBO BB/  
PULPROG zg30  
TD 65536  
SOLVENT MeOD  
NS 16  
DS 2  
SWH 10000.000 Hz  
FIDRES 0.152588 Hz  
AQ 3.2767999 sec  
RG 127.68  
DW 50.000 usec  
DE 6.50 usec  
TE 303.0 K  
D1 1.00000000 sec  
TD0 1

==== CHANNEL f1 =====  
SFO1 500.2030889 MHz  
NUC1 1H  
P1 10.00 usec  
PLW1 22.00000000 W

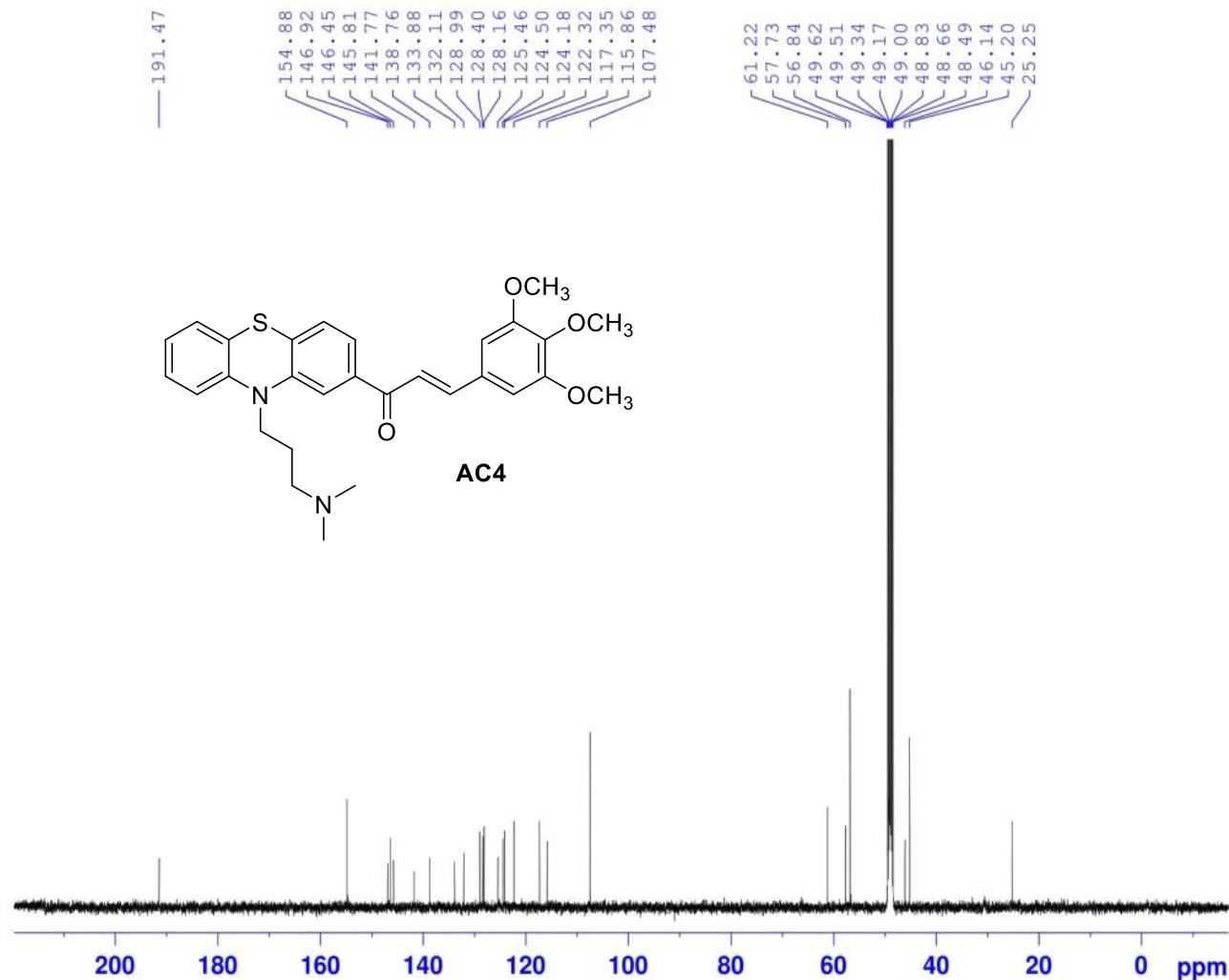
F2 - Processing parameters  
SI 65536  
SF 500.2000001 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

# <sup>1</sup>H-NMR



# $^{13}\text{C}$ -NMR

C4-MeOD-C13CPD



Current Data Parameters  
NAME 113D\_C4  
EXPNO 2  
PROCNO 1

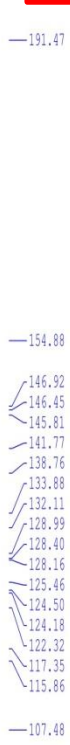
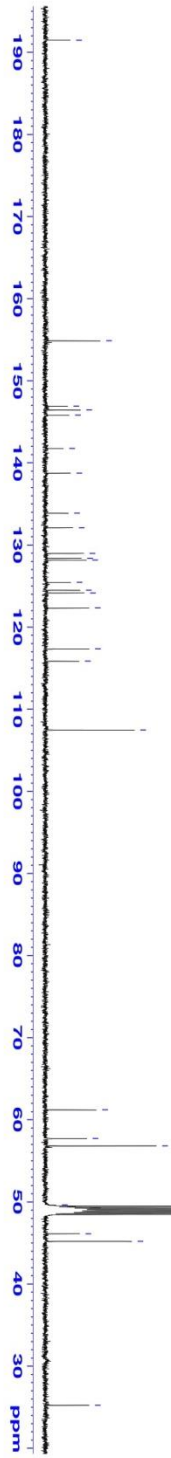
F2 - Acquisition Parameters  
Date\_ 20170606  
Time 19.38  
INSTRUM spect  
PROBHD 5 mm PABBO BB/  
PULPROG zgpg30  
TD 65536  
SOLVENT MeOD  
NS 2048  
DS 4  
SWH 29761.904 Hz  
FIDRES 0.454131 Hz  
AQ 1.1010048 sec  
RG 198.57  
DW 16.800 usec  
DE 6.50 usec  
TE 303.0 K  
D1 2.00000000 sec  
D11 0.03000000 sec  
TD0 1

----- CHANNEL f1 -----  
SFO1 125.7879670 MHz  
NUC1 13C  
P1 10.00 usec  
PLW1 88.00000000 W

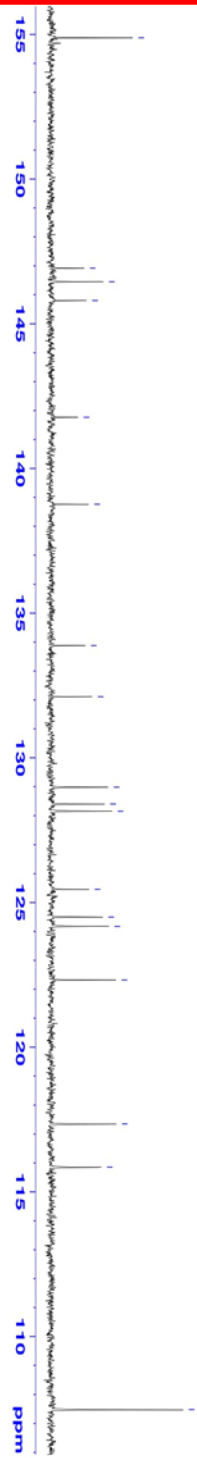
----- CHANNEL f2 -----  
SFO2 500.2020008 MHz  
NUC2 1H  
CPDPRG[2] waltz16  
PCPD2 80.00 usec  
PLW2 22.00000000 W  
PLW12 0.34375000 W  
PLW13 0.22000000 W

F2 - Processing parameters  
SI 32768  
SF 125.7752144 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40

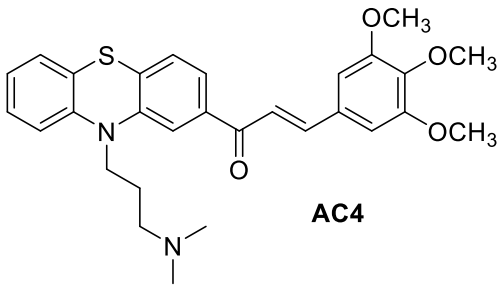
# $^{13}\text{C}$ -NMR



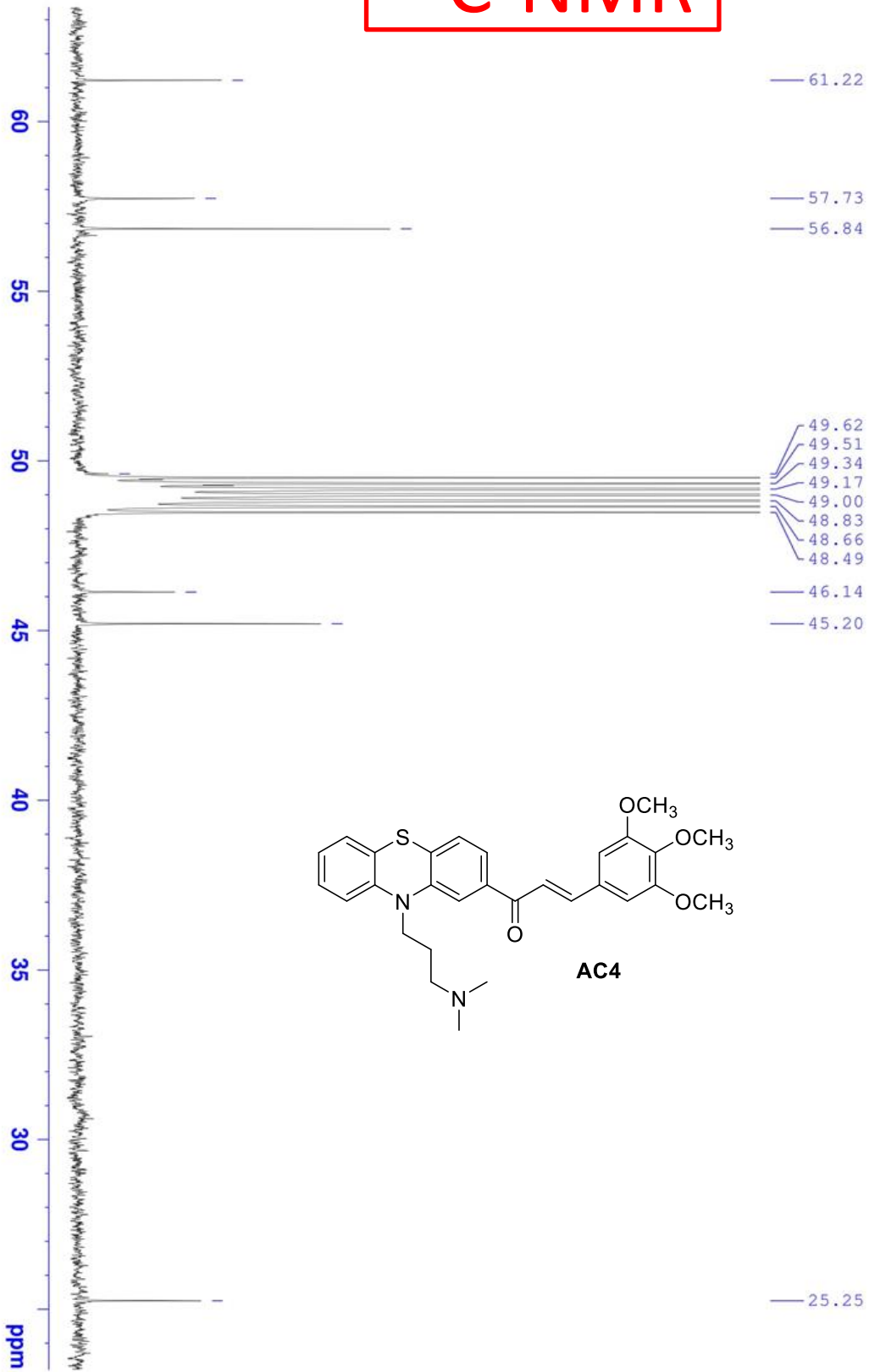
**C4-MeOD-C13CPD**



**C4-MeOD-C13CPD**



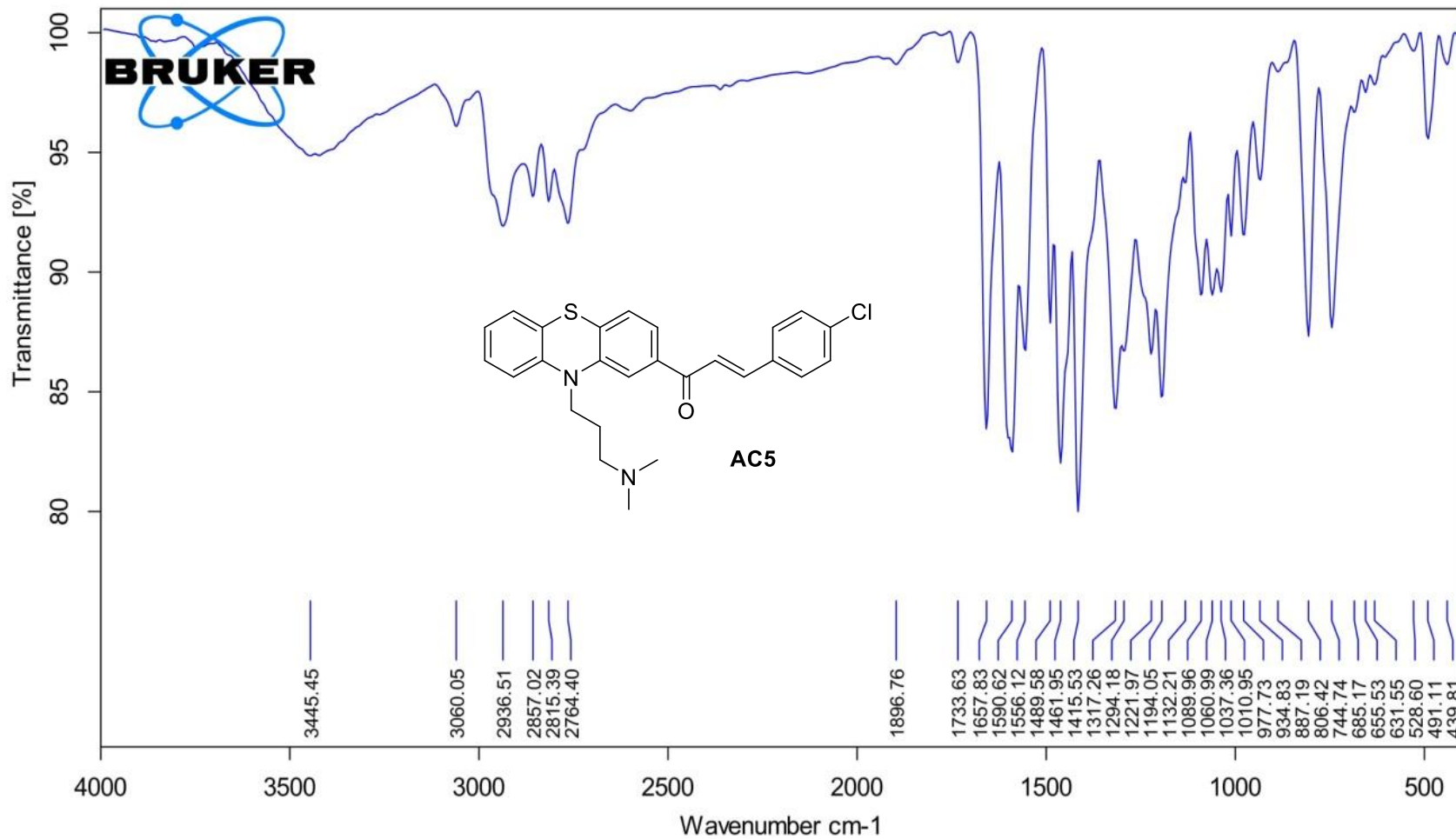
# $^{13}\text{C}$ -NMR



C4-MeOD-C13CPD

BRUKER

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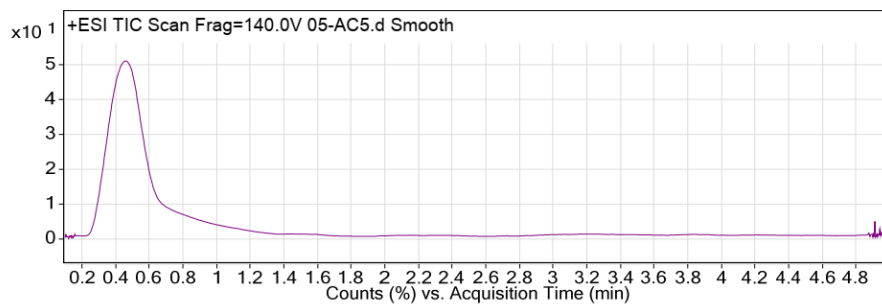
# MS

## Qualitative Analysis Report

<b>Data Filename</b>	05-AC5.d	<b>Sample Name</b>	05-AC5
<b>Sample Type</b>	Sample	<b>Position</b>	P2-B4
<b>Instrument Name</b>	Instrument 1	<b>User Name</b>	
<b>Acq Method</b>	Cot ngan - MSMS_Pos.m	<b>Acquired Time</b>	22/08/2020 10:38:45 AM
<b>IRM Calibration Status</b>	Success	<b>DA Method</b>	COTNGAN.M.m
<b>Comment</b>			
<b>Sample Group</b>		<b>Info.</b>	
<b>Stream Name</b>	LC 1	<b>Acquisition SW</b>	6200 series TOF/6500 series
		<b>Version</b>	Q-TOF B.06.01 (B6172 SP1)

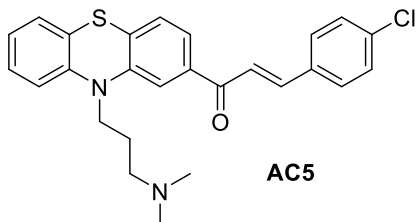
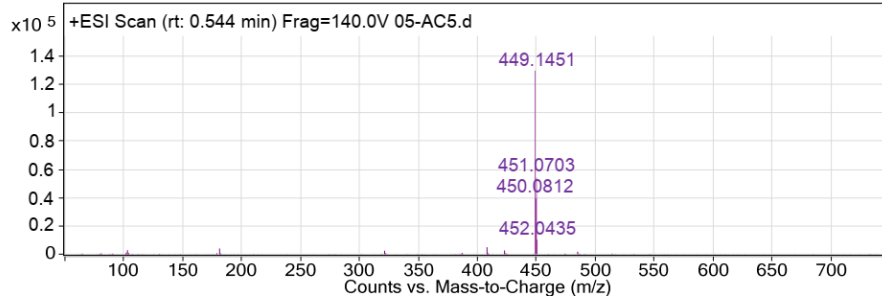
### User Chromatograms

Fragmentor Voltage 140 Collision Energy 0 Ionization Mode ESI



### User Spectra

Fragmentor Voltage 140 Collision Energy 0 Ionization Mode ESI

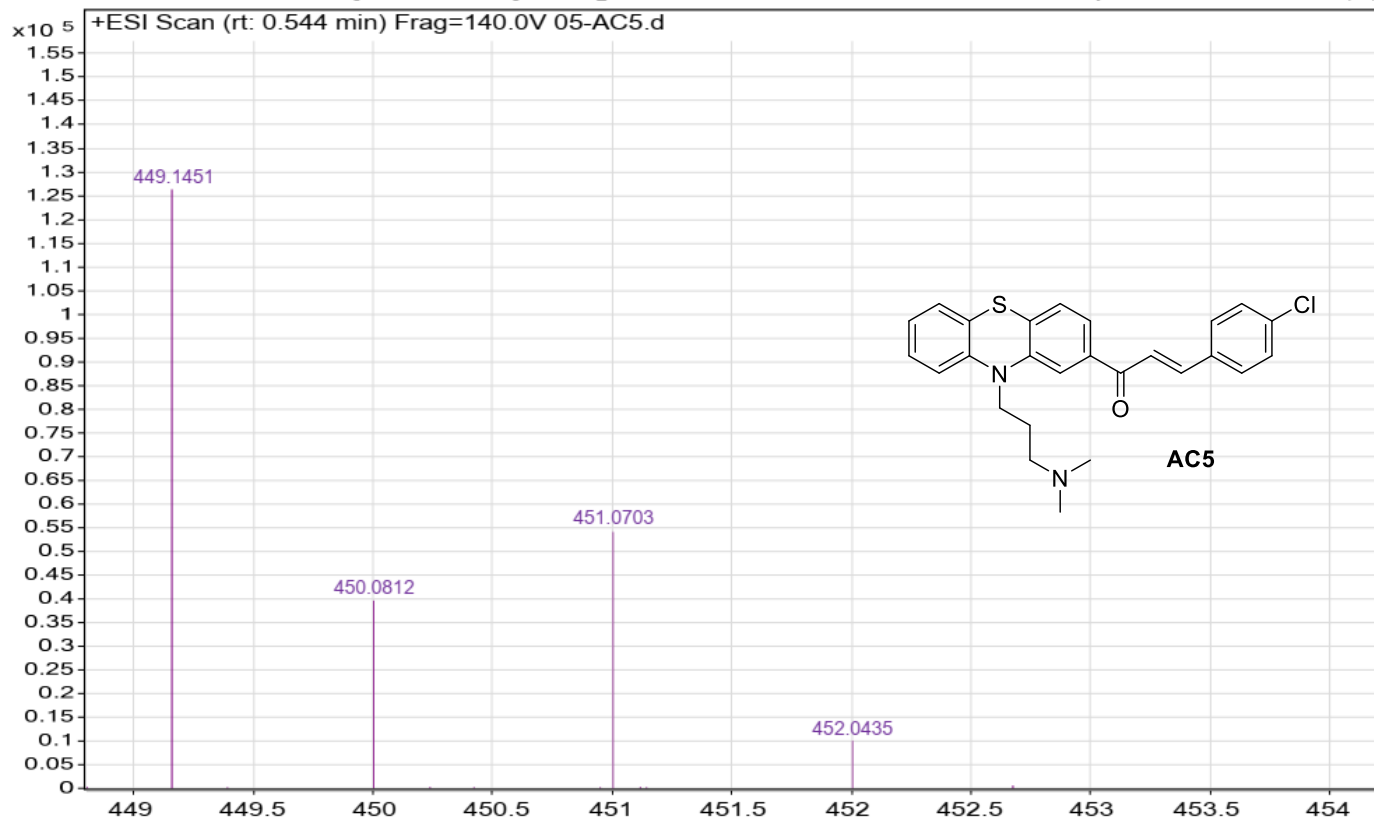


--- End Of Report ---



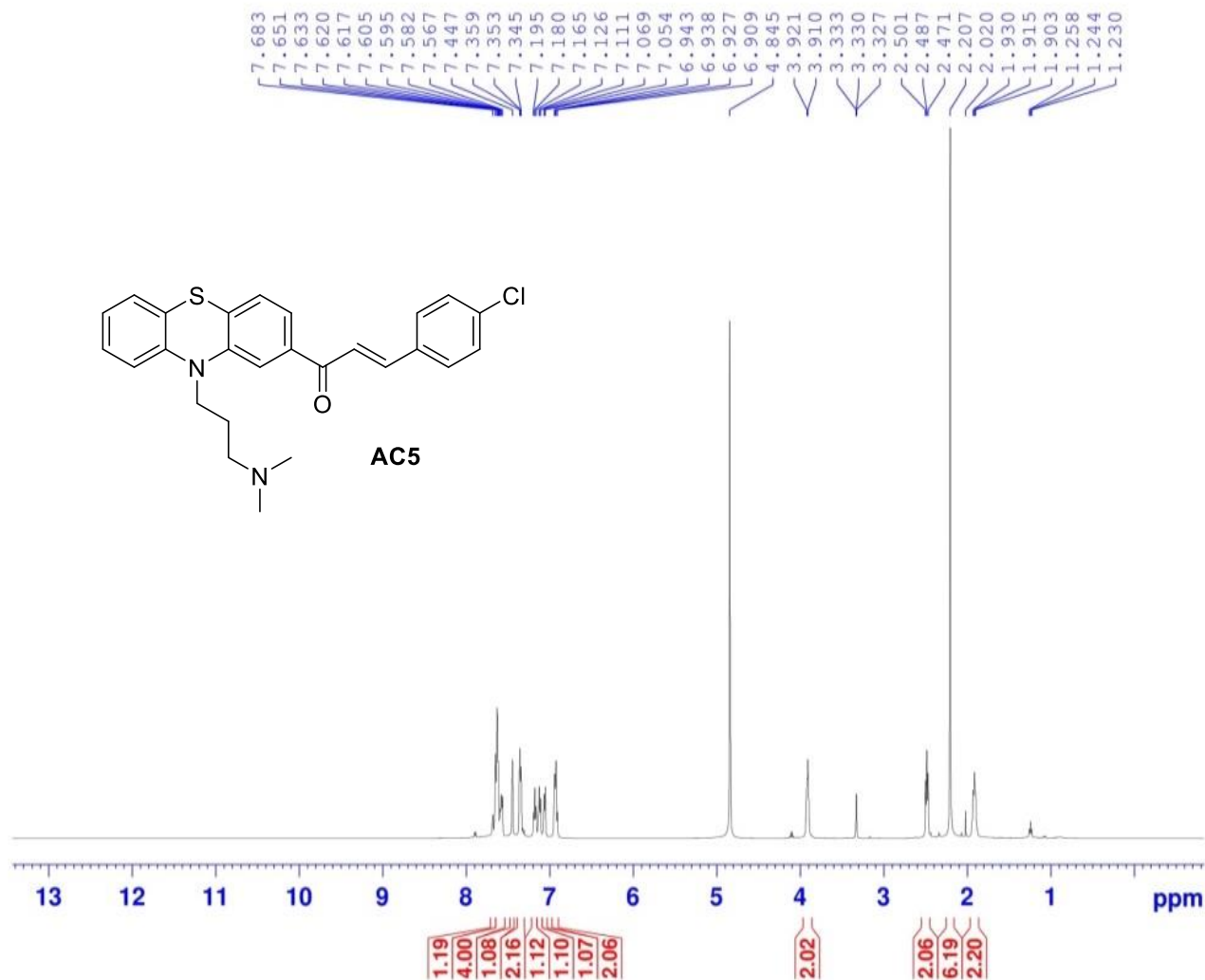
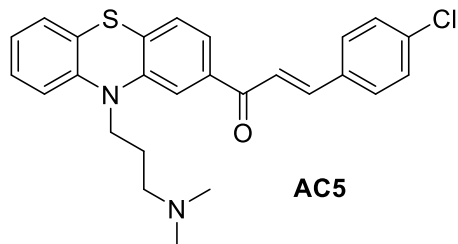
# MS

Sample Name	05-AC5	Position	P2-B4	Instrument Name	Instrument 1	User Name	
Inj Vol	2	InjPosition		SampleType	Sample	IRM Calibration Status	Success
Data Filename	05-AC5.d	ACQ Method	Cot ngan - MSMS_Pos.	Comment		Acquired Time	22/08/2020 10:38:45 AM



# <sup>1</sup>H-NMR

C9-MeOD-1H



Current Data Parameters  
NAME 113DAO\_C9  
EXPNO 1  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20170704  
Time 11.15  
INSTRUM spect  
PROBHD 5 mm PABBO BB/  
PULPROG zg30  
TD 65536  
SOLVENT MeOD  
NS 16  
DS 2  
SWH 10000.000 Hz  
FIDRES 0.152588 Hz  
AQ 3.2767999 sec  
RG 30.85  
DW 50.000 usec  
DE 6.50 usec  
TE 303.0 K  
D1 1.00000000 sec  
TD0 1

===== CHANNEL f1 =====  
SF01 500.2030889 MHz  
NUC1 1H  
P1 10.00 usec  
PLW1 22.00000000 W

F2 - Processing parameters  
SI 65536  
SF 500.2000013 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

# <sup>1</sup>H-NMR

C9-MeOD-1H



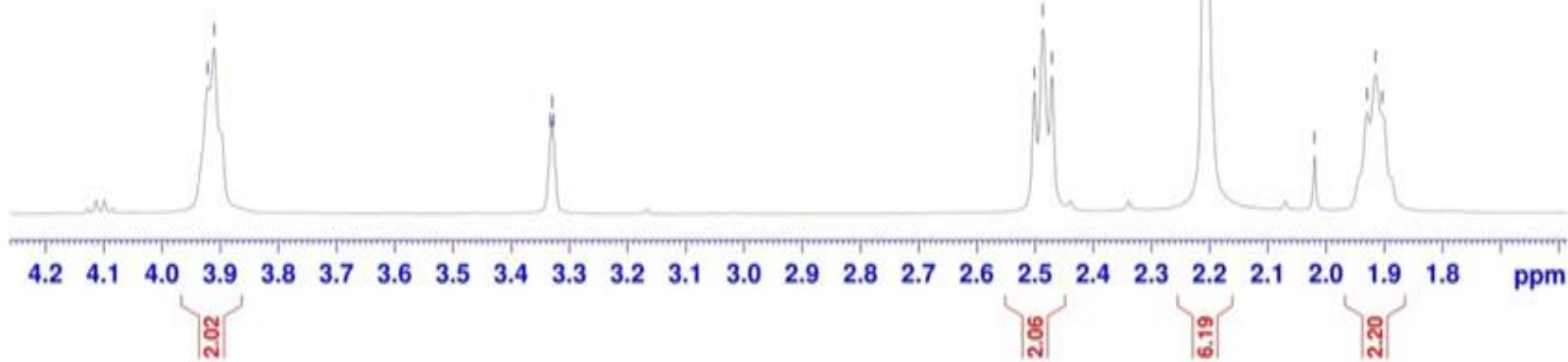
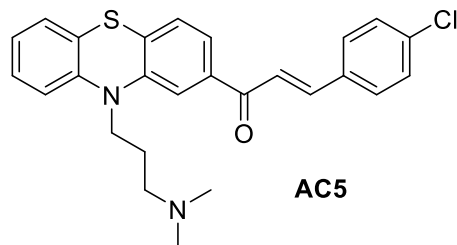
3.921  
3.910

3.333  
3.330  
3.327

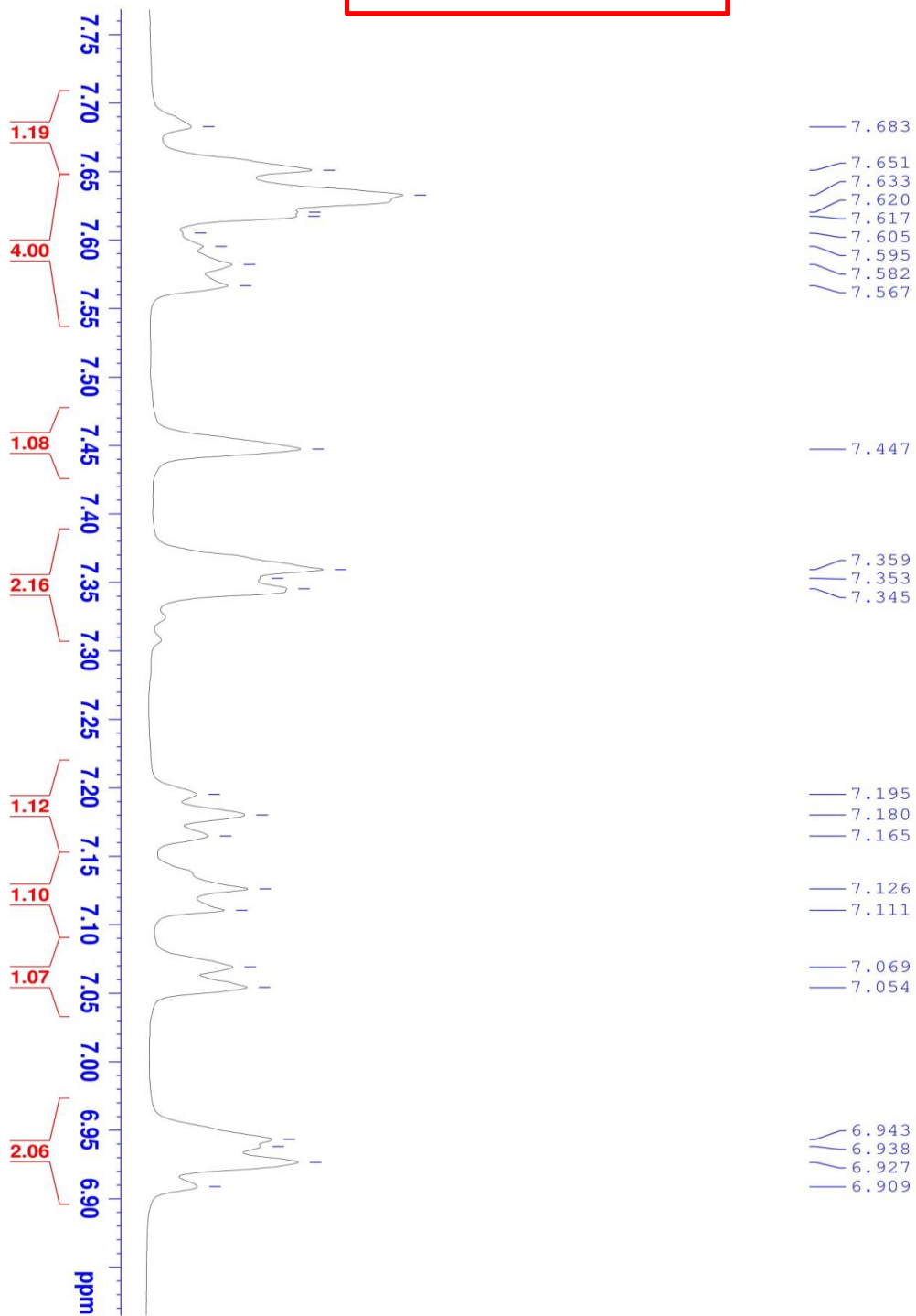
2.501  
2.487  
2.471

2.207

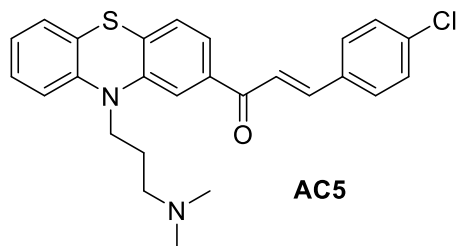
2.020  
1.930  
1.915  
1.903



# <sup>1</sup>H-NMR



C9-MeOD-1H



# <sup>13</sup>C-NMR

C9-MeOD-C13CPD



Current Data Parameters  
NAME 113DA0\_C9  
EXPNO 2  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20170704  
Time 16.32  
INSTRUM spect  
PROBHD 5 mm PABBO BB/  
PULPROG zgpg30  
TD 65536  
SOLVENT MeOD  
NS 128  
DS 4  
SWH 29761.904 Hz  
FIDRES 0.454131 Hz  
AQ 1.1010048 sec  
RG 198.57  
DW 16.800 usec  
DE 6.50 usec  
TE 303.0 K  
D1 2.00000000 sec  
D11 0.03000000 sec  
TD0 1

----- CHANNEL f1 -----  
SFO1 125.7879670 MHz  
NUC1 13C  
P1 10.00 usec  
PLW1 88.00000000 W

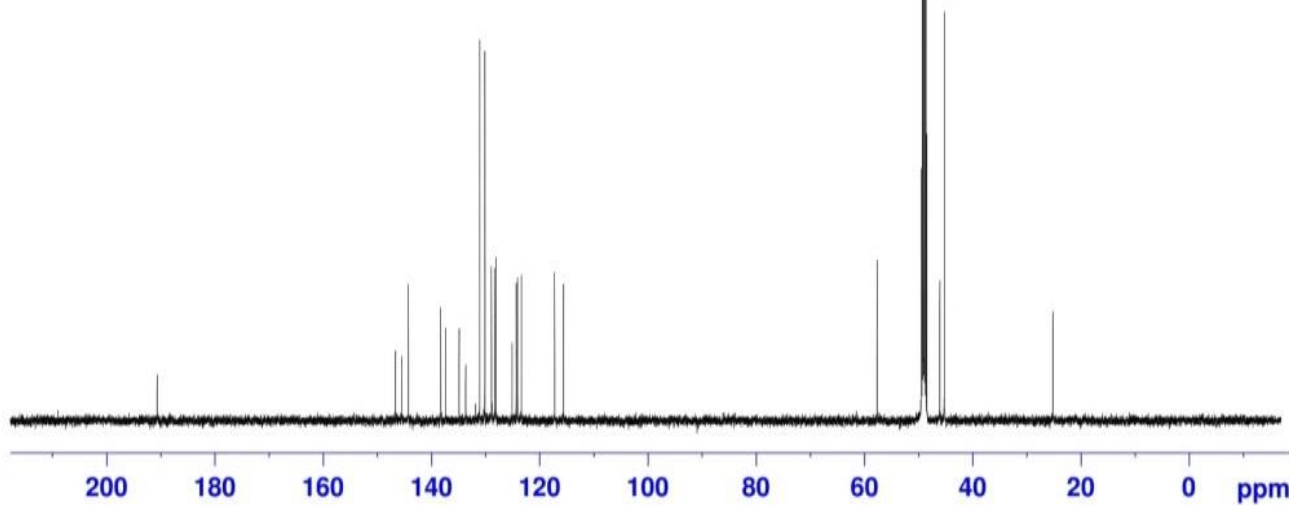
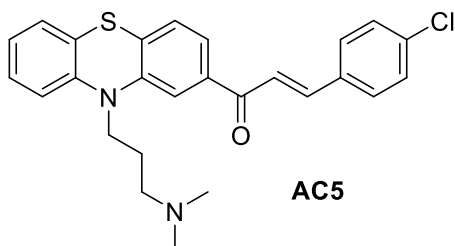
----- CHANNEL f2 -----  
SFO2 500.2020008 MHz  
NUC2 1H  
CPDPRG[2] waltz16  
PCPD2 80.00 usec  
PLW2 22.00000000 W  
PLW12 0.34375000 W  
PLW13 0.22000000 W

F2 - Processing parameters  
SI 32768  
SF 125.7752207 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40

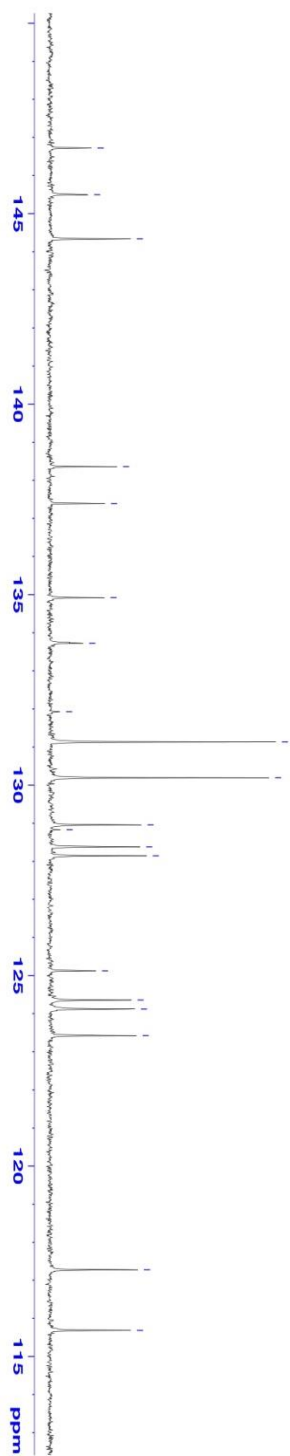
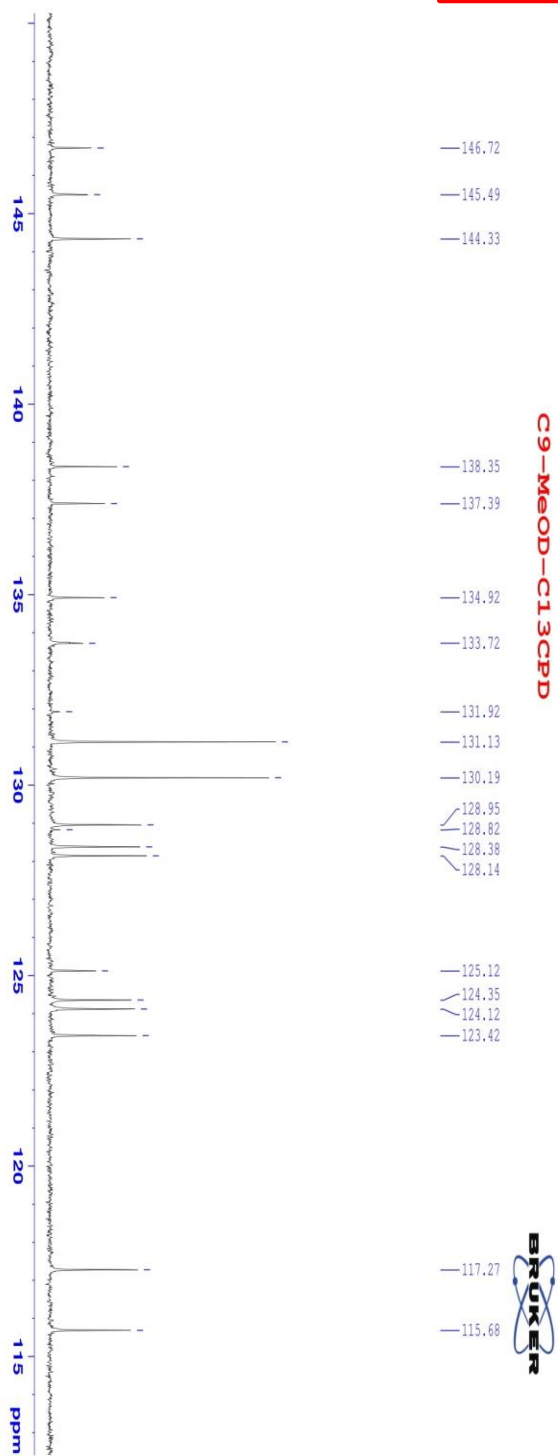
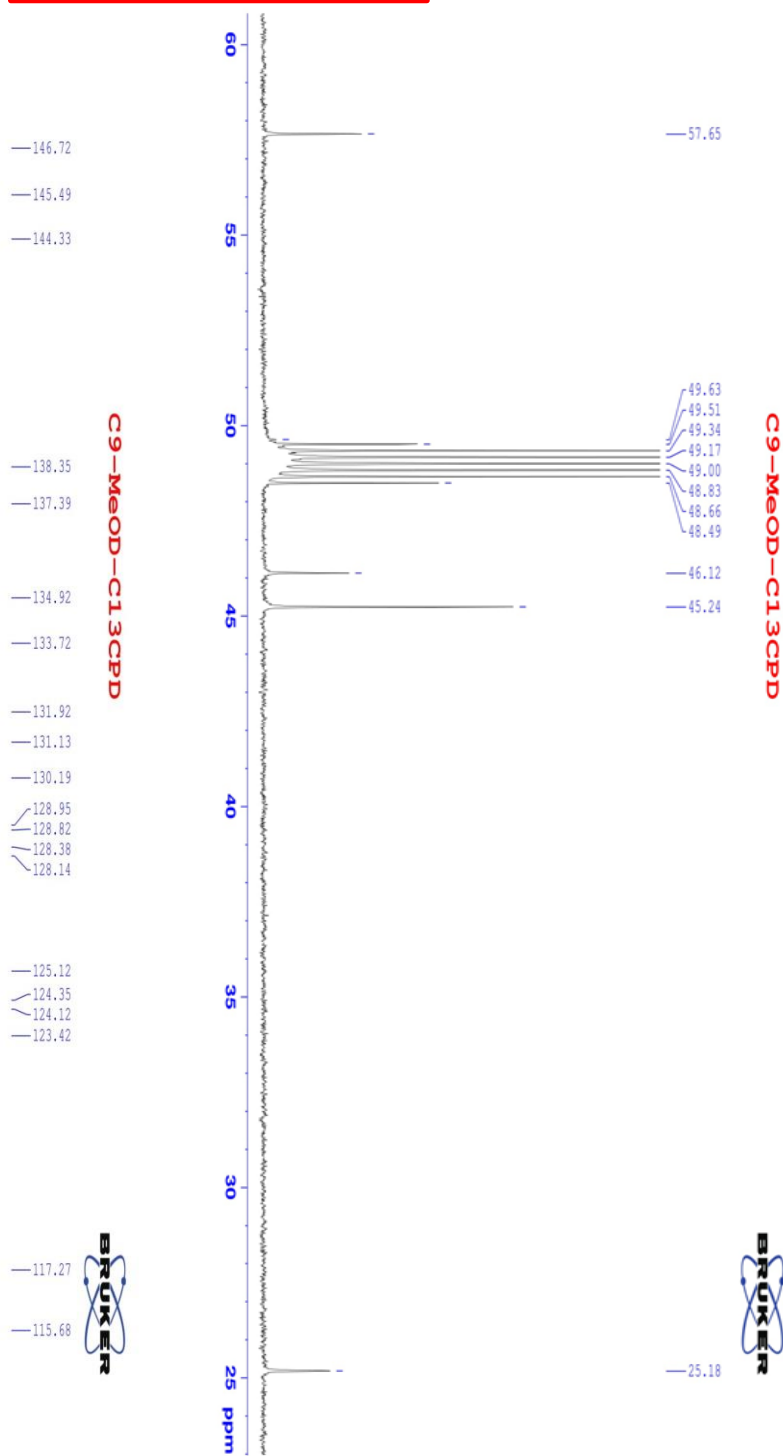
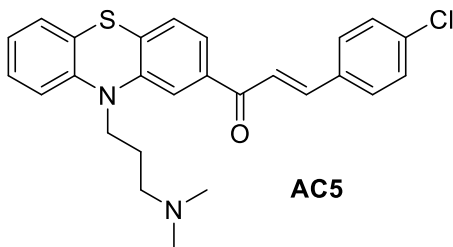
190.69

146.72  
145.49  
144.33  
138.35  
137.39  
134.92  
133.72  
131.92  
131.13  
130.19  
128.95  
128.82  
128.38  
128.14  
125.12  
124.35  
124.12  
123.42  
117.27  
115.68

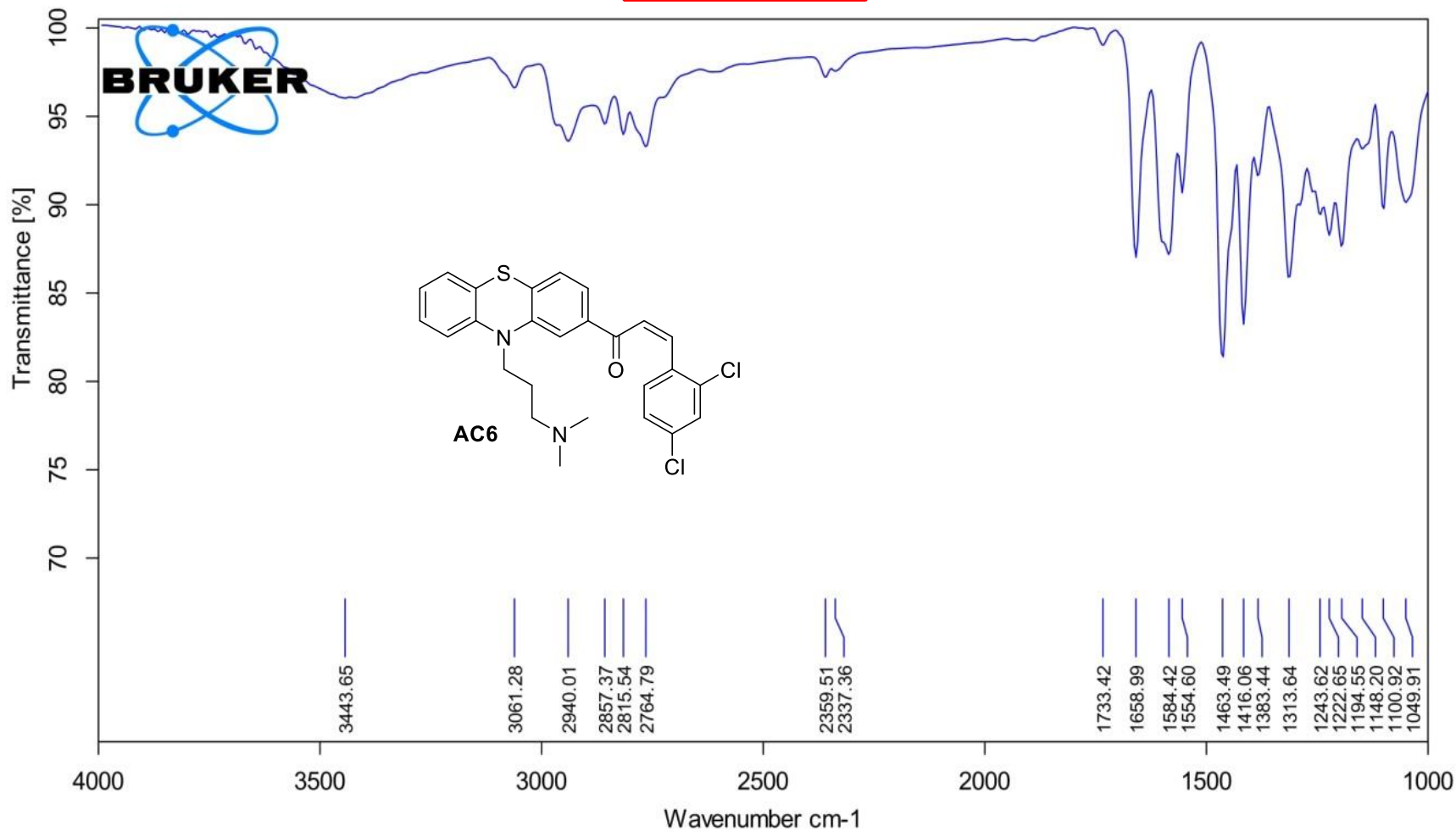
57.65  
49.63  
49.51  
49.34  
49.17  
49.00  
48.83  
48.66  
48.49  
46.12  
45.24  
25.18



# $^{13}\text{C}$ -NMR



IR



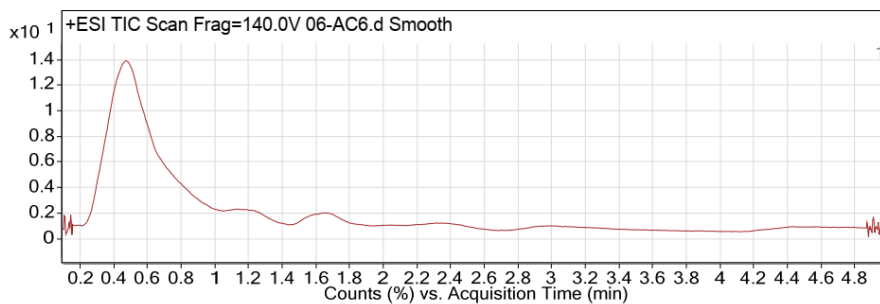
# MS

## Qualitative Analysis Report

<b>Data Filename</b>	06-AC6.d	<b>Sample Name</b>	06-AC6
<b>Sample Type</b>	Sample	<b>Position</b>	P2-B3
<b>Instrument Name</b>	Instrument 1	<b>User Name</b>	
<b>Acq Method</b>	Cot ngan - MSMS_Pos.m	<b>Acquired Time</b>	22/08/2020 10:43:11 AM
<b>IRM Calibration Status</b>	Success	<b>DA Method</b>	COTNGAN.M.m
<b>Comment</b>			
<b>Sample Group</b>		<b>Info.</b>	
<b>Stream Name</b>	LC 1	<b>Acquisition SW</b>	6200 series TOF/6500 series
		<b>Version</b>	Q-TOF B.06.01 (B6172 SP1)

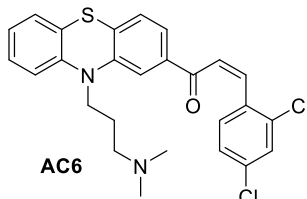
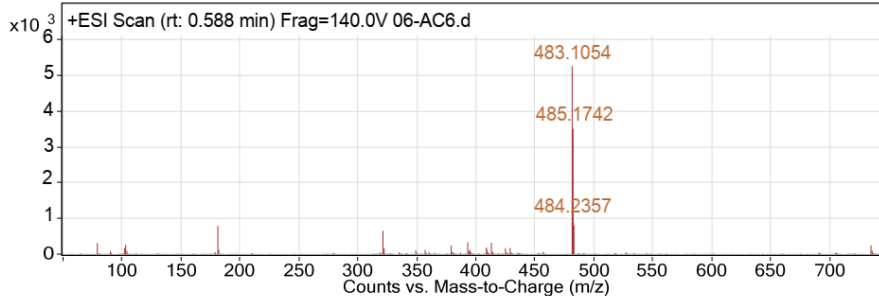
### User Chromatograms

Fragmentor Voltage 140 Collision Energy 0 Ionization Mode ESI



### User Spectra

Fragmentor Voltage 140 Collision Energy 0 Ionization Mode ESI

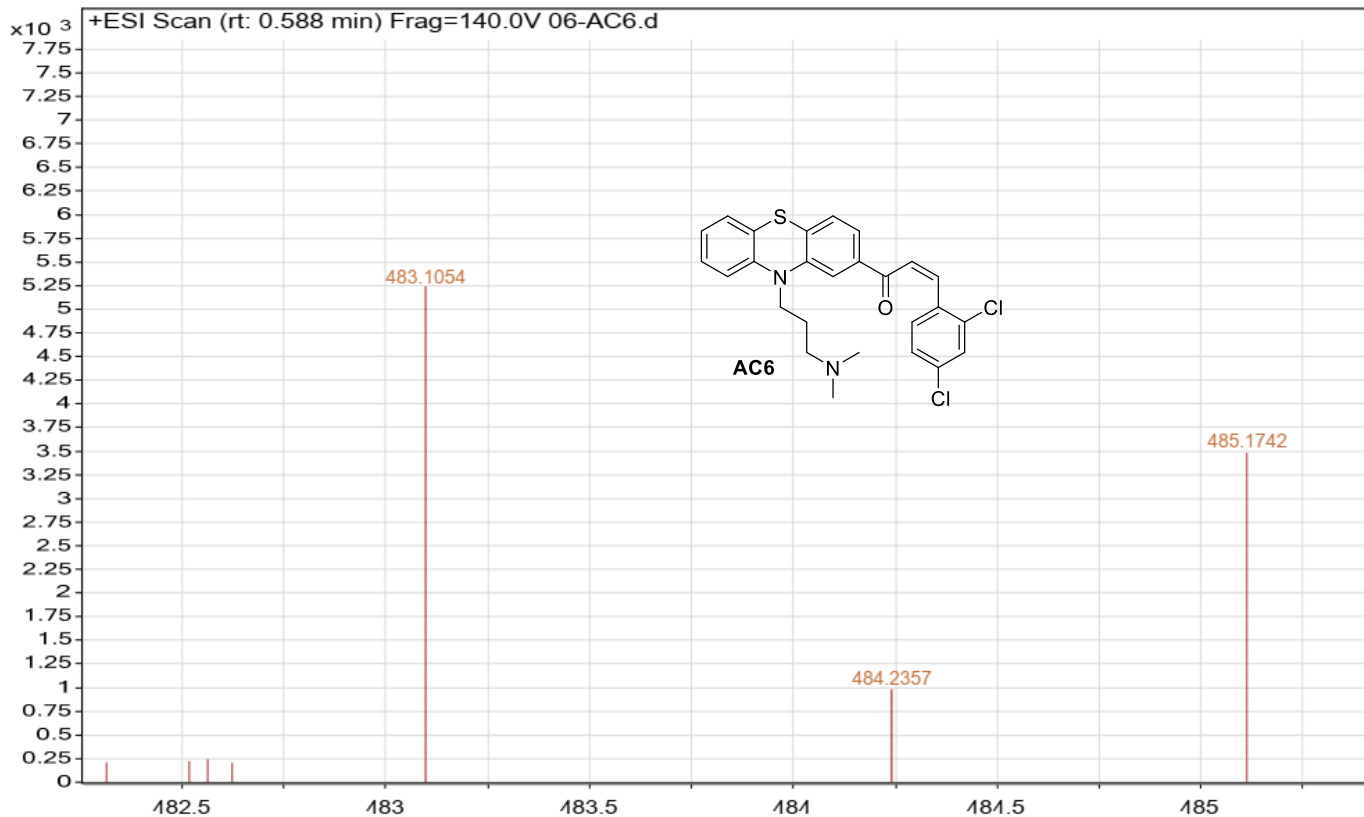


--- End Of Report ---



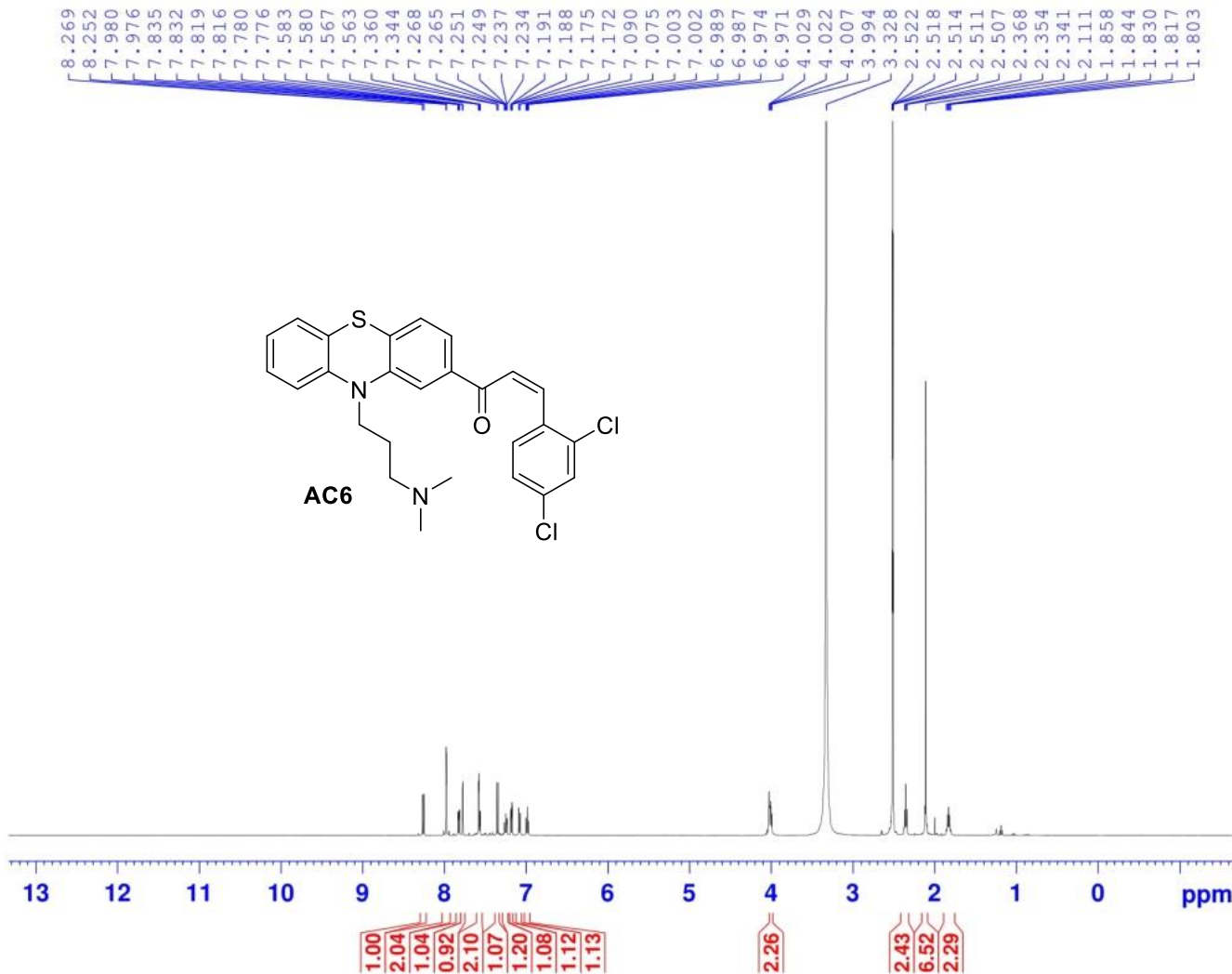
# MS

Sample Name	06-AC6	Position	P2-B3	Instrument Name	Instrument 1	User Name	
Inj Vol	2	InjPosition		SampleType	Sample	IRM Calibration Status	Success
Data Filename	06-AC6.d	ACQ Method	Cot ngan - MSMS_Pos.	Comment		Acquired Time	22/08/2020 10:43:11 AM



# <sup>1</sup>H-NMR

C8-DMSO-1H



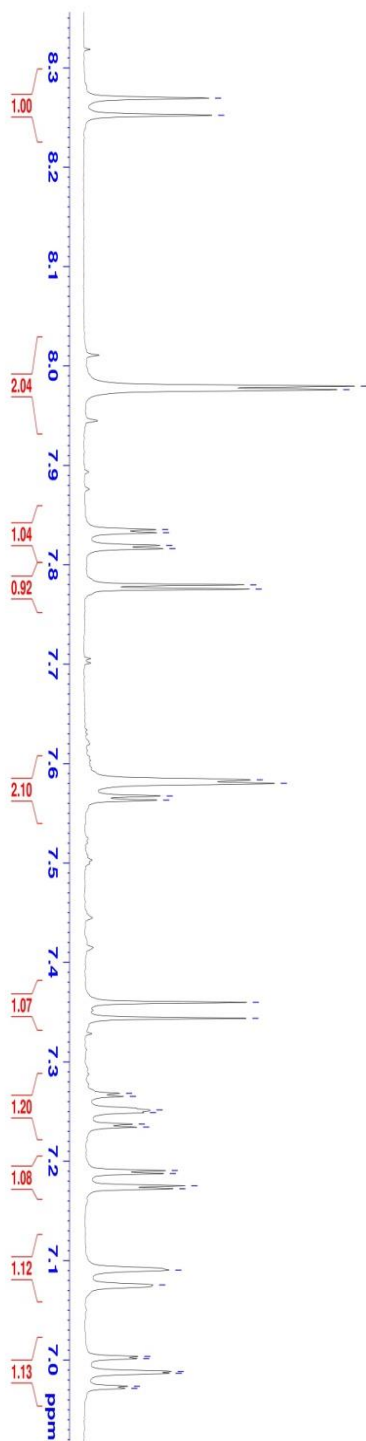
Current Data Parameters  
NAME 113D\_C8  
EXPNO 1  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20170606  
Time 16.22  
INSTRUM spect  
PROBHD 5 mm PABBO BB/  
PULPROG zg30  
TD 65536  
SOLVENT DMSO  
NS 16  
DS 2  
SWH 10000.000 Hz  
FIDRES 0.152588 Hz  
AQ 3.2767999 sec  
RG 157.35  
DW 50.000 usec  
DE 6.50 usec  
TE 303.0 K  
D1 1.00000000 sec  
TD0 1

----- CHANNEL f1 -----  
SFO1 500.2030889 MHz  
NUC1 1H  
P1 10.00 usec  
PLW1 22.00000000 W

F2 - Processing parameters  
SI 65536  
SF 500.2000000 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

# <sup>1</sup>H-NMR



8.269  
8.252

7.980  
7.976

7.835  
7.832  
7.819  
7.816  
7.780  
7.776

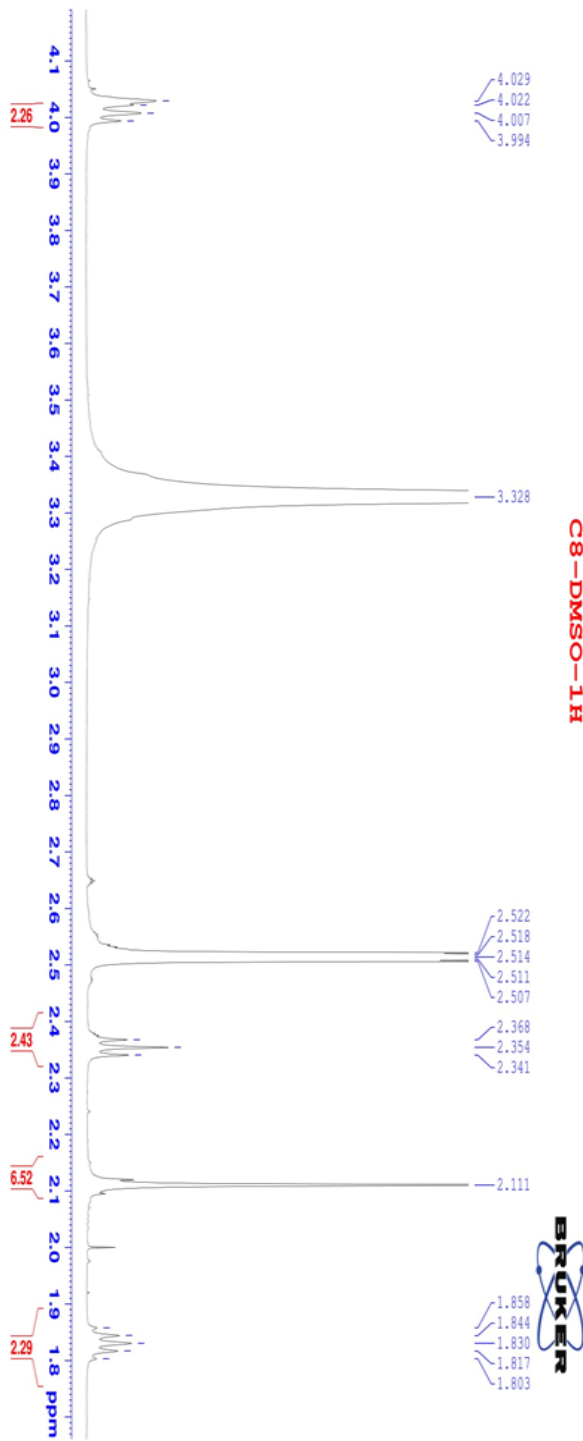
**CDCl<sub>3</sub>-1H**

7.583  
7.580  
7.567  
7.563

7.360  
7.344  
7.268  
7.265  
7.251  
7.249  
7.237  
7.234  
7.234  
7.191  
7.188  
7.175  
7.172

**BRUKER**

7.090  
7.075  
7.003  
7.002  
6.989  
6.987  
6.974  
6.971



4.029  
4.022  
4.007  
3.994

2.26

3.328

**DMSO-d<sub>6</sub>-1H**

2.522  
2.518  
2.514  
2.511  
2.507

2.43

2.368  
2.354  
2.341

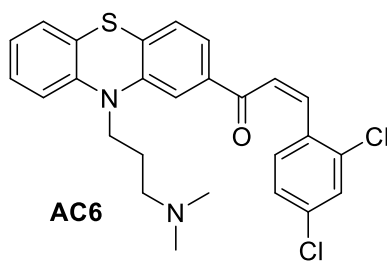
6.52

2.111

1.858  
1.844  
1.830  
1.817  
1.803

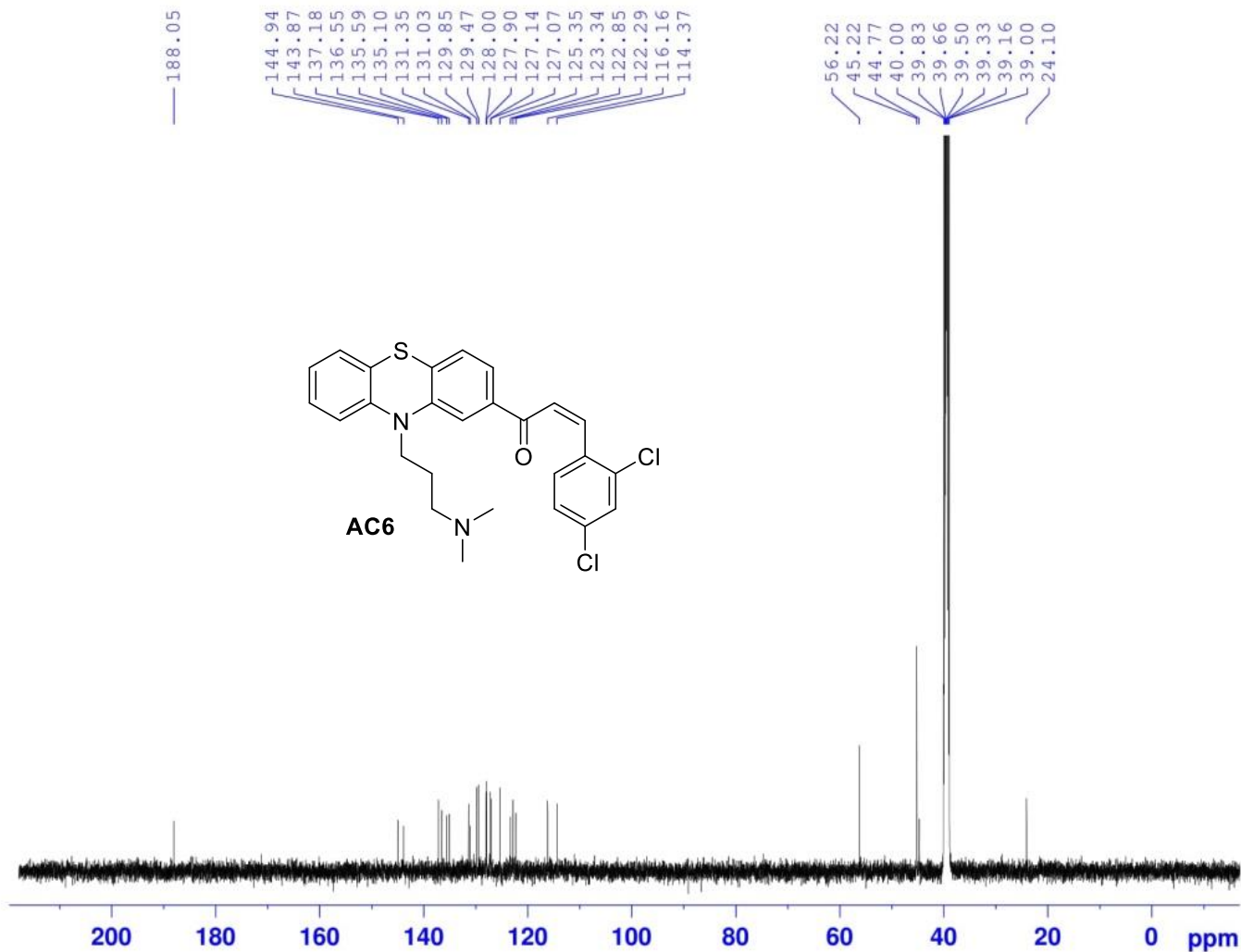
**BRUKER**

2.29



# <sup>13</sup>C-NMR

C8-DMSO-C13CPD



Current Data Parameters  
NAME 113D\_C8  
EXPNO 2  
PROCNO 1

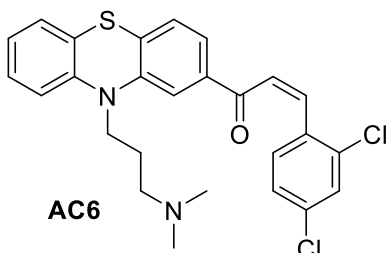
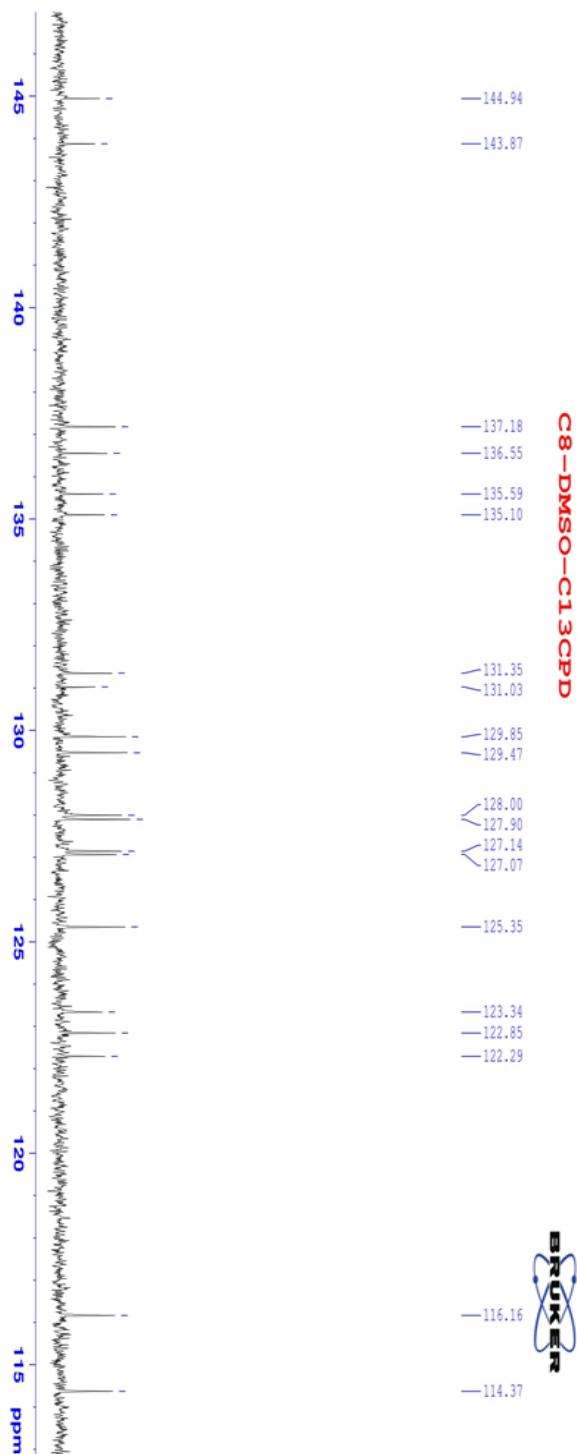
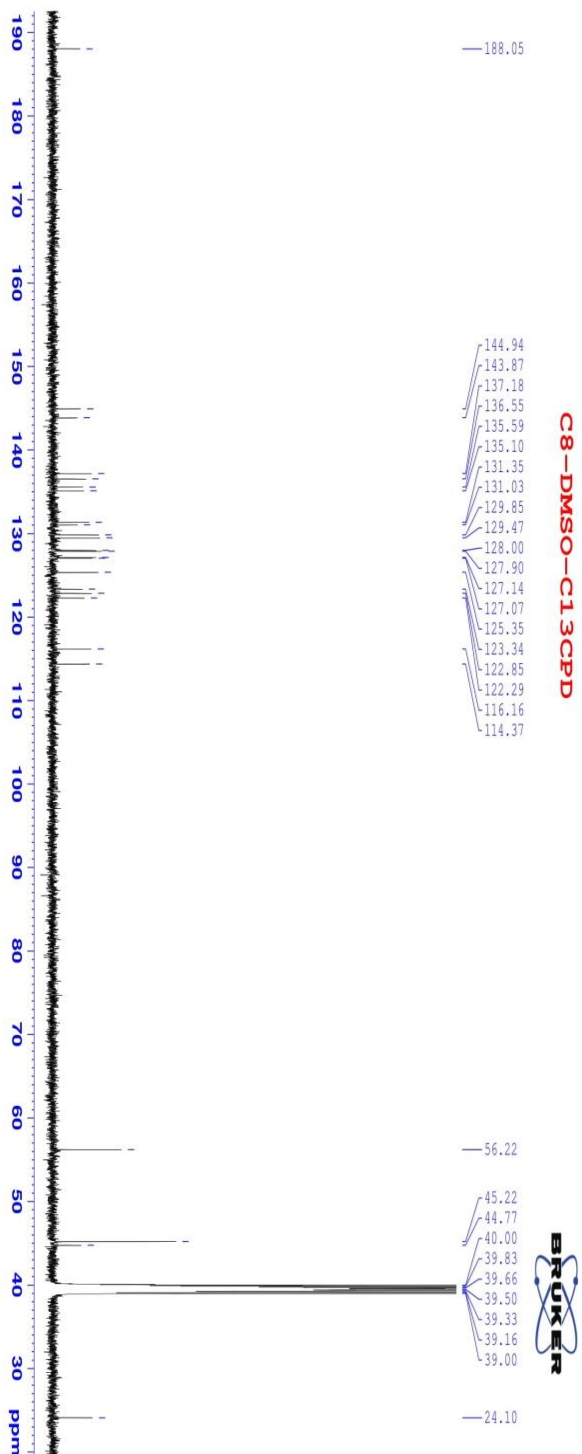
F2 - Acquisition Parameters  
Date\_ 20170607  
Time 4.54  
INSTRUM spect  
PROBHD 5 mm PABBO BB/  
PULPROG zgpg30  
TD 65536  
SOLVENT DMSO  
NS 4096  
DS 4  
SWH 29761.904 Hz  
FIDRES 0.454131 Hz  
AQ 1.1010048 sec  
RG 198.57  
DW 16.800 usec  
DE 6.50 usec  
TE 303.0 K  
D1 2.00000000 sec  
D11 0.03000000 sec  
TD0 1

----- CHANNEL f1 -----  
SFO1 125.7879670 MHz  
NUC1 13C  
P1 10.00 usec  
PLW1 88.00000000 W

----- CHANNEL f2 -----  
SFO2 500.2020008 MHz  
NUC2 1H  
CPDPRG[2] waltz16  
PCPD2 80.00 usec  
PLW2 22.00000000 W  
PLW12 0.34375000 W  
PLW13 0.22000000 W

F2 - Processing parameters  
SI 32768  
SF 125.7754565 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40

# $^{13}\text{C}$ -NMR



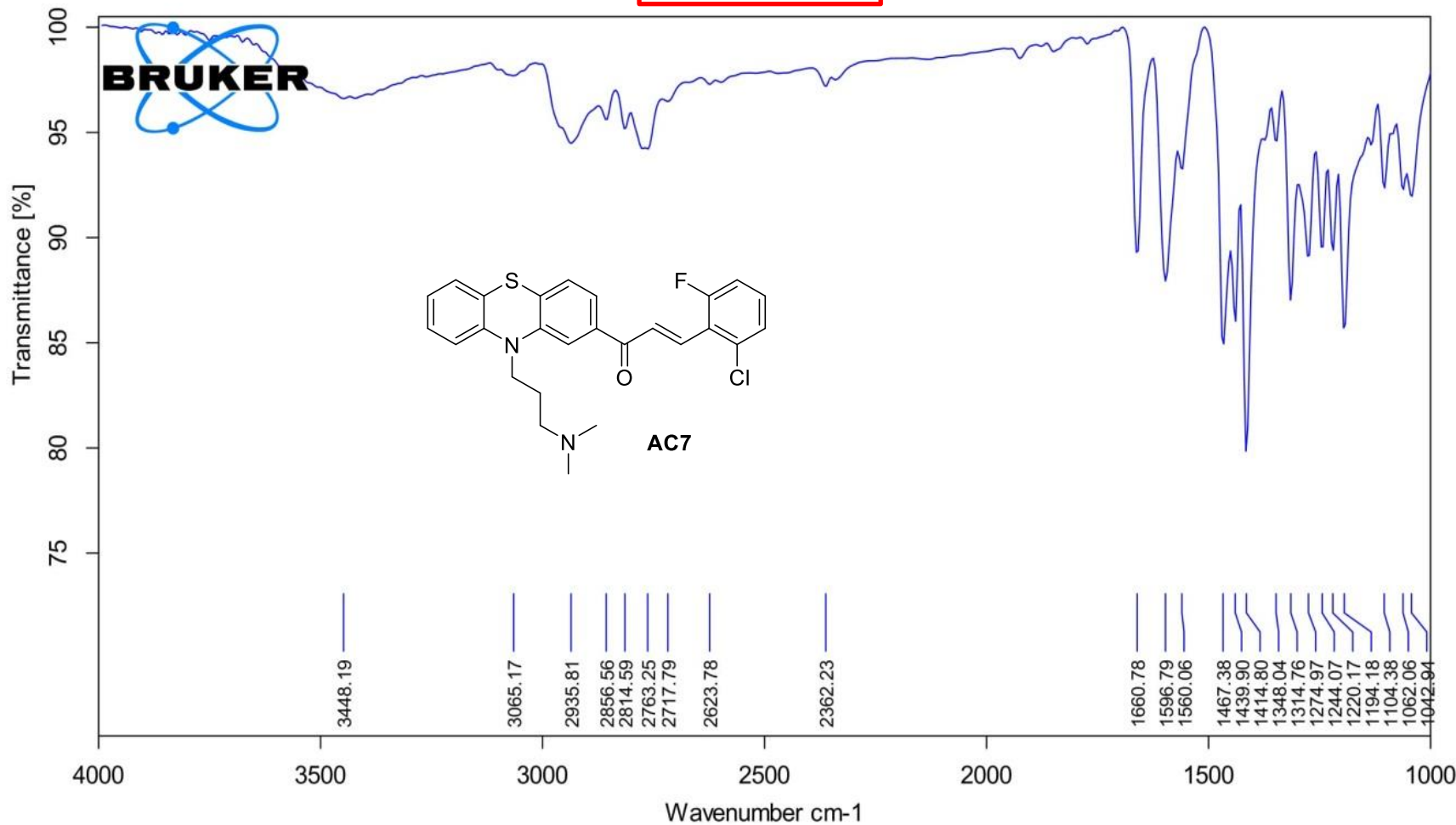
# $^{13}\text{C}$ -NMR



C8-DMSO-C13CPD



IR



E:\OPUS 7\2017\THANG 6\20170606\C6.0	C6	SENSOR 27 - BRUKER - GERMANY	6/8/2017
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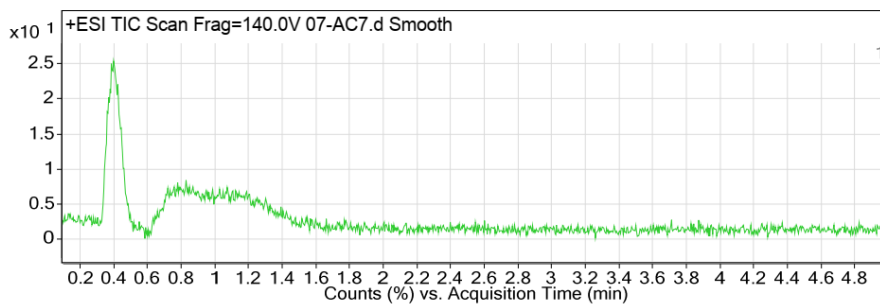
# MS

## Qualitative Analysis Report

<b>Data Filename</b>	07-AC7.d	<b>Sample Name</b>	07-AC7
<b>Sample Type</b>	Sample	<b>Position</b>	P2-B7
<b>Instrument Name</b>	Instrument 1	<b>User Name</b>	
<b>Acq Method</b>	Cot ngan - MSMS_Pos.m	<b>Acquired Time</b>	22/08/2020 10:55:28 AM
<b>IRM Calibration Status</b>	Success	<b>DA Method</b>	COTNGAN.M.m
<b>Comment</b>			
<b>Sample Group</b>		<b>Info.</b>	
<b>Stream Name</b>	LC 1	<b>Acquisition SW</b>	6200 series TOF/6500 series
		<b>Version</b>	Q-TOF B.06.01 (B6172 SP1)

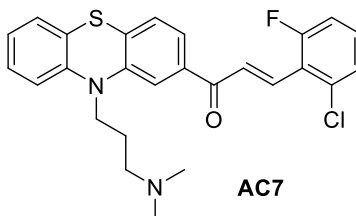
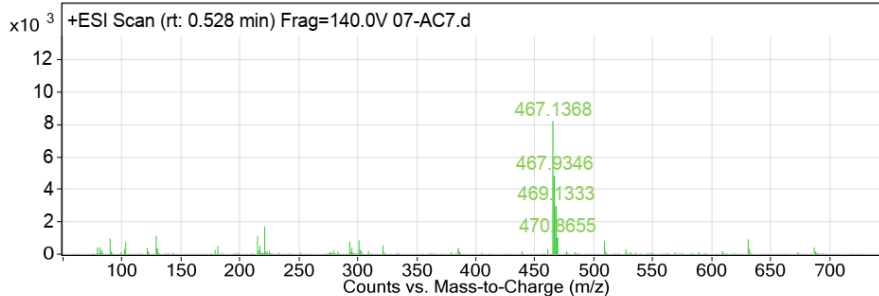
### User Chromatograms

Fragmentor Voltage 140 Collision Energy 0 Ionization Mode ESI



### User Spectra

Fragmentor Voltage 140 Collision Energy 0 Ionization Mode ESI

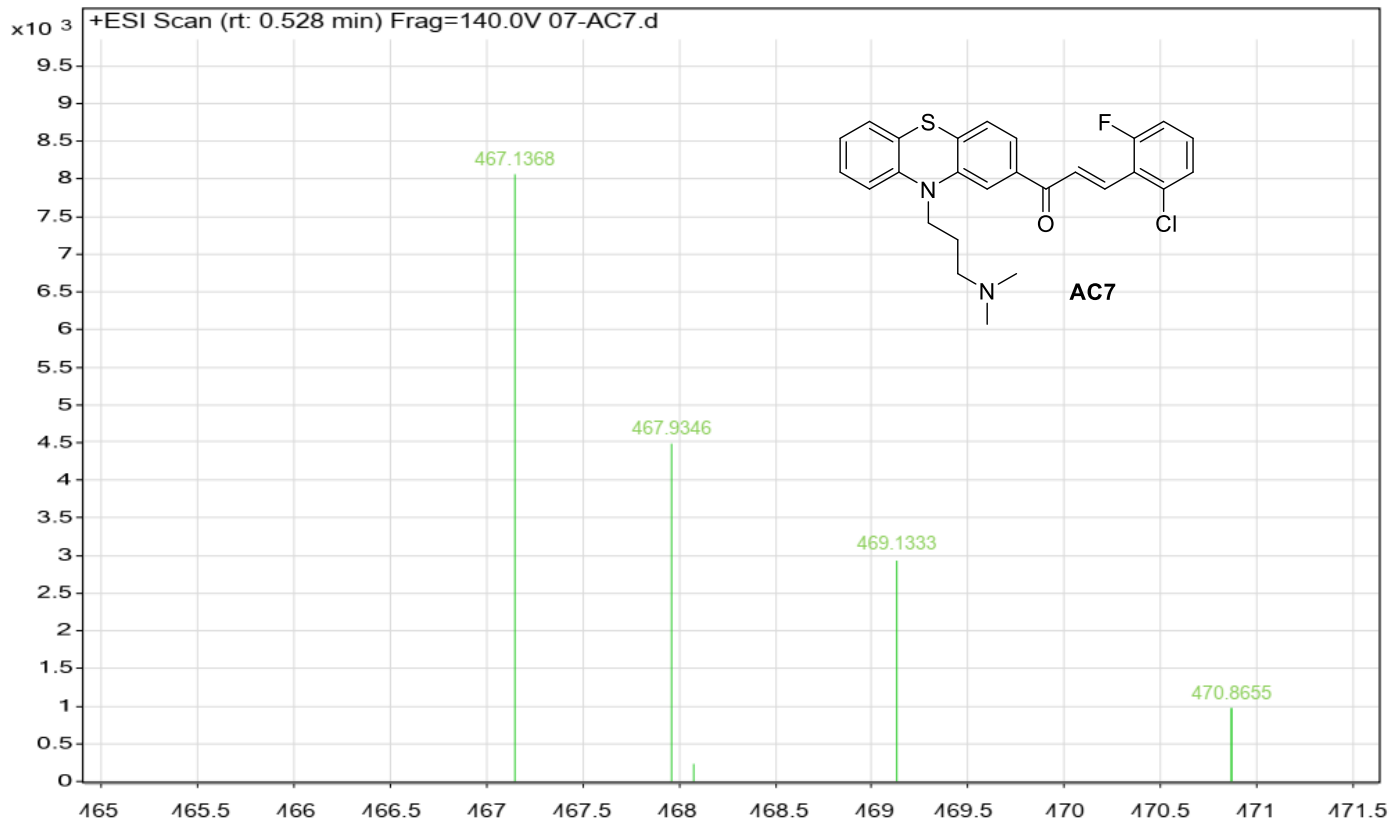


--- End Of Report ---



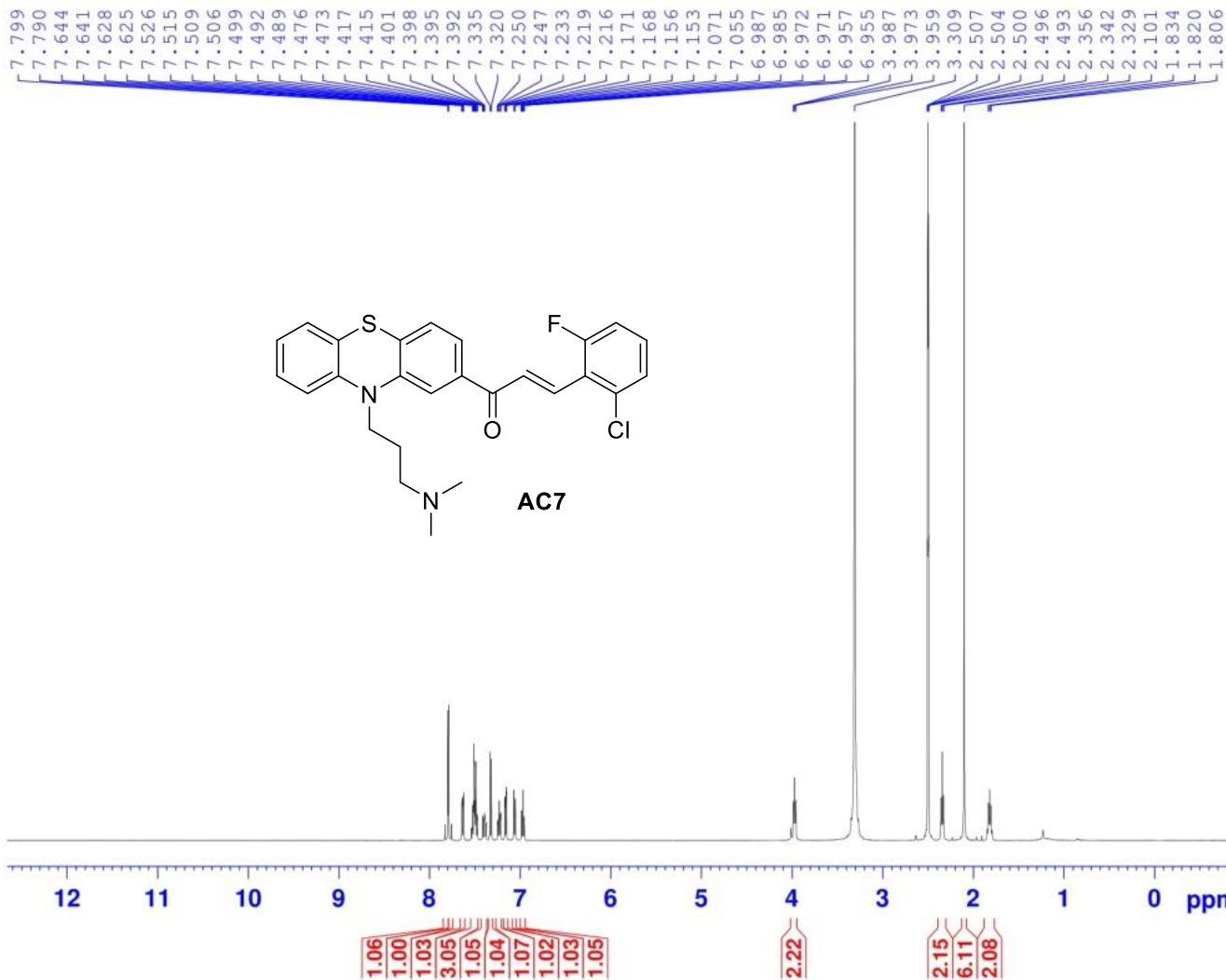
# MS

Sample Name	07-AC7	Position	P2-B7	Instrument Name	Instrument 1	User Name	
Inj Vol	2	InjPosition		SampleType	Sample	IRM Calibration Status	Success
Data Filename	07-AC7.d	ACQ Method	Cot ngan - MSMS_Pos.	Comment		Acquired Time	22/08/2020 10:55:28 AM

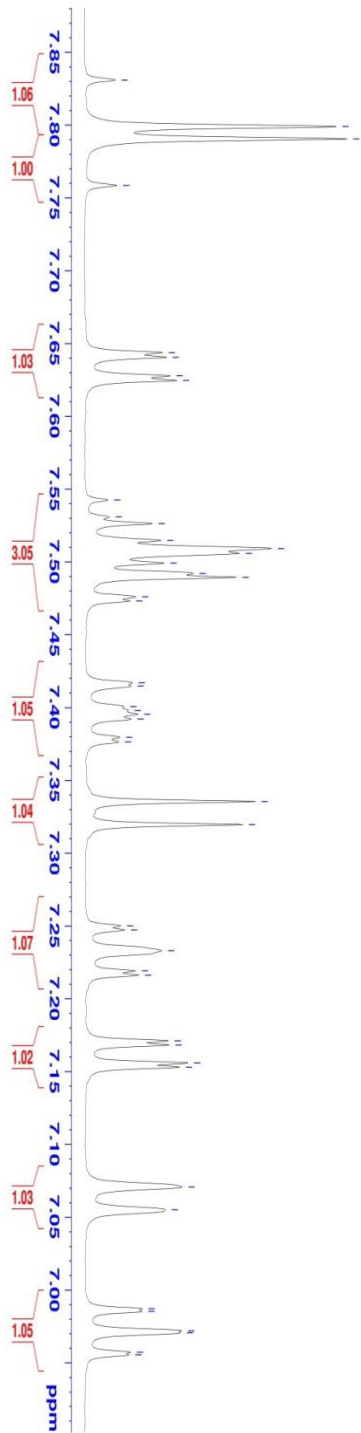


# <sup>1</sup>H-NMR

C6-DMSO-1H



# $^1\text{H-NMR}$



7.831  
7.799  
7.790  
7.759

7.644  
7.641  
7.628  
7.625

7.543  
7.531  
7.526  
7.515  
7.509  
7.506  
7.499  
7.492  
7.489  
7.476  
7.473  
7.417  
7.415  
7.401  
7.398  
7.395  
7.392  
7.380  
7.376  
7.335  
7.320

7.250  
7.247  
7.233  
7.219  
7.216  
7.171  
7.168  
7.156  
7.153

7.071  
7.055

6.987  
6.985  
6.972  
6.971  
6.957  
6.955

C6-DMSO- $\text{d}_6$

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3.987  
3.973  
3.959

3.309

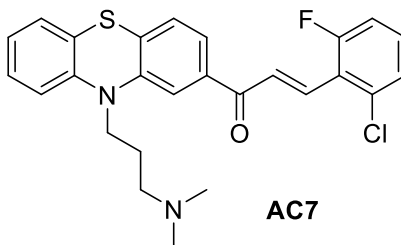
2.507  
2.504  
2.500  
2.496  
2.493  
2.356  
2.342  
2.329

2.101

1.847  
1.834  
1.820  
1.806  
1.793

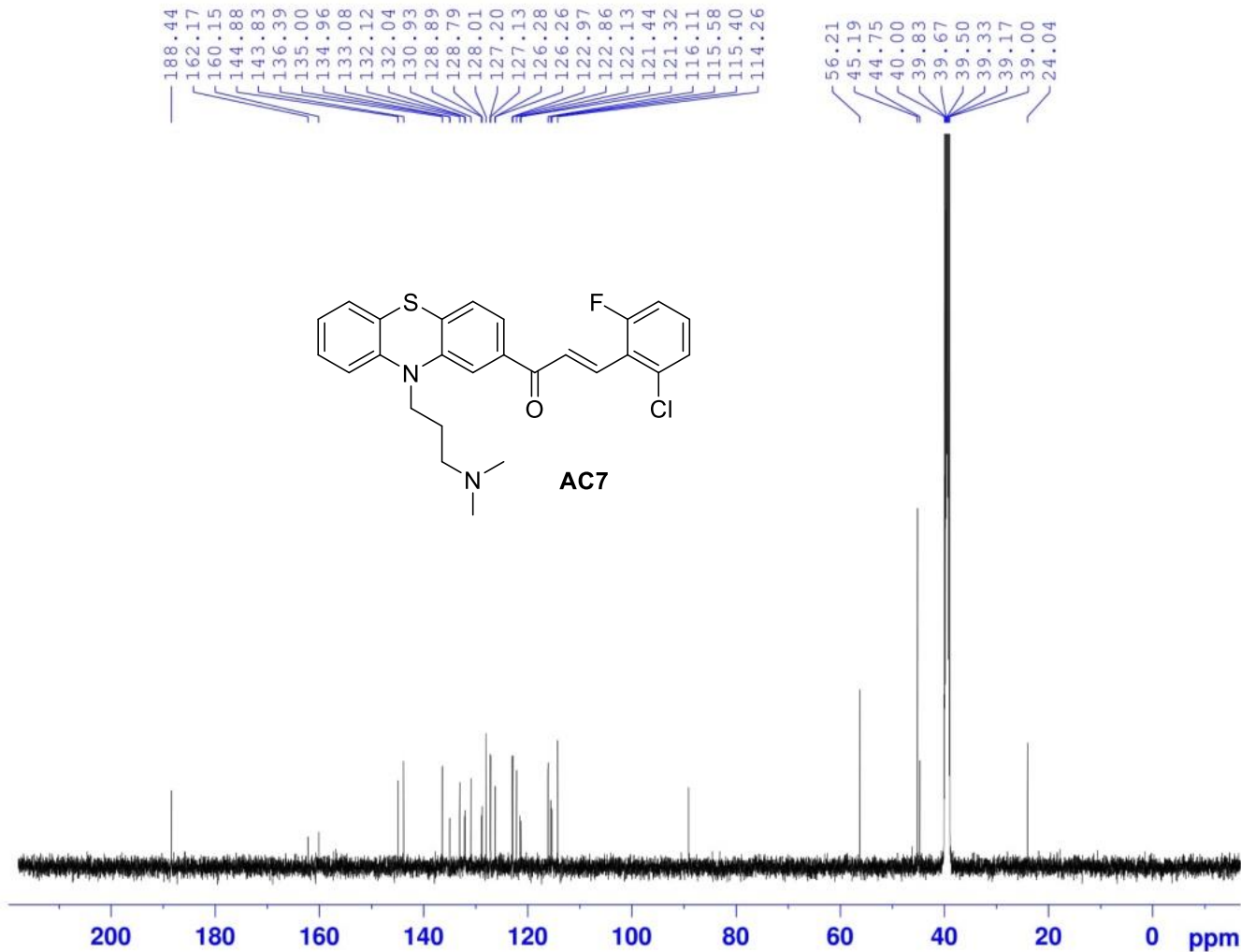
C6-DMSO- $\text{d}_6$

BRUKER



# <sup>13</sup>C-NMR

C6-DMSO-C13CPD



Current Data Parameters  
NAME 113D\_C6  
EXPNO 2  
PROCNO 1

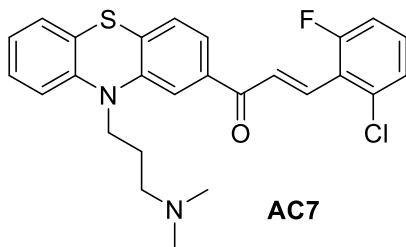
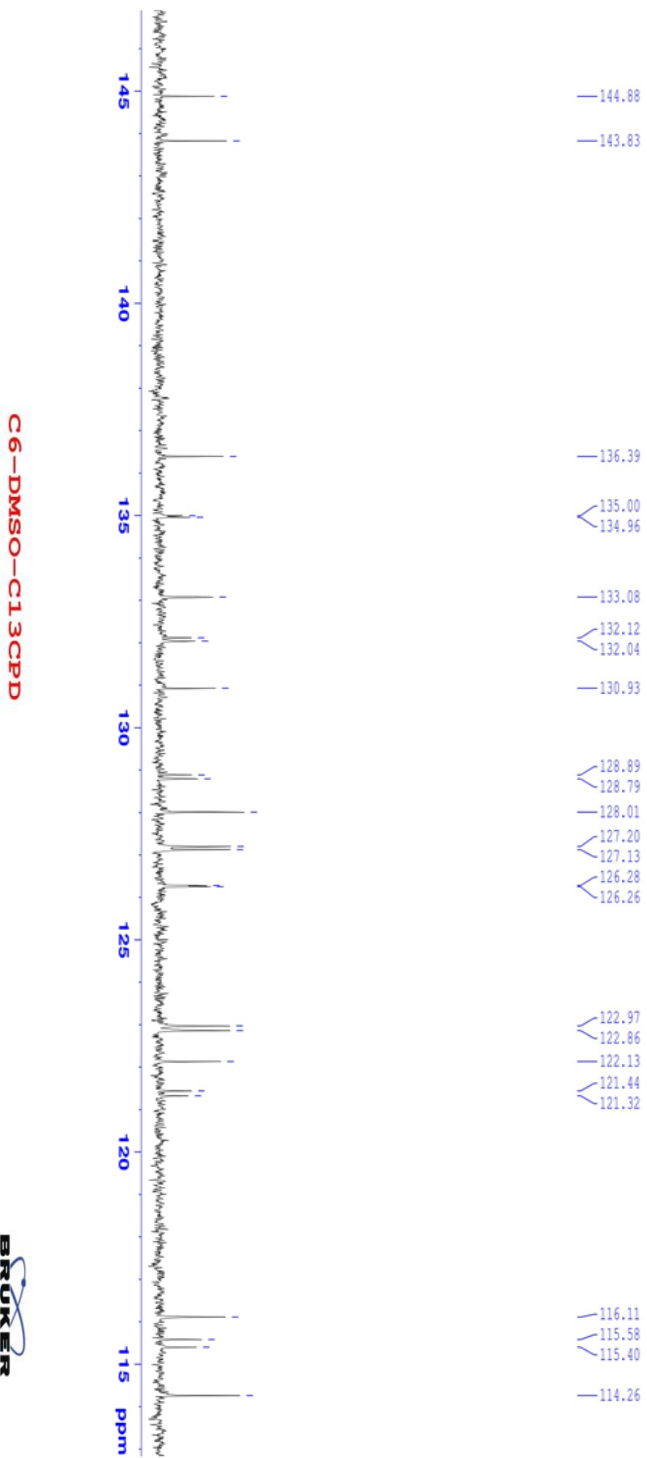
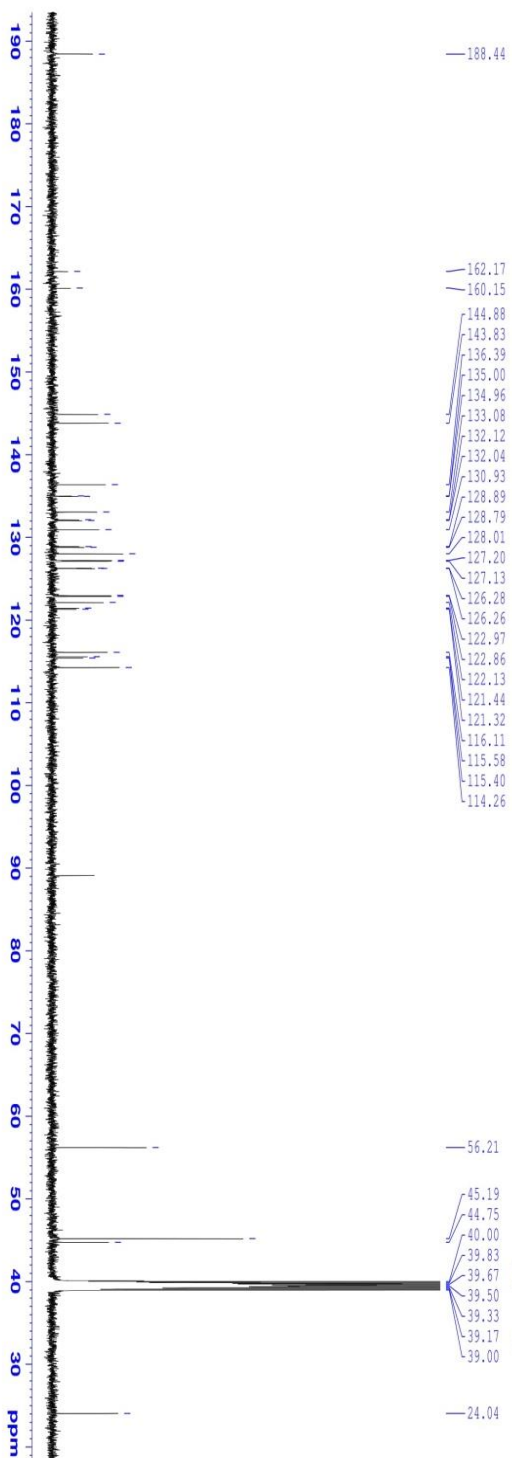
F2 - Acquisition Parameters  
Date\_ 20170607  
Time 1.15  
INSTRUM spect  
PROBHD 5 mm PABBO BB/  
PULPROG zgpg30  
TD 65536  
SOLVENT DMSO  
NS 3072  
DS 4  
SWH 29761.904 Hz  
FIDRES 0.454131 Hz  
AQ 1.1010048 sec  
RG 198.57  
DW 16.800 usec  
DE 6.50 usec  
TE 303.0 K  
D1 2.00000000 sec  
D11 0.03000000 sec  
TD0 1

===== CHANNEL f1 =====  
SFO1 125.7879670 MHz  
NUC1 13C  
P1 10.00 usec  
PLW1 88.00000000 W

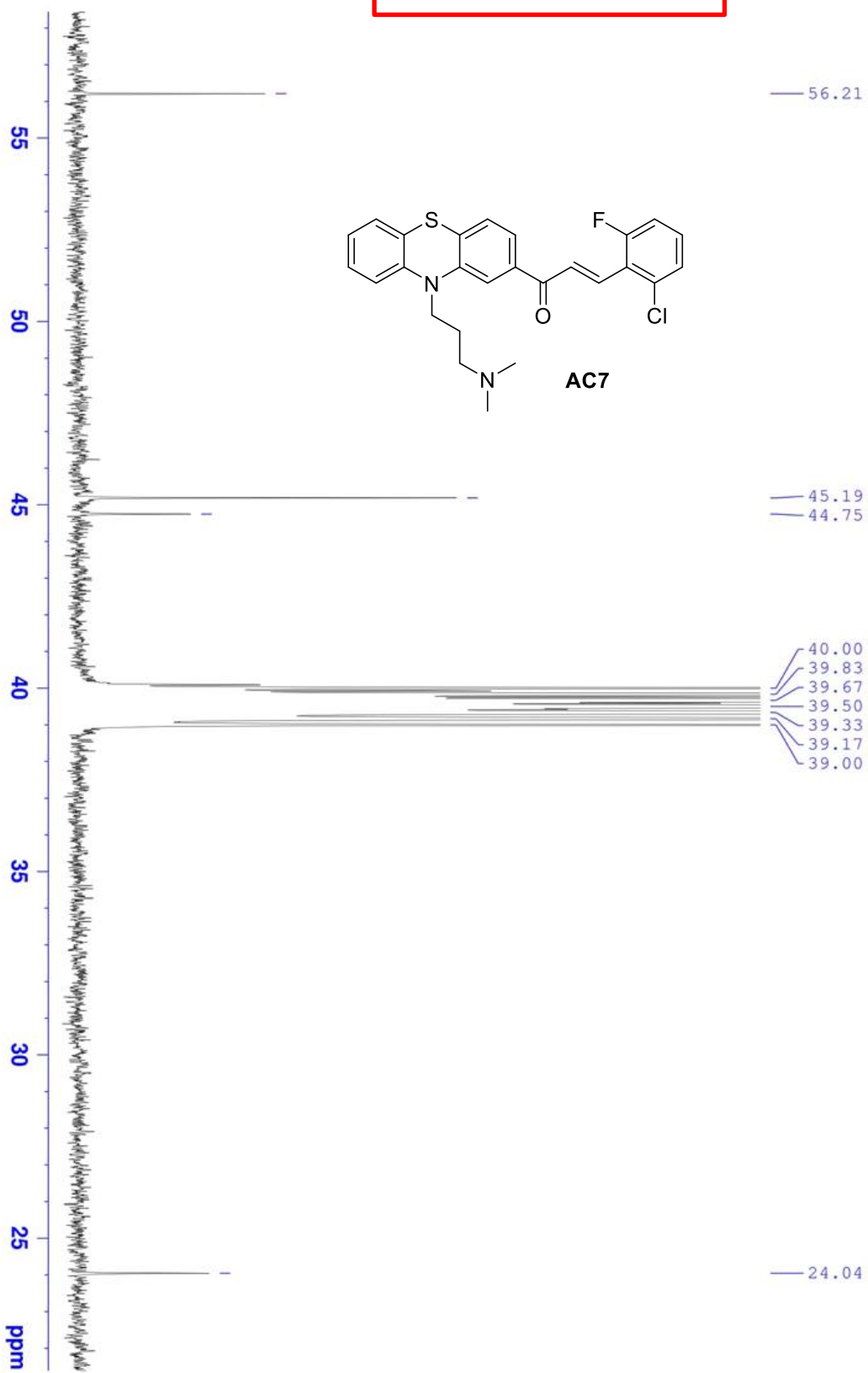
===== CHANNEL f2 =====  
SFO2 500.2020008 MHz  
NUC2 1H  
CPDPRG[2] waltz16  
PCPD2 80.00 usec  
PLW2 22.00000000 W  
PLW12 0.34375000 W  
PLW13 0.22000000 W

F2 - Processing parameters  
SI 32768  
SF 125.7754563 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40

# $^{13}\text{C}$ -NMR



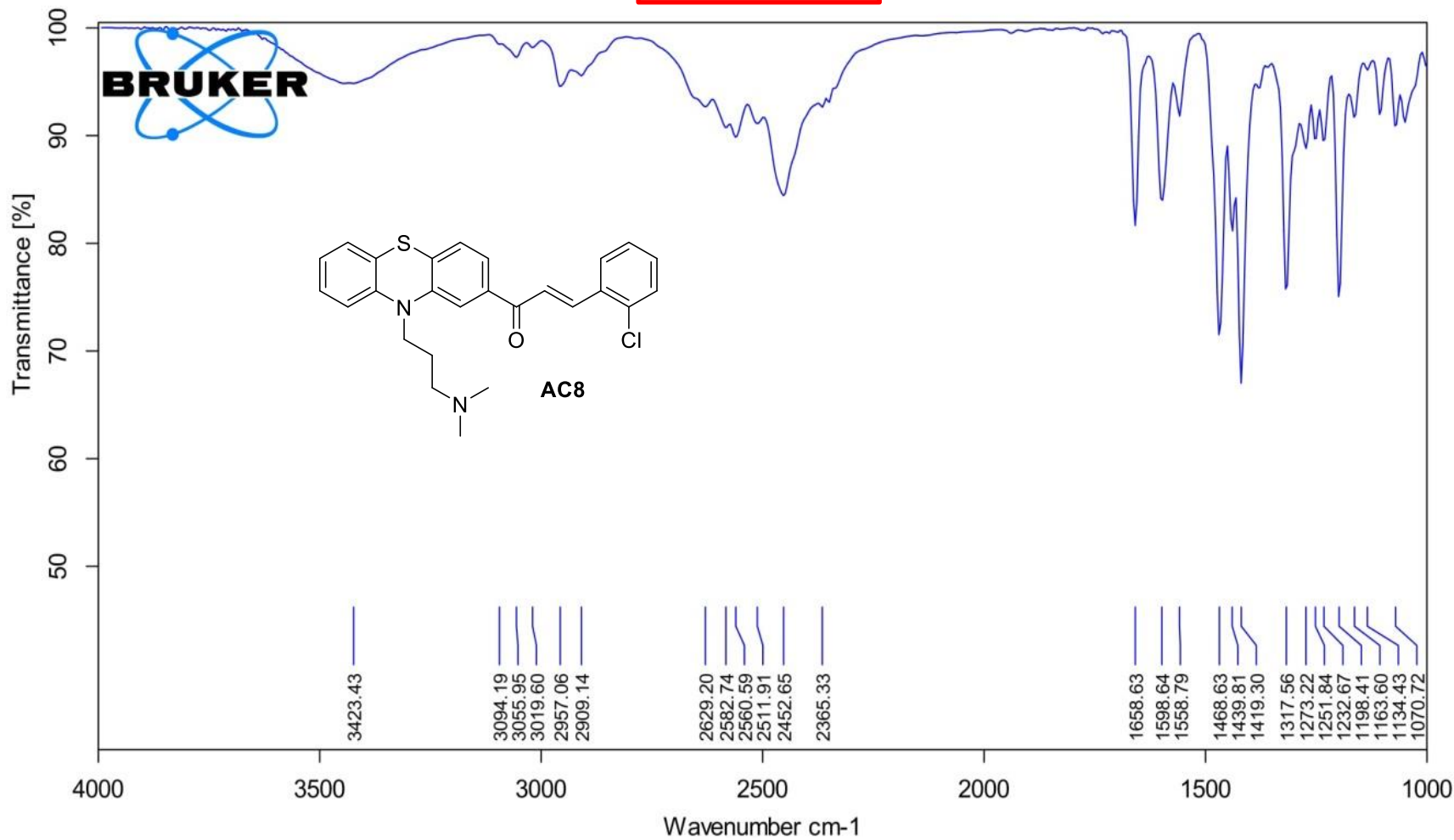
# $^{13}\text{C-NMR}$



C6-DMSO-C13CPD



IR



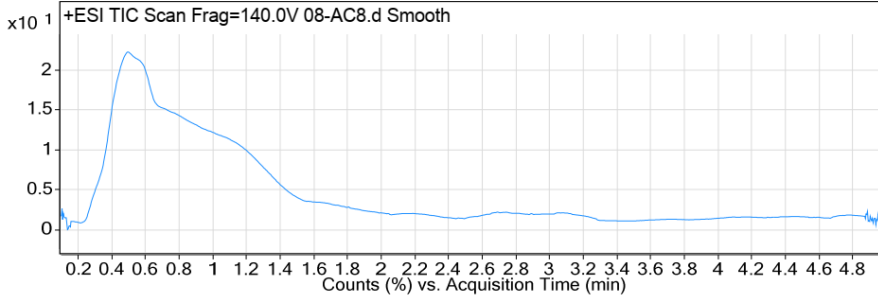
# MS

## Qualitative Analysis Report

<b>Data Filename</b>	08-AC8.d	<b>Sample Name</b>	08-AC8
<b>Sample Type</b>	Sample	<b>Position</b>	P2-C5
<b>Instrument Name</b>	Instrument 1	<b>User Name</b>	
<b>Acq Method</b>	Cot ngan - MSMS_Pos.m	<b>Acquired Time</b>	22/08/2020 11:14:25 AM
<b>IRM Calibration Status</b>	Success	<b>DA Method</b>	COTNGAN.M.m
<b>Comment</b>			
<b>Sample Group</b>		<b>Info.</b>	
<b>Stream Name</b>	LC 1	<b>Acquisition SW</b>	6200 series TOF/6500 series
		<b>Version</b>	Q-TOF B.06.01 (B6172 SP1)

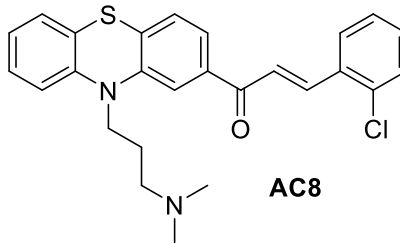
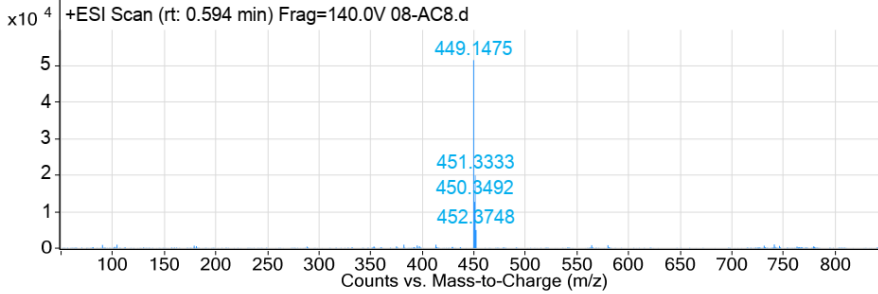
### User Chromatograms

Fragmentor Voltage 140 Collision Energy 0 Ionization Mode ESI



### User Spectra

Fragmentor Voltage 140 Collision Energy 0 Ionization Mode ESI

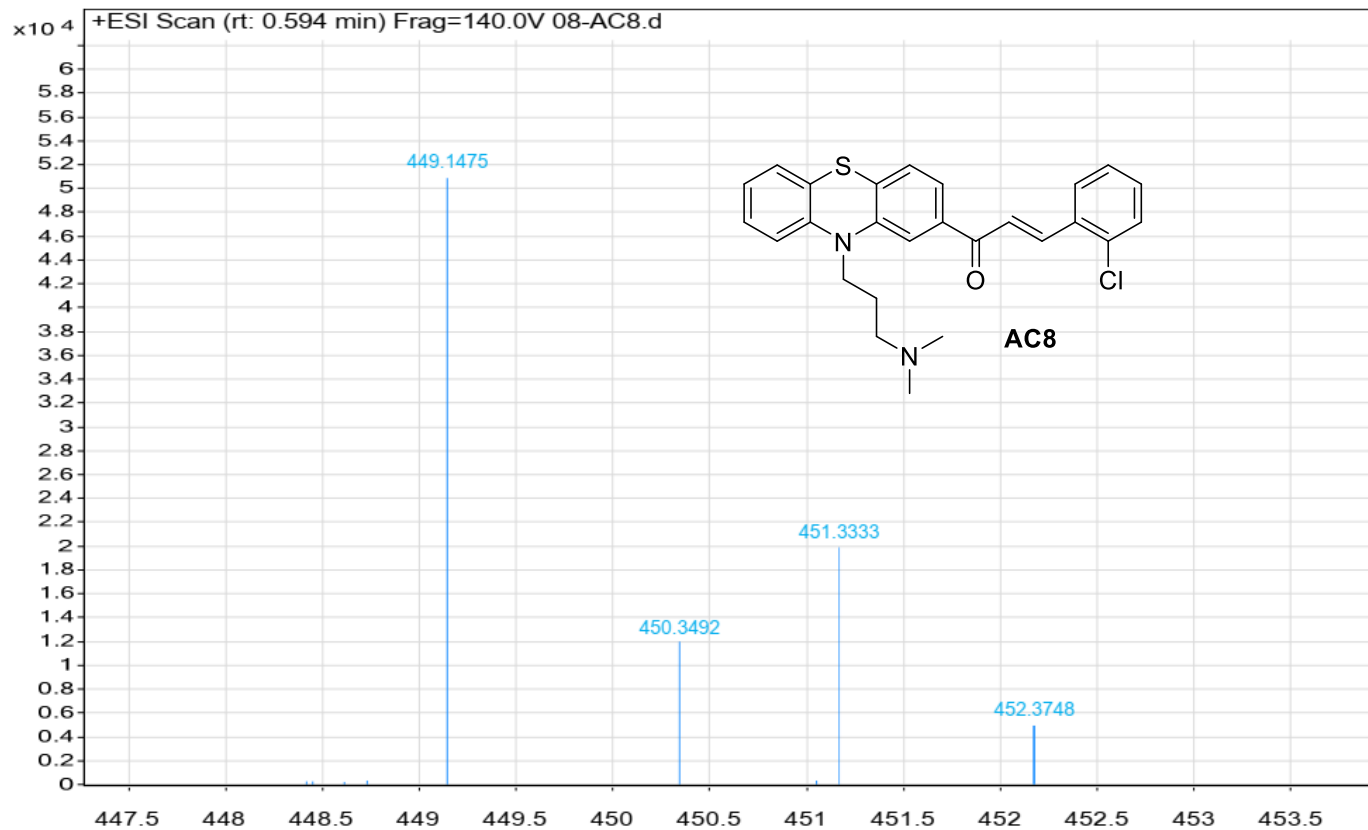


--- End Of Report ---



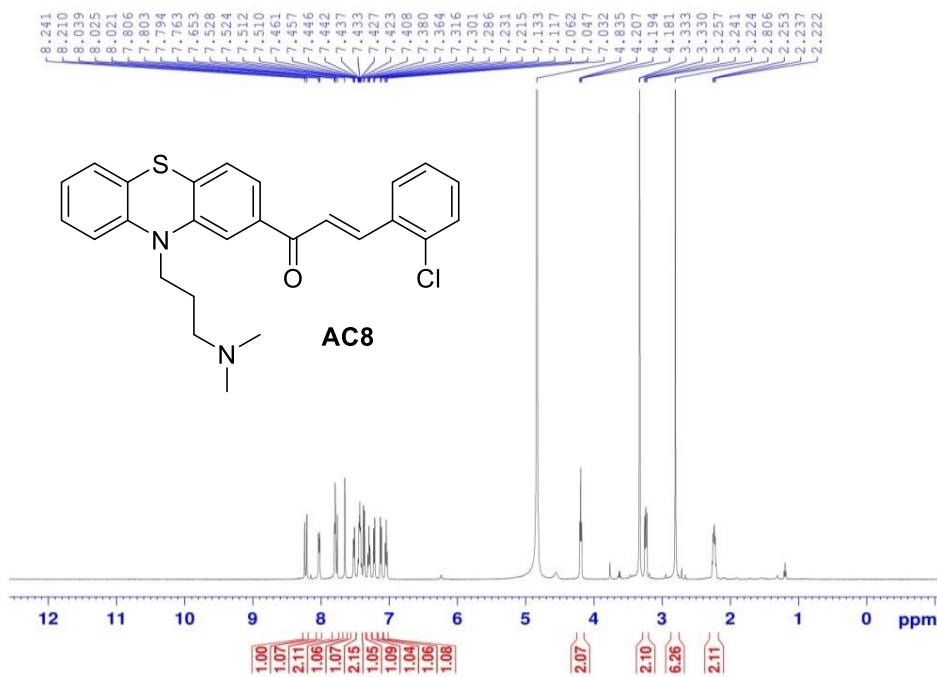
# MS

Sample Name	08-AC8	Position	P2-C5	Instrument Name	Instrument 1	User Name	
Inj Vol	2	InjPosition		SampleType	Sample	IRM Calibration Status	Success
Data Filename	08-AC8.d	ACQ Method	Cot ngan - MSMS_Pos.	Comment		Acquired Time	22/08/2020 11:14:25 AM



# <sup>1</sup>H-NMR

C1-MeOD-1H



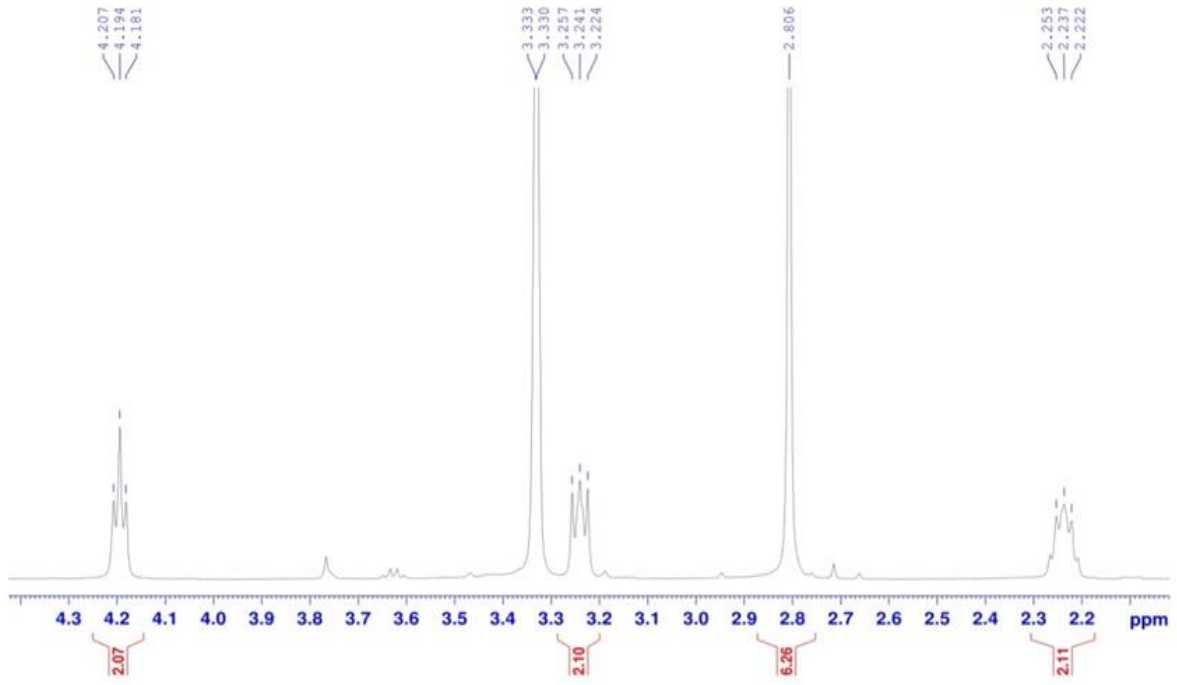
Current Data Parameters  
NAME 113D\_C1  
EXPNO 1  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20170506  
Time 15.08  
INSTRUM spect  
PROBHD 5 mm PABBO BB/  
PULPROG zg30  
TD 65536  
SOLVENT MeOD  
NS 16  
DS 2  
SWH 10000.000 Hz  
FIDRES 0.152588 Hz  
AQ 3.2767999 sec  
RG 157.35  
DW 50.000 usec  
DE 6.50 usec  
TE 303.0 K  
D1 1.00000000 sec  
TDO 1

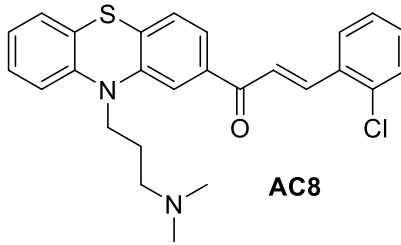
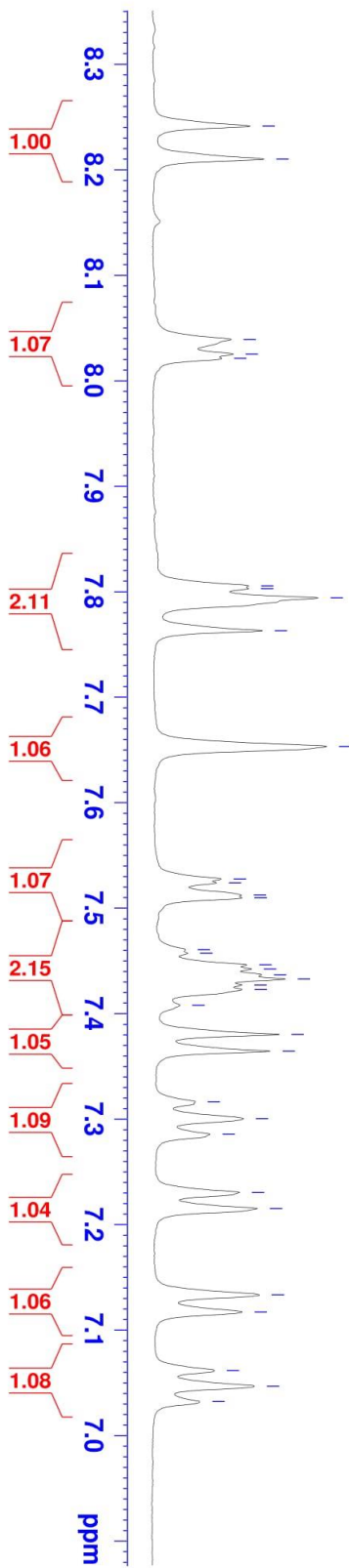
===== CHANNEL f1 =====  
SFO1 500.2030889 MHz  
NUC1 1H  
P1 10.00 usec  
PLW1 22.00000000 W

F2 - Processing parameters  
SI 65536  
SF 500.2000008 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

C1-MeOD-1H



# <sup>1</sup>H-NMR



8.241

8.210

8.039

8.025

8.021

7.806

7.803

7.794

7.763

7.653

7.528

7.524

7.512

7.510

7.461

7.457

7.446

7.442

7.437

7.433

7.427

7.423

7.408

7.380

7.364

7.316

7.301

7.286

7.231

7.215

7.133

7.117

7.062

7.047

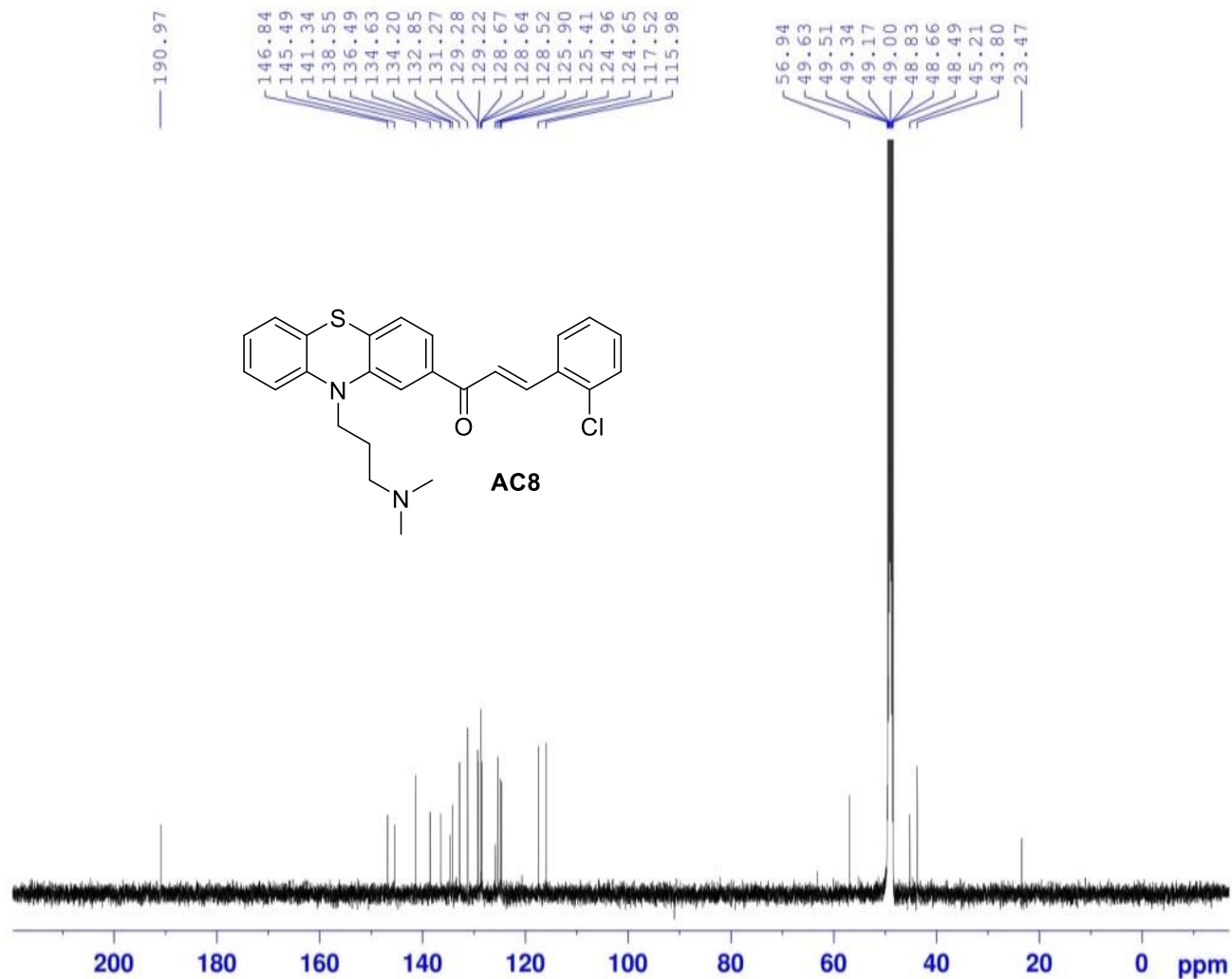
7.032

CDCl<sub>3</sub>-MeOD-1H



# $^{13}\text{C}$ -NMR

C1-MeOD-C13CPD



Current Data Parameters  
NAME 113D\_C1  
EXPNO 2  
PROCNO 1

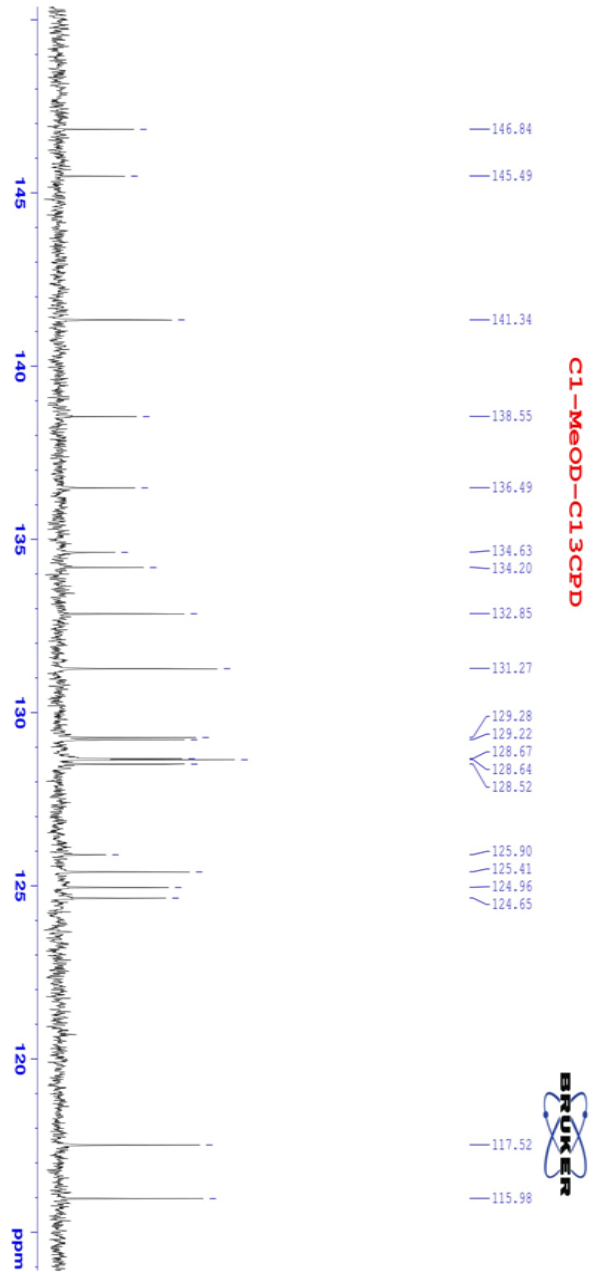
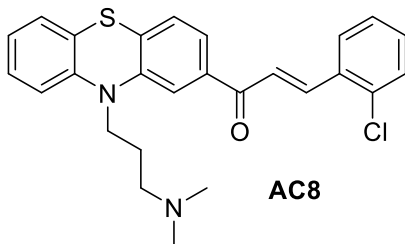
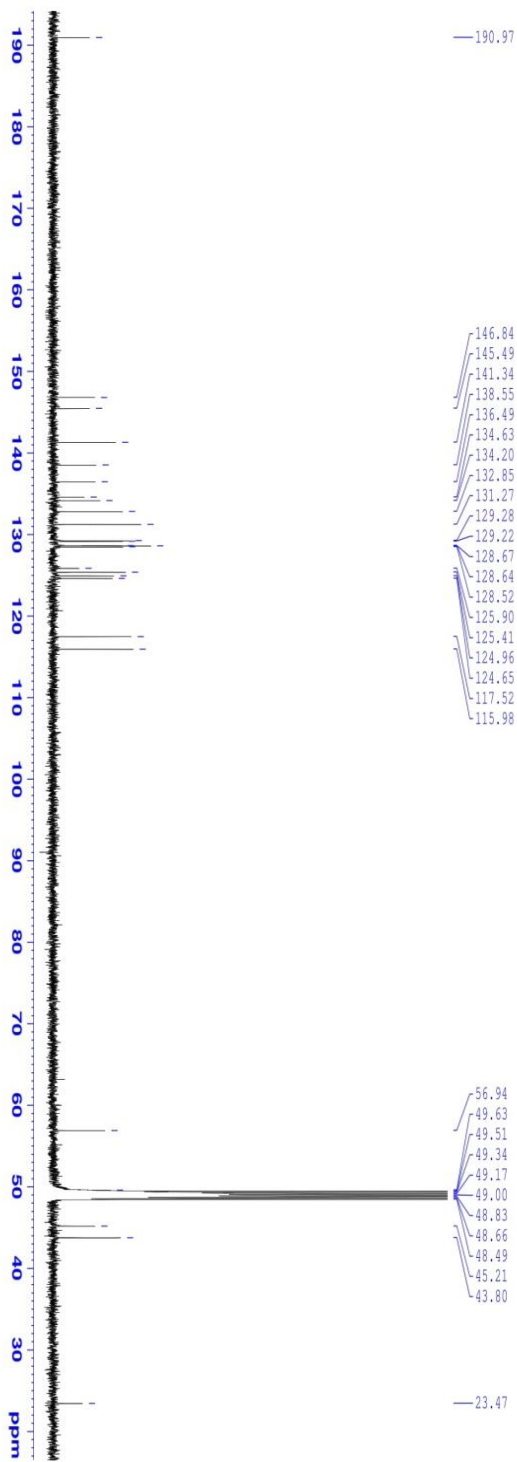
F2 - Acquisition Parameters  
Date\_ 20170607  
Time 7.46  
INSTRUM spect  
PROBHD 5 mm PABBO BB/  
PULPROG zgpg30  
TD 65536  
SOLVENT MeOD  
NS 2048  
DS 4  
SWH 29761.904 Hz  
FIDRES 0.454131 Hz  
AQ 1.1010048 sec  
RG 198.57  
DW 16.800 usec  
DE 6.50 usec  
TE 303.0 K  
D1 2.00000000 sec  
D11 0.03000000 sec  
TD0 1

----- CHANNEL f1 -----  
SFO1 125.7879670 MHz  
NUC1 13C  
P1 10.00 usec  
PLW1 88.00000000 W

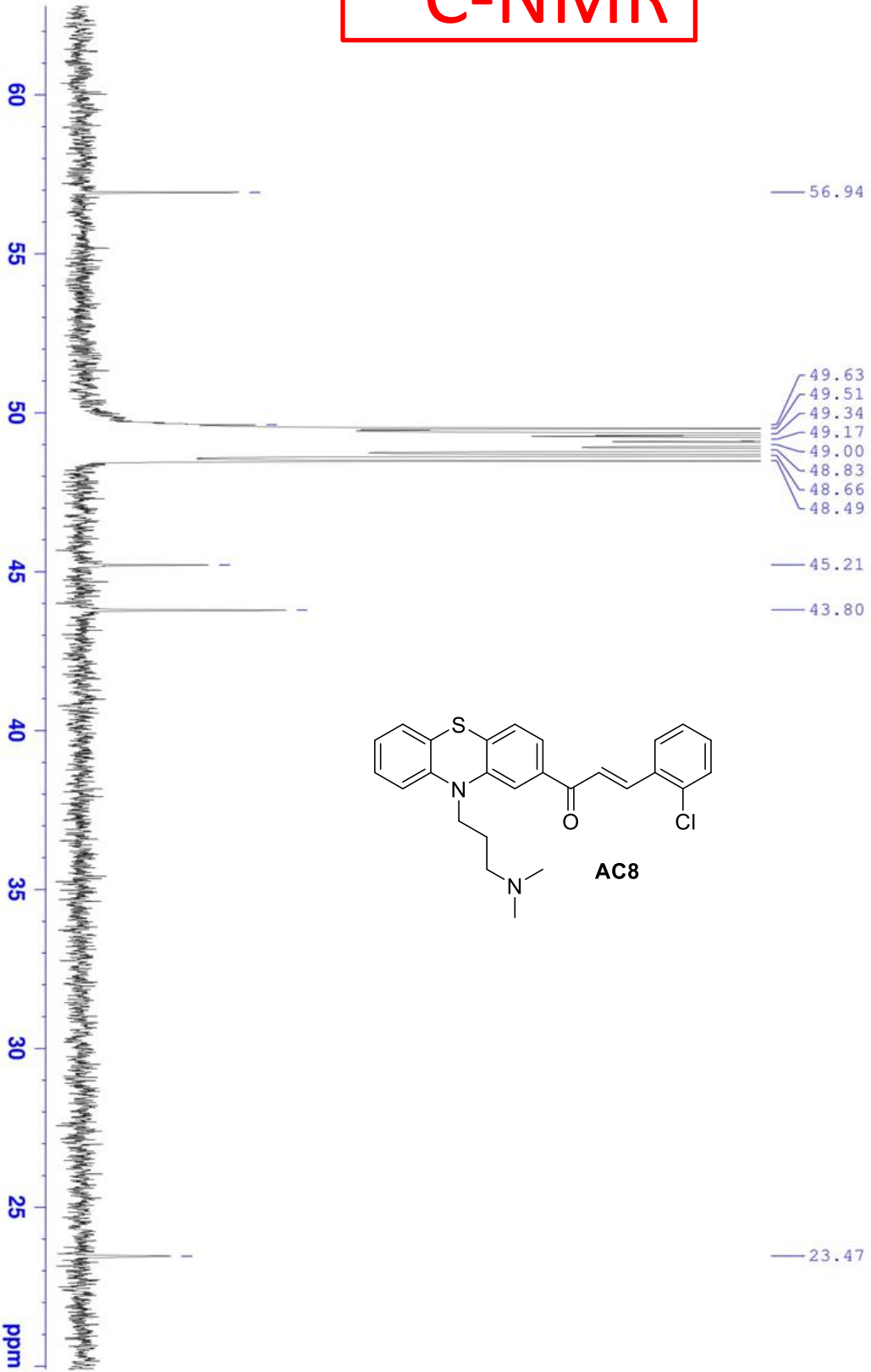
----- CHANNEL f2 -----  
SFO2 500.2020008 MHz  
NUC2 1H  
CPDPRG[2] waltz16  
PCPD2 80.00 usec  
PLW2 22.00000000 W  
PLW12 0.34375000 W  
PLW13 0.22000000 W

F2 - Processing parameters  
SI 32768  
SF 125.7752134 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40

# $^{13}\text{C}$ -NMR



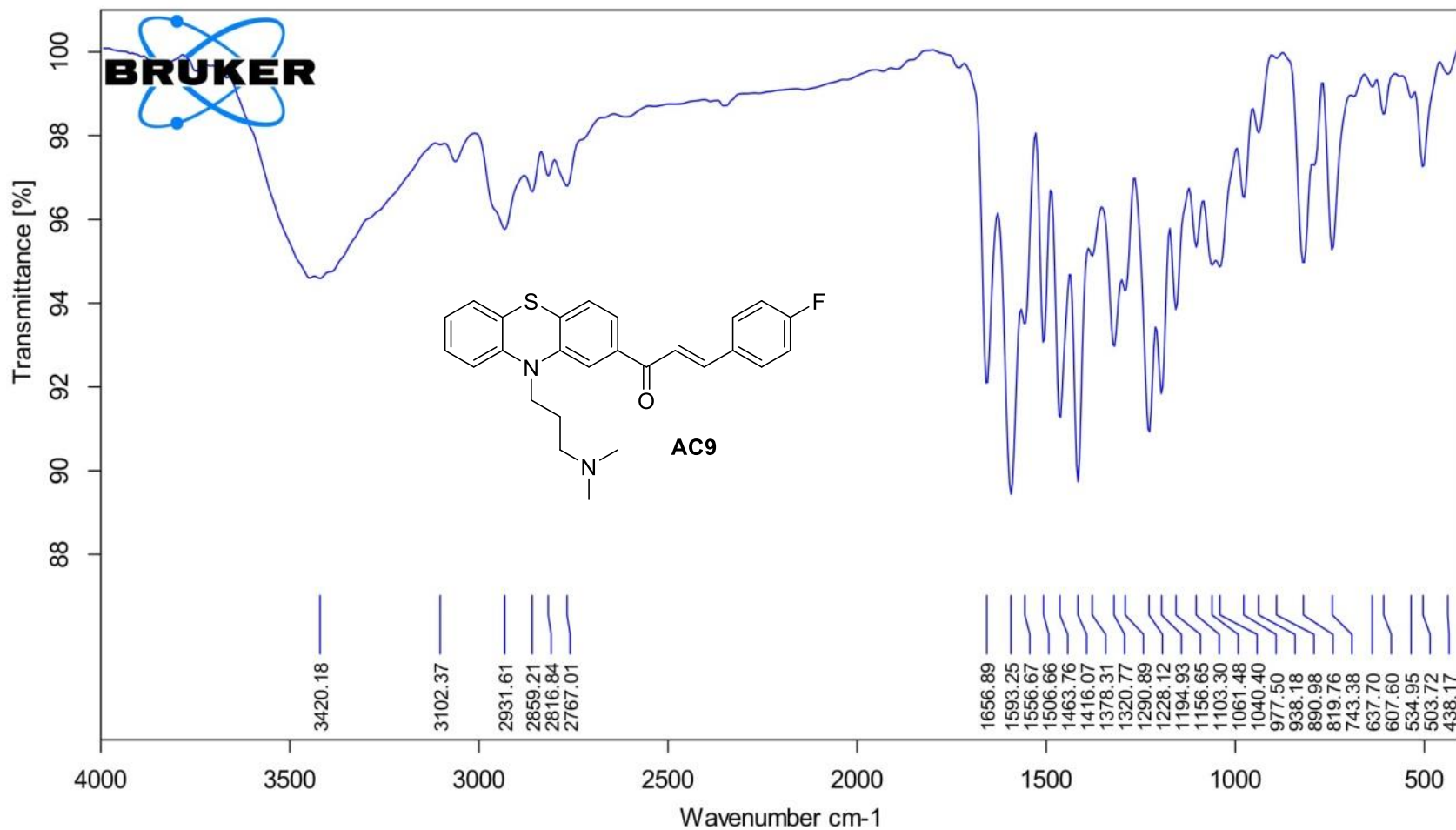
# $^{13}\text{C-NMR}$



CDCl<sub>3</sub>-MeOD-CDCl<sub>3</sub>



# IR





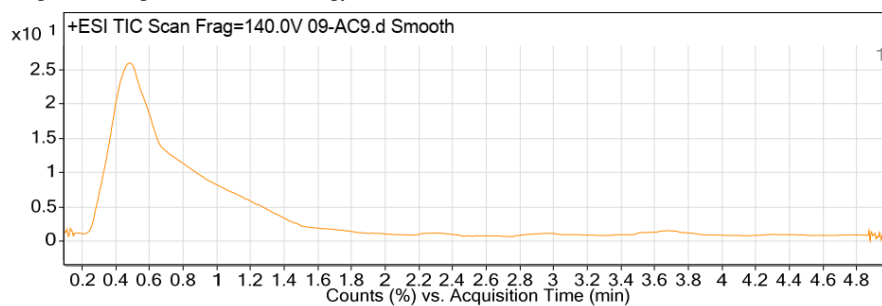
# MS

## Qualitative Analysis Report

<b>Data Filename</b>	09-AC9.d	<b>Sample Name</b>	09-AC9
<b>Sample Type</b>	Sample	<b>Position</b>	P2-B2
<b>Instrument Name</b>	Instrument 1	<b>User Name</b>	
<b>Acq Method</b>	Cot ngan - MSMS_Pos.m	<b>Acquired Time</b>	22/08/2020 11:22:38 AM
<b>IRM Calibration Status</b>	Success	<b>DA Method</b>	COTNGAN.M.m
<b>Comment</b>			
<b>Sample Group</b>		<b>Info.</b>	
<b>Stream Name</b>	LC 1	<b>Acquisition SW</b>	6200 series TOF/6500 series
		<b>Version</b>	Q-TOF B.06.01 (B6172 SP1)

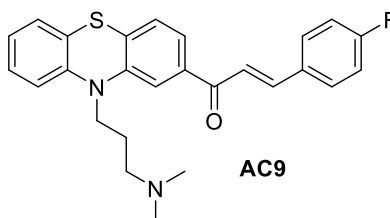
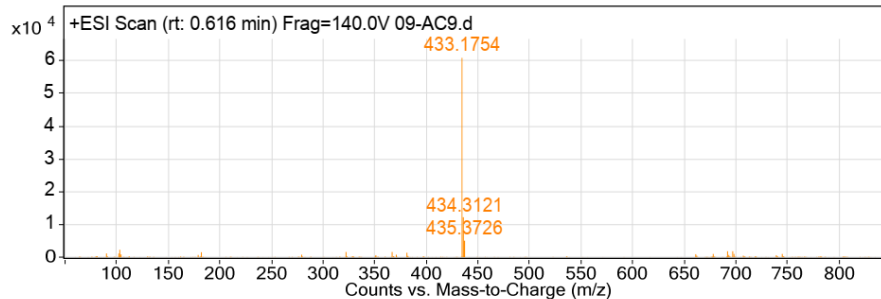
### User Chromatograms

Fragmentor Voltage 140 Collision Energy 0 Ionization Mode ESI



### User Spectra

Fragmentor Voltage 140 Collision Energy 0 Ionization Mode ESI

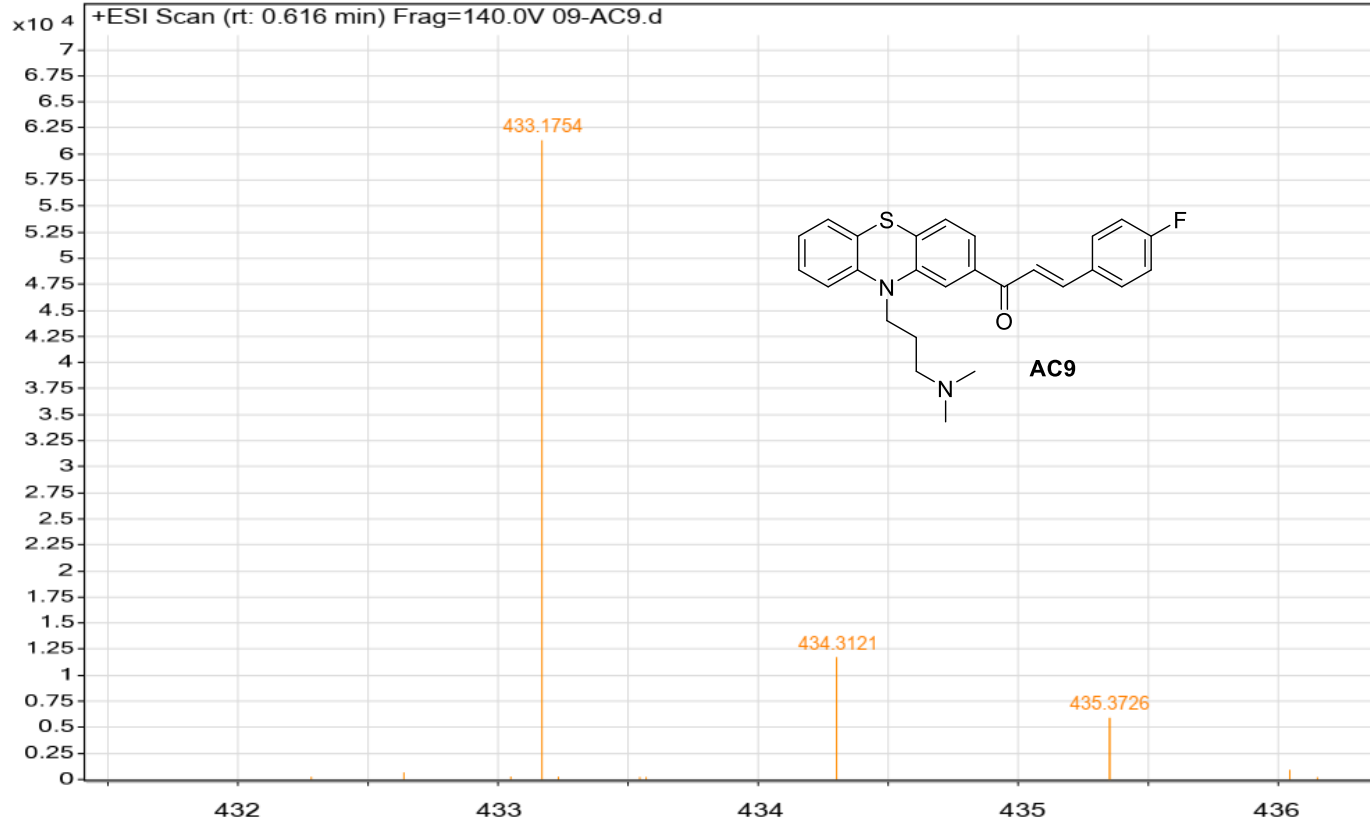


--- End Of Report ---

# MS

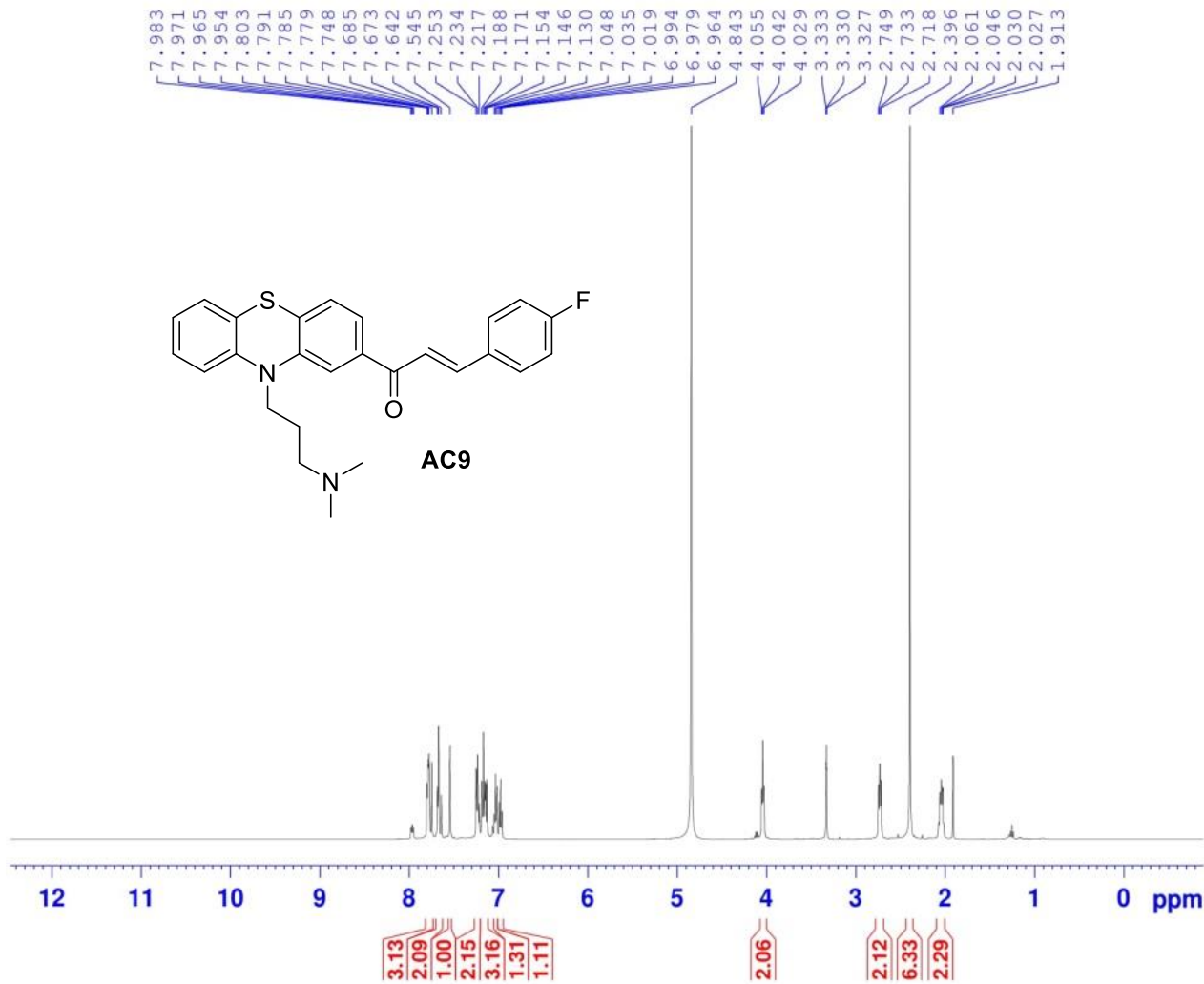
+ESI Scan (rt: 0.616 min) Fraa=140.0V BDE 4.d

Sample Name	09-AC9	Position	P2-B2	Instrument Name	Instrument 1	User Name	
Inj Vol	2	InjPosition		SampleType	Sample	IRM Calibration Status	Success
Data Filename	09-AC9.d	ACQ Method	Cot ngan - MSMS_Pos.	Comment		Acquired Time	22/08/2020 11:22:38 AM



# <sup>1</sup>H-NMR

C11-MeOD-1H



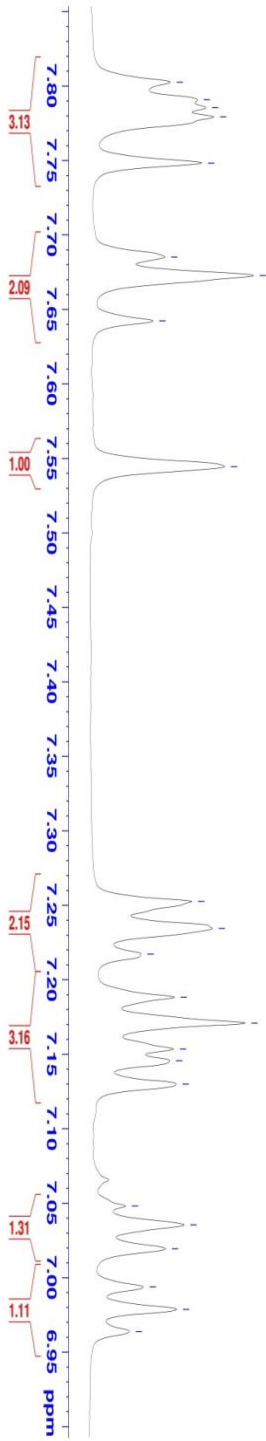
Current Data Parameters  
NAME 114DAO\_C11  
EXPNO 1  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20170704  
Time 11.25  
INSTRUM spect  
PROBHD 5 mm PABBO BB/  
PULPROG zg30  
TD 65536  
SOLVENT MeOD  
NS 16  
DS 2  
SWH 10000.000 Hz  
FIDRES 0.152588 Hz  
AQ 3.2767999 sec  
RG 79.36  
DW 50.000 usec  
DE 6.50 usec  
TE 303.0 K  
D1 1.00000000 sec  
TD0 1

===== CHANNEL f1 =====  
SFO1 500.2030889 MHz  
NUC1 1H  
P1 10.00 usec  
PLW1 22.00000000 W

F2 - Processing parameters  
SI 65536  
SF 500.2000016 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

# <sup>1</sup>H-NMR



7.803  
7.791  
7.785  
7.779  
7.748

7.685  
7.673  
7.642

7.545

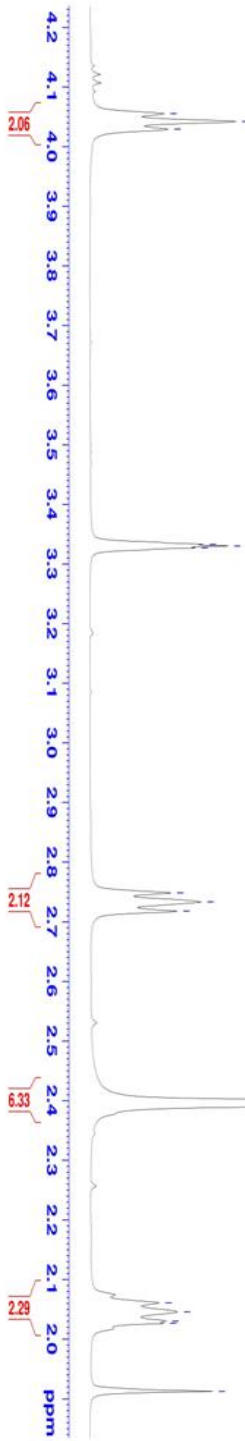
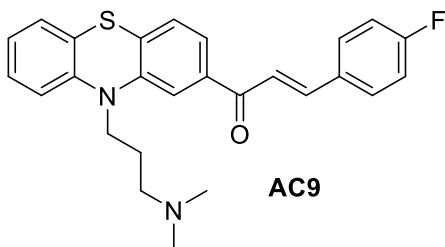
C11-MeOD-1H

7.253  
7.234  
7.217

7.188  
7.171  
7.154  
7.146  
7.130

7.048  
7.035  
7.019  
6.994  
6.979  
6.964

BRUKER



4.055  
4.042  
4.029

3.333  
3.330  
3.327

C11-MeOD-1H

2.749  
2.733  
2.718

2.396

2.061  
2.046  
2.030  
2.027

1.913

BRUKER

# <sup>13</sup>C-NMR

C11-MeOD-C13CPD



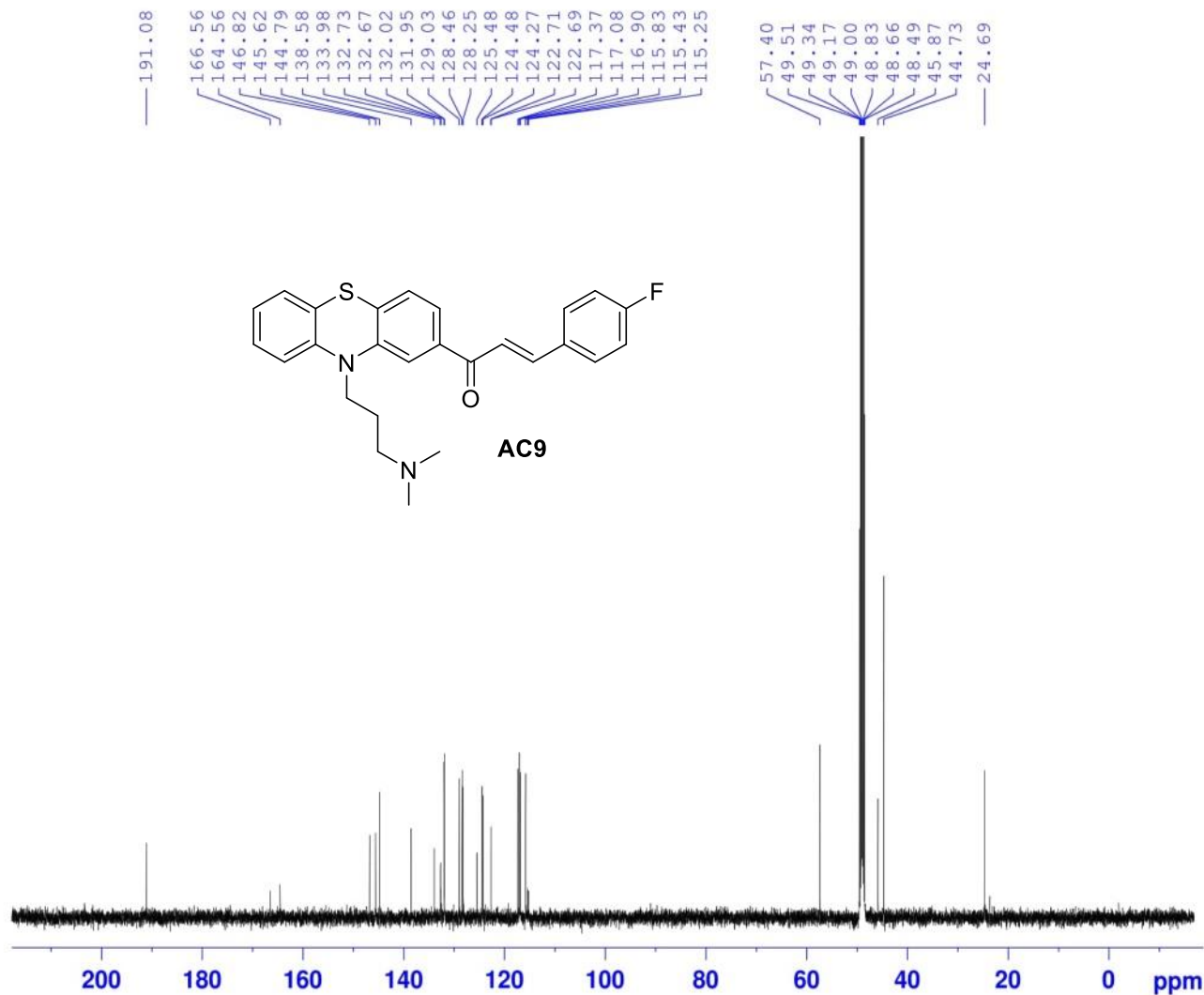
Current Data Parameters  
NAME 113DAO\_C11  
EXPNO 2  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20170705  
Time 14.39  
INSTRUM spect  
PROBHD 5 mm PABBO BB/  
PULPROG zgpg30  
TD 65536  
SOLVENT MeOD  
NS 128  
DS 4  
SWH 29761.904 Hz  
FIDRES 0.454131 Hz  
AQ 1.1010048 sec  
RG 198.57  
DW 16.800 usec  
DE 6.50 usec  
TE 303.0 K  
D1 2.00000000 sec  
D11 0.03000000 sec  
TD0 1

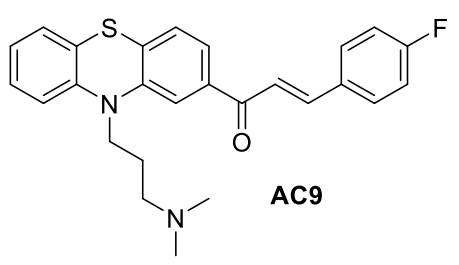
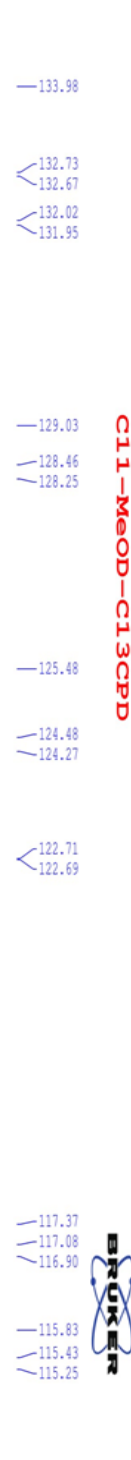
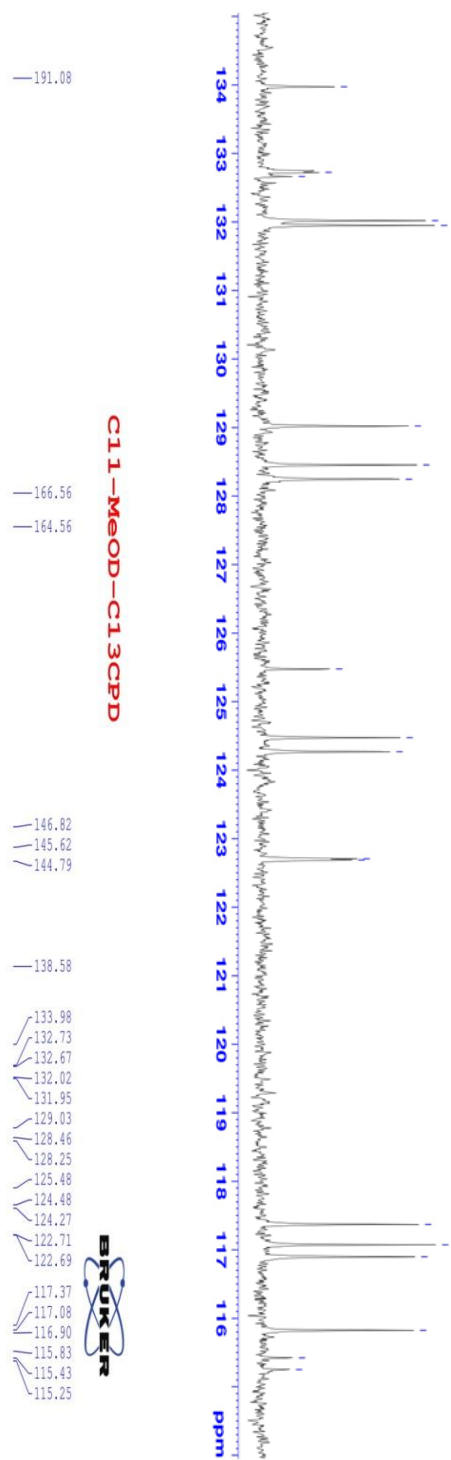
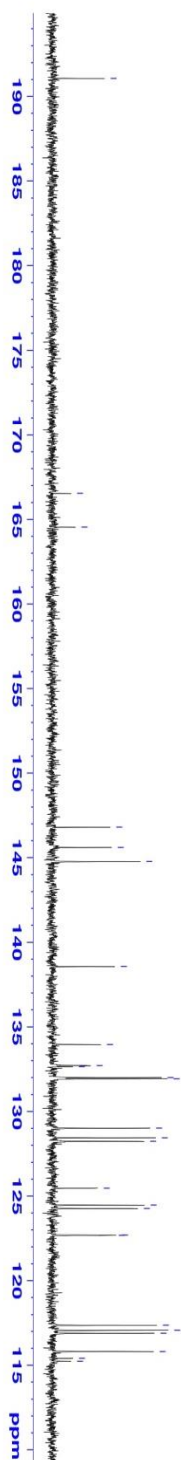
===== CHANNEL f1 =====  
SFO1 125.7879670 MHz  
NUC1 13C  
P1 10.00 usec  
PLW1 88.00000000 W

===== CHANNEL f2 =====  
SFO2 500.2020008 MHz  
NUC2 1H  
CPDPRG[2] waltz16  
PCPD2 80.00 usec  
PLW2 22.00000000 W  
PLW12 0.34375000 W  
PLW13 0.22000000 W

F2 - Processing parameters  
SI 32768  
SF 125.7752168 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40



# $^{13}\text{C}$ -NMR



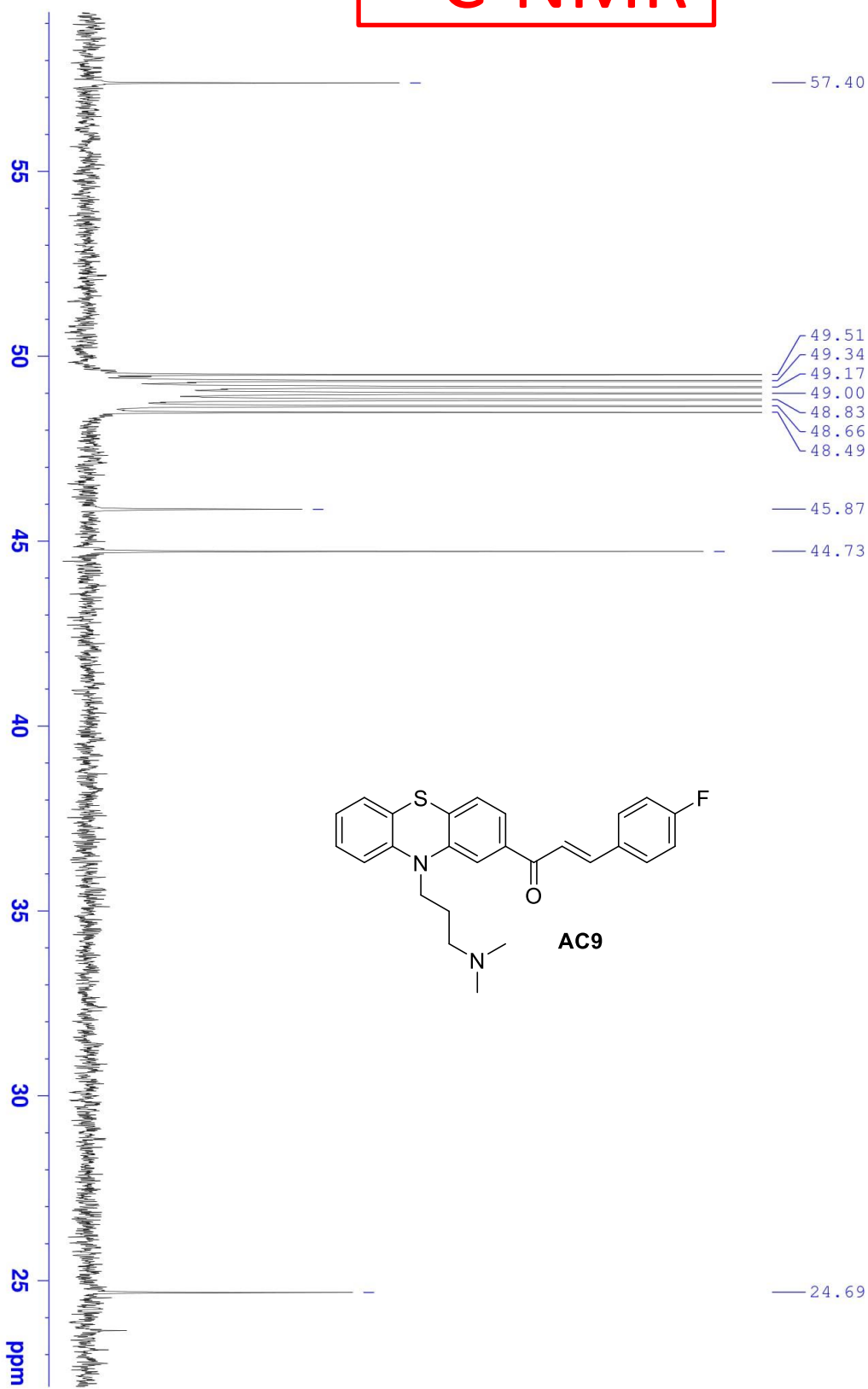
C11-MeOD-C13CPD

C11-MeOD-C13CPD

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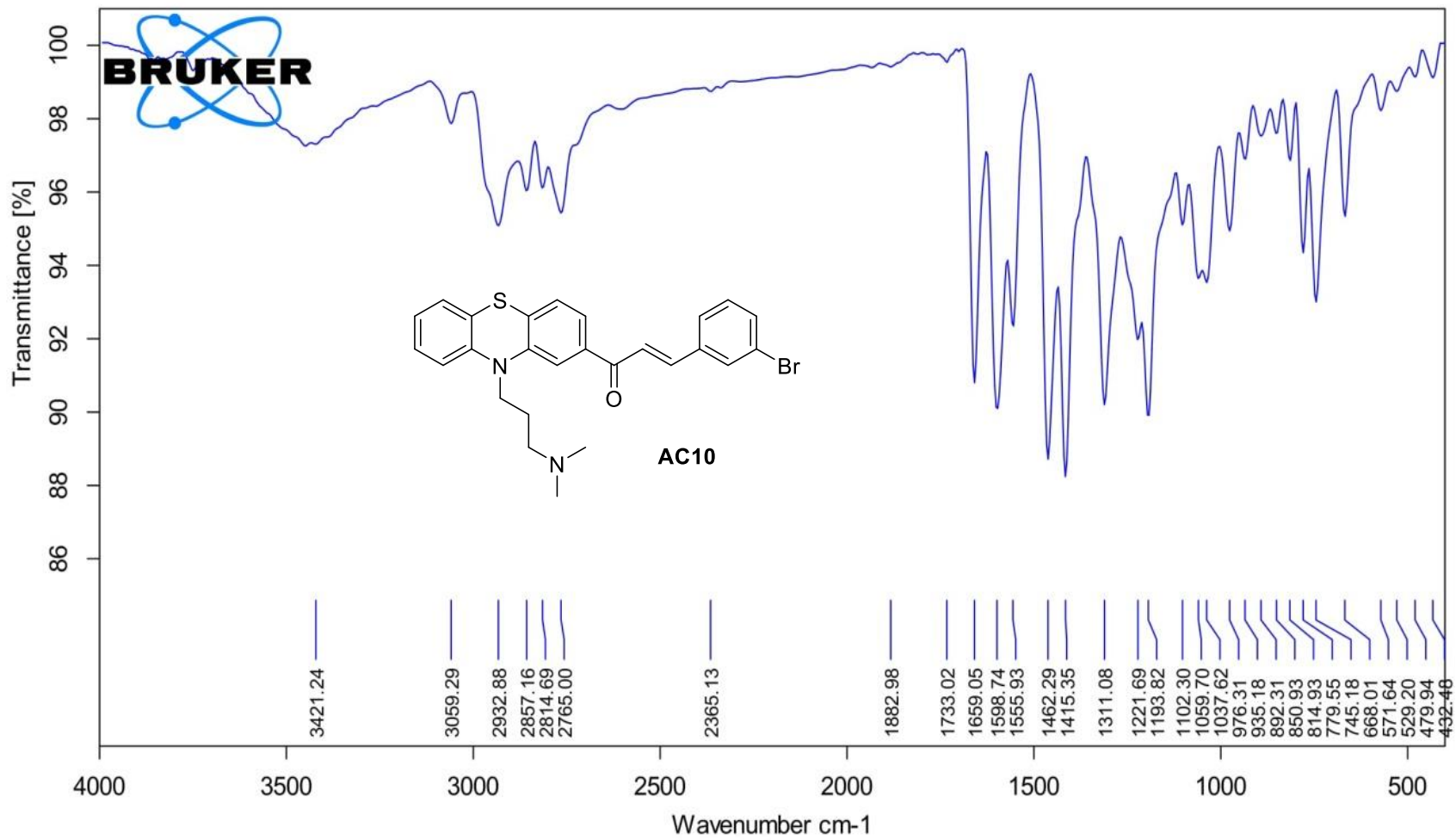
# $^{13}\text{C}$ -NMR



C11-MeOD-C13CPD



IR



E:\OPUS 7\2017\THANG 7\20170714\C14.0	C14	Tensor 27 - Bruker - Germany	7/14/2017
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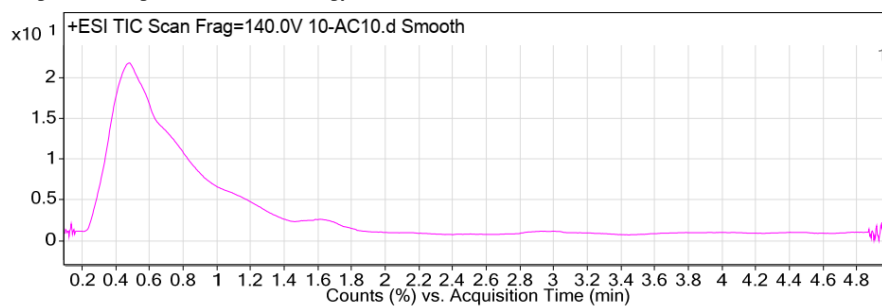
# MS

## Qualitative Analysis Report

<b>Data Filename</b>	10-AC10.d	<b>Sample Name</b>	10-AC10
<b>Sample Type</b>	Sample	<b>Position</b>	P2-C1
<b>Instrument Name</b>	Instrument 1	<b>User Name</b>	
<b>Acq Method</b>	Cot ngan - MSMS_Pos.m	<b>Acquired Time</b>	22/08/2020 11:30:09 AM
<b>IRM Calibration Status</b>	Success	<b>DA Method</b>	COTNGAN.M.m
<b>Comment</b>			
<b>Sample Group</b>		<b>Info.</b>	
<b>Stream Name</b>	LC 1	<b>Acquisition SW</b>	6200 series TOF/6500 series
		<b>Version</b>	Q-TOF B.06.01 (B6172 SP1)

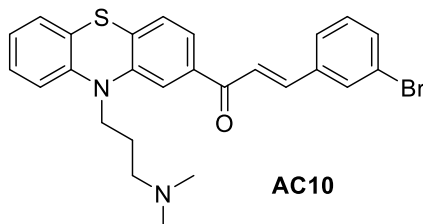
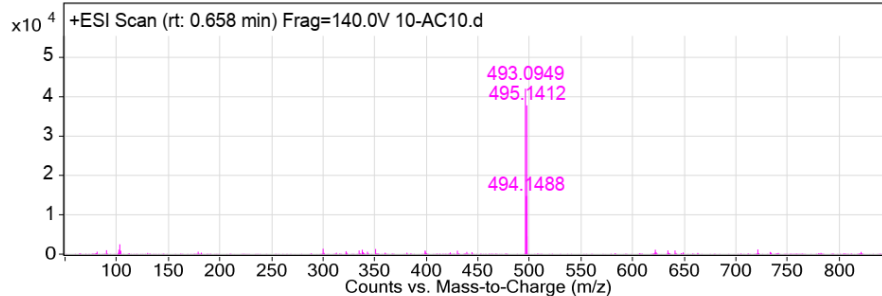
### User Chromatograms

Fragmentor Voltage 140 Collision Energy 0 Ionization Mode ESI



### User Spectra

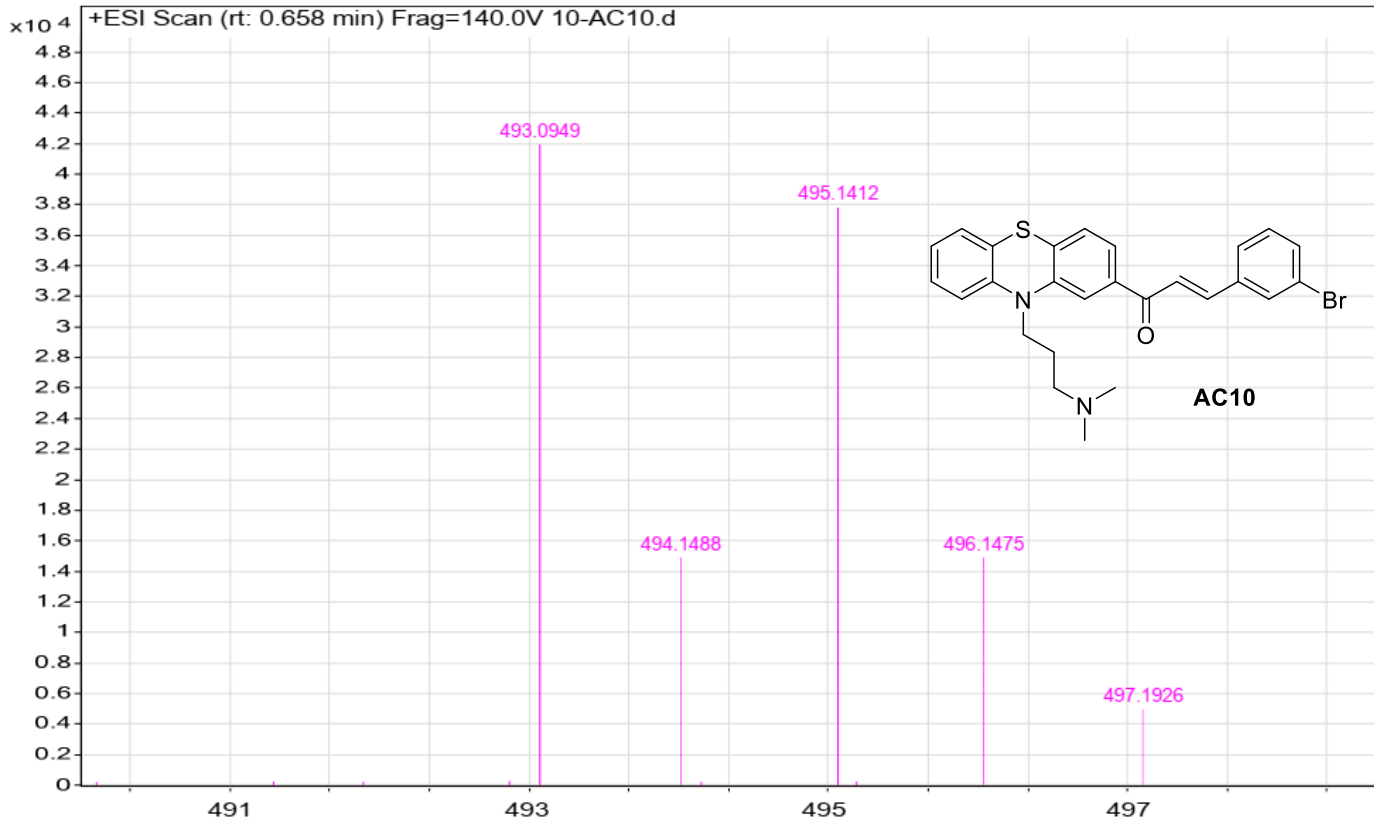
Fragmentor Voltage 140 Collision Energy 0 Ionization Mode ESI



--- End Of Report ---

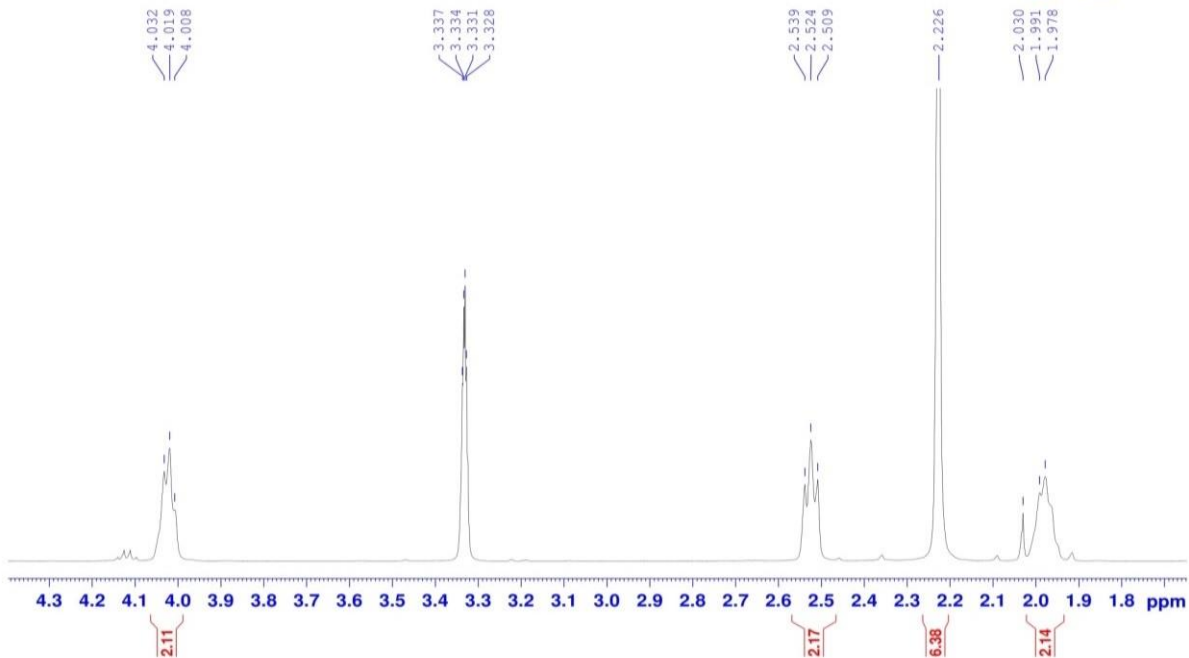
# MS

Sample Name	10-AC10	Position	P2-C1	Instrument Name	Instrument 1	User Name	
Inj Vol	2	InjPosition		SampleType	Sample	IRM Calibration Status	Success
Data Filename	10-AC10.d	ACQ Method	Cot ngan - MSMS_Pos.	Comment		Acquired Time	22/08/2020 11:30:09 AM

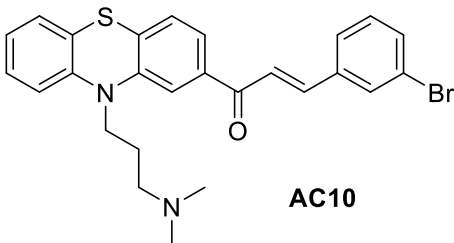
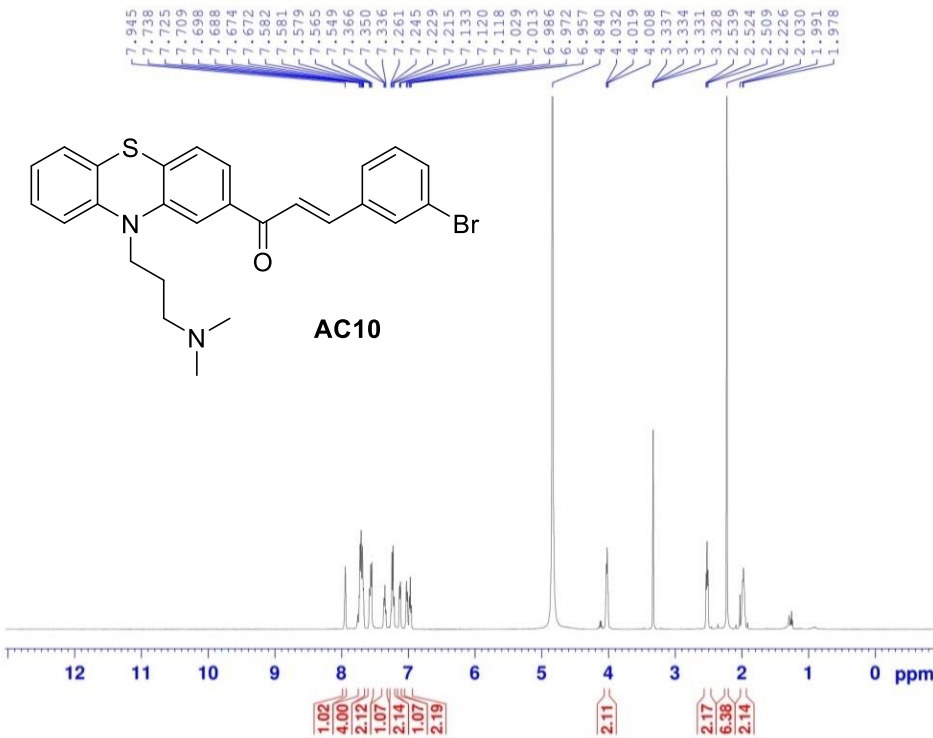


# <sup>1</sup>H-NMR

C14-MeOD-1H



C14-MeOD-1H



Current Data Parameters

NAME 114DAO\_C14  
 EXPNO 1  
 PROCNO 1

F2 - Acquisition Parameters

Date\_ 20170704  
 Time 11.35  
 INSTRUM spect  
 PROBHD 5 mm PABBO BB/  
 PULPROG zg30  
 TD 65536  
 SOLVENT MeOD  
 NS 16  
 DS 2  
 SWH 10000.000 Hz  
 FIDRES 0.152588 Hz  
 AQ 3.2767999 sec  
 RE 97.76  
 DW 50.000 usec  
 DE 6.50 usec  
 TE 303.0 K  
 D1 1.00000000 sec  
 TDO 1

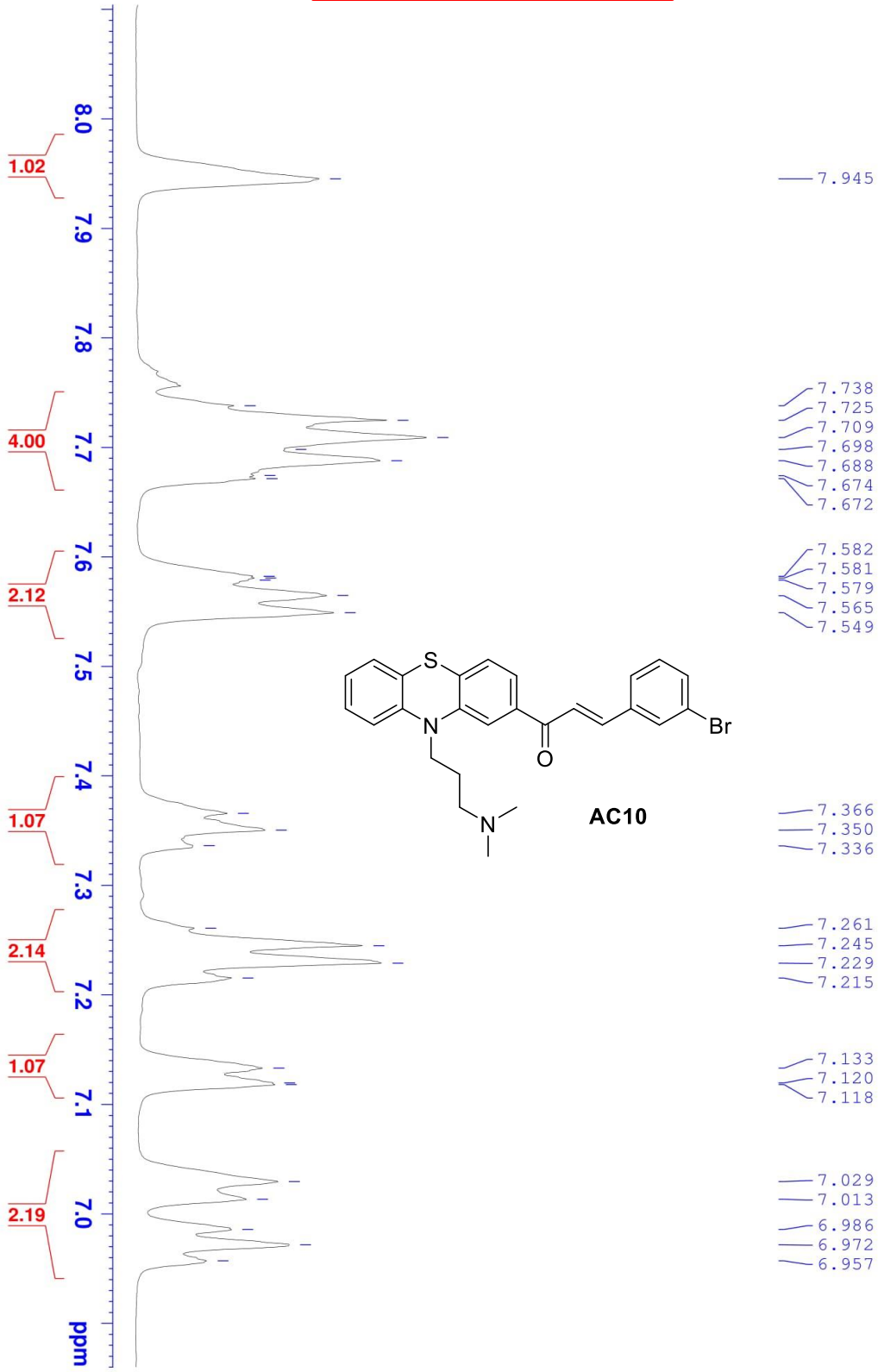
===== CHANNEL f1 =====

SFO1 500.2030889 MHz  
 NUC1 1H  
 P1 10.00 usec  
 PLW1 22.00000000 W

F2 - Processing parameters

SI 65536  
 SF 500.2000000 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 1.00

# <sup>1</sup>H-NMR



C14-MeOD-1H



# <sup>13</sup>C-NMR

C14-MeOD-C13CPD



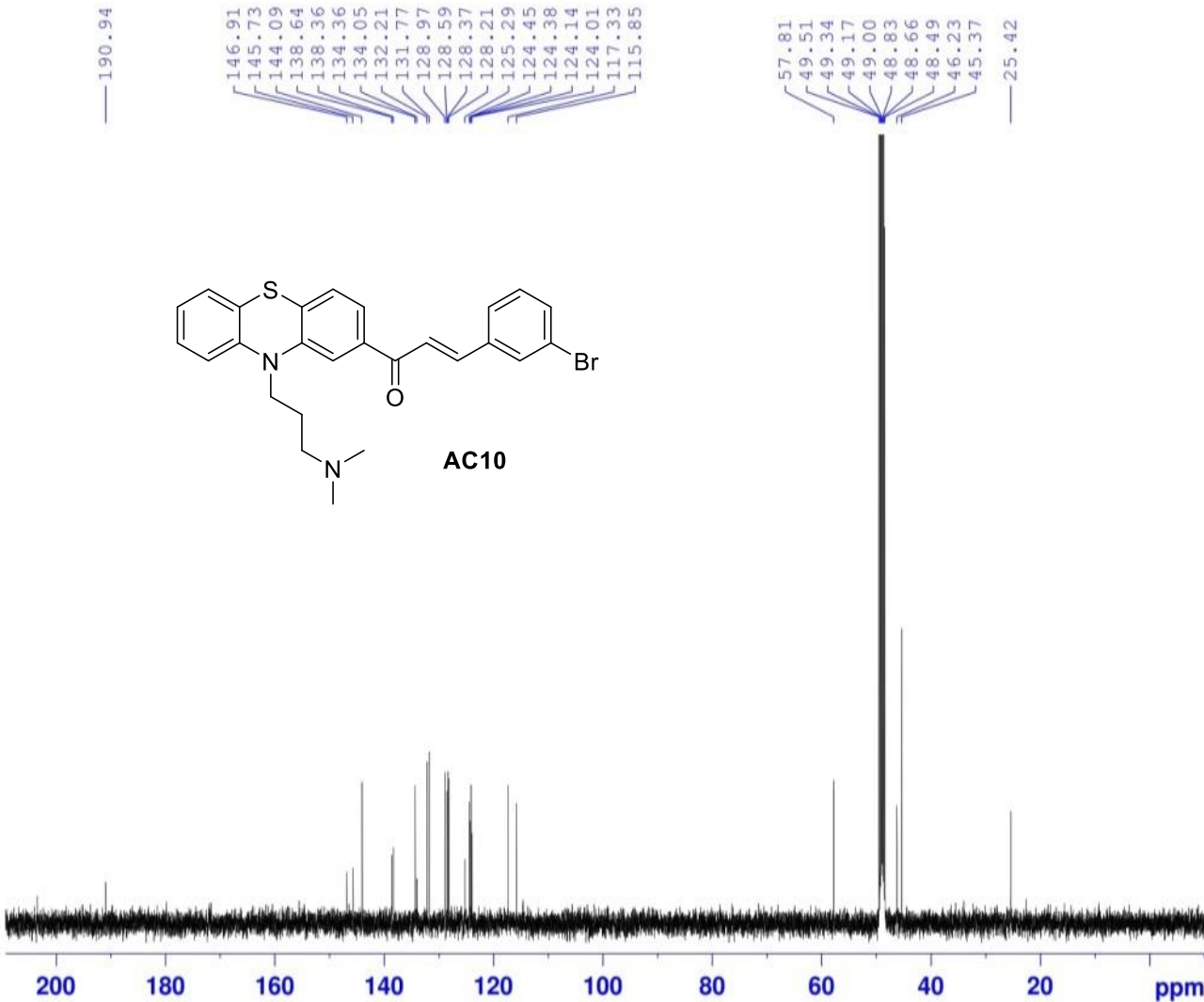
Current Data Parameters  
NAME 114DAO\_C14  
EXPNO 2  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20170705  
Time 15.02  
INSTRUM spect  
PROBHD 5 mm PABBO BB/  
PULPROG zgpg30  
TD 65536  
SOLVENT MeOD  
NS 256  
DS 4  
SWH 29761.904 Hz  
FIDRES 0.454131 Hz  
AQ 1.1010048 sec  
RG 198.57  
DW 16.800 usec  
DE 6.50 usec  
TE 303.0 K  
D1 2.00000000 sec  
D11 0.03000000 sec  
TD0 1

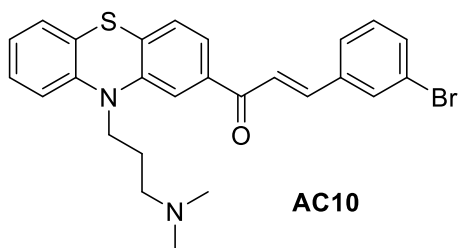
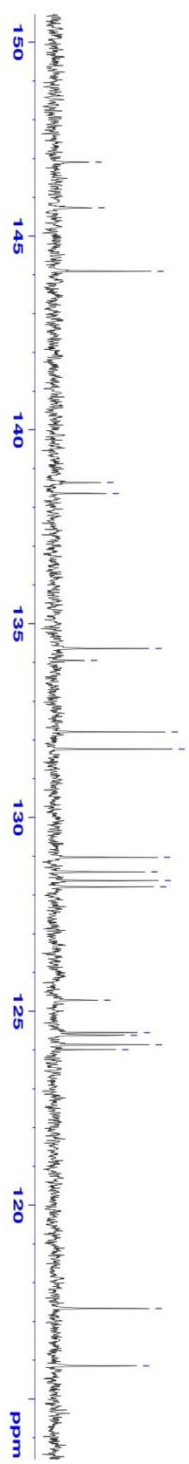
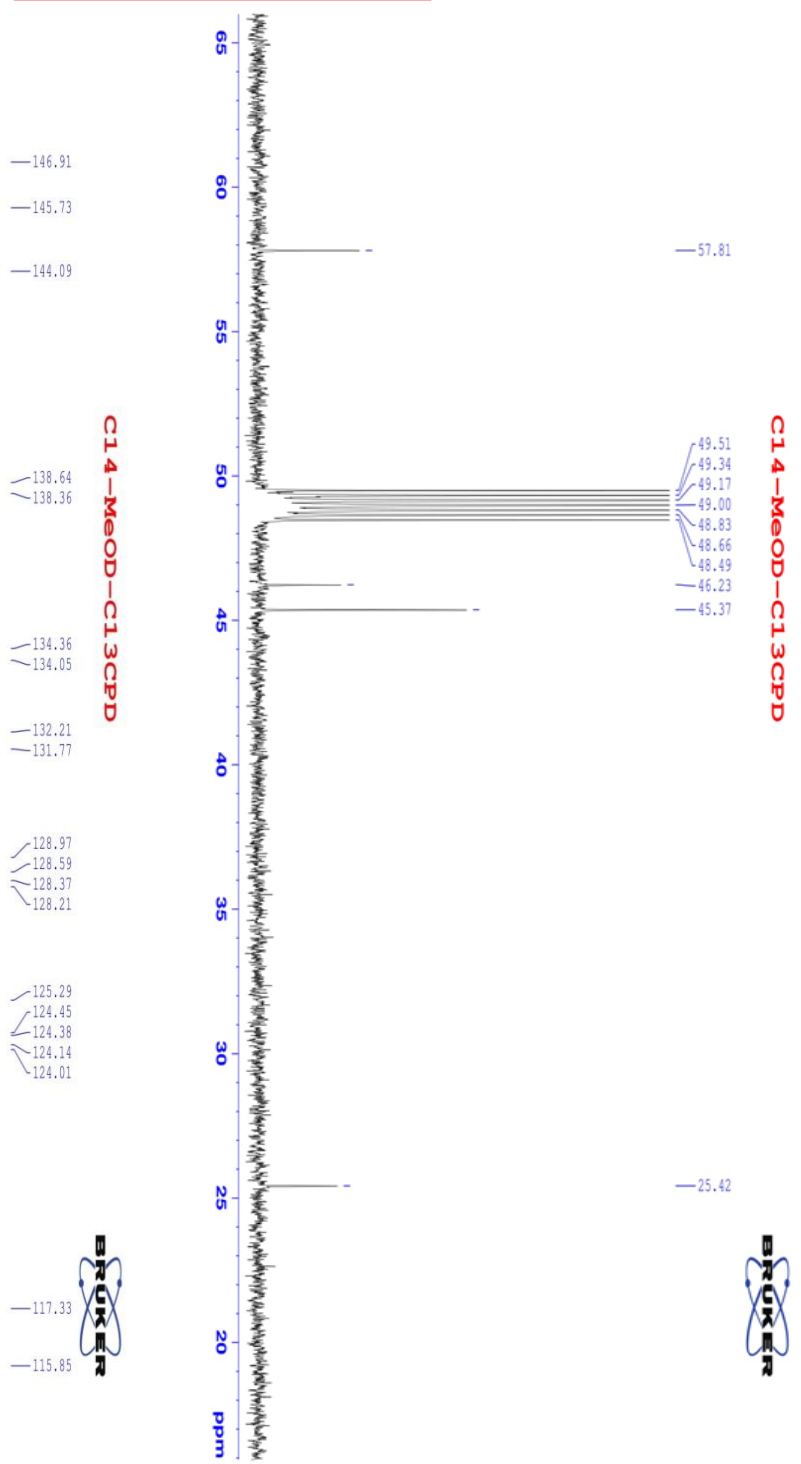
----- CHANNEL f1 -----  
SFO1 125.7879670 MHz  
NUC1 13C  
P1 10.00 usec  
PLW1 88.00000000 W

----- CHANNEL f2 -----  
SFO2 500.2020008 MHz  
NUC2 1H  
CPDPRG[2] waltz16  
PCPD2 80.00 usec  
PLW2 22.00000000 W  
PLW12 0.34375000 W  
PLW13 0.22000000 W

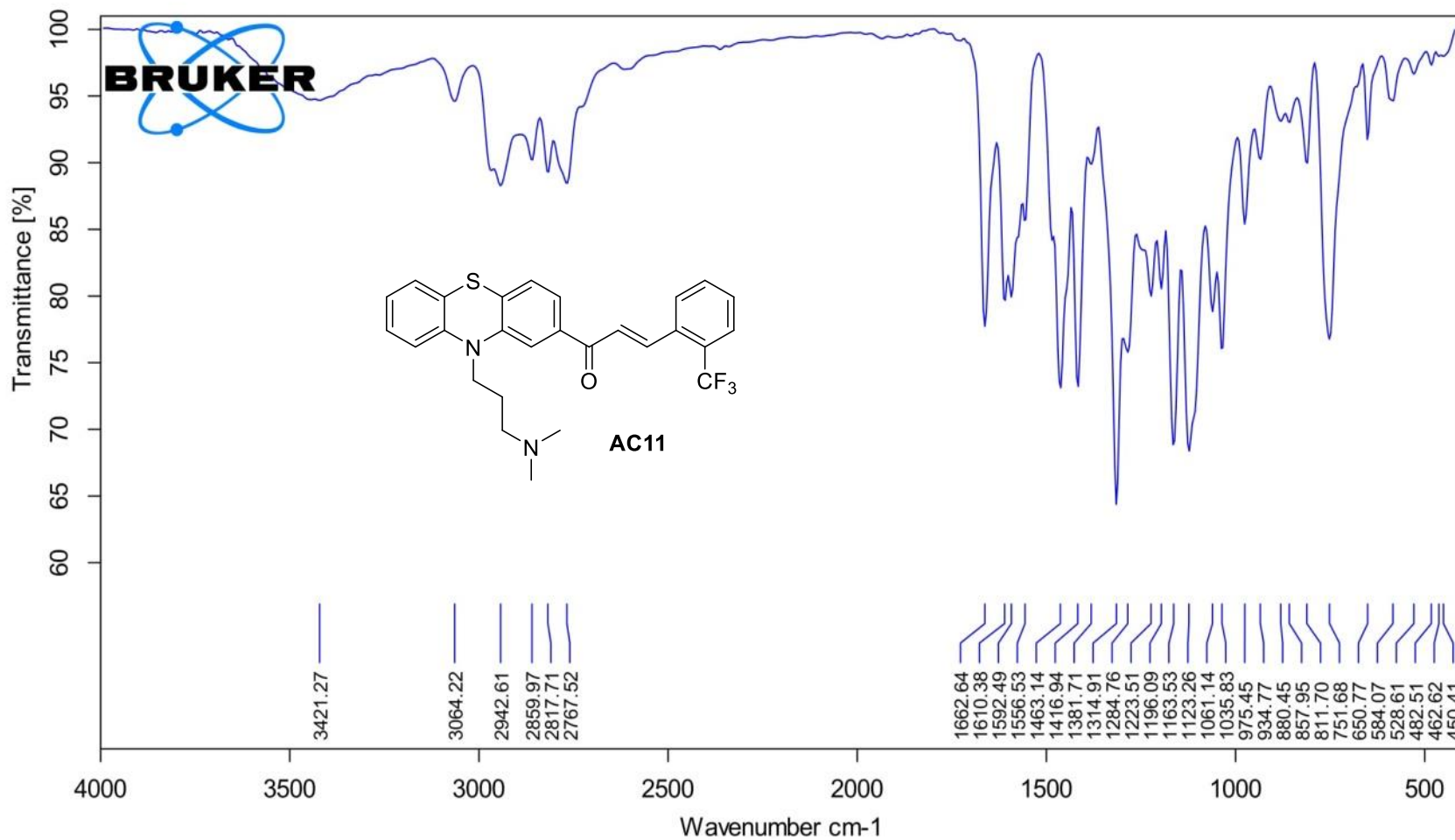
F2 - Processing parameters  
SI 32768  
SF 125.7752156 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40



# $^{13}\text{C}$ -NMR



IR



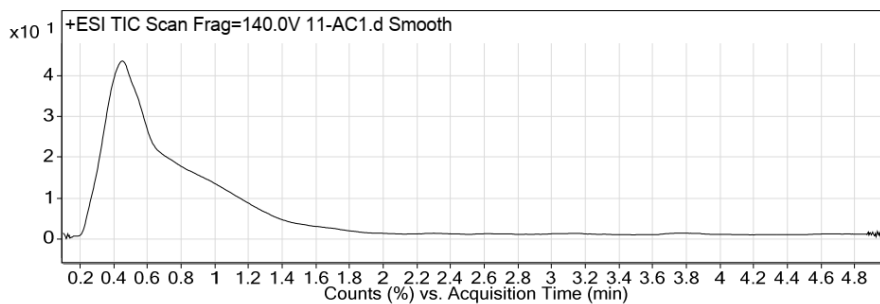
# MS

## Qualitative Analysis Report

<b>Data Filename</b>	11-AC11.d	<b>Sample Name</b>	11-AC11
<b>Sample Type</b>	Sample	<b>Position</b>	P2-B1
<b>Instrument Name</b>	Instrument 1	<b>User Name</b>	
<b>Acq Method</b>	Cot ngan - MSMS_Pos.m	<b>Acquired Time</b>	22/08/2020 11:40:04 AM
<b>IRM Calibration Status</b>	Success	<b>DA Method</b>	COTNGAN.M.m
<b>Comment</b>			
<b>Sample Group</b>		<b>Info.</b>	
<b>Stream Name</b>	LC 1	<b>Acquisition SW</b>	6200 series TOF/6500 series
		<b>Version</b>	Q-TOF B.06.01 (B6172 SP1)

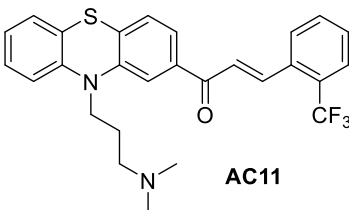
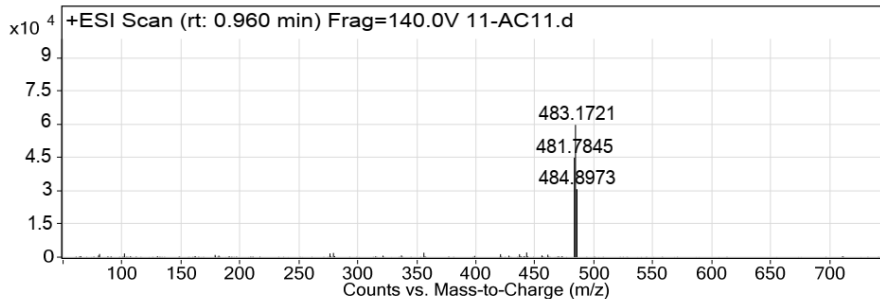
### User Chromatograms

Fragmentor Voltage 140 Collision Energy 0 Ionization Mode ESI



### User Spectra

Fragmentor Voltage 140 Collision Energy 0 Ionization Mode ESI

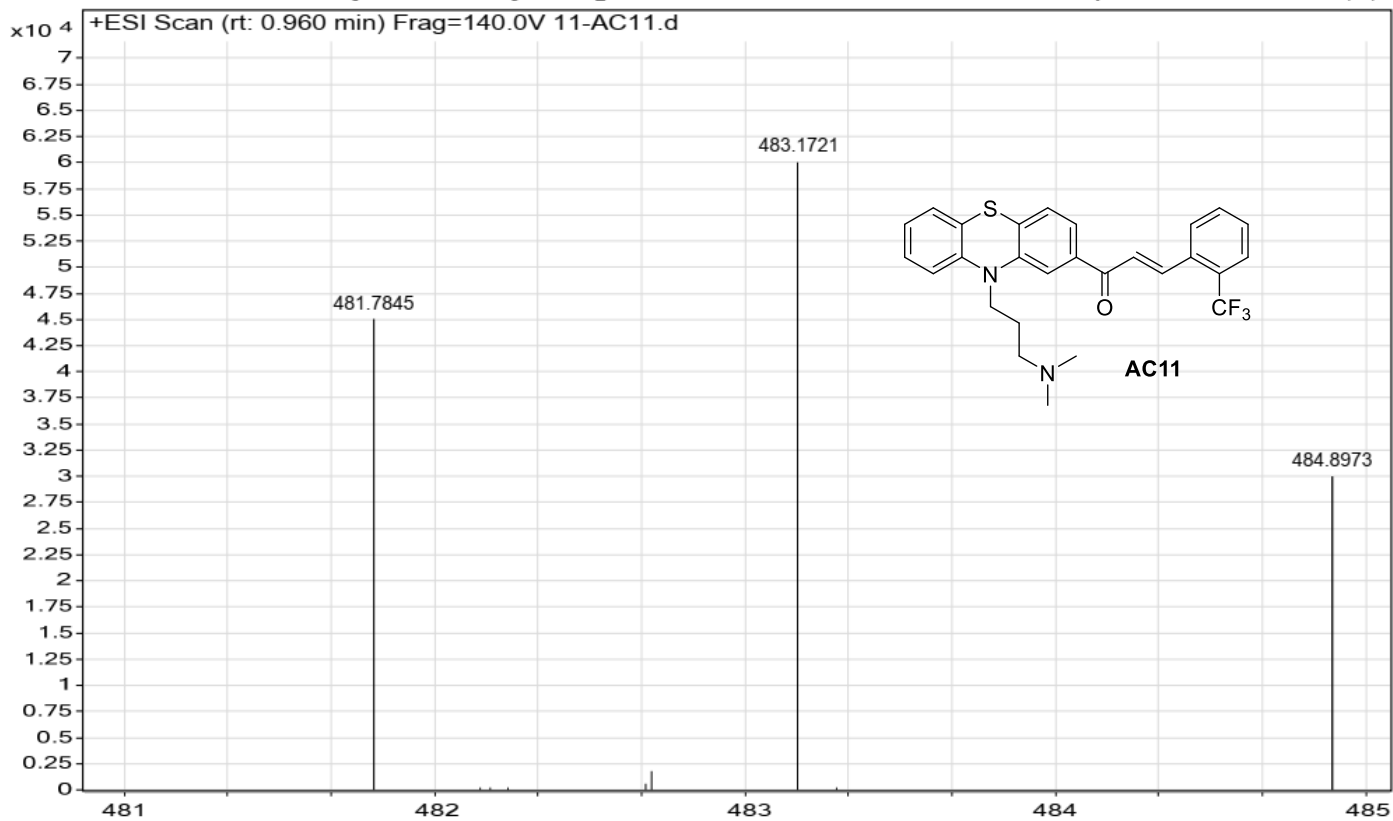


--- End Of Report ---



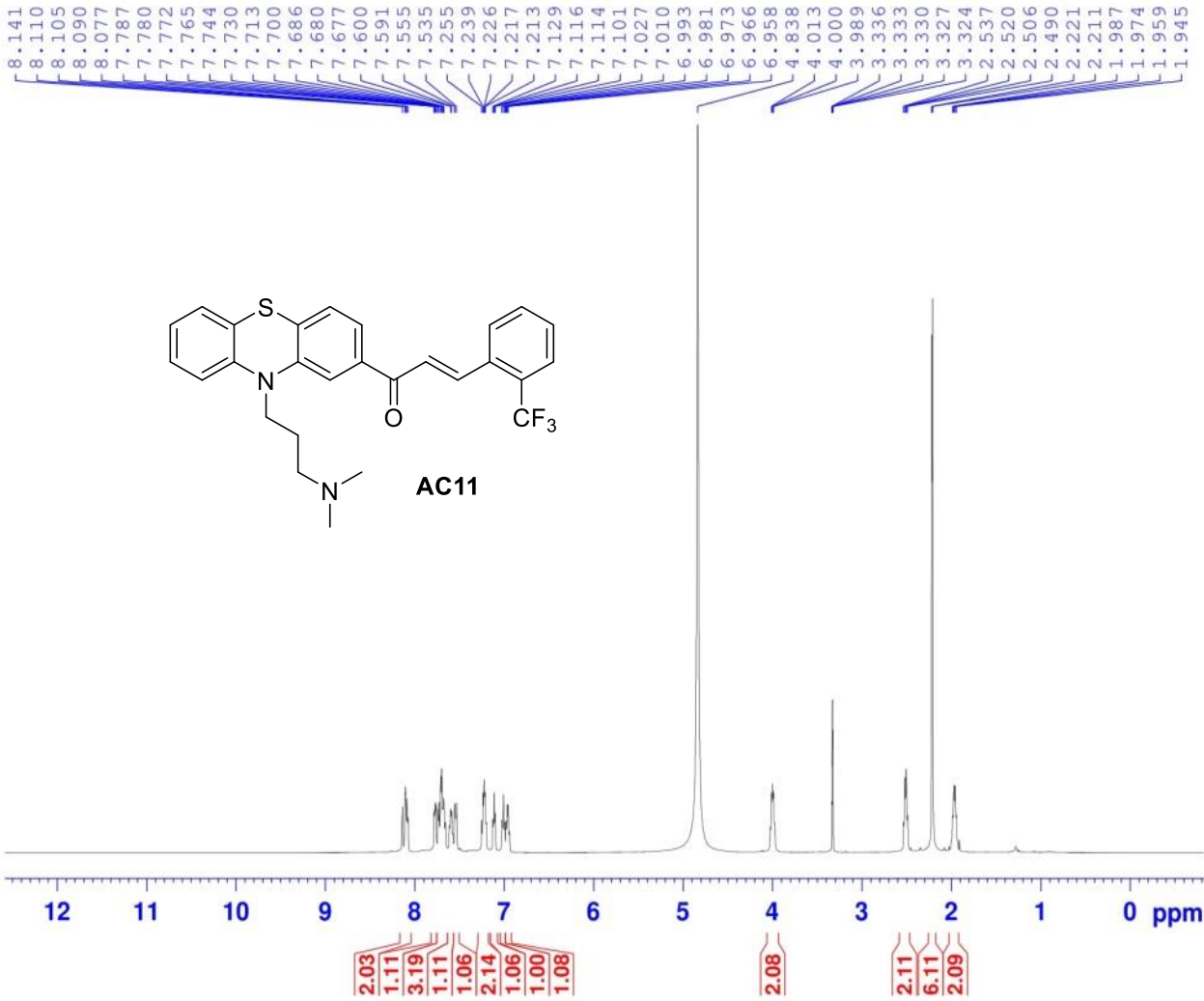
# MS

Sample Name	11-AC11	Position	P2-B1	Instrument Name	Instrument 1	User Name	
Inj Vol	2	InjPosition		SampleType	Sample	IRM Calibration Status	Success
Data Filename	11-AC11.d	ACQ Method	Cot ngan - MSMS_Pos.	Comment		Acquired Time	22/08/2020 11:40:04 AM



# <sup>1</sup>H-NMR

C15-MeOD-1H



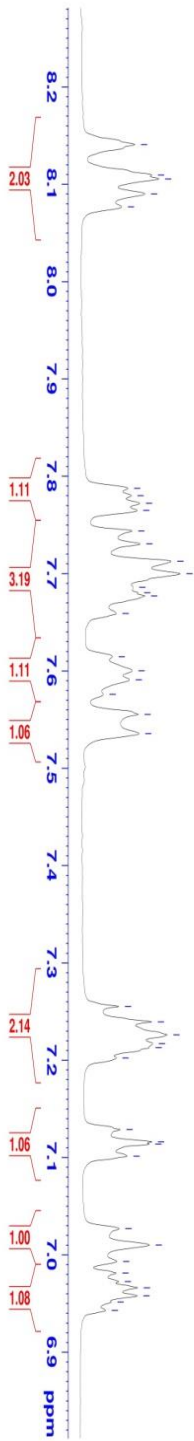
Current Data Parameters  
NAME 113DAO\_C15  
EXPNO 1  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20170705  
Time 15.07  
INSTRUM spect  
PROBHD 5 mm PABBO BB/  
PULPROG zg30  
TD 65536  
SOLVENT MeOD  
NS 16  
DS 2  
SWH 10000.000 Hz  
FIDRES 0.152588 Hz  
AQ 3.2767999 sec  
RG 79.36  
DW 50.000 usec  
DE 6.50 usec  
TE 303.0 K  
D1 1.00000000 sec  
TD0 1

==== CHANNEL f1 =====  
SFO1 500.2030889 MHz  
NUC1 1H  
P1 10.00 usec  
PLW1 22.00000000 W

F2 - Processing parameters  
SI 65536  
SF 500.2000011 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

# <sup>1</sup>H-NMR



8.141  
8.110  
8.105  
8.090  
8.077

7.787  
7.780  
7.772  
7.765  
7.744  
7.730  
7.713  
7.700  
7.686  
7.680  
7.677  
7.659  
7.615  
7.600  
7.591  
7.576  
7.555  
7.535

C15-MeOD-1H

7.255  
7.239  
7.226  
7.217  
7.213  
7.202

7.129  
7.116  
7.114  
7.101

7.027  
7.010  
6.993  
6.981  
6.973  
6.966  
6.958  
6.951  
6.943

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4.026  
4.013  
4.000  
3.989  
3.977

3.336  
3.333  
3.330  
3.327  
3.324

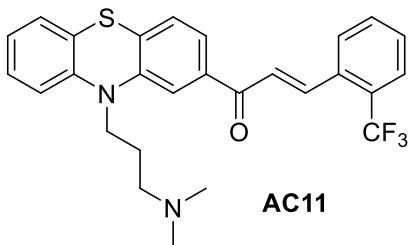
2.537  
2.520  
2.506  
2.490

2.221  
2.211

1.987  
1.974  
1.959  
1.945

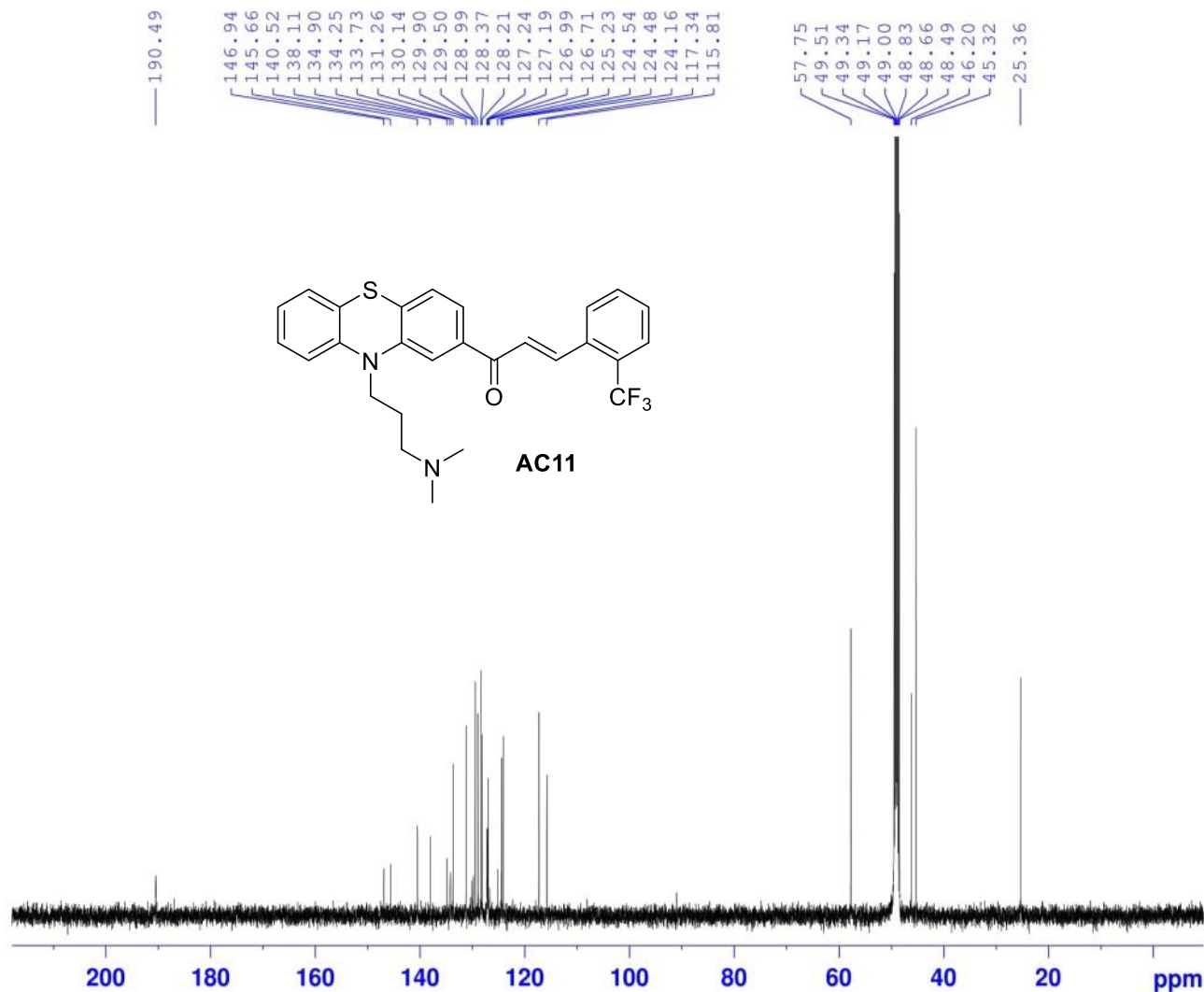
C15-MeOD-1H

BRUKER



# <sup>13</sup>C-NMR

C15-MeOD-C13CPD



Current Data Parameters  
NAME 113DAO\_C15  
EXPNO 2  
PROCNO 1

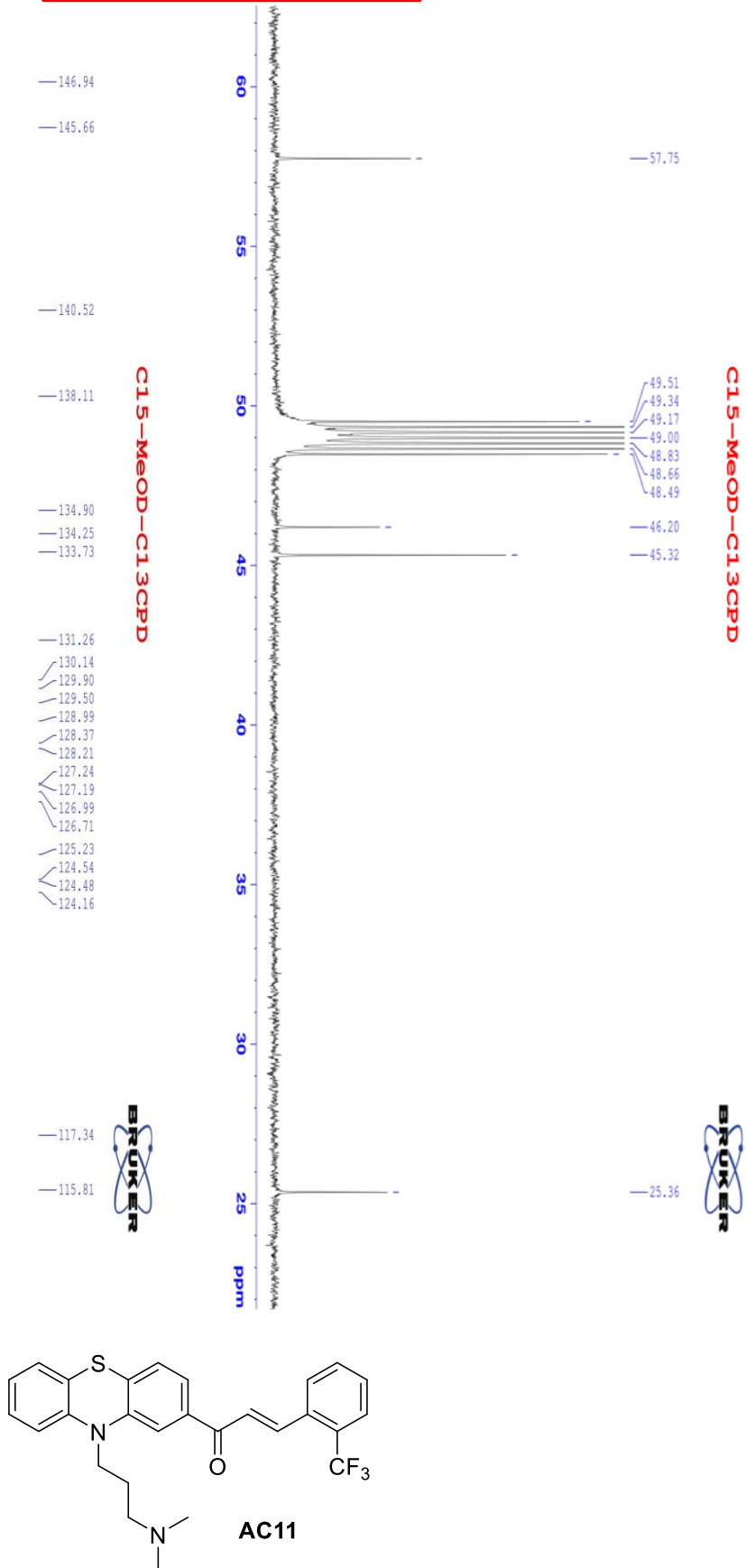
F2 - Acquisition Parameters  
Date\_ 20170705  
Time 15.23  
INSTRUM spect  
PROBHD 5 mm PABBO BB/  
PULPROG zgpg30  
TD 65536  
SOLVENT MeOD  
NS 256  
DS 4  
SWH 29761.904 Hz  
FIDRES 0.454131 Hz  
AQ 1.1010048 sec  
RG 198.57  
DW 16.800 usec  
DE 6.50 usec  
TE 303.0 K  
D1 2.00000000 sec  
D11 0.03000000 sec  
TD0 1

==== CHANNEL f1 =====  
SFO1 125.7879670 MHz  
NUC1 13C  
P1 10.00 usec  
PLW1 88.00000000 W

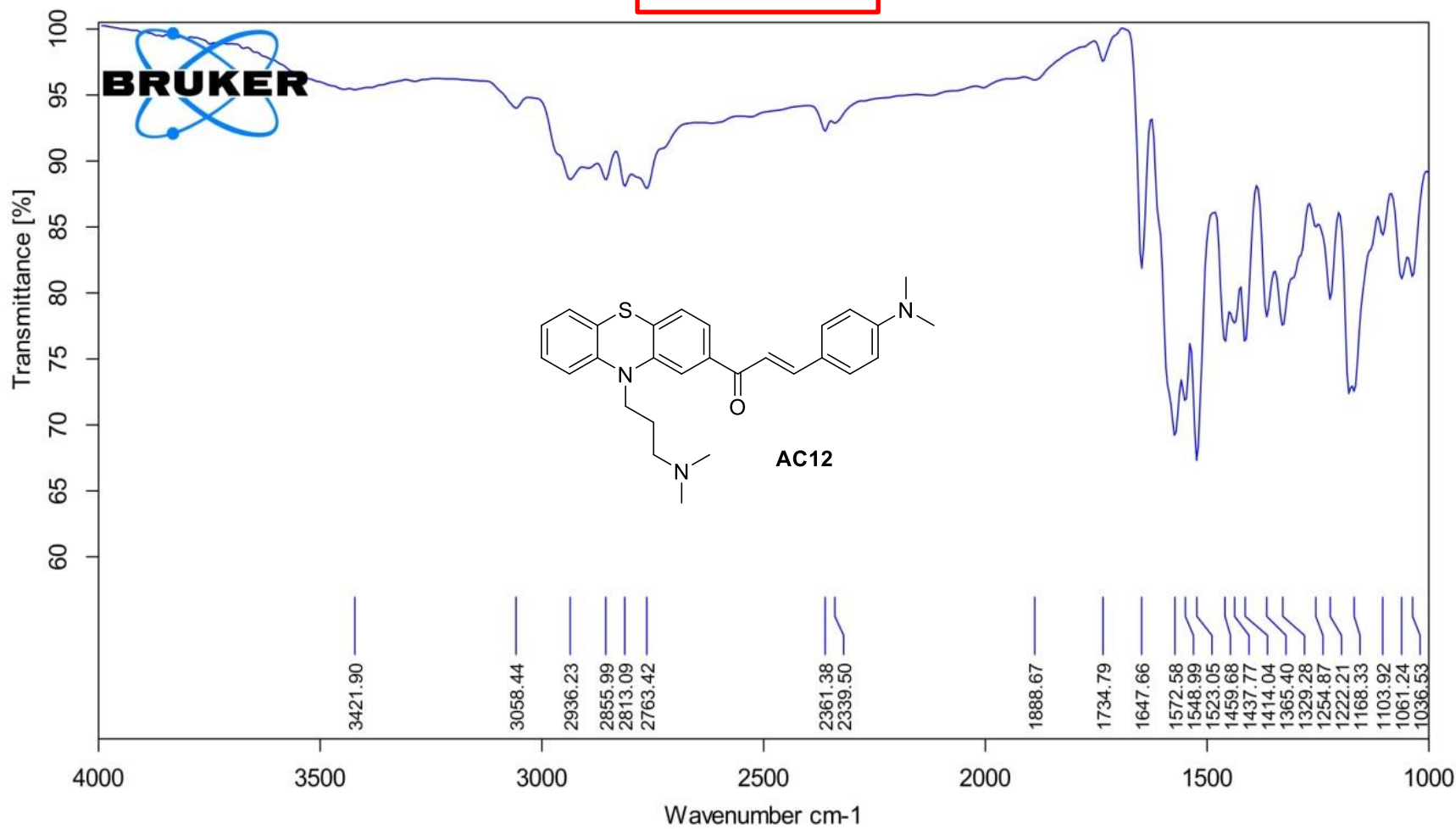
==== CHANNEL f2 =====  
SFO2 500.2020008 MHz  
NUC2 1H  
CPDPRG[2] waltz16  
PCPD2 80.00 usec  
PLW2 22.00000000 W  
PLW12 0.34375000 W  
PLW13 0.22000000 W

F2 - Processing parameters  
SI 32768  
SF 125.7752168 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40

# $^{13}\text{C}$ -NMR



IR



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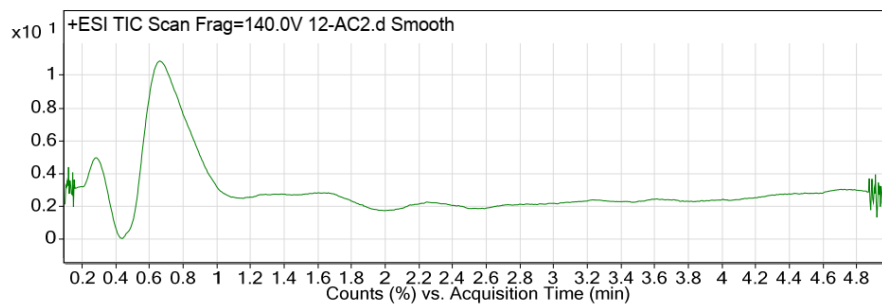
# MS

## Qualitative Analysis Report

<b>Data Filename</b>	12-AC12.d	<b>Sample Name</b>	12-AC12
<b>Sample Type</b>	Sample	<b>Position</b>	P2-B9
<b>Instrument Name</b>	Instrument 1	<b>User Name</b>	
<b>Acq Method</b>	Cot ngan - MSMS_Pos.m	<b>Acquired Time</b>	22/08/2020 11:50:36 AM
<b>IRM Calibration Status</b>	Success	<b>DA Method</b>	COTNGAN.M.m
<b>Comment</b>			
<b>Sample Group</b>		<b>Info.</b>	
<b>Stream Name</b>	LC 1	<b>Acquisition SW</b>	6200 series TOF/6500 series
		<b>Version</b>	Q-TOF B.06.01 (B6172 SP1)

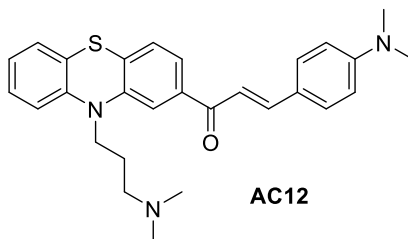
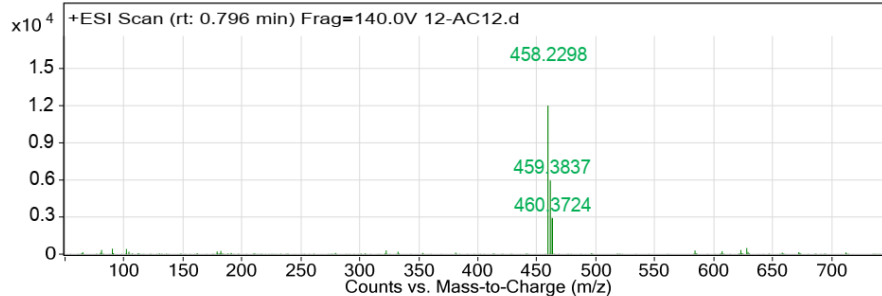
### User Chromatograms

Fragmentor Voltage 140 Collision Energy 0 Ionization Mode ESI



### User Spectra

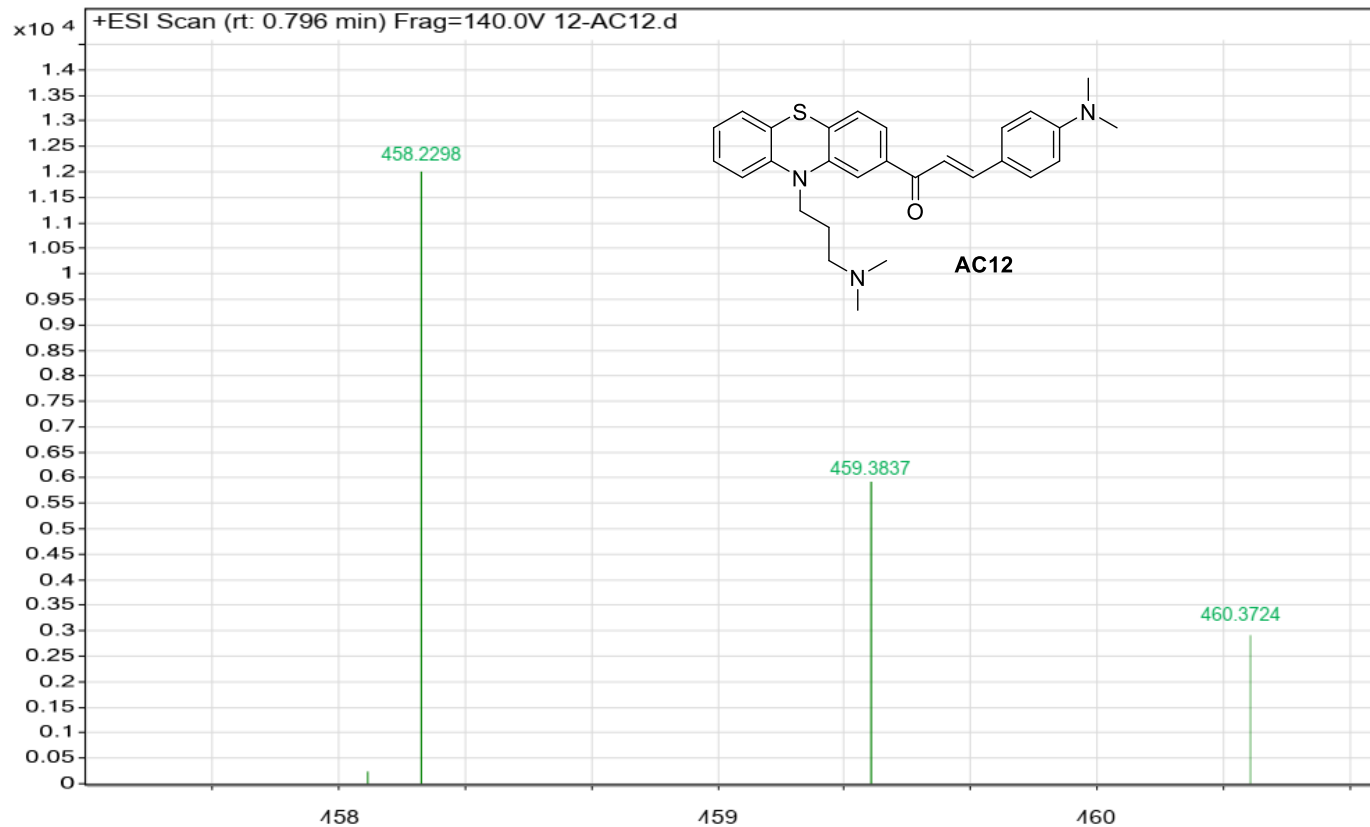
Fragmentor Voltage 140 Collision Energy 0 Ionization Mode ESI



--- End Of Report ---

# MS

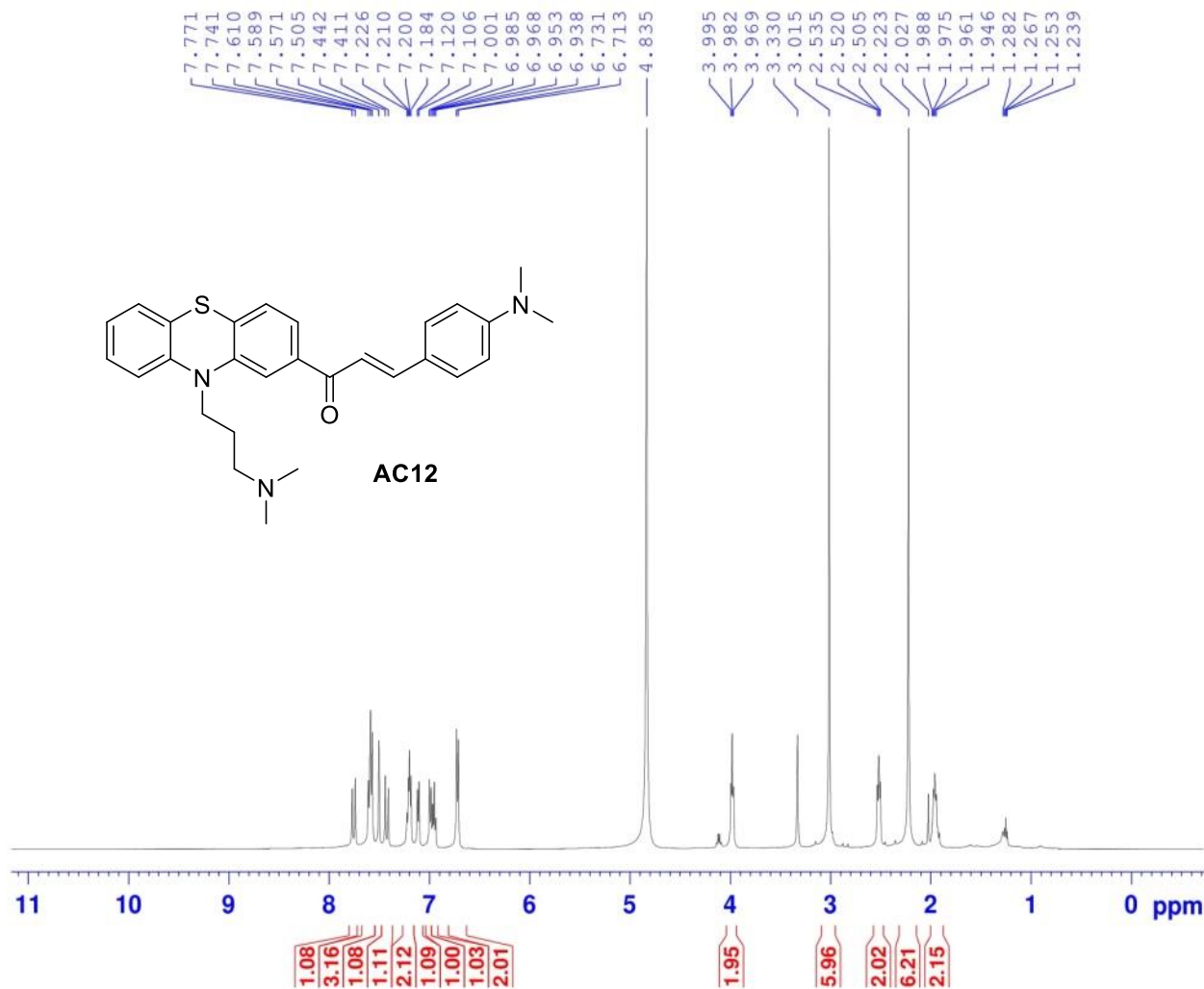
Sample Name	12-AC12	Position	P2-B9	Instrument Name	Instrument 1	User Name	
Inj Vol	2	InjPosition		SampleType	Sample	IRM Calibration Status	Success
Data Filename	12-AC12.d	ACQ Method	Cot ngan - MSMS_Pos.	Comment		Acquired Time	22/08/2020 11:50:36 AM





# <sup>1</sup>H-NMR

C7-MeOD-1H



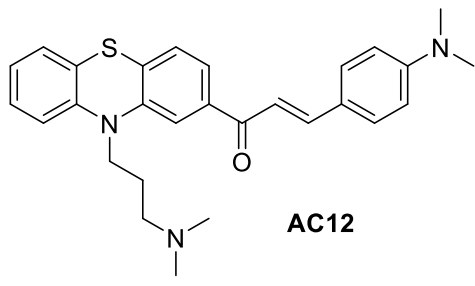
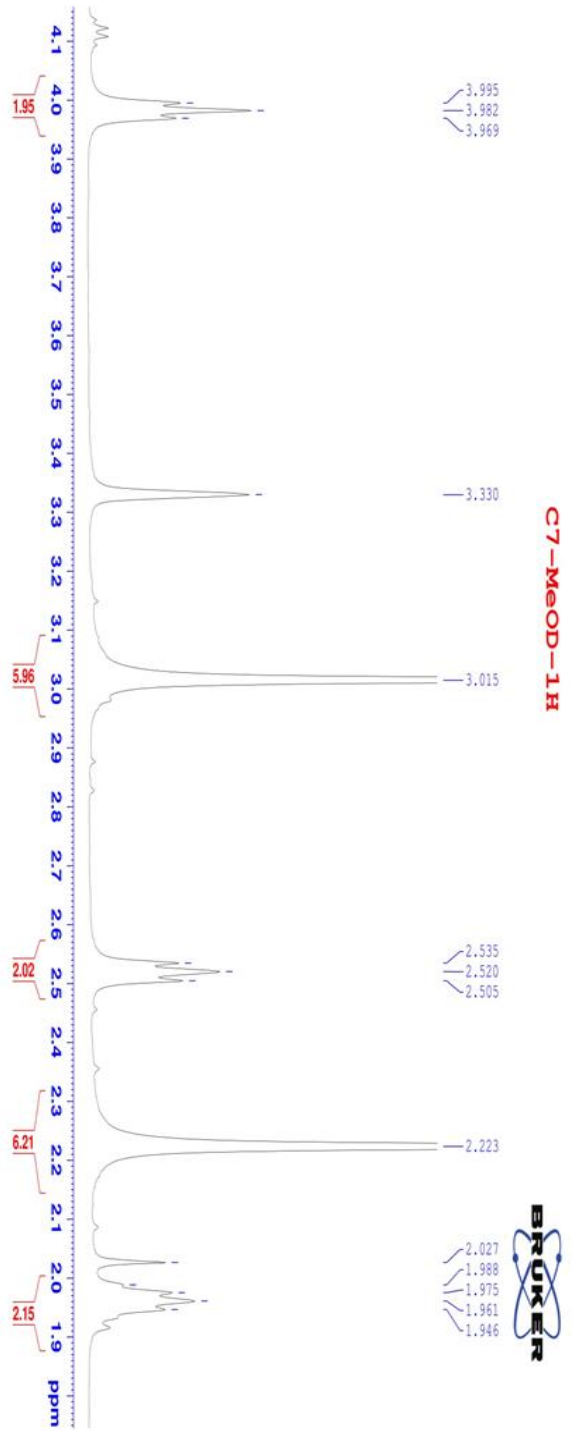
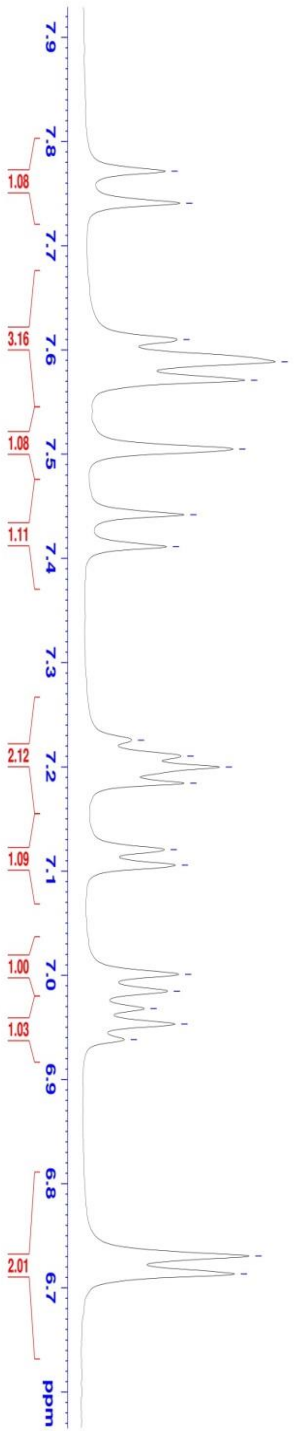
Current Data Parameters  
NAME 113DAO\_C7  
EXPNO 1  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20170704  
Time 16.50  
INSTRUM spect  
PROBHD 5 mm PABBO BB/  
PULPROG zg30  
TD 65536  
SOLVENT MeOD  
NS 16  
DS 2  
SWH 10000.000 Hz  
FIDRES 0.152588 Hz  
AQ 3.2767999 sec  
RG 79.36  
DW 50.000 usec  
DE 6.50 usec  
TE 303.0 K  
D1 1.00000000 sec  
TD0 1

----- CHANNEL f1 -----  
SFO1 500.2030889 MHz  
NUC1 1H  
P1 10.00 usec  
PLW1 22.00000000 W

F2 - Processing parameters  
SI 65536  
SF 500.2000020 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

# <sup>1</sup>H-NMR



# <sup>13</sup>C-NMR

C7-MeOD-C13CPD



Current Data Parameters  
NAME 113DAO\_C7  
EXPNO 2  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20170704  
Time 17.00  
INSTRUM spect  
PROBHD 5 mm PABBO BB/  
PULPROG zgpg30  
TD 65536  
SOLVENT MeOD  
NS 128  
DS 4  
SWH 29761.904 Hz  
FIDRES 0.454131 Hz  
AQ 1.1010048 sec  
RG 198.57  
DW 16.800 usec  
DE 6.50 usec  
TE 303.0 K  
D1 2.00000000 sec  
D11 0.03000000 sec  
TDO 1

===== CHANNEL f1 =====  
SFO1 125.7879670 MHz  
NUC1 13C  
P1 10.00 usec  
PLW1 88.00000000 W

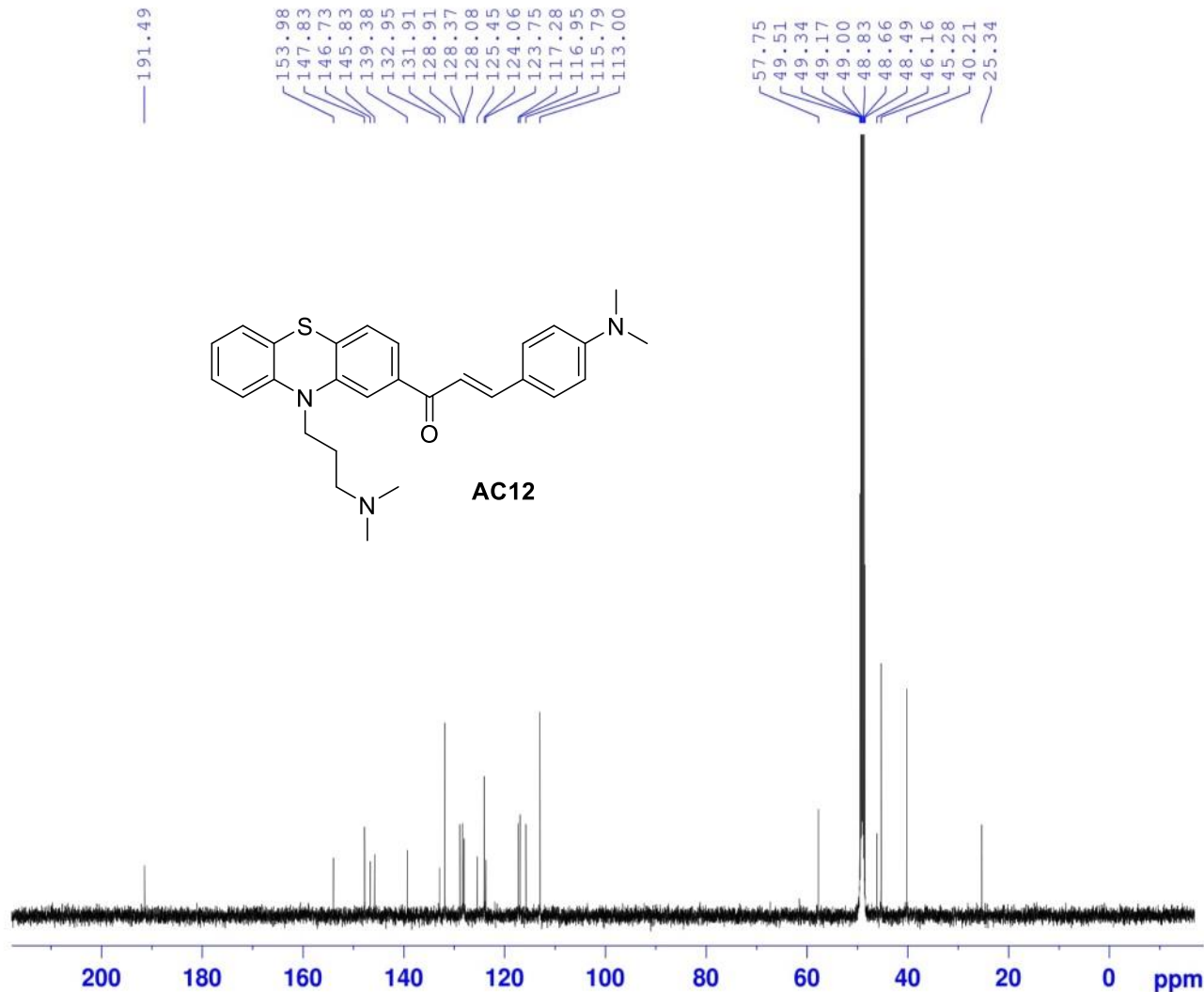
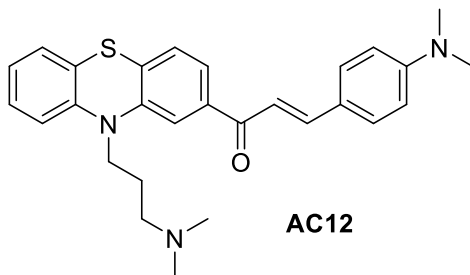
===== CHANNEL f2 =====  
SFO2 500.2020008 MHz  
NUC2 1H  
CPDPRG[2] waltz16  
PCPD2 80.00 usec  
PLW2 22.00000000 W  
PLW12 0.34375000 W  
PLW13 0.22000000 W

F2 - Processing parameters  
SI 32768  
SF 125.7752164 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40

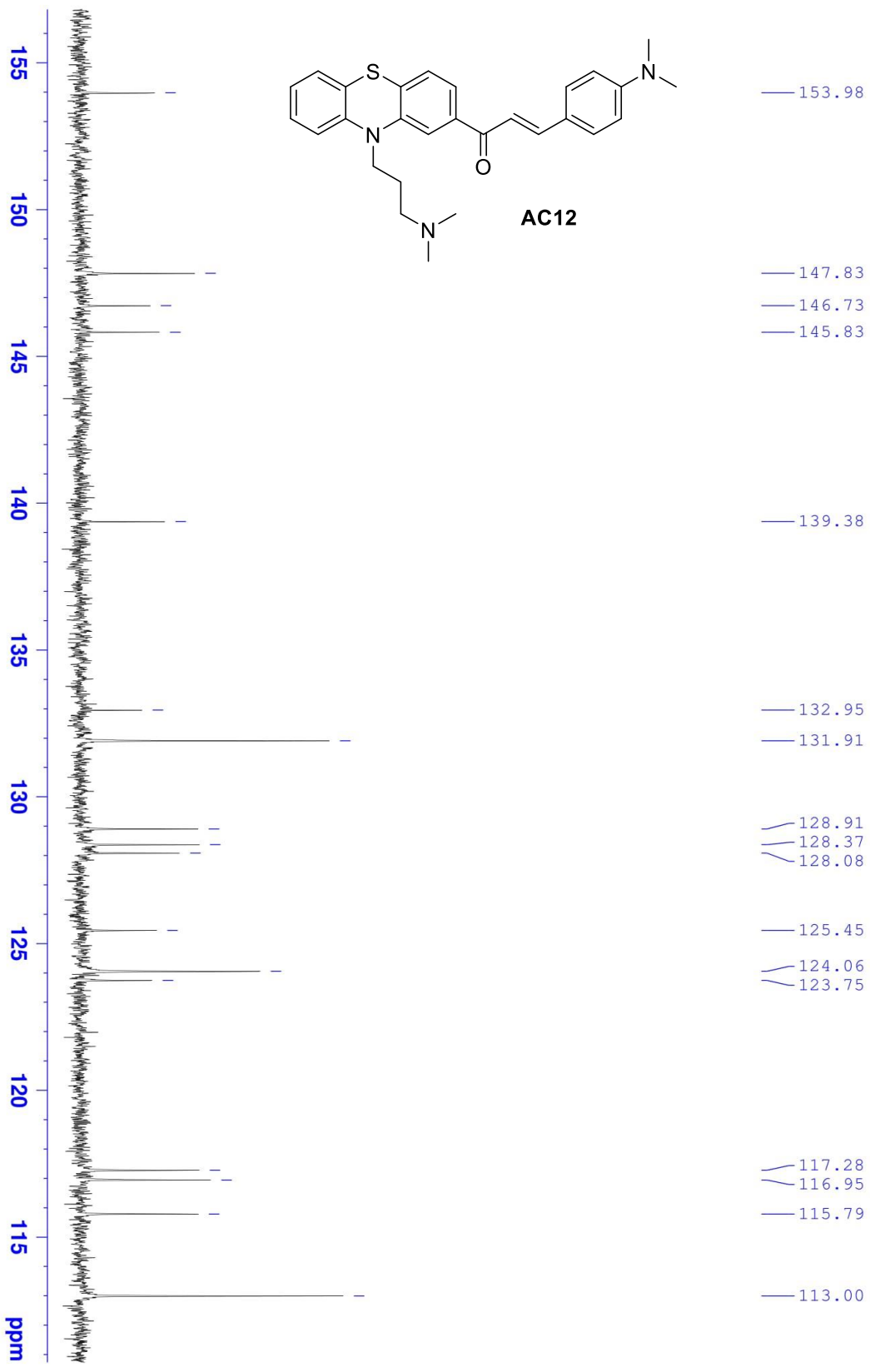
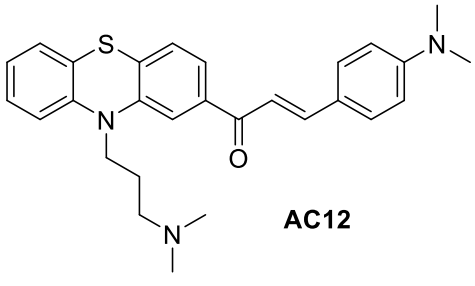
191.49

153.98  
147.83  
146.73  
145.83  
139.38  
132.95  
131.91  
128.91  
128.37  
128.08  
125.45  
124.06  
123.75  
117.28  
116.95  
115.79  
113.00

57.75  
49.51  
49.34  
49.17  
49.00  
48.83  
48.66  
48.49  
46.16  
45.28  
40.21  
25.34



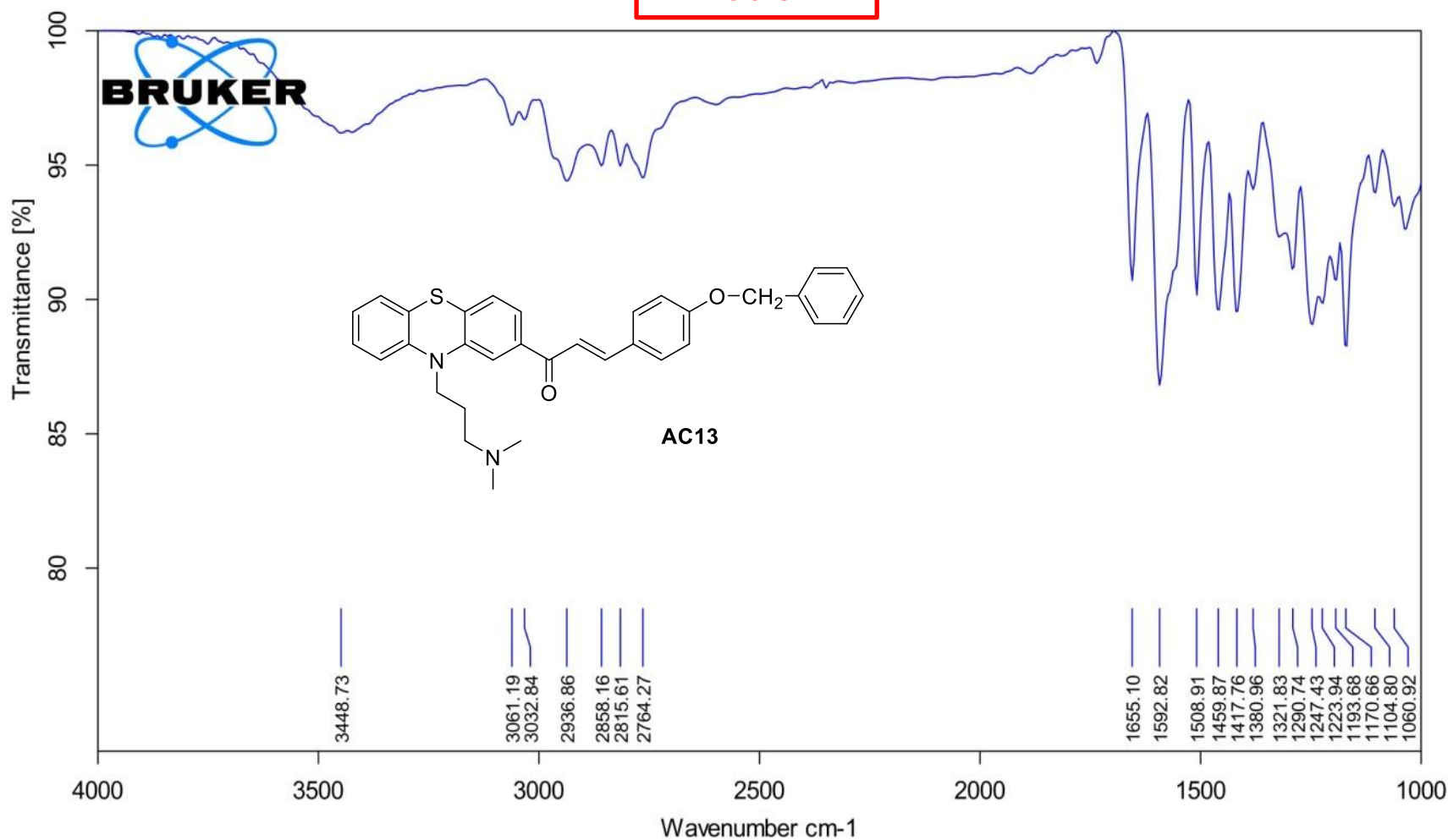
# $^{13}\text{C}$ -NMR



C7-MeOD-C13CPD



IR



# MS

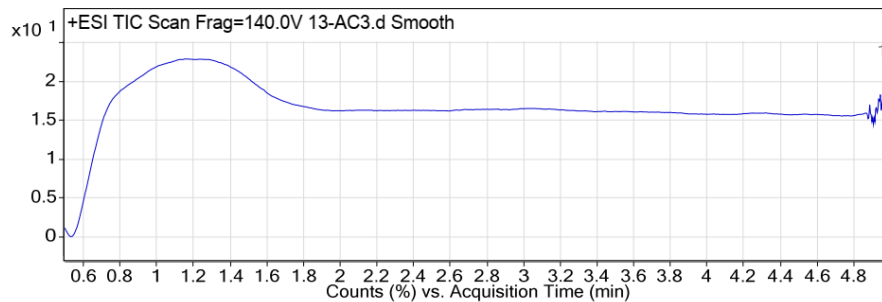
## Qualitative Analysis Report

<b>Data Filename</b>	13-AC13.d	<b>Sample Name</b>	13-AC13
<b>Sample Type</b>	Sample	<b>Position</b>	P2-C2
<b>Instrument Name</b>	Instrument 1	<b>User Name</b>	
<b>Acq Method</b>	Cot ngan - MSMS_Pos.m	<b>Acquired Time</b>	22/08/2020 12:00:43 PM
<b>IRM Calibration Status</b>	Success	<b>DA Method</b>	COTNGAN.M.m
<b>Comment</b>			
<b>Sample Group</b>		<b>Info.</b>	
<b>Stream Name</b>	LC 1	<b>Acquisition SW</b>	6200 series TOF/6500 series
		<b>Version</b>	Q-TOF B.06.01 (B6172 SP1)

+ESI TIC Scan Frag=140.0V BTBZ 9.d Smooth

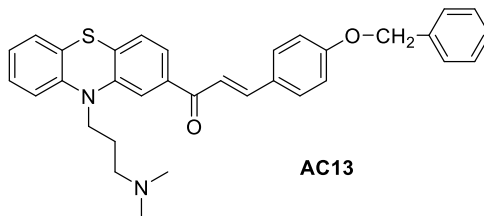
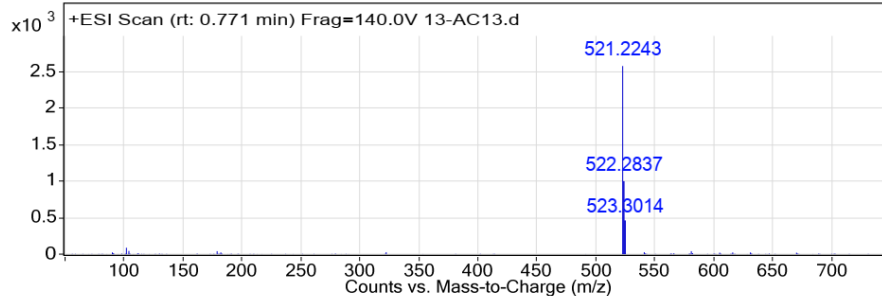
### User Chromatograms

Fragmentor Voltage 140 Collision Energy 0 Ionization Mode ESI



### User Spectra

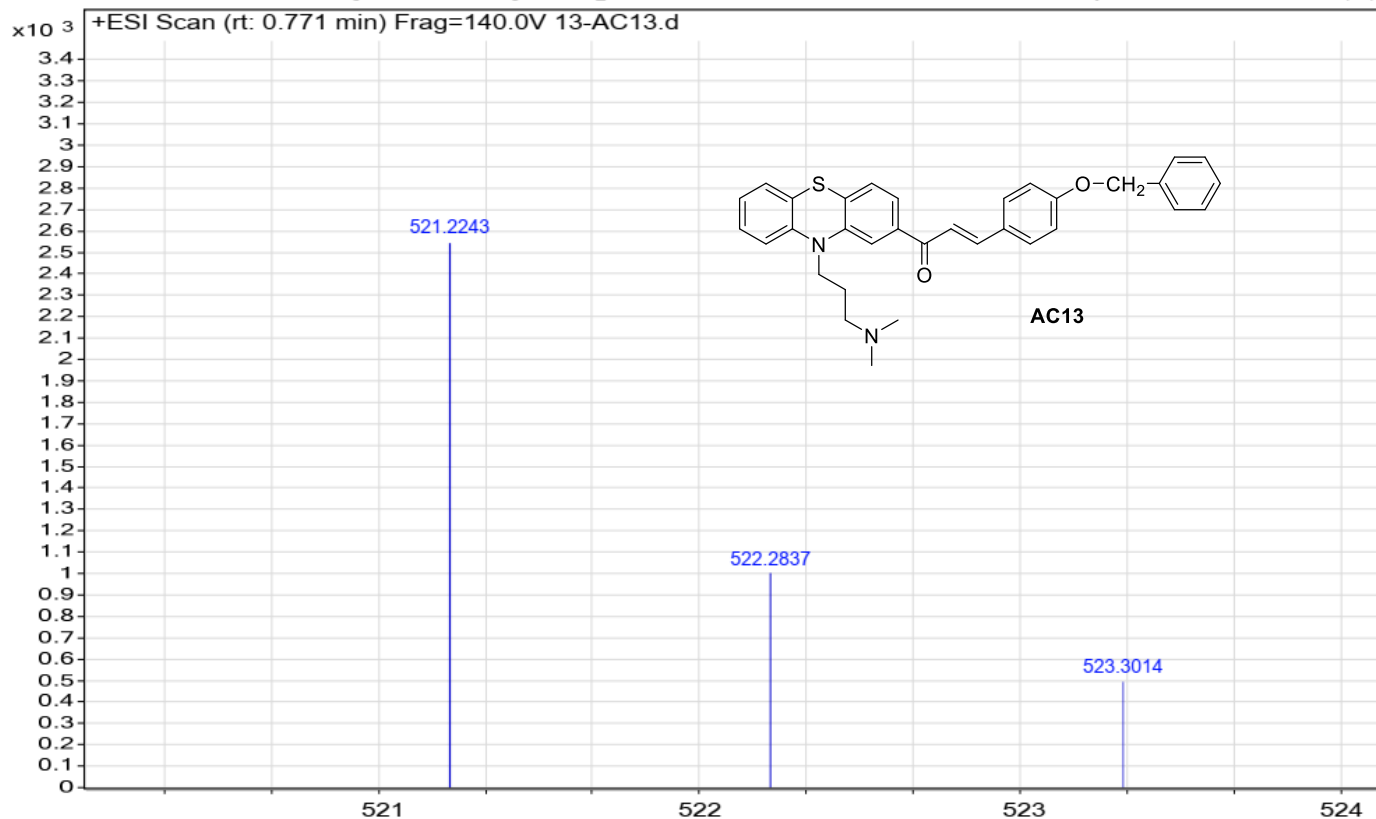
Fragmentor Voltage 140 Collision Energy 0 Ionization Mode ESI



--- End Of Report ---

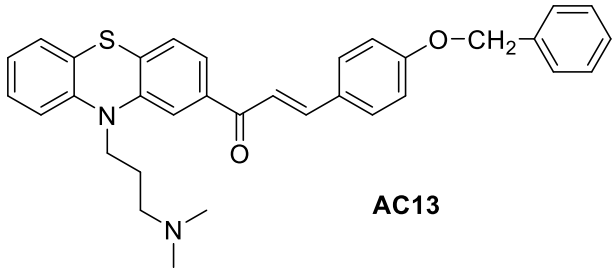
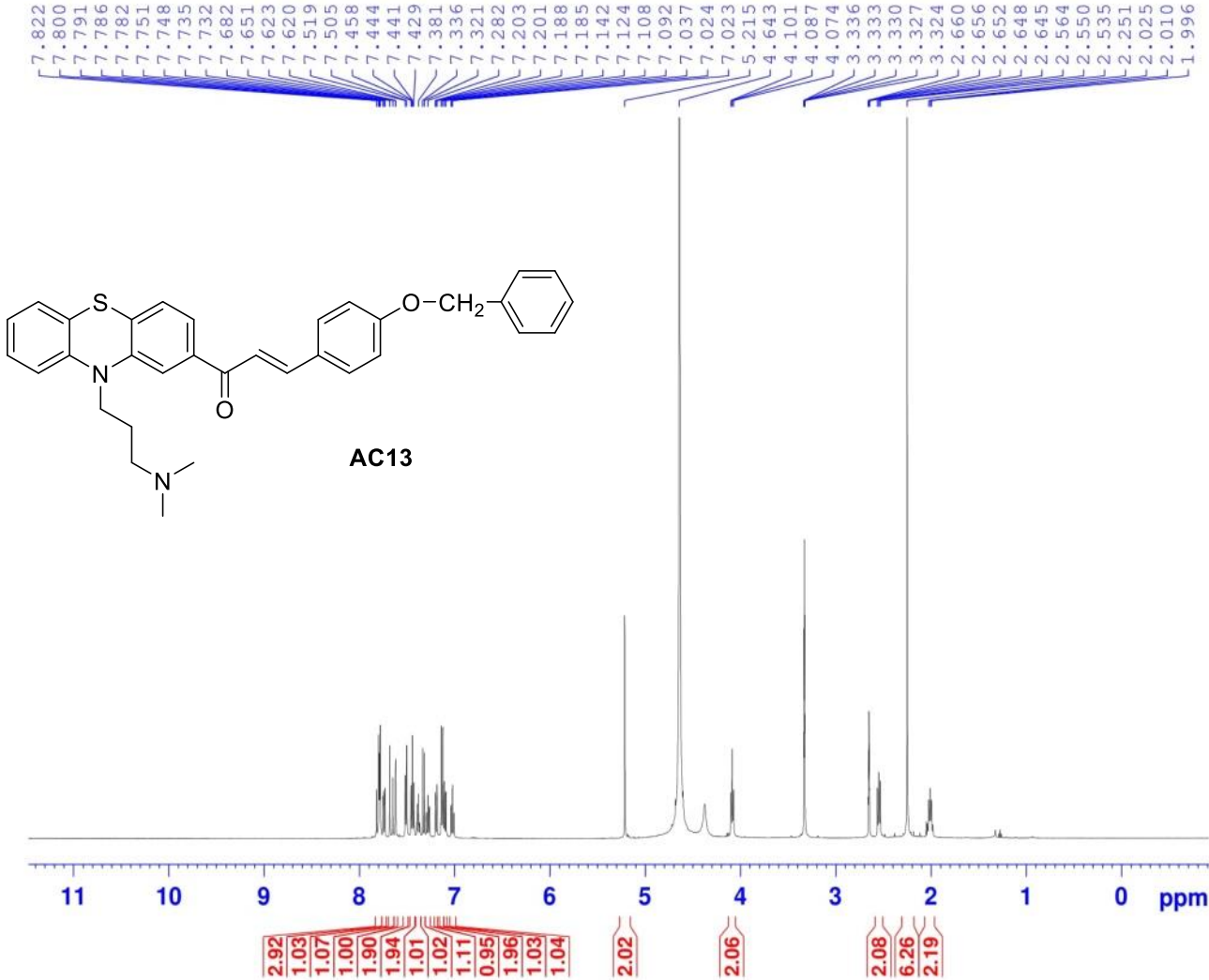
# MS

Sample Name	13-AC13	Position	P2-C2	Instrument Name	Instrument 1	User Name	
Inj Vol	2	InjPosition		SampleType	Sample	IRM Calibration Status	Success
Data Filename	13-AC13.d	ACQ Method	Cot ngan - MSMS_Pos.	Comment		Acquired Time	22/08/2020 12:00:43 PM



# <sup>1</sup>H-NMR

C5-MeOD+DMSO-1H



Current Data Parameters  
NAME 113D\_C5  
EXPNO 1  
PROCNO 1

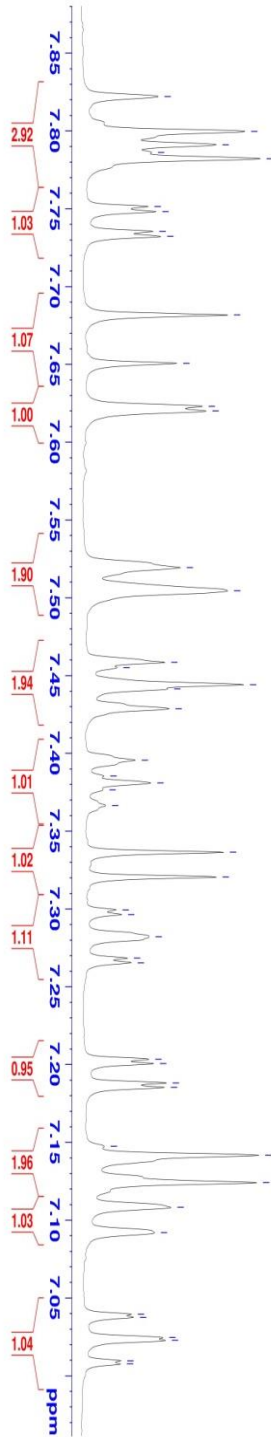
F2 - Acquisition Parameters  
Date\_ 20170607  
Time 10.47  
INSTRUM spect  
PROBHD 5 mm PABBO BB/  
PULPROG zg30  
TD 65536  
SOLVENT MeOD  
NS 16  
DS 2  
SWH 10000.000 Hz  
FIDRES 0.152588 Hz  
AQ 3.2767999 sec  
RG 111.09  
DW 50.000 usec  
DE 6.50 usec  
TE 303.0 K  
D1 1.00000000 sec  
TD0 1

==== CHANNEL f1 =====  
SFO1 500.2030889 MHz  
NUC1 1H  
P1 10.00 usec  
PLW1 22.00000000 W

F2 - Processing parameters  
SI 65536  
SF 500.2000003 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00



# <sup>1</sup>H-NMR



7.822  
7.800  
7.791  
7.786  
7.782  
7.751  
7.748  
7.735  
7.732

7.682  
7.651  
7.623  
7.620

7.519  
7.505

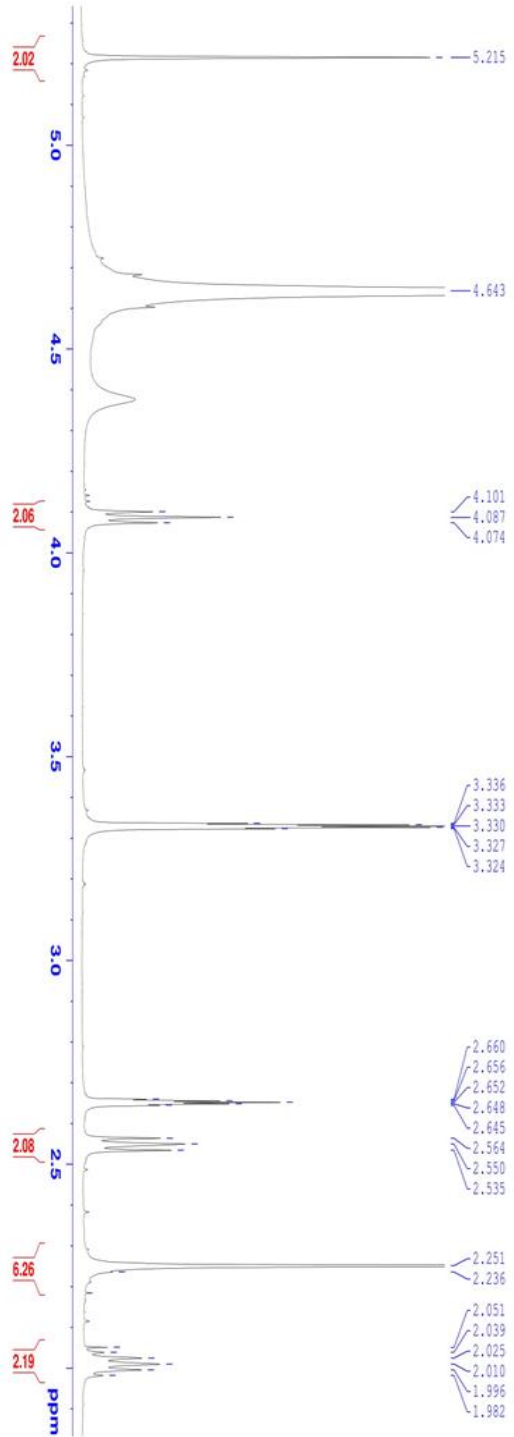
7.458  
7.455  
7.444  
7.441  
7.429  
7.396  
7.385  
7.381  
7.377  
7.366  
7.336  
7.321  
7.299  
7.296  
7.282  
7.268  
7.265

7.203  
7.201  
7.188  
7.185

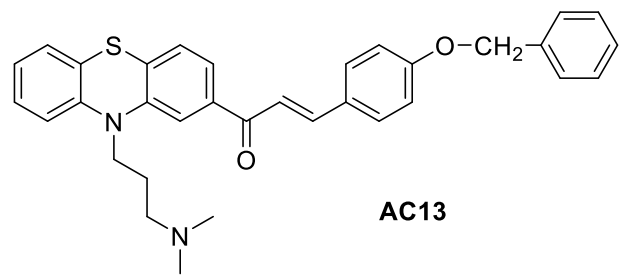
7.147  
7.142  
7.124  
7.108  
7.092

7.040  
7.037  
7.024  
7.023  
7.010  
7.008

C5-MeOD+DMSO-1H

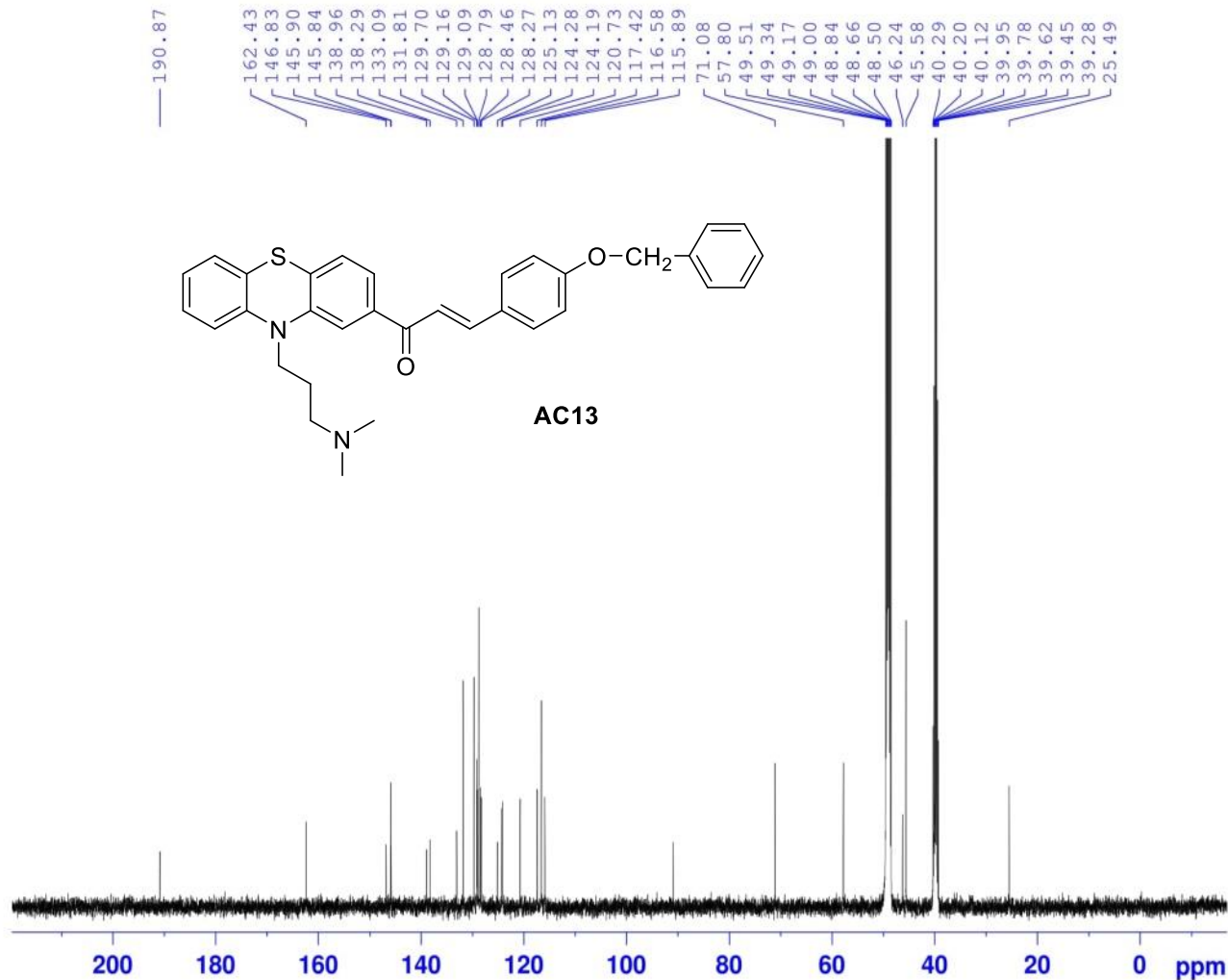


C5-MeOD+DMSO-1H



# <sup>13</sup>C-NMR

C5-MeOD+DMSO-C13CPD



Current Data Parameters  
NAME 113D\_C5  
EXPNO 2  
PROCNO 1

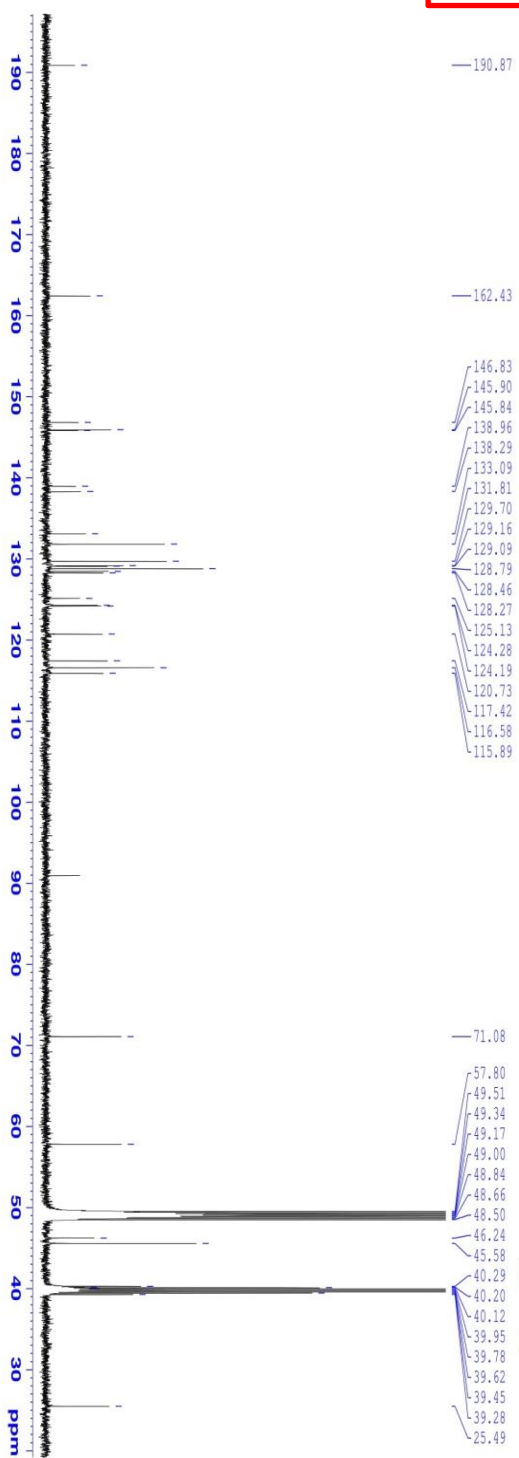
F2 - Acquisition Parameters  
Date\_ 20170607  
Time 13.48  
INSTRUM spect  
PROBHD 5 mm PABBO BB/  
PULPROG zgpg30  
TD 65536  
SOLVENT MeOD  
NS 2048  
DS 4  
SWH 29761.904 Hz  
FIDRES 0.454131 Hz  
AQ 1.1010048 sec  
RG 198.57  
DW 16.800 usec  
DE 6.50 usec  
TE 303.0 K  
D1 2.00000000 sec  
D11 0.03000000 sec  
TD0 1

===== CHANNEL f1 =====  
SFO1 125.7879670 MHz  
NUC1 13C  
P1 10.00 usec  
PLW1 88.00000000 W

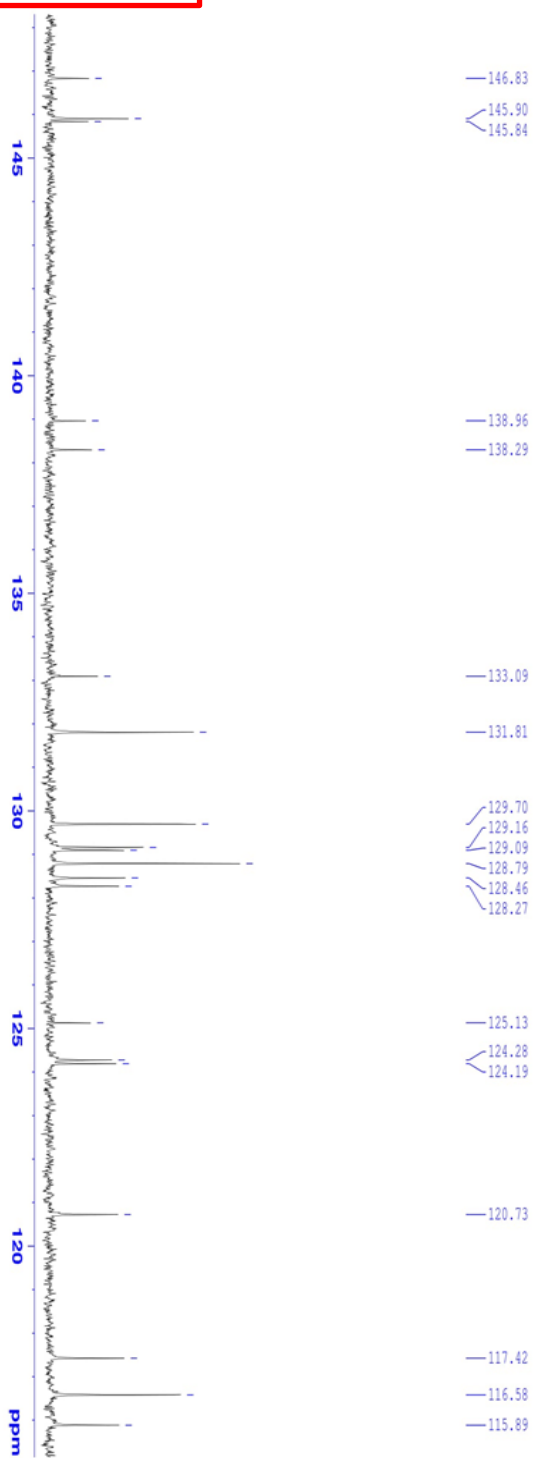
===== CHANNEL f2 =====  
SFO2 500.2020008 MHz  
NUC2 1H  
CPDPRG[2] waltz16  
PCPD2 80.00 usec  
PLW2 22.00000000 W  
PLW12 0.34375000 W  
PLW13 0.22000000 W

F2 - Processing parameters  
SI 32768  
SF 125.7753900 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40

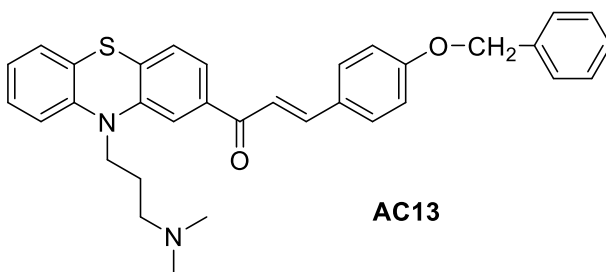
# $^{13}\text{C}$ -NMR



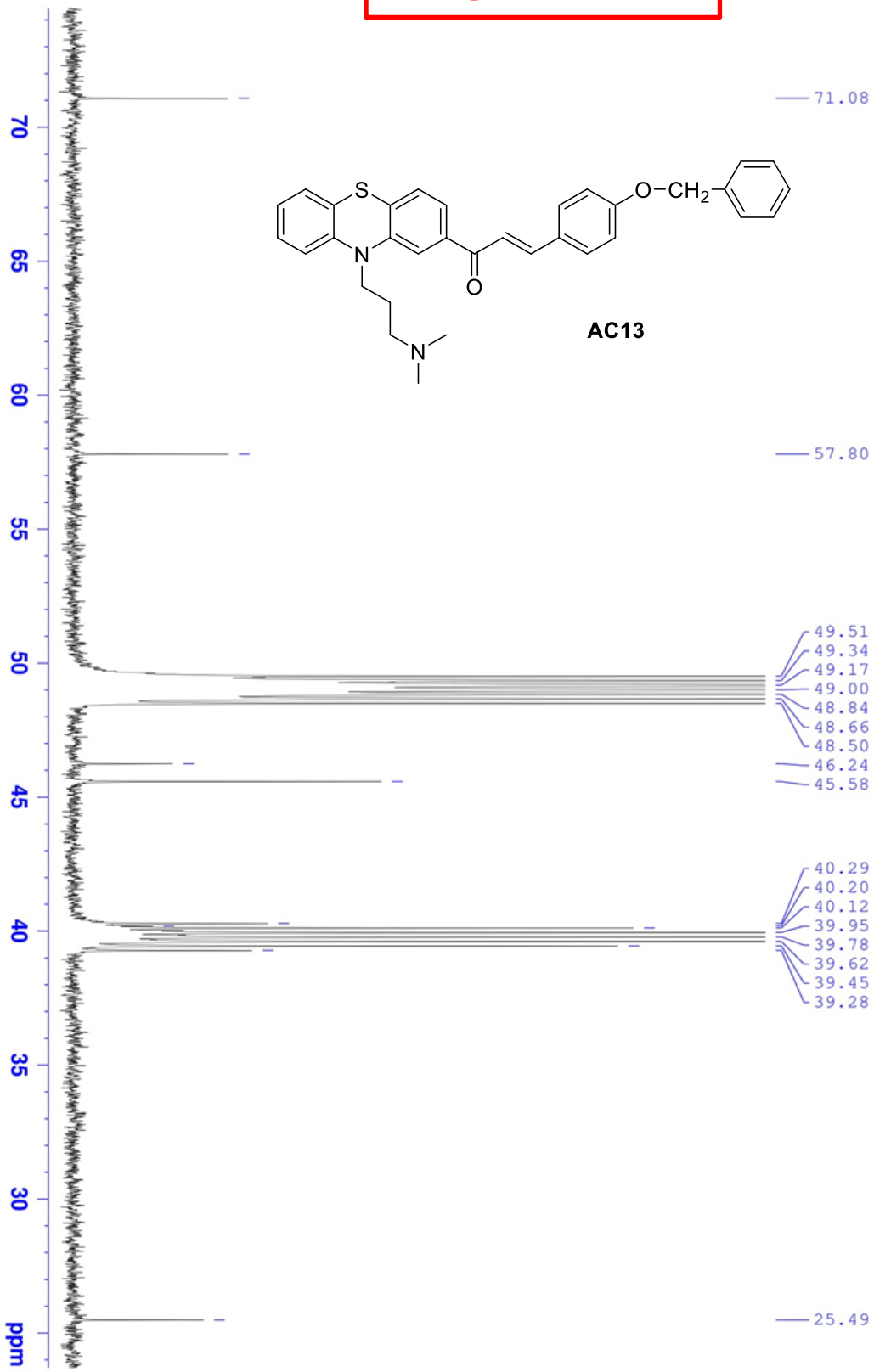
C5-MeOD+DMSO-C13CPD



C5-MeOD+DMSO-C13CPD



# $^{13}\text{C}$ -NMR



CDCl<sub>3</sub>-MeOD+DMSO-C13CPD



## Reference

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4. Oprea, T.I. Property distribution of drug-related chemical databases. *Journal of computer-aided molecular design* **2000**, *14*, 251-264, doi:10.1023/a:1008130001697.