



**Table S1.** Prevalence of apramycin- vs. RMTase resistance gene annotations in human clinical isolates deposited in the NCBI National Database of Antibiotic Resistant Organisms (NDARO) as of June 24, 2020.

	Clinical Isolate Resistance Gene Annotations										
	Total	<i>aac(3)-IV</i>		<i>apmA</i>		<i>npmA</i>		<i>kamB</i>		<i>RMTase</i>	
	<i>n</i>	<i>n</i>	%	<i>n</i>	%	<i>n</i>	%	<i>n</i>	%	<i>n</i>	%
<b>All genomes</b>	182,405	1,265	0.69%	2	0.00%	2	0.00%	0	0.00%	4,941	2.71%
<b>Gram-negatives</b>											
<i>A. baumannii</i>	5,728	1	0.02%	0	0.00%	0	0.00%	0	0.00%	2,150	37.53%
<i>P. aeruginosa</i>	4,940	3	0.06%	0	0.00%	0	0.00%	0	0.00%	47	0.95%
<i>E. coli / Shigella</i>	37,283	298	0.80%	0	0.00%	0	0.00%	0	0.00%	300	0.80%
<i>K. pneumoniae</i>	14,999	485	3.23%	0	0.00%	0	0.00%	0	0.00%	2,250	15.00%
<i>K. oxytoca</i>	478	0	0.00%	0	0.00%	0	0.00%	0	0.00%	14	2.93%
<i>Enterobacter</i> spp.	2,352	6	0.26%	0	0.00%	0	0.00%	0	0.00%	117	4.97%
<i>S. marcescens</i>	786	1	0.13%	0	0.00%	0	0.00%	0	0.00%	7	0.89%
<i>C. freundii</i>	472	1	0.21%	0	0.00%	0	0.00%	0	0.00%	26	5.51%
<i>M. morgani</i>	47	2	4.26%	0	0.00%	0	0.00%	0	0.00%	3	6.38%
<i>S. enterica</i>	68,418	451	0.66%	0	0.00%	0	0.00%	0	0.00%	11	0.02%
<i>C. jejuni</i>	9,681	0	0.00%	2	0.02%	0	0.00%	0	0.00%	0	0.00%
<b>Gram-positives</b>											
<i>C. difficile</i>	2,319	0	0.00%	0	0.00%	2	0.09%	0	0.00%	0	0.00%
<i>E. faecium</i>	7,215	0	0.00%	0	0.00%	0	0.00%	0	0.00%	0	0.00%

**Table S2.** Prevalence of apramycin- vs. RMTase resistance gene annotations in the carbapenemase-positive subpopulation of human clinical isolates deposited in the NCBI National Database of Antibiotic Resistant Organisms (NDARO) as of June 24, 2020.

	Carbapenemase-Positive Clinical Isolate Resistance Gene Annotations										
	Total	<i>aac(3)-IV</i>		<i>apmA</i>		<i>npmA</i>		<i>kamB</i>		<i>RMTase</i>	
	<i>n</i>	<i>n</i>	%	<i>n</i>	%	<i>n</i>	%	<i>n</i>	%	<i>n</i>	%
<b>All CP genomes</b>	21,195	495	2.34%	0	0.00%	0	0.00%	0	0.00%	4,551	21.47%
<b>Gram-negatives</b>											
<i>A. baumannii</i>	5,465	0	0.00%	0	0.00%	0	0.00%	0	0.00%	2,145	39.25%
<i>P. aeruginosa</i>	2,724	1	0.04%	0	0.00%	0	0.00%	0	0.00%	44	1.62%
<i>E. coli / Shigella</i>	1,022	47	4.60%	0	0.00%	0	0.00%	0	0.00%	210	20.55%
<i>K. pneumoniae</i>	8,535	433	5.07%	0	0.00%	0	0.00%	0	0.00%	1,992	23.34%
<i>K. oxytoca</i>	109	0	0.00%	0	0.00%	0	0.00%	0	0.00%	13	11.93%
<i>Enterobacter</i> spp.	1,036	4	0.39%	0	0.00%	0	0.00%	0	0.00%	96	9.27%
<i>S. marcescens</i>	167	0	0.00%	0	0.00%	0	0.00%	0	0.00%	7	4.19%
<i>C. freundii</i>	236	1	0.42%	0	0.00%	0	0.00%	0	0.00%	25	10.59%
<i>M. morgani</i>	13	1	7.69%	0	0.00%	0	0.00%	0	0.00%	3	23.08%
<i>S. enterica</i>	19	1	5.26%	0	0.00%	0	0.00%	0	0.00%	2	10.53%
<i>C. jejuni</i>	618	0	0.00%	0	0.00%	0	0.00%	0	0.00%	0	0.00%
<b>Gram-positives</b>											
<i>C. difficile</i>	0	0	n/a	0	n/a	0	n/a	0	n/a	0	n/a
<i>E. faecium</i>	1	0	0.00%	0	0.00%	0	0.00%	0	0.00%	0	0.00%

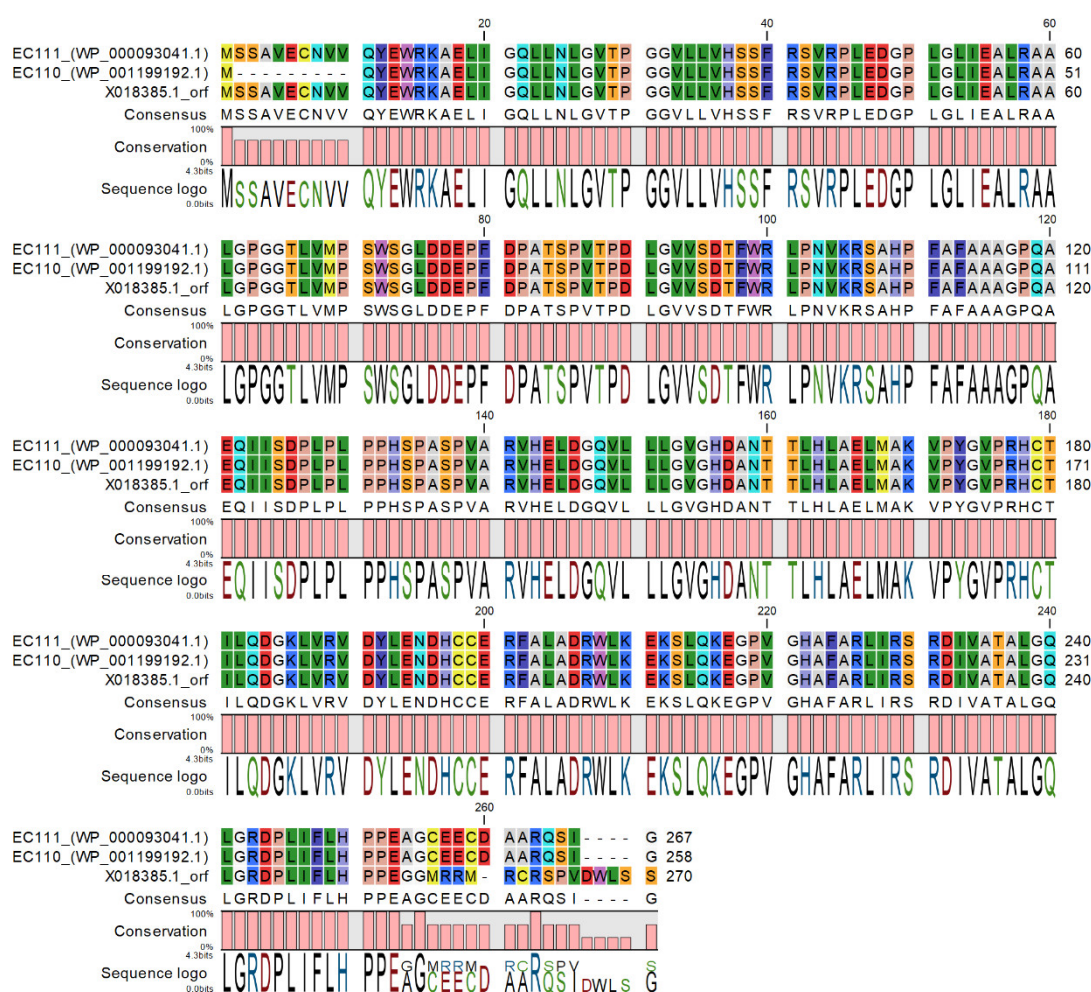
**Table S3.** Strains used in this study.

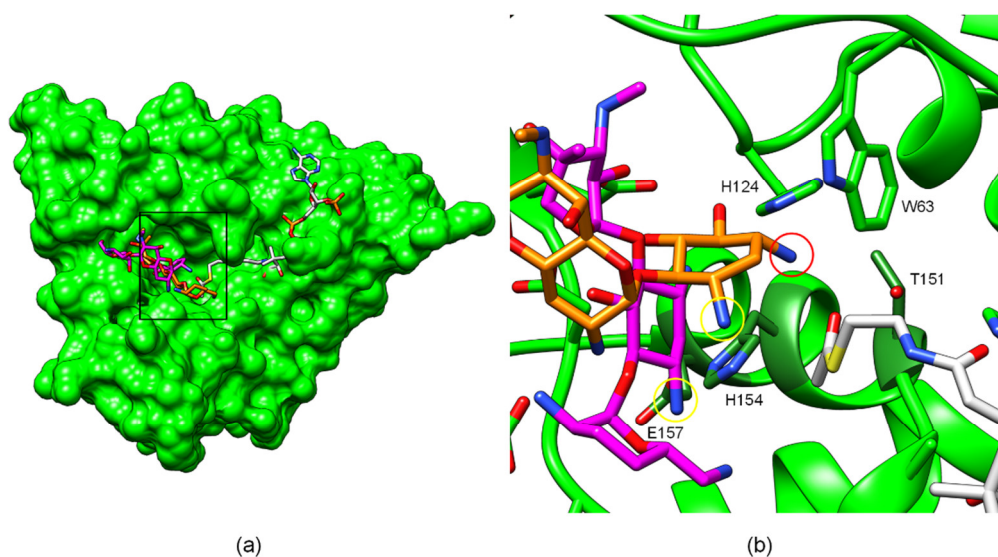
Strain	Aminoglycoside Resistance Gene	Gene Variation	Promoter Strength <sup>a</sup>
Clinical isolates			
ATCC 25922	none	-	-
AG173	<i>aac(3)-IV</i>	None (native)	-
AG380	<i>aac(3)-IV</i>	None (native)	-
AG381	<i>aac(3)-IV</i>	None (native)	-
Recombinant			
DH5 $\alpha$	none	-	-
EC118	<i>aac(3)-IV</i>	None (native)	+
EC111	<i>aac(3)-IV</i>	None (native)	++
EC119	<i>aac(3)-IV</i>	None (native)	+++
EC116	<i>aac(3)-IV</i>	Truncated <sup>b</sup>	+
EC110	<i>aac(3)-IV</i>	Truncated <sup>b</sup>	++
EC117	<i>aac(3)-IV</i>	Truncated <sup>b</sup>	+++
EC109	<i>aac(3)-IV</i>	H154A	++
EC274	<i>aac(3)-IV</i>	H124Y	+
EC275	<i>aac(3)-IV</i>	D67A	+
EC276	<i>aac(3)-IV</i>	E185A	+
EC277	<i>aac(3)-IV</i>	D187A	+
EC278	<i>aac(3)-IV</i>	E249A	+
EC279	<i>aac(3)-IV</i>	E248A E249A	+
EC280	<i>aac(3)-IV</i>	C247A C250A	+
EC281	<i>aac(3)-IV</i>	C247S C250S	+
EC282	<i>aac(3)-IV</i>	W63A	+
EC283	<i>aac(3)-IV</i>	W63L	+

<sup>a</sup>Insulated constitutive promoters as described by [37]; <sup>b</sup>N-terminal truncation by 9 amino acids corresponding to the reference gene (see Figure S1).

**Table S4.** Aminoglycoside susceptibility of recombinant *E. coli* strains constitutively expressing N-terminally truncated wild-type or mutant *aac(3)-IV* under defined promoter control.

	WT	WT	WT	H154A
Strain	EC116	EC110	EC117	EC109
Promoter	+	++	+++	++
9-aa leader	no	no	no	no
MIC (mg/L)				
Apramycin	128-256	>512	>512	1-2
Gentamicin	16	64	128-256	0-125-0.25
Tobramycin	32	128	512	0.125
Sisomicin	4	32	128	0.125
Netilmicin	16-32	32-64	128	0.125-0.25
Paromomycin	4-8	16	64-128	0.5
Amikacin	0.5	0.5	0.25-0.5	0.5
Plazomicin	0.125	0.25	0.25	0.125





**Figure S2.** AAC(3)-IV activity model showing apramycin (magenta), gentamicin (orange) and acetyl-co-enzyme-A (light grey). This model is based on the published AAC(3)-IV\* H154A models (PDB ID: 6MN3, 6MN4 and 6MN5). The histidine 154 rotamer rotation was modeled based on a AAC(3)-IIIb model (PDB accession: 6mb9) and the acetyl-co-enzyme-A was modeled according to a AAC(3)-VI model (PDB ID: 6BC4). (A) general overview. (B) Magnified view of the active site. The supposed catalytic triade (T151, H154 and E157) is colored in dark green (based on ref [40]). The 3-amino groups of apramycin and gentamicin are indicated by yellow circles. Modification of the 1-amino-group (indicated by a red circle) most likely results in steric hindrance that prevents proper positioning of the 3-amino group for acetylation, drug susceptibility is therefore maintained as observed for 1-*N*-modified amikacin and plazomicin in this study.