

SUPPLEMENTAL TABLES

Supplemental Table 1. Patient characteristics and identified pathogen in the respective sample. ^a Age in months. ^b Pathogen identified by clinical microbiological analyses (culturing and/or 16S DNA. Abbreviations: RME, right middle ear effusion; LME, left middle ear effusion; NPH, nasopharynx sample; neg, negative; WR, without remark *i.e.*, no previous history of breached tympanic membrane; Myringot, previous myringotomy without ventilation tube insertion; VT, previous myringotomy with ventilation tube insertion; Perf AOM, documented history of perforated tympanic membrane due to acute otitis media; HI, *Haemophilus influenzae*; Spn, *Streptococcus pneumoniae*; Mrx, *Moraxella catarrhalis*; S.aur, *Staphylococcus aureus*; GAS, *Streptococcus pyogenes*; CoNS, coagulase negative staphylococci; A.otit, *Alloiococcus otitidis*; Staph, *Staphylococcus* spp and Neiss, *Neisseria* spp.

<i>Patient characteristics</i>			<i>RME</i>		<i>LME</i>		<i>NPH</i>
Patient	Gender	Age ^a	Pathogen identified ^b	Comment	Pathogen identified ^b	Comment	Pathogen identified ^b
Pat 2	Male	110	neg	WR	neg	WR	neg
Pat 3	Male	75	HI	WR	neg	WR	Spn. Mrx. HI
Pat 4	Male	51	neg	WR	neg	WR	Mrx
Pat 6	Male	52	neg	WR	neg	WR	Mrx
Pat 8	Female	43	neg	Myringot.	neg	WR	Mrx
Pat 9	Male	92	neg	VT	neg	V.T.	neg
Pat 10	Male	60	neg	WR	neg	WR	Mrx
Pat 12	Female	71	neg	WR	neg	WR	HI
Pat 13	Female	74	S.aur. Mrx	WR	A. otit	WR	S.aur
Pat 14	Female	57	HI spec	WR	neg	WR	GAS. Mrx
Pat 15	Male	48	GAS, CoNS	Myringot.	neg	Myringot.	GAS. Mrx
Pat 16	Female	58	A.otit	WR	A.otit	WR	Mrx
Pat 17	Female	72	neg	WR	A.otit	WR	Mrx
Pat 19	Female	61	neg	WR	neg	WR	Mrx
Pat 20	Female	86	neg	VT	Staph. CoNS	VT	HI. Mrx
Pat 21	Female	24	HI. CoNS	WR	CoNS	WR	HI. GAS
Pat 22	Female	59	HI	WR	neg	WR	Mrx. HI
Pat 25	Male	103	neg	VT	neg	VT	Mrx
Pat 27	Male	68	neg	VT	neg	VT	Mrx
Pat 28	Female	38	neg	WR	neg	WR	Spn. HI
Pat 29	Male	47	HI	VT	neg	VT	HI. Mrx
Pat 30	Male	74	S.aur	WR	S.aur	WR	neg
Pat 31	Female	35	neg	WR	neg	WR	Spn. Mrx
Pat 33	Male	42	neg	WR	neg	WR	Spn. Mrx
Pat 34	Female	46	HI. S.aur	WR	HI	WR	HI
Pat 35	Male	32	neg	Perf AOM	neg	WR	HI. Neiss
Pat 37	Male	44	neg	WR	neg	WR	Mrx

Supplemental Table 2. Correlation coefficient of nMDS plot from Figure 1B, with *P*- and *q*-values indicated.

<i>Comparison</i>	<i>R</i> ²	<i>P</i> -value	<i>q</i> -value
<i>EEC vs MEE</i>	0.0442	0.0619	0.0619
<i>EEC vs NPH</i>	0.2515	0.0010 ^{***}	0.0015
<i>MEE vs NPH</i>	0.2163	0.0010 ^{***}	0.0015

Supplemental Table 3. Leukocyte populations and inflammatory mediators in MEEs and NPH samples. ^a Values are median percentage \pm SEM (*n*) of leukocyte subpopulation of all viable cells. ^b Exact *P*-value by Mann-Whitney Wilcoxon test. ^c percentage (%) of naïve or memory cells of all CD4⁺ or CD8⁺ T lymphocytes as determined by flow cytometry. ^d [% CD4⁺ cells of all viable cells / % CD8⁺ cells of all viable cells]. ^e Median concentration of indicated mediator in pg/mL \pm SEM (*n*), as assessed by cytometric cytokine bead array (CBA).

<i>Leukocyte subpopulation</i> ^a	<i>MEE</i>	<i>NPH</i>	<i>P-value</i> ^b
<i>CD66b</i> ⁺ neutrophils	49.80 \pm 5.84 (28)	16.76 \pm 6.35 (17)	0.110
<i>CD14</i> ⁺ <i>HLA-DR</i> ⁺ monocytes/macrophages	1.31 \pm 0.55 (28)	1.17 \pm 1.35 (17)	0.890
<i>CD3</i> ⁺ T lymphocytes	4.65 \pm 3.61 (28)	15.00 \pm 2.88 (17)	0.185
<i>CD4</i> ⁺ T-helper cells	5.18 \pm 1.51 (20)	6.71 \pm 1.72 (14)	0.568
% <i>CD45RA</i> ⁺ naïve <i>CD4</i> ⁺ cells ^c	3.84 \pm 8.21 (12)	4.95 \pm 1.2 (9)	0.638
% <i>CD45RO</i> ⁺ memory <i>CD4</i> ⁺ cells ^c	87.35 \pm 8.82 (12)	89.10 \pm 9.73 (9)	0.931
<i>CD8</i> ⁺ cytotoxic T cells	1.76 \pm 0.83 (20)	6.35 \pm 2.15 (15)	0.002 ^{**}
% <i>CD45RA</i> ⁺ naïve <i>CD8</i> ⁺ cells ^c	26.45 \pm 5.66 (12)	8.07 \pm 2.03 (10)	0.014 [*]
% <i>CD45RO</i> ⁺ memory <i>CD8</i> ⁺ cells ^c	63.00 \pm 6.36 (13)	75.45 \pm 5.66 (10)	0.455
<i>CD4/CD8</i> T cell ratio ^d	2.52 \pm 0.54 (20)	0.86 \pm 0.76 (15)	0.542
<i>CD19</i> ⁺ B lymphocytes	0.98 \pm 0.66 (28)	23.90 \pm 6.57 (17)	<0.0001 ^{***}
<i>CD56</i> ⁺ <i>CD3</i> ⁻ NK cells	0.065 \pm 0.37 (28)	0.00 \pm 0.07 (16)	0.244
<i>CD56</i> ⁺ <i>CD3</i> ⁺ NKT cells	0.00 \pm 0.06 (27)	0.00 \pm 0.03 (17)	>0.999
<i>Inflammatory mediator</i> ^e	<i>MEE</i>	<i>NPH</i>	<i>P-value</i> ^b
<i>IL-6</i>	155.43 \pm 348.36 (50)	44.83 \pm 29.93 (27)	0.063
<i>IL-8</i>	1963.34 \pm 603.07 (50)	2141.41 \pm 677.84 (27)	0.666
<i>IL-10</i>	2.95 \pm 4.29 (50)	2.46 \pm 2.31 (27)	0.549
<i>TNF</i>	0.73 \pm 4.32 (50)	3.61 \pm 1.89 (27)	0.013 [*]
<i>IL-1β</i>	9.43 \pm 68.49 (50)	94.90 \pm 43.09 (27)	0.002 ^{**}

Supplemental Table 4. Leukocyte populations and inflammatory mediators in MEEs and NPH samples. Samples were stratified according to age at myringotomy (cut-off median age of patients; 58 months). ^a Values are median percentage \pm SEM (*n*) of leukocyte subpopulation of all viable cells. ^b Exact *P*-value by Mann-Whitney Wilcoxon test. ^c percentage (%) of naïve or memory cells of all CD4⁺ or CD8⁺ T lymphocytes as determined by flow cytometry. ^d [% CD4⁺ cells of all viable cells / % CD8⁺ cells of all viable cells]. ^e Median concentration of indicated mediator in pg/mL \pm SEM (*n*), as assessed by cytometric cytokine bead array (CBA).

<i>Leukocyte subpopulation</i> ^a	<i>MEE</i>			<i>NPH</i>		
	<58mo	>58mo	<i>P</i> -value ^b	<58mo	>58mo	<i>P</i> -value ^b
<i>CD66b</i> ⁺ neutrophils	75.1 \pm 9.42 (13)	44.69 \pm 6.16 (15)	0.022*	32.56 \pm 9.58 (8)	5.96 \pm 8.24 (9)	0.277
<i>CD14</i> ⁺ <i>HLA-DR</i> ⁺ monocytes/macrophages	2.61 \pm 0.81 (13)	0.6 \pm 0.78 (15)	0.768	2.08 \pm 1.34 (8)	0 \pm 2.35 (9)	0.162
<i>CD3</i> ⁺ T lymphocytes	2.77 \pm 2.09 (13)	4.96 \pm 6.06 (15)	0.085	13.9 \pm 2.52 (8)	25.9 \pm 4.72 (9)	0.144
<i>CD4</i> ⁺ T-helper cells	5.18 \pm 1.39 (6)	4.99 \pm 2.08 (14)	>0.999	8.75 \pm 2.57 (6)	5.30 \pm 2.43 (8)	0.573
% <i>CD45RA</i> ⁺ naïve <i>CD4</i> ⁺ cells ^c	2.33 \pm 13.73 (5)	10.8 \pm 10.86 (7)	0.399	9.76 \pm 1.82 (5)	4.65 \pm 1.11 (4)	0.286
% <i>CD45RO</i> ⁺ memory <i>CD4</i> ⁺ cells ^c	94.2 \pm 14.37 (5)	75.6 \pm 11.41 (7)	0.213	88.2 \pm 2.79 (5)	89.4 \pm 21.77 (4)	0.905
<i>CD8</i> ⁺ cytotoxic T cells	1.58 \pm 0.19 (6)	1.96 \pm 1.15 (14)	0.534	6.35 \pm 2.93 (7)	6.90 \pm 3.28 (8)	0.955
% <i>CD45RA</i> ⁺ naïve <i>CD8</i> ⁺ cells ^c	8.33 \pm 9.78 (5)	33.3 \pm 5.61 (7)	0.073	8.62 \pm 1.55 (5)	7.14 \pm 4.01 (5)	0.691
% <i>CD45RO</i> ⁺ memory <i>CD8</i> ⁺ cells ^c	75.0 \pm 8.65 (5)	59.4 \pm 8.65 (8)	0.210	75.0 \pm 9.34 (5)	83.0 \pm 5.85 (5)	0.222
<i>CD4/CD8</i> T cell ratio ^d	3.4 \pm 0.64 (6)	2.19 \pm 0.74 (13)	0.124	1.34 \pm 1.12 (7)	0.86 \pm 1.21 (7)	0.463
<i>CD19</i> ⁺ B lymphocytes	1.08 \pm 0.57 (13)	0.3 \pm 1.13 (15)	0.708	25.7 \pm 10.04 (8)	21.6 \pm 9.2 (9)	0.838
<i>CD56</i> ⁺ <i>CD3</i> ⁻ NK cells	0.0 \pm 0.17 (13)	0.56 \pm 0.64 (15)	0.289	0.0 \pm 0.06 (7)	0.18 \pm 0.11 (9)	0.306
<i>CD56</i> ⁺ <i>CD3</i> ⁺ NKT cells	0.0 \pm 0.0 (12)	0.0 \pm 0.10 (15)	0.226	0.0 \pm 0.07 (8)	0.0 \pm 0.0 (9)	0.206
<i>Inflammatory mediator</i> ^e	<58mo	>58mo	<i>P</i> -value ^b	<58mo	>58mo	<i>P</i> -value ^b
<i>IL-6</i>	103.6 \pm 589.9 (28)	105.9 \pm 159.6 (26)	0.602	44.6 \pm 20.5 (14)	44.8 \pm 58.9 (13)	0.905
<i>IL-8</i>	2272.2 \pm 959.3 (28)	1149.7 \pm 526.5 (26)	0.265	1883.6 \pm 478.2 (14)	2344.9 \pm 1320.3 (13)	0.607
<i>IL-10</i>	2.9 \pm 6.5 (28)	2.3 \pm 4.5 (26)	0.721	2.1 \pm 1.8 (14)	2.5 \pm 4.4 (13)	0.838
<i>TNF</i>	0.7 \pm 7.6 (28)	0.2 \pm 1.2 (26)	0.340	3.7 \pm 3.2 (14)	3.2 \pm 1.9 (13)	0.693
<i>IL-1β</i>	4.9 \pm 115.3 (28)	8.5 \pm 40.3 (26)	0.515	108.8 \pm 60.3 (14)	76.8 \pm 63.3 (13)	0.519

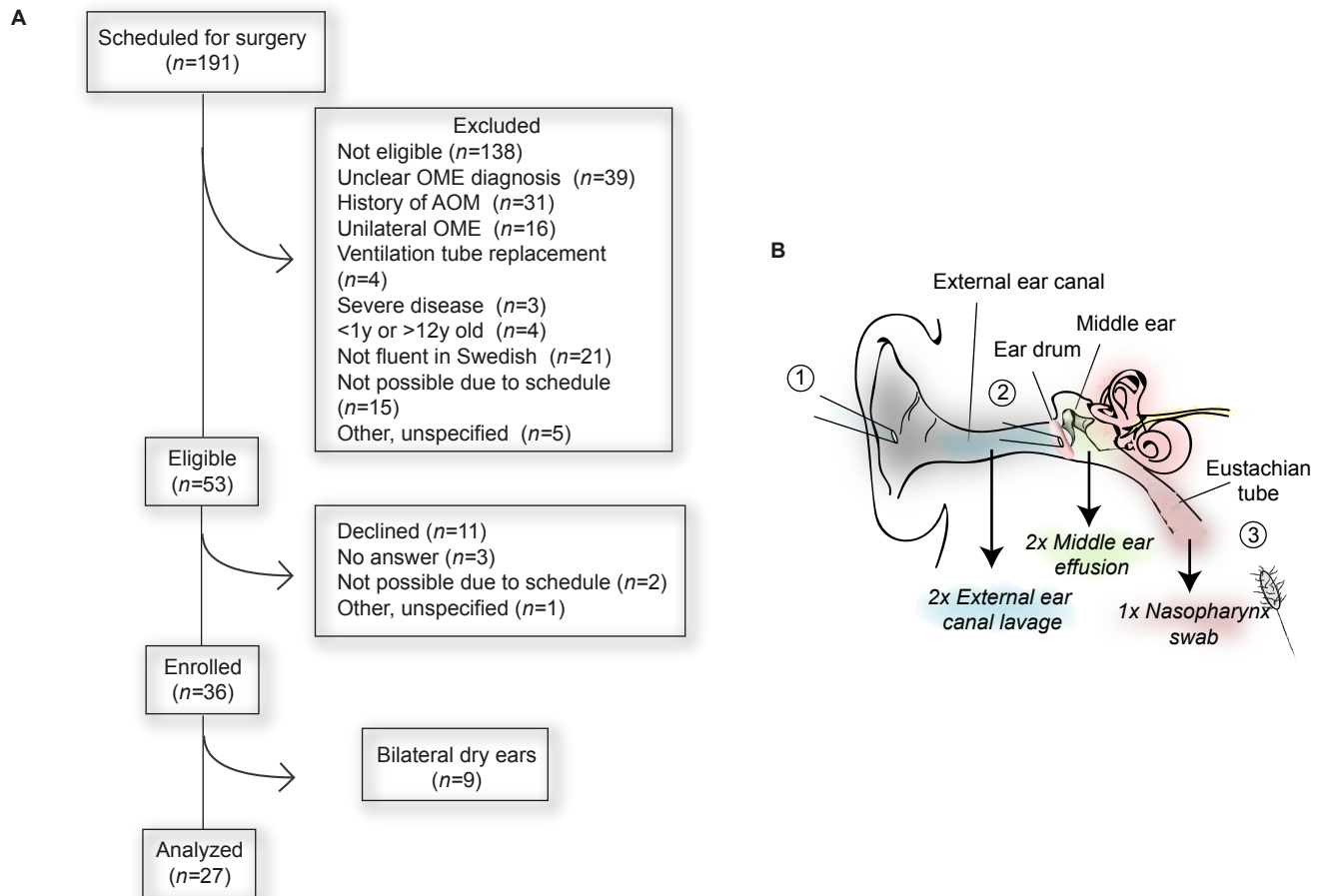
Supplemental Table 5. Spearman correlation of inflammatory cells and mediators in NPH samples. The potential correlation between inflammatory cells and mediators in NPH samples was assessed by Spearman r correlation. R values and significance is indicated. * $P < 0.05$. Abbreviations; Neu, neutrophils (CD66b⁺); Mo-Macr, monocytes/macrophages (CD14⁺HLA-DR⁺); T cells (CD3⁺); CD8⁺ cytotoxic T cells; CD4⁺ T-helper cells; CD4/CD8, %CD4⁺ cells / % CD8⁺ cells; B cells (CD19⁺); NK cells (CD56⁺CD3⁻) and NKT cells (CD56⁺CD3⁺).

<i>Mediator</i>	<i>Leukocyte population</i>								
	<i>Neu</i>	<i>Mo-Macr</i>	<i>T cells</i>	<i>CD8 Tc</i>	<i>CD4 Th</i>	<i>CD4/C D8</i>	<i>Bc</i>	<i>NK</i>	<i>NKT</i>
<i>IL-8</i>	0.284	0.210	0.105	-0.248	0.179	-0.314	-0.179	0.149	0.053
<i>IL-1β</i>	0.135	0.134	0.006	0.108	-0.207	0.068	0.107	0.035	0.486*
<i>IL-6</i>	0.032	-0.125	0.278	0.090	0.089	-0.103	0.153	-0.041	-0.053
<i>IL-10</i>	-0.055	-0.014	0.084	-0.097	-0.170	-0.150	0.086	-0.011	-0.272
<i>TNFα</i>	0.248	-0.147	0.177	-0.152	0.089	-0.262	-0.071	-0.093	0.280
<i>IL10/TNF</i>	-0.121	0.276	0.013	-0.181	-0.046	-0.170	0.075	0.265	-0.401
<i>IL-1ra</i>	-0.262	0.029	0.332	-0.033	-0.060	-0.253	-0.007	0.209	-0.103
<i>IL-2</i>	0.064	-0.007	0.095	0.014	0.014	-0.185	-0.113	-0.137	0.381
<i>IL-4</i>	-0.009	-0.079	0.150	0.146	-0.061	-0.091	-0.013	-0.026	0.310
<i>IL-5</i>	0.012	-0.241	0.156	0.302	0.067	0.123	0.006	0.258	0.480
<i>IL-7</i>	-0.242	0.014	0.260	0.151	-0.008	-0.132	0.108	0.220	0.276
<i>IL-9</i>	0.015	0.452	-0.024	-0.264	0.082	-0.368	-0.011	-0.030	0.172
<i>IL-12</i>	0.084	0.014	0.135	0.006	0.299	-0.246	-0.042	-0.107	0.381
<i>IL-13</i>	-0.164	0.102	0.284	0.028	0.163	-0.213	0.049	0.268	0.208
<i>IL-15</i>	0.215	-0.155	0.069	0.208	0.054	0.101	-0.110	0.029	0.474
<i>IL-17</i>	0.196	0.155	-0.046	0.038	-0.104	-0.104	-0.095	-0.134	0.310
<i>Eotaxin</i>	-0.037	-0.088	0.345	0.236	-0.115	0.077	0.121	0.448	-0.241
<i>FGF basic</i>	0.068	0.065	0.112	0.440	-0.297	0.473	0.398	0.149	0.378
<i>G-CSF</i>	-0.253	0.209	0.297	-0.011	0.203	-0.253	0.051	0.134	0.310
<i>GM-CSF</i>	-0.123	-0.057	0.123	0.311	-0.183	0.128	0.234	0.209	0.384
<i>IFNγ</i>	-0.284	-0.038	0.327	-0.214	0.011	-0.390	-0.020	0.078	0.034
<i>IP-10</i>	0.083	0.329	0.196	-0.100	0.078	-0.228	-0.002	0.577	-0.106
<i>MCP-1</i>	0.187	0.025	0.130	-0.071	0.071	-0.264	-0.156	0.123	0.172
<i>MIP-1α</i>	0.244	0.178	0.064	-0.220	0.269	-0.473	-0.288	-0.131	-0.103
<i>PDGF-bb</i>	0.071	0.262	-0.281	0.147	-0.050	0.047	-0.129	-0.111	0.420
<i>MIP-1β</i>	0.393	0.187	0.055	-0.192	0.242	-0.363	-0.222	-0.123	-0.172
<i>RANTES</i>	-0.108	0.299	0.099	-0.181	0.071	-0.302	0.077	0.011	0.310
<i>VEGF</i>	0.187	0.107	0.147	0.011	0.072	-0.138	-0.111	0.103	0.243
<i>M-CSF</i>	0.219	0.401	-0.100	-0.074	-0.130	-0.091	-0.069	-0.040	0.173
<i>IL-18</i>	0.125	-0.250	0.191	0.489	-0.401	0.588	0.231	0.198	0.172

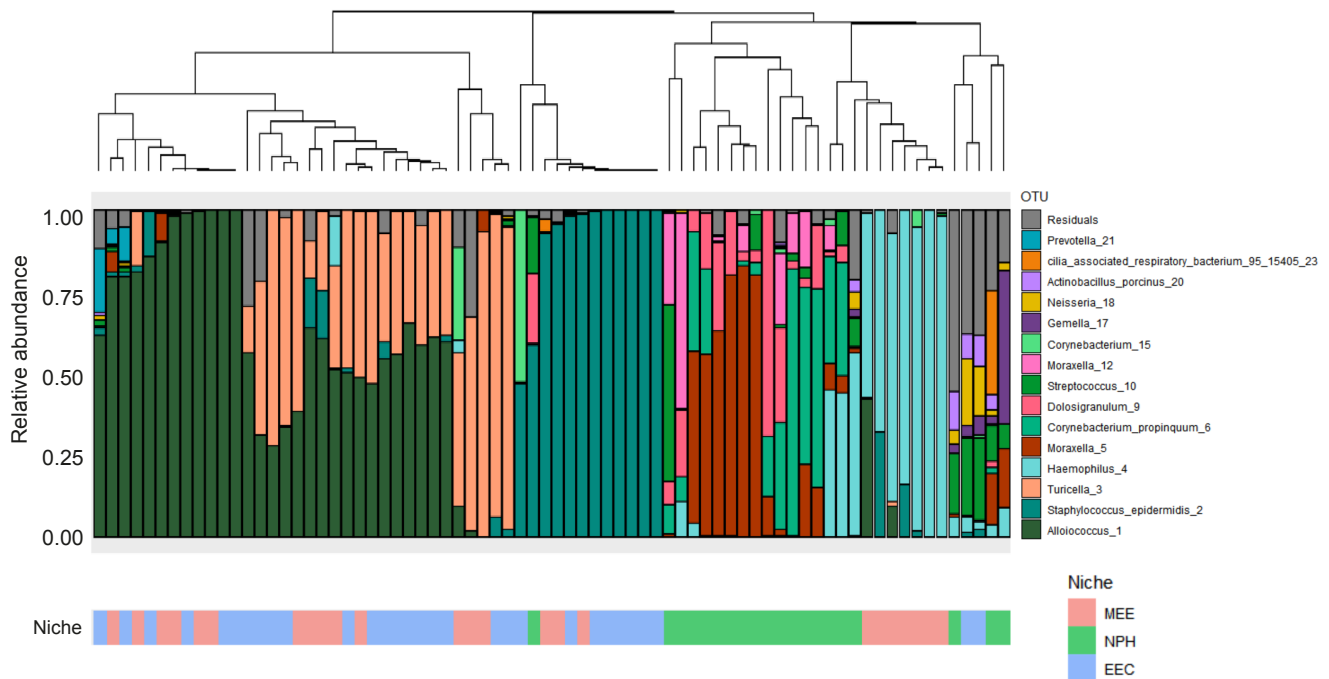
Supplemental Table 6. PerMANOVA analysis of β -diversity and clinical parameters.

<i>All samples</i>				
<i>Parameter</i>	<i>Number of samples</i>	<i>Degrees of freedom</i>	<i>R2</i>	<i>P-value</i>
<i>Niche (NPH, MEE, EEC)</i>	74	2	0.225	0.001 ***
<i>Age (<58 mo, >58 mo)</i>	74	1	0.019	0.110
<i>Gender (male, female)</i>	74	1	0.009	0.507
<i>Affected ears (uni-, bilateral)</i>	74	1	0.012	0.342
<i>Inflammation type (neutrophil / lymphocyte predominance)</i>	26	2	0.048	0.267
<i>Middle ear effusion samples</i>				
<i>Parameter</i>	<i>Number of samples</i>	<i>Degrees of freedom</i>	<i>R2</i>	<i>P-value</i>
<i>Age (<58 mo, >58 mo)</i>	24	1	0.065	0.220
<i>Gender (male, female)</i>	24	1	0.042	0.386
<i>Affected ears (uni-, bilateral)</i>	24	1	0.067	0.176
<i>Nasopharyngeal samples</i>				
<i>Parameter</i>	<i>Number of samples</i>	<i>Degrees of freedom</i>	<i>R2</i>	<i>P-value</i>
<i>Age (<58 mo, >58 mo)</i>	20	1	0.025	0.870
<i>Gender (male, female)</i>	20	1	0.053	0.422
<i>Affected ears (uni-, bilateral)</i>	20	1	0.054	0.420

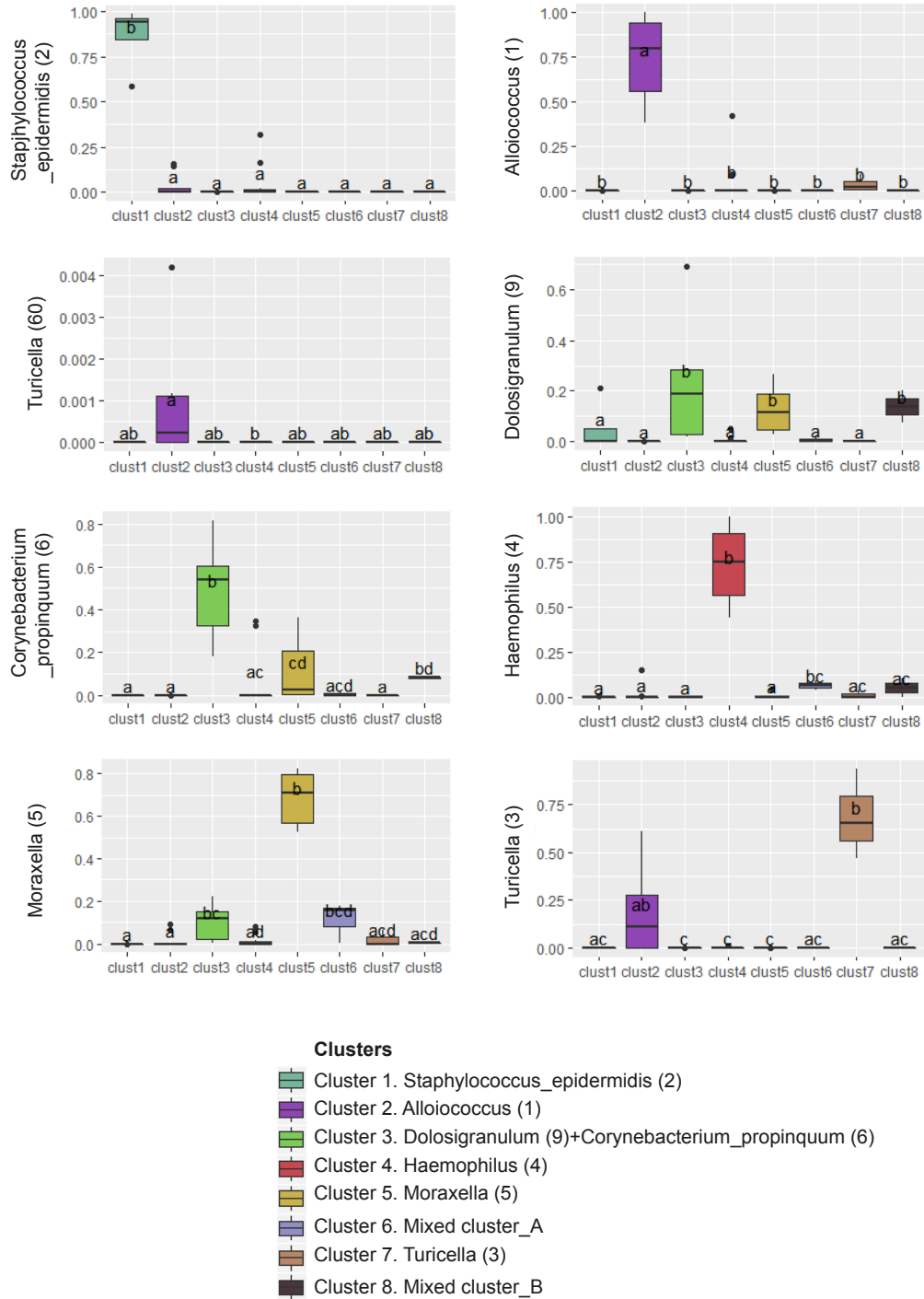
SUPPLEMENTAL FIGURES



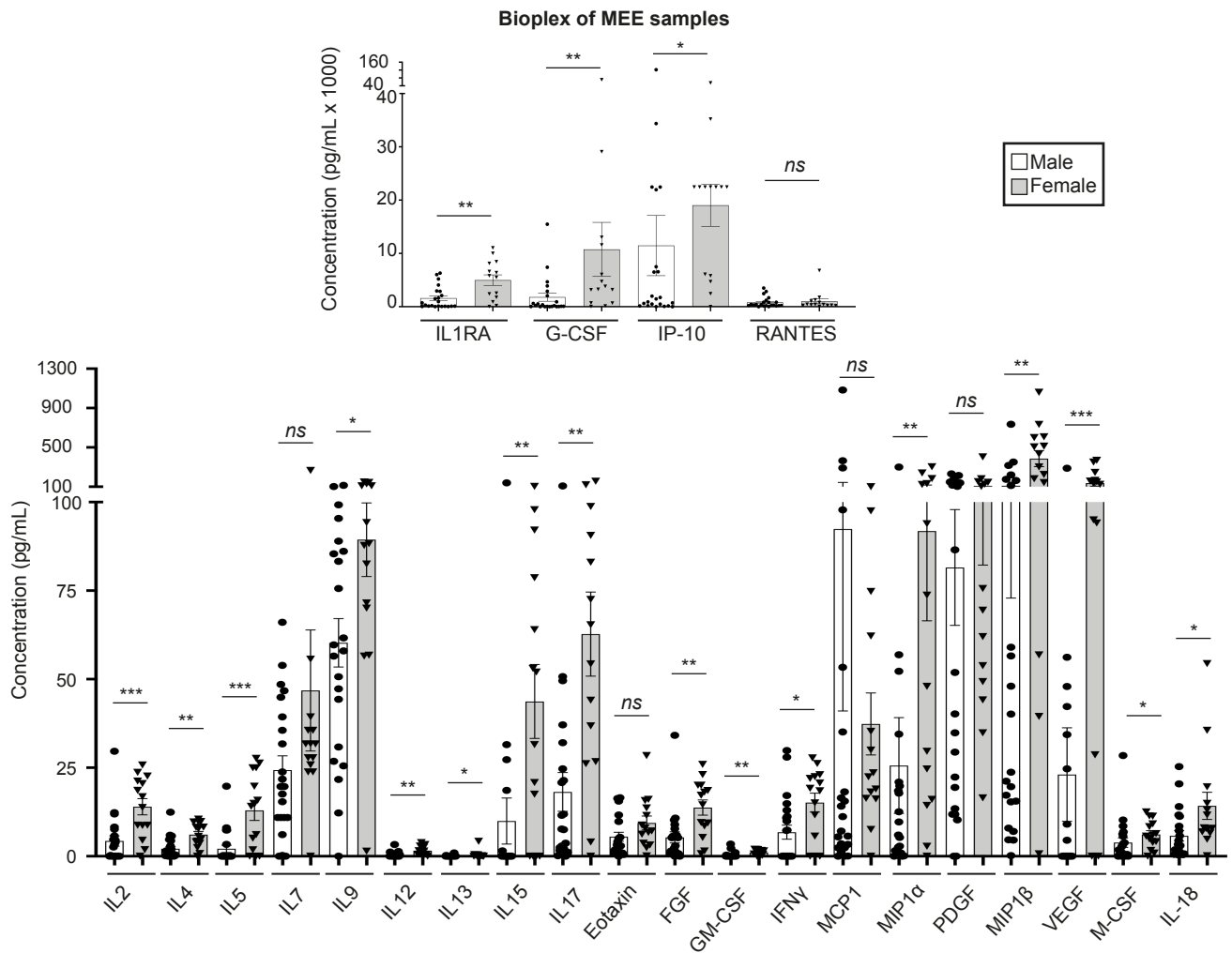
Supplemental Figure 1. Overview of the patient inclusion and sample collection. A. A schematic summary of the patient inclusion. Of 191 patients scheduled for surgery, 36 patients were enrolled in the study. Nine patients were excluded due to bilateral dry ears. The remaining 27 patients were analyzed for inflammatory parameters and microbiota composition. **B.** Sample collection was performed as follows: 1) The external ear canals of the enrolled patients were first washed with sterile saline for analyses of the external ear canal (EEC) microbiota composition. 2) After a subsequent wash with 70% denatured alcohol, the middle ear effusions (MEEs) were collected and analyzed for inflammatory cells and mediators as well as microbiota composition. 3) Nasopharyngeal (NPH) swabs from all patients were reconstituted in sterile saline and analyzed for inflammatory cells and mediators as well as microbiota composition.



Supplemental Figure 2. Niche-based hierarchical clustering of microbiota profiles. Dendrogram visualizing an average linkage hierarchical clustering of all study samples on the basis of the Bray–Curtis dissimilarity matrix. Stacked bar charts show the relative abundance of the 15 highest-ranked operational taxonomic units (OTUs) and of residual bacteria. Niche identity is depicted adjacent to the branch ends. NPH samples mainly clusters separately and are dominated by *Moraxella* (5), *Corynebacterium propinquum* (6), *Streptococcus* (10), *Haemophilus* (4) and *Dolosigranulum* (9). MEEs and EEC lavages mainly clusters together, although MEEs also shared some features with NPH samples with a high relative abundance of *Alloiooccus* (1), followed by *Haemophilus* (4), *Turicella* (3) and *Staphylococcus epidermidis* (2), whereas EEC lavages displayed high relative abundance of *Alloiooccus* (1), *S. epidermidis* (2) and *Turicella* (3).



Supplemental Figure 3. Biomarker species stratified according to the identified clusters. Hierarchical clustering and random forest analysis identified eight clusters and biomarker species. Boxplot showing relative abundance of the identified biomarker species, stratified by microbiome clusters. Multivariable linear regression modeling was used to assess the association between the log-transformed biomarker abundance and the clusters corrected by niche with Tukey post hoc test, values with unlike letters differed significantly.



Supplemental Figure 4. Bio-plex analyses of MEE samples. MEE samples were analyzed by Bio-plex multiple cytokine assay for inflammatory mediators and stratified according to gender. Concentration (pg/mL) of indicated inflammatory mediators in individual samples with bars representing mean concentration \pm SEM shown. Male MEEs (black circles, white bars) $n=22$ and female MEEs (black triangles, grey bars) $n=14$. All statistics by Mann Whitney U test * $P < 0.05$, ** $P < 0.01$ and *** $P < 0.001$.