

Supporting Information

Synthesis and Characterization of Heterodimers and Fluorescent Nisin Species by Incorporation of Methionine Analogues and Subsequent Click Chemistry

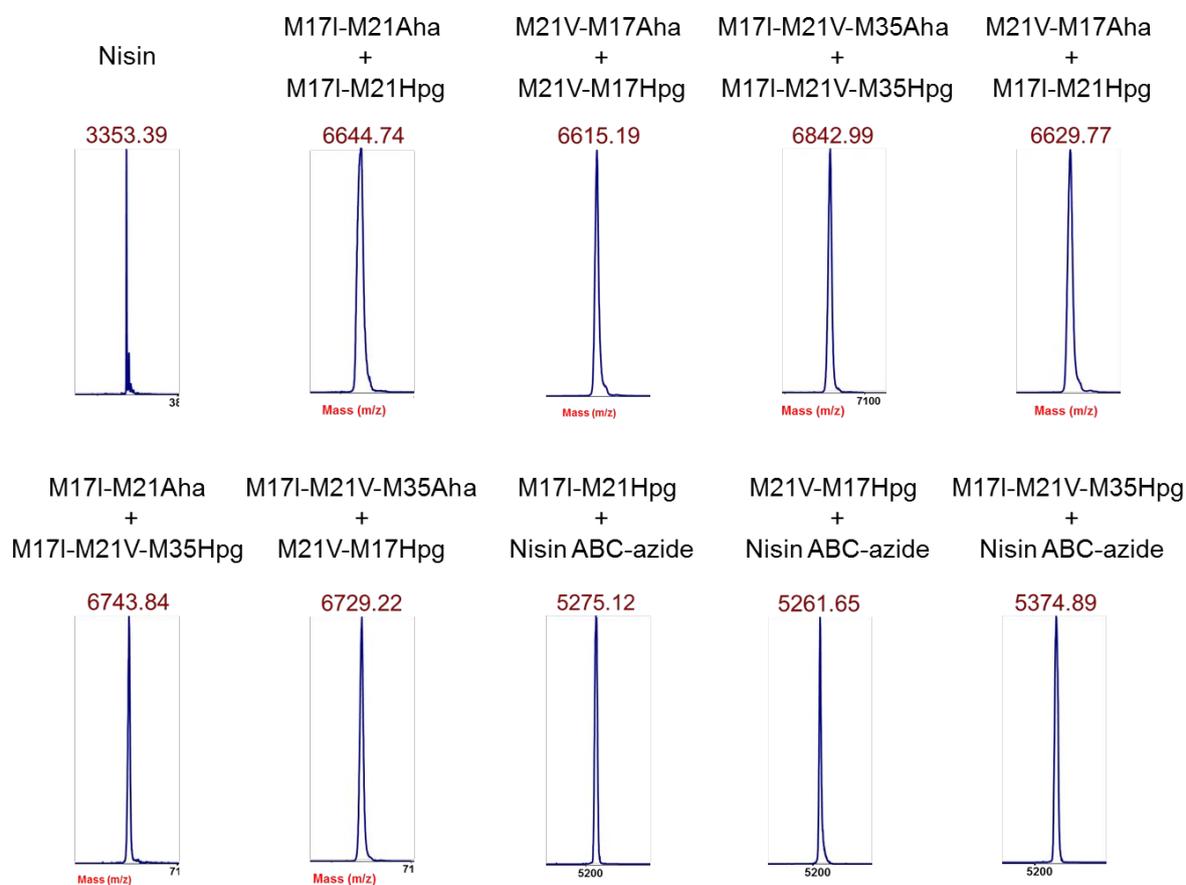
Jingjing Deng, Jakob H. Viel, Jingqi Chen and Oscar P. Kuipers*

Department of Molecular Genetics, University of Groningen, Nijenborgh 7, 9747 AG
Groningen, The Netherlands.

* Corresponding author: Email: o.p.kuipers@rug.nl

Supplementary Figures

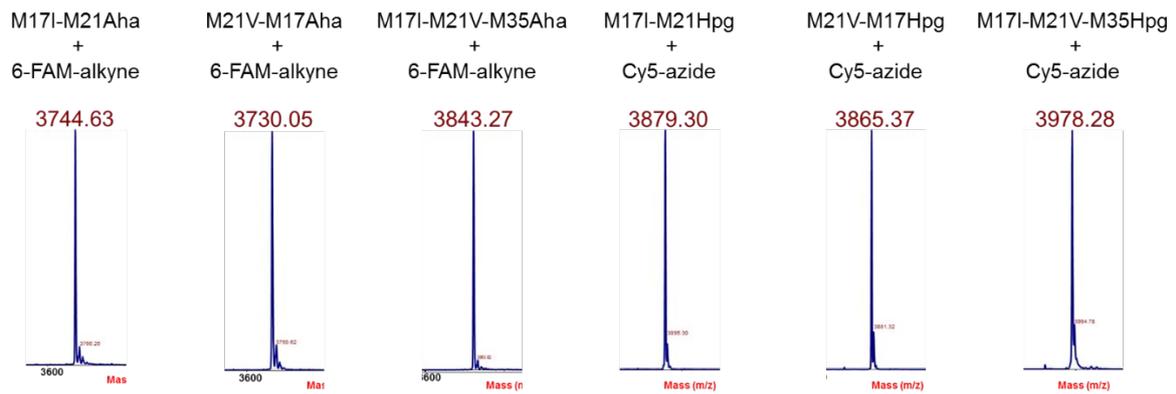
A



B

Peptides	Predicted mass (Da)	Measured Mass (Da)
Nisin	3353.03	3353.39
M17I-M21Aha + M17I-M21Hpg	6644.97	6644.74
M21V-M17Aha + M21V-M17Hpg	6616.93	6615.19
M17I-M21V-M35Aha + M17I-M21V-M35Hpg	6843.23	6842.99
M21V-M17Aha + M17I-M21Hpg	6630.95	6629.77
M17I-M21Aha + M17I-M21V-M35Hpg	6744.10	6743.84
M17I-M21V-M35Aha + M21V-M17Hpg	6730.08	6729.22
M17I-M21Hpg + Nisin ABC-azide	5276.39	5275.12
M21V-M17Hpg + Nisin ABC-azide	5262.37	5261.65
M17I-M21V-M35Hpg + Nisin ABC-azide	5375.52	5374.89

Supplementary Figure 1. A). MALDI-TOF analysis of nisin, dimeric nisin constructs and nisin hybrids. B). Predicted and measured mass of nisin, dimeric nisin constructs and nisin hybrids.

A**B**

Peptides	Predicted mass (Da)	Measured Mass (Da)
M17I-M21Aha + 6-FAM-alkyne	3744.36	3744.63
M21V-M17Aha + 6-FAM-alkyne	3730.34	3730.05
M17I-M21V-M35Aha + 6-FAM-alkyne	3843.49	3843.27
M17I-M21Hpg + Cy5-azide	3879.36	3879.30
M21V-M17Hpg + Cy5-azide	3865.34	3865.37
M17I-M21V-M35Hpg + Cy5-azide	3978.49	3978.28

Supplementary Figure 2. A). MALDI-TOF analysis of fluorescently labeled nisin variants. B). Predicted and measured mass of fluorescently-labeled nisin variants.

Supplementary Tables

Supplementary Table 1. Strains and plasmids used in this study.

Strains or Plasmids	Characteristics	References
Strains		
<i>Lactococcus lactis</i> NZ9000	<i>pepN::nisRK</i> ; Expression host strain	1
Indicator strains		
<i>Micrococcus flavus</i>		Lab collection
<i>Staphylococcus aureus</i> CAL	Methicillin resistant (MRSA)	The University Medical Center Groningen, The Netherlands
<i>Staphylococcus aureus</i> MW2	Methicillin resistant (MRSA)	The University Medical Center Groningen, The Netherlands
<i>Enterococcus faecium</i> LMG 16003	Avoparcin and vancomycin resistant (VRE)	Laboratory of Microbiology, Gent, Belgium
<i>Enterococcus faecalis</i> LMG 16216	Vancomycin resistant (VRE)	Laboratory of Microbiology, Gent, Belgium
<i>Bacillus cereus</i> ATCC 14579		2
<i>Listeria monocytogenes</i> LMG 10470		3
Plasmids		
pIL3EryBTC	EryR, <i>nisBTC</i> , modification and transport of lantibiotics	4
pCZ-nisA	CmR, <i>nisA</i> , encoding NisA, under the control of PczcD promoter	5
pCZ-nisA-M17I	Point mutant of pCZ-nisA, with the Met 17 of nisin changed to Ile	This work
pCZ-nisA-M21V	Point mutant of pCZ-nisA, with the Met 21 of nisin changed to Val	This work
pCZ-nisA-M17I-M21V-M35	Point mutant of pCZ-nisA, with the Met 17 and 21 of nisin changed to Ile and Val, respectively, with Met 35	This work
pCZ-nisA-I4M-M17I-M21V	Point mutant of pCZ-nisA, with the Ile 4, Met 17 and Met 21 of nisin changed to Met, Ile and Val, respectively	This work
pNZnisP8H	CmR, <i>nisP</i> , encoding NisP mutant, with 8 histines	6

Supplementary Table 2. Primers used in this study

Mutants	Primer	Sequence
M17I	pCZ-F	aacagtagtggcctcgtagc
	M17I-Rev	gctgttttcatgttacaaccaatcagagctcctgttttac
	M17I-Fwd	gtaaaacaggagctctgattggtgtaacatgaaaacagc
	pCZ-R	tagtctcggacattctgctc
M21V	pCZ-F	aacagtagtggcctcgtagc
	M21V-Rev	tacaatgacaagttgctgttttacgttacaacccatcagagctc
	M21V-Fwd	agctctgatgggtgtaacgtaaaaacagcaactgtcattgtag
	pCZ-R	tagtctcggacattctgctc
M17I- M21V- M35	NheI-For	atcagctagcacggaatagacatggtgttc
	M35-Rev1	ctacaatgacaagttgctgttttacgttacaaccaatcagagctcctgttttac
	M35-Rev2	taccgatgcctgcaggcttacatttgcttacgtgaatactacaatgacaagttg
I4M- M17I- M21V	NheI-For	atcagctagcacggaatagacatggtgttc
	I4-Rev1	cagagctcctgttttacaaccgggtgtacatagcgacatacttgaatgctgggtg
	I4-Rev2	acaatgacaagttgctgttttacgttacaaccaatcagagctcctgttttac
	I4-Rev3	taccgatgcctgcaggcttatttgcttacgtgaatactacaatgacaagttgctg

References

- (1) Kuipers, O. P., de Ruyter, P. G. G. A., Kleerebezem, M. and de Vos, W. M. (1997) Controlled overproduction of proteins by lactic acid bacteria. *Trends biotechnol.* 15, 135-140.
- (2) Ivanova, N., Sorokin, A., Anderson, I., Galleron, N., Candelon, B., Kapatral, V., Bhattacharyya, A., Reznik, G., Mikhailova, N., Lapidus, A., Chu, L., Mazur, M., Goltsman, E., Larsen, N., D'Souza, M., Walunas, T., Grechkin, Y., Pusch, G., Haselkorn, R., Fonstein, M., Ehrlich, S. D., Overbeek, R. and Kyrpides, N. (2003) Genome sequence of *Bacillus cereus* and comparative analysis with *Bacillus anthracis*. *Nature* 423, 87-91.
- (3) Enan, G. (2006) Control of the regrowing bacteriocin resistant variants of *Listeria monocytogenes* LMG 10470 *in vitro* and in food by nisinplantaricin UG1 mixture. *Biotechnology* 5, 143-147.
- (4) van Heel, A. J., Mu, D., Montalban-Lopez, M., Hendriks, D., and Kuipers, O. P. (2013) Designing and producing modified, new-to-nature peptides with antimicrobial activity by use of a combination of various lantibiotic modification enzymes. *ACS synth. biol.* 2, 397-404.
- (5) Mu, D., Montalban-Lopez, M., Masuda, Y., and Kuipers, O. P. (2013) Zirex: a novel zinc-regulated expression system for *Lactococcus lactis*. *Appl. Environ. Microbiol.* 79, 4503-4508.
- (6) Montalban-Lopez, M., Deng, J., van Heel, A. J., and Kuipers, O. P. (2018) Specificity and application of the lantibiotic protease NisP. *Front. Microbiol.* 9, 160.