

Hypothalamic CDK4 regulates thermogenesis by modulating sympathetic innervation of adipose tissues

Judit Castillo-Armengol, Valentin Burquissau, Sarah Geller, Honglei Ji, Ilenia Severi, Wiebe Venema, Eric-Aria Fernandez, Catherine Moret, Katharina Huber, Lucia Leal-Esteban, Anita Nasrallah, Laia Martinez-Carreres, Guy Niederhäuser, Patricia Seoane Collazo, Sylviane Lagarrigue, Miguel Lopez, Antonio Giordano, Sophie Croisier, Bernard Thorens, Isabel Lopez-Mejia, and Lluis Fajas **DOI: 10.15252/embr.201949807**

Corresponding author(s): Lluis Fajas (lluis.fajas@unil.ch)

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Transaction Report:

(Note: With the exception of the correction of typographical or spelling errors that could be a source of ambiguity, letters and reports are not edited. Depending on transfer agreements, referee reports obtained elsewhere may or may not be included in this compilation. Referee reports are anonymous unless the Referee chooses to sign their reports.)

Dear Lluis,

Thank you for the submission of your research manuscript to our journal, which was now seen by two referees, whose reports are copied below.

As you can see, the referees express interest in the analysis. However, they also raise a number of concerns that need to be addressed to consider publication here. In particular, referees require better support for beta-3-receptor activation (ref #1, point 2), better characterization/quantification of TH levels of CDK4 depleted mice (ref #1 points 3, 4 and ref #2 point 3) and calorimetry data (ref #2 point 1), and more support into neuronal specificity of CDK4 role (ref #2 point 4). I find the reports informed and constructive, and believe that addressing the concerns raised will significantly strengthen the manuscript.

Given these constructive comments, we would like to invite you to revise your manuscript with the understanding that the referee concerns (as in their reports) must be fully addressed and their suggestions taken on board. Please address all referee concerns in a complete point-by-point response. Acceptance of the manuscript will depend on a positive outcome of a second round of review. It is EMBO reports policy to allow a single round of revision only and acceptance or rejection of the manuscript will therefore depend on the completeness of your responses included in the next, final version of the manuscript.

We generally allow three months as standard revision time. As a matter of policy, competing manuscripts published during this period will not negatively impact on our assessment of the conceptual advance presented by your study. However, we request that you contact the editor as soon as possible upon publication of any related work, to discuss how to proceed. Should you foresee a problem in meeting this three-month deadline, please let us know in advance and we may be able to grant an extension.

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2) individual production quality figure files as .eps, .tif, .jpg (one file per figure).

3) a .docx formatted letter INCLUDING the reviewers' reports and your detailed point-by-point responses to their comments. As part of the EMBO Press transparent editorial process, the point-by-point response is part of the Review Process File (RPF), which will be published alongside your paper. For more details on our Transparent Editorial Process, please visit our website: https://www.embopress.org/page/journal/14693178/authorguide#transparentprocess You are able to opt out of this by letting the editorial office know (emboreports@embo.org). If you do opt out, the Review Process File link will point to the following statement: "No Review Process File is available with this article, as the authors have chosen not to make the review process public in this case."

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- Additional Tables/Datasets should be labeled and referred to as Table EV1, Dataset EV1, etc. Legends have to be provided in a separate tab in case of .xls files. Alternatively, the legend can be supplied as a separate text file (README) and zipped together with the Table/Dataset file.

7) We would also encourage you to include the source data for figure panels that show essential data.

Numerical data should be provided as individual .xls or .csv files (including a tab describing the data). For blots or microscopy, uncropped images should be submitted (using a zip archive if multiple images need to be supplied for one panel). Additional information on source data and instruction on how to label the files are available .

8) Regarding data quantification, please ensure to specify the name of the statistical test used to generate error bars and P values, the number (n) of independent experiments underlying each data point (not replicate measures of one sample), and the test used to calculate p-values in each figure

legend. Discussion of statistical methodology can be reported in the materials and methods section, but figure legends should contain a basic description of n, P and the test applied. Please note that error bars and statistical comparisons may only be applied to data obtained from at least three independent biological replicates.

Please also include scale bars in all microscopy images.

We would also welcome the submission of cover suggestions, or motifs to be used by our Graphics Illustrator in designing a cover.

I look forward to seeing a revised version of your manuscript when it is ready. Please let me know if you have questions or comments regarding the revision.

Kind regards,

Deniz

Deniz Senyilmaz Tiebe, PhD Editor EMBO Reports

Referee #1:

The paper describes the role of Cdk4 in the regulation of brown adipocyte function. The authors demonstrate that global loss of Cdk4 leads to increased browning, cold resistance as well as enhanced energy expenditure. The authors nicely show that the effect is due to beta-adrenergic signaling but seems to be non-cell autonomous since specific ablation of Cdk4 in BAT does not phenocopy the global knockout mouse. In a VMH specific Cdk4 ko mouse the higher fiber density observed in the global ko mice is phenocopied suggesting that Cdk4 controls BAT activity by regulation of sympathetic innervation.

There are several points that need to be addressed:

1. The introduction is guite long, the authors should shorten it, especially the summary at the end which is in essence the abstract

2. The regulation of beta-3-receptor mRNA in the inguinal WAT should be treated with caution. Data indicates that prolonged activation leads to a reduction rather than an increase in the receptor mRNA. If the authors want to base some arguments on this regulation the protein levels and localizations would need to be shown.

3. Fig.4 and Fig. 5 are TH protein levels increased in ingWAT and scWAT?

4. The data from Fig. 5 is a bit counter-intuitive. The authors observe the same increase in TH fibers but no change in thermogenesis. This obviously makes it difficult to reconcile the observations. Did they check in ingWAT, possibly under stimulated conditions if thermogenesis is altered. Are the compensatory reductions, which could explain the lack of a response (betareceptors for example). The argument of embryonic deletion does not really fit as the global knockout would probably recombine even earlier than the VMH specific one.

Referee #2:

In the manuscript "Hypothalamic CDK4 regulates thermogenesis by modulating the sympathetic innervation and activation of brown adipose tissue," Castillo-Armengol et al. propose a role for CDK4 in the regulation of thermogenesis via SF1 neurons of the ventromedial hypothalamus. They demonstrate that CDK4 mice have higher energy expenditure under standard housing conditions and respond better to cold challenge. This is explained by increased activity of the brown adipose tissue. While knockout of CDK4 in BAT does not phenocopy whole body knockout, knockout of CDK4 in SF1 neurons in the VMH is able to partially reproduce the phenotype, suggesting a role of CKD4 in hypothalamic control of metabolism.

This work demonstrates a previously unknown role for CDK4, more thoroughly studied as a cellcycle regulatory protein, in control of metabolism. Although there are publications exploring the role of CDK4 in muscle and adipose tissue, this is the first to demonstrate role for this protein in CNS control of metabolism. These observations will be of interest to those following the increasing number of publications regarding this topic. However, there are still some issues that must be addressed.

1. In the analysis of indirect calorimetry data from the CLAMS system, the authors use averages of VO2, EE, and RER to compare groups. While the differences they observe are valid and statistically significant, a more nuanced and complete analysis could be done using ANCOVA. This would be easily done with publicly available software such as CaIR.

2. The Western blots in the paper are normalized to Ponceau stain, which is less sensitive than a loading control such as Actin or GAPDH. Such a loading controls should be included.

3. In figures 4 and 5, the authors demonstrate that CDK4 knockout results in an increase in THpositive fibers in BAT. Although the IHC images are accompanied by a quantitation of these fibers, the validity of this data is still subject to the sections of tissue chosen for analysis. A bulk analysis of the BAT depot using Western blot should be performed to show an increase in TH overall. It would also be informative to demonstrate an increase in phosphorylated hormone sensitive lipase by Western, as would be expected with an increase in adrenergic signaling in this tissue.

4. Finally, the authors should more thoroughly address the issue that knockout of CDK4 in the SF1 neurons of the VMH does not fully recapitulate the whole-body knockout phenotype. While the SF1-Cre mice show resistance to cold exposure, they do not exhibit the differences in BAT morphology or gene expression or the changes in energy expenditure, food intake, and indirect calorimetry measurements that the whole-body CDK4 knockout mice exhibit. While the authors hypothesize in the discussion that this could due to developmental effects, it seems more likely that SF1 neurons of the VMH are only partially responsible for the phenotype. Other regions of the CNS, for example, could also contribute, as could elements of the peripheral nervous system. This should at the very least be addressed in the discussion. Experiments with a pan-neuronal Cre, such as SynI Cre, would effectively address the question of whether neuronal CDK4 is entirely responsible for the phenotypes observed in the global CDK4 knockout. Alternatively, injection of AAV-Cre into specific brain regions could be used to identify other nuclei involved in regulation of thermogenesis by CDK4.

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The paper describes the role of Cdk4 in the regulation of brown adipocyte function. The authors demonstrate that global loss of Cdk4 leads to increased browning, cold resistance as well as enhanced energy expenditure. The authors nicely show that the effect is due to beta-adrenergic signaling but seems to be non-cell autonomous since specific ablation of Cdk4 in BAT does not phenocopy the global knockout mouse. In a VMH specific Cdk4 ko mouse the higher fiber density observed in the global ko mice is phenocopied suggesting that Cdk4 controls BAT activity by regulation of sympathetic innervation.

We thank this reviewer for the comments and suggestions. We have performed several new experiments to prove the physiological relevance of our findings. We have addressed the major concerns and suggestions as follows:

There are several points that need to be addressed:

1. The introduction is quite long, the authors should shorten it, especially the summary at the end which is in essence the abstract.

Following the reviewer's suggestion, we have now reviewed the introduction and done the appropriate changes to shorten and improve it.

2. The regulation of beta-3-receptor mRNA in the inguinal WAT should be treated with caution. Data indicates that prolonged activation leads to a reduction rather than an increase in the receptor mRNA. If the authors want to base some arguments on this regulation the protein levels and localizations would need to be shown.

To address this question, and following this reviewer suggestion, we have done two experiments. First, whole tissue lysates of scWAT were analyzed by Western blotting to assess the beta-3-adrenergic receptor (ADRB3) levels of expression. This experiment indicated no significant differences between scWAT of $Cdk4^{+/+}$ and $Cdk4^{-/-}$ mice (Appendix Fig S1A-B). On the other hand, we have performed a tissue subcellular fractionation of the same samples to determine the location of ADRB3 protein. This experiment clearly showed that ADRB3 was located at the plasma membrane in the scWAT of both $Cdk4^{+/+}$ and $Cdk4^{+/-}$ mice (Appendix Fig S1C). In conclusion, despite our previous experiment showed that Adrb3 gene expression was significantly increased in the scWAT of $Cdk4^{+/-}$ mice, no differences were found at the level of protein expression. The lack of differences, together with our results showing no differences in TH protein expression in the tissue (see next question), suggest that the adrenergic pathway is not changed in the white adipose tissue of the $Cdk4^{-/-}$ mice.

Appendix Figure S1



3. Fig.4 and Fig. 5 are TH protein levels increased in ingWAT and scWAT?

We have performed immunohistochemical analysis of TH protein in the scWAT of both models of *Cdk4* deletion (*Cdk4-/-* and *Cdk4flox/flox Sf1-Cre* mice). We have observed no differences in TH protein levels in the scWAT of *Cdk4-/-* mice (Appendix Fig S2A-B). In contrast, a higher amount of TH positive fibers was found in the *Cdk4flox/flox Sf1-Cre* compared to the control *Cdk4flox/flox* scWAT (Fig EV5B-C). This result was confirmed by the protein quantification resulting from a western blot analysis of TH (Fig EV5D-E). These new findings suggest that in the *Cdk4flox/flox Sf1-Cre* there is an overall increase in the sympathetic innervation in adipose tissue and that both BAT and scWAT are contributing to the enhanced cold resistance of these animals.

Appendix Figure S2



Figure EV5



4. The data from Fig. 5 is a bit counter-intuitive. The authors observe the same increase in TH fibers but no change in thermogenesis. This obviously makes it difficult to reconcile the observations. Did they check in ingWAT, possibly under stimulated conditions if thermogenesis is altered. Are the compensatory reductions, which could explain the lack of a response (beta-receptors for example). The argument of embryonic deletion does not really fit as the global knockout would probably recombine even earlier than the VMH specific one.

We agree with this reviewer that increased innervation of the tissue (TH) should result in changes in gene expression in BAT. We could not find, however differences in the expression of thermogenesis genes in this tissue. But consistent with increased TH, these mice were more resistant to cold. As this reviewer proposes, changes in WAT could, at least partially, explain this paradox. Following reviewer's suggestion, we have analyzed the expression levels of thermogenic genes in the scWAT of the $Cdk4^{flox/flox}Sf1$ -Cre mice in basal conditions. We found a tendency to increase thermogenic gene markers such as Ucp1 or Dio2, and a significant increase in the expression of the thermogenic gene marker Prdm16, essential for brown and beige adipocyte identity and function (Fig EV5A). Most important is the finding that the protein expression of UCP1 was highly increased in the scWAT of the $Cdk4^{flox/flox}Sf1$ -Cre mice indicating and confirming that these animals present a recruitment of thermogenic adipocytes in the scWAT depot in order to defeat thermal stress (Fig EV5D, F).





Referee #2:

In the manuscript "Hypothalamic CDK4 regulates thermogenesis by modulating the sympathetic innervation and activation of brown adipose tissue," Castillo-Armengol et al. propose a role for CDK4 in the regulation of thermogenesis via SF1 neurons of the ventromedial hypothalamus. They demonstrate that CDK4 mice have higher energy expenditure under standard housing conditions and respond better to cold challenge. This is explained by increased activity of the brown adipose tissue. While knockout of CDK4 in BAT does not phenocopy whole body knockout, knockout of CDK4 in SF1 neurons in the VMH is able to partially reproduce the phenotype, suggesting a role of CKD4 in hypothalamic control of metabolism.

This work demonstrates a previously unknown role for CDK4, more thoroughly studied as a cell-cycle regulatory protein, in control of metabolism. Although there are publications exploring the role of CDK4 in muscle and adipose tissue, this is the first to demonstrate role for this protein in CNS control of metabolism. These observations will be of interest to those following the increasing number of publications regarding this topic. However, there are still some issues that must be addressed.

We would like to thank the reviewer for the positive evaluation of our manuscript.

1. In the analysis of indirect calorimetry data from the CLAMS system, the authors use averages of VO2, EE, and RER to compare groups. While the differences they observe are valid and statistically significant, a more nuanced and complete analysis could be done using ANCOVA. This would be easily done with publicly available software such as CalR.

As suggested by the reviewer, we have now used the Cal R software (Mina, LeClair et al., 2018) to use the generalized linear model (GLM), utilizing body mass as a covariate to model mass-dependent metabolic parameters obtained with indirect calorimetry. The results of these novel analysis are now found in Figure 1G-I, and figure EV4I-J. Both the "genotype effect" (or "group effect") and the "mass effect" are now shown. A clear significant genotype difference was identified for energy balance, with *Cdk4*-/- mice clearly showing a negative energy balance at room temperature, in agreement with their increased oxidative capacity, thermogenesis and sympathetic tone, and a near significant energy balance difference between *Cdk4*+/+ mice and *Cdk4*-/- mice at 6°C; and this despite a modest sample size (N=6-5).

Figure 1



Figure EV4



	24°C		
Effect	Mass	Group	
Oxygen Consumption (ml/hr)	0.8159	0.2338	
Carbon Dioxide Production (ml/hr)	0.6008	0.4934	GLM
Energy Expenditure (kcal/hour)	0.7562	0.2788	
Energy Balance (kcal/hour)	0.2474	0.7553	
	24°C		4
Effect		Group	l S
Respiratory Exchange Ratio		0.4689	ANG

RER, a mass independent metabolic parameter was analyzed using ANOVA. A *p*-value of 0.2058 was obtained at room temperature and a p-value of <0.001 *** was obtained at 6°C.

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However, no significant differences were identified using GLM when total body mass was used a covariate for oxygen consumption and energy expenditure.

We have chosen to depict these mass-dependent metabolic variables normalized to body weight, due to dramatic differences of body mass observed between $Cdk4^{+/+}$ mice and $Cdk4^{+/-}$ mice ($Cdk4^{-/-}$ mice weigh on average 9 grams less than $Cdk4^{+/+}$ mice, meaning roughly 30% less).

2. The Western blots in the paper are normalized to Ponceau stain, which is less sensitive than a loading control such as Actin or GAPDH. Such a loading controls should be included.

We have performed additional western blot analysis to add the appropriate loading controls for each experiment. HSP90, GAPDH or TUBULIN have been used for this purpose. The newly included loading controls can be found in Fig 2D, Fig 3F, Fig 6H, Fig EV3C, Fig EV5D.

3. In figures 4 and 5, the authors demonstrate that CDK4 knockout results in an increase in THpositive fibers in BAT. Although the IHC images are accompanied by a quantitation of these fibers, the validity of this data is still subject to the sections of tissue chosen for analysis. A bulk analysis of the BAT depot using Western blot should be performed to show an increase in TH overall. It would also be informative to demonstrate an increase in phosphorylated hormone sensitive lipase by Western, as would be expected with an increase in adrenergic signaling in this tissue.

We thank this reviewer for this suggestion. We have now analyzed the protein expression of TH in BAT protein lysates of equivalent protein concentration and found, surprisingly, a decrease between *Cdk4-/-* and *Cdk4+/+* mice (Appendix Fig S3A-B). *Appendix Fig S3*



This result is in conflict with the previous reported observations of increase TH positive fibers by immunohistochemistry performed in the same tissues. It is important to mention that our immunohistochemistry studies are 10 representative photos of 2-3 sections for each mouse, covering a representative surface of the whole tissue. Also, the IHC quantification of TH positive fibers is normalized to 100 adipocytes, meaning that the increase in the sympathetic fiber content of the *Cdk4-/-* mice is a measure of the stimulation that the thermogenic cells can receive (Fig 4A-B). We then asked for the underlying reason that could explain the discrepancy between western blot (WB) and immunohistochemistry (IHC) results. Indeed, the morphology and structure of the BAT of *Cdk4+/+* and Cdk4-/- mice is very different. As seen on images of Figure 4A, lipid content is strikingly decreased in *Cdk4+/-* mice, leading to a higher amount of proteins per amount of tissue (Appendix Fig S3E). WB analyses are normalized by tissue protein content, whereas IHC results are normalized by number of adipocytes. Hence, considering one similar nerve fiber innervating one adipocyte, the ratio of TH content to the protein will be artificially increased in $Cdk4^{+/+}$ mice and decreased in $Cdk4^{+/-}$ mice. In order to overcome this issue, we have analyzed the expression of TH protein normalized by the tissue weight (i.e., we loaded each well with the amount of proteins corresponding to the same amount of tissue). In these conditions, we found a clear increase in TH expression in the BAT of $Cdk4^{+/-}$ mice when compared to an equal amount of BAT from the $Cdk4^{+/+}$ mice (Appendix Fig S3F-G). Similar results were found for the synaptic protein PSD95 (Appendix Fig S3F, H), as well as for UCP1 (Fig S3F, I).

Appendix Fig S3



It is important to take into account that the IHC quantification of TH positive fibers is normalized to 100 adipocytes, meaning that the increase in the sympathetic fiber content of the Cdk4-/- mice is a measure of the stimulation that the thermogenic cells can receive (Fig 4A-B). According to this experiment, the brown adipocytes in the BAT of Cdk4-/- mice are more prone to be stimulated because they have more sympathetic fibers in their proximity compared to the adipocytes of the BAT from the Cdk4+/+ mice. In summary, our data shows that the BAT of Cdk4-/-, considered as a whole tissue, is more innervated and has an increased TH content than the BAT of Cdk4+/+ mice.

As requested by the reviewer, we have also analyzed the expression levels of the phosphorylated hormone-sensitive lipase protein (p-HSL^{S660}) in BAT protein lysates of equivalent protein concentration. We observed no significant differences in the levels of expression of p-HSL^{S660} (Appendix Fig S3C-D). This controversial result can be explained by the fact that as the BAT of the *Cdk*4-/- is delipidated in basal conditions, there might be a compensatory inhibition of HSL activity.

Appendix Figure S3



We have also analyzed the same parameters in the $Cdk4^{flox/flox}$ Sf1-Cre mice. First, we have explored the TH protein expression by western blot in the BAT of these animals in basal conditions. The expression of TH protein was increased in the $Cdk4^{flox/flox}$ Sf1-Cre mice, in accordance with the immunohistochemical analysis performed before (Appendix Fig S4A-B). Interestingly, the morphology and structure of the BAT of these mice is identical to the BAT of control mice.

Appendix Figure S4



Nevertheless, the analysis of phosphorylated hormone sensitive lipase (p-HSL^{S660}) in the same animals showed a big variability in the samples and no differences in the expression of this protein (Appendix Fig S4C-D). This controversial result could be explained by the fact that the animals had a minimum thermal stress in these conditions (RT).

Appendix Figure S4



4. Finally, the authors should more thoroughly address the issue that knockout of CDK4 in the SF1 neurons of the VMH does not fully recapitulate the whole-body knockout phenotype. While the SF1-Cre mice show resistance to cold exposure, they do not exhibit the differences in BAT morphology or gene expression or the changes in energy expenditure, food intake, and indirect

calorimetry measurements that the whole-body CDK4 knockout mice exhibit. While the authors hypothesize in the discussion that this could due to developmental effects, it seems more likely that SF1 neurons of the VMH are only partially responsible for the phenotype. Other regions of the CNS, for example, could also contribute, as could elements of the peripheral nervous system. This should at the very least be addressed in the discussion. Experiments with a pan-neuronal Cre, such as SynI Cre, would effectively address the question of whether neuronal CDK4 is entirely responsible for the phenotypes observed in the global CDK4 knockout. Alternatively, injection of AAV-Cre into specific brain regions could be used to identify other nuclei involved in regulation of thermogenesis by CDK4.

We particularly thank this reviewer for this suggestion. We agree that it is important to determine if *Cdk4* deletion modulates neuronal activity in other thermogenic brain nuclei which could explain why the phenotype of the *Cdk4*^{flox/flox} *Sf1-Cre* mice does not exactly replicate the phenotype of the *Cdk4*^{-/-} mice. To address this hypothesis, we have evaluated the neuronal activity in the brain of *Cdk4*-/- mice by assessing the expression of c-FOS, which is a transcription factor whose expression is enhanced upon neuronal depolarization and it has been broadly used as a marker of neuronal activation (Bullitt, 1990, West, Griffith et al., 2002). Therefore, we quantified the number of c-FOS immunoreactive cells in the brain of *Cdk*4^{+/+} and *Cdk*4^{+/+} mice in basal conditions paying special attention to the medial preoptic area (mPOA) (Morrison, 2016, Morrison, Madden et al., 2014, Yoshida, Nakamura et al., 2003), the arcuate (ARC) (Rahmouni & Morgan, 2007), the ventromedial hypothalamus (VMH) (Kim, Zhao et al., 2011, Seoane-Collazo, Roa et al., 2018), the dorsomedial hypothalamus (DMH) (Hogan, Coscina et al., 1982, Monge-Roffarello, Labbe et al., 2014) and the lateral hypothalamic area (LH) (Martins, Seoane-Collazo et al., 2016, Sellayah, Bharaj et al., 2011); all well-known areas involved in the response to thermal stress. Immunofluorescence analysis showed that $Cdk4^{-/-}$ mice have a significant increment of c-FOS positive cells in the VMH when compared to the *Cdk*4^{+/+} control mice. This data is consistent with our previous findings in the *Cdk4*^{flox/flox} *Sf1-Cre* mice which indicated that CDK4 is an important regulator of the VMH activity.

Interestingly, a significant higher staining of c-FOS positive cells was also found in the ARC and LHA and it is important to mention that a high tendency to an increased c-FOS expression was also found in the mPOA of the Cdk4-/- mice. This is a very exciting finding, not only because indicates that the phenotype of the Cdk4-/- mice could be explained by the relevant participation of CDK4 in other brain regions that might be also regulating thermogenesis, but also because it points to a new role of CDK4 in general homeostasis in the brain.

The representative images and quantification of this experiment have been added to the manuscript in new Figure 5. These new findings were also added in the discussion of the revised version of this manuscript.

New figure 6



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Dear Lluis,

Thank you for submitting the revised version of your manuscript. It has now been seen by both of the original referees.

As you can see, the referees find that the study is significantly improved during revision and recommend publication. Before I can accept the manuscript, I need you to address some minor points below:

• Please comment on the overexposure issue of the tubulin blot of EV Figure 2A in the figure legend. Please upload the image exported from the Western Blot developer that shows the saturation points as source data and refer to it in the figure legend, as well.

• As per our guidelines, please add a 'Data Availability Section', where you state that no data were deposited in a public database.

- We noted that Wiebe Venema's initials are misspelled as VB in Author Contributions.
- We realized that the funding information is incomplete in the manuscript submission system.
- We noted that the appendix figures are currently not called out in the text.

• Papers published in EMBO Reports include a 'Synopsis' to further enhance discoverability. Synopses are displayed on the html version of the paper and are freely accessible to all readers. The synopsis includes a short standfirst summarizing the study in 1 or 2 sentences that summarize the key findings of the paper and are provided by the authors and streamlined by the handling editor. I would therefore ask you to include your synopsis blurb.

• In addition, please provide an image for the synopsis. This image should provide a rapid overview of the question addressed in the study but still needs to be kept fairly modest since the image size cannot exceed 550x400 pixels.

• Our production/data editors have asked you to clarify several points in the figure legends (see attached document). Please incorporate these changes in the attached word document and return it with track changes activated.

Thank you again for giving us to consider your manuscript for EMBO Reports, I look forward to your minor revision.

Kind regards,

Deniz

Deniz Senyilmaz Tiebe, PhD Editor EMBO Reports

Referee #1:

The authors adressed all my concerns very convincingly and I would suggest to accept the paper

Referee #2:

The revised paper is acceptable for publication.

Dear Deniz,

First of all, I want to thank you for the overall process of review of our manuscript. We appreciate the comments of both the reviewers and the editor. We are now sending a revised version addressing the minor points raised in the last review.

• Please comment on the overexposure issue of the tubulin blot of EV Figure 2A in the figure legend. Please upload the image exported from the Western Blot developer that shows the saturation points as source data and refer to it in the figure legend, as well.

We explain now in the figure legend the issue of saturation. We refer to the file from the imager that we uploaded in the data source folder.

• As per our guidelines, please add a 'Data Availability Section', where you state that no data were deposited in a public database.

We added the Data availability section

• We noted that Wiebe Venema's initials are misspelled as VB in Author Contributions.

This has been corrected

• We realized that the funding information is incomplete in the manuscript submission system.

We included all funding information in the online submission system

• We noted that the appendix figures are currently not called out in the text.

We are now referring to the appendix figures in the text.

• Papers published in EMBO Reports include a 'Synopsis' to further enhance discoverability. Synopses are displayed on the html version of the paper and are freely accessible to all readers. The synopsis includes a short standfirst summarizing the study in 1 or 2 sentences that summarize the key findings of the paper and are provided by the authors and streamlined by the handling editor. I would therefore ask you to include your synopsis blurb.

We have uploaded the synopsis blurb

• In addition, please provide an image for the synopsis. This image should provide a rapid overview of the question addressed in the study but still needs to be kept fairly modest since the image size cannot exceed 550x400 pixels.

We have uploaded a synopsis image

• Our production/data editors have asked you to clarify several points in the figure legends (see attached document). Please incorporate these changes in the attached word document and return it with track changes activated.

We clarified and addressed the points raised by the production editors.

Dear Lluis,

Thank you for submitting your revised manuscript. I have now looked at everything and all looks fine. Therefore I am very pleased to accept your manuscript for publication in EMBO Reports.

Congratulations on a nice work!

Kind regards,

Deniz

Deniz Senyilmaz Tiebe, PhD Editor EMBO Reports

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