Supplementary Information

## A Cu(II)-ATP complex efficiently catalyses enantioselective Diels-Alder reactions

Wang et al.

## **Supplementary Tables**

Entry	Cofactor	Conversion (%)	Endo/exo	ee ( <b>3a</b> %, <i>exo</i> )	ee ( <b>3a</b> %, <i>endo</i> )
1	Cu(OTf) <sub>2</sub>	98	91:9	79	72
2	CuSO <sub>4</sub>	95	90:10	77	67
3	CuCl <sub>2</sub>	98	91:9	74	62
4	Cu(NO <sub>3</sub> ) <sub>2</sub>	90	91:9	74	65

Supplementary	/ Table 1	Enantiosoloctivo	Diols-Aldor	reaction cat	alvzod by	Cu <sup>2+</sup> . ATP wit	h different	conner(II)	ealte a
Supplemental	y lable	Enantioselective	Diels-Aluel	reaction cat	alyzeu by '		n umerent	copper(ii) :	saits.

<sup>a</sup> Reaction conditions: **1a** (1 mM), **2** (200 mM), ATP (250 μM), Cu(II) salt (50 μM), MES buffer (20 mM, pH 5.5), 4 °C, 24 h.

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Entry	ATP (μM)	Cu(OTf) <sub>2</sub> (μM)	Conversion (%)	Endo/exo	ee ( <b>3a</b> %, <i>endo</i> )
1	500	50	92	91:9	66
2	250	50	98	91:9	72
3	100	50	67	91:9	57
4	50	50	63	91:9	46
5	25	50	83	91:9	20
6	10	50	78	91:9	7
7	5	50	73	92:8	1
8	250	25	72	89:11	66
9	250	50	98	91:9	72
10	250	125	79	91:9	62
11	250	250	67	91:9	52
12	250	500	88	92:8	46

### Supplementary Table 2 Enantioselective Diels-Alder reaction catalyzed by Cu<sup>2+</sup> ATP with different molar ratios.<sup>a</sup>

<sup>a</sup> Reaction conditions: **1a** (1 mM), **2** (200 mM), ATP (5-500 μM), Cu(OTf)<sub>2</sub> (25-500 μM), MES buffer (20 mM, pH 5.5), 4 °C, 24 h.

Entry	Additive	Concentration (mM)	Conversion (%)	Endo/exo	ee ( <b>3a</b> %, <i>endo</i> )
1	none	none	98	91:9	72
2	NaCl	10	74	90:10	65
3	KCI	10	87	90:10	67
4	NH₄CI	10	75	90:10	65
5	MgCl <sub>2</sub>	10	90	91:9	53
6	MgCl <sub>2</sub>	50	67	91:9	42
7	MgCl <sub>2</sub>	100	92	90:10	31
8	MgCl <sub>2</sub>	250	94	93:7	18
9	MgCl <sub>2</sub>	500	85	93:7	4
10	MgCl <sub>2</sub>	1000	87	93:7	0

### Supplementary Table 3 Enantioselective Diels-Alder reaction catalyzed by Cu<sup>2+</sup> ATP with different additives.<sup>a</sup>

<sup>a</sup> Reaction conditions: **1a** (1 mM), **2** (200 mM), additive, ATP (250 μM), Cu(OTf)<sub>2</sub> (50 μM), MES buffer (20 mM, pH 5.5), 4 °C, 24 h.

Supplementary Table 4 Enantioselective Diels-Alder reaction	n catalyzed by Cu <sup>2+</sup> ·ATP at different temperatures. <sup>a</sup>
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Entry	<i>T</i> (°C)	Conversion (%)	Endo/exo	ee ( <b>3a</b> %, <i>endo</i> )
1	0	87	91:9	66
2	4	98	91:9	72
3	15	98	90:10	62
4	25	96	91:9	56
5	37	71	89:11	44

<sup>a</sup> Reaction conditions: **1a** (1 mM), **2** (200 mM), ATP (250 μM), Cu(OTf)<sub>2</sub> (50 μM), MES buffer (20 mM, pH 5.5), 24 h.

Entry	Buffer	рН	Conversion (%)	Endo/exo	ee ( <b>3a</b> %, <i>endo</i> )
1	MES	5.0	80	90:10	63
2	MES	5.5	98	91:9	72
3	MES	6.0	90	90:10	65
4	MES	6.5	74	90:10	64
5	MOPS	6.5	93	89:11	69
6	MOPS	7.0	89	89:11	67
7	MOPS	7.4	87	88:12	69
8	Tris-HCI	7.4	27	86:14	55
9	PBS	7.4	63	89:11	65

Supplementary Table 5 Enantioselective Diels-Alder reaction catalyzed by Cu<sup>2+</sup>·ATP in different buffers.<sup>a</sup>

 $^a$  Reaction conditions: 1a (1 mM), 2 (200 mM), ATP (250  $\mu$ M), Cu(OTf)\_2 (50  $\mu$ M), buffer (20 mM), 4 °C, 24 h.

# Supplementary Table 6 Comparable catalytic performances of enantioselective Diels-Alder reactions catalyzed by various metallo-biohybrid catalysts.

Reference	Publication year	Biological scaffold	Metal species	Conversion (%)	Endo/exo	ee ( <i>endo</i> , %)
This work	Not yet	ATP	Cu(OTf) <sub>2</sub>	85	84:16	84
1	1998	L-abrine	Cu(NO <sub>3</sub> ) <sub>2</sub>	>90	>90:10	74
2	2000	Ribozyme	NaCl and MgCl <sub>2</sub>	Not mentioned	95:5	95
3	2005	Double-stranded DNA	Cu(NO <sub>3</sub> ) <sub>2</sub> and 9-aminoacridine derivative	>80	91:9	53
4	2006	Double-stranded DNA	Cu(NO <sub>3</sub> ) <sub>2</sub> and 4,4'-dimethyl-2,2'- bipyridine	>80	99:1	99
5	2006	Bovine serum albumin	Copper phthalocyanine- 3,4',4'',4'''-tetrasulfonic acid tetrasodium salt	91	91:9	98
6	2009	Bovine pancreatic polypeptide	Cu(NO <sub>3</sub> ) <sub>2</sub>	73	>95:5	83
7	2010	tHisF	CuSO <sub>4</sub>	73	93:7	46
8	2010	G-quadruplex DNA	Cu(NO <sub>3</sub> ) <sub>2</sub> and 4,4'-dimethyl-2,2'- bipyridine	>85	95:5	34
9	2012	G-quadruplex DNA	Cu(NO <sub>3</sub> ) <sub>2</sub>	99	98:2	74
10	2012	LmrR_M89C-Phen	Cu(NO <sub>3</sub> ) <sub>2</sub>	93	95:5	97
11	2013	SCP-2LV83C	Cu(NO <sub>3</sub> ) <sub>2</sub> and phenanthroline derivative	20	88:22	25
12	2013	G-quadruplex DNA	5,10,15,20-Tetrakis(1- methylpyridinium-4- yl)porphyrinatocopper(II) tetraperchlorate	94	97:3	69
13	2014	Cyclic peptides	Cu(NO <sub>3</sub> ) <sub>2</sub>	85	96:4	96
14	2015	Cucurbit[8]uril	Cu(NO <sub>3</sub> ) <sub>2</sub>	99	97:3	92
15	2015	Lipase	Cu(NO <sub>3</sub> ) <sub>2</sub> and phenanthroline derivative	98	94:6	92
16	2015	G-quadruplex DNA	$Cu(NO_3)_2$ and tertpyridine	>99	98:2	99
17	2016	β-Barrel protein nitrobindin	Cu(II) terpyridyl complexes	22	90:10	Not determined
18	2016	FhuA-terpyridyl	Cu(NO <sub>3</sub> ) <sub>2</sub>	69	96:4	0
19	2016	Neocarzinostatin	Cu(NO <sub>3</sub> ) <sub>2</sub> and phenanthroline derivative	98	84:16	0
20	2016	G-tripex DNA	Cu(NO <sub>3</sub> ) <sub>2</sub>	99	99:1	64
21	2017	DNA hairpins	Cu(NO <sub>3</sub> ) <sub>2</sub> and 4,4'-dimethyl-2,2'- bipyridine	99	99:1	96
22	2017	β-Barrel protein nitrobindin	Cu(NO <sub>3</sub> ) <sub>2</sub>	56	95:5	69
23	2017	αRep A3	Cu(NO <sub>3</sub> ) <sub>2</sub> and phenanthroline derivative	11	86:14	62
24	2017	Double-stranded RNA	Cu(NO <sub>3</sub> ) <sub>2</sub> and 4,4'-dimethyl-2,2'- bipyridine	11	91:9	10
25	2018	HEK-A <sub>24</sub>	Cu(NO <sub>3</sub> ) <sub>2</sub> and phenanthroline derivative	22	80:20	28
26	2019	mTFP	Cu(NO <sub>3</sub> ) <sub>2</sub>	97	93:7	26
27	2019	αRep (A3_A3')F119C	Terpy-Cu(NO <sub>3</sub> ) <sub>2</sub>	15	93:7	52
28	2019	ACCO	Cu(NO <sub>3</sub> ) <sub>2</sub>	100	99:1	>99
29	2020	c-di-AMP	Cu(OTf) <sub>2</sub>	99	97:3	80

## **Supplementary Figures**



Supplementary Fig. 1 CD spectra of ATP (250 µM) with different copper(II) cofactors (50 µM) in MES buffer (20 mM, pH 5.5).



Supplementary Fig. 2 CD spectra of ATP (250 µM) with different divalent metal salts (50 µM) in MES buffer (20 mM, pH 5.5).



Supplementary Fig. 3 CD spectra of ATP (250 µM) with different copper(II) salts (50 µM) in MES buffer (20 mM, pH 5.5).



**Supplementary Fig. 4** CD spectra of ATP (250 µM) and Cu(OTf)<sub>2</sub> (50 µM) in the presence of different additive ions (10 mM) in MES buffer (20 mM, pH 5.5).



**Supplementary Fig. 5** CD spectra of ATP (250  $\mu$ M) and Cu(OTf)<sub>2</sub> (50  $\mu$ M) with different concentration of MgCl<sub>2</sub> (10-1000 mM) in MES buffer (20 mM, pH 5.5).



**Supplementary Fig. 6 a** HPLC analysis of the reaction medium of  $Cu^{2+}$ ·ATP-catalyzed Diels-Alder reaction. Reaction conditions: **1a** (1 mM), **2** (200 mM), ATP (250  $\mu$ M), Cu(OTf)<sub>2</sub> (50  $\mu$ M), MES buffer (20 mM, pH 5.5), 4 °C. **b** The stability of ATP in MES buffer (20 mM, pH 5.5) at different reaction time points.



Supplementary Fig. 7 CD spectra of different ATP analogues (250 µM) with Cu(OTf)<sub>2</sub> (50 µM) in MES buffer (20 mM, pH 5.5).



Supplementary Fig. 8 CD spectra of different NTPs (250 µM) with Cu(OTf)<sub>2</sub> (50 µM) in MES buffer (20 mM, pH 5.5).



Supplementary Fig. 9 CD spectra of ATP (60 µM) with different amounts of Cu(OTf)<sub>2</sub> (0-100 µM) in MES buffer (20 mM, pH 5.5).



**Supplementary Fig. 10** UV-Vis spectra of ATP (20 µM) with different amounts of Cu(OTf)<sub>2</sub> (0-200 µM) in MES buffer (20 mM, pH 5.5).



**Supplementary Fig. 11** UV-Vis spectra of ATP (20 μM) and Cu(OTf)<sub>2</sub> (20 μM) in the presence of different additives (1 mM) in MES buffer (20 mM, pH 5.5).



Supplementary Fig. 12 UV-Vis spectra of different NTPs (20 µM) with Cu(OTf)<sub>2</sub> (20 µM) in MES buffer (20mM, pH 5.5).



**Supplementary Fig. 13** Determination of the molar extinction coefficient of **1a**. The absorbance of **1a** at 326 nm was determined with the fixed concentration of **1a** of 10, 20, 30, 40, 50, 60 and 70  $\mu$ M. The molar extinction coefficient of **1a** ( $\epsilon_{1a} = 17470 \text{ M}^{-1} \text{ cm}^{-1}$ ) was estimated from the slope of the fitting curve (R<sup>2</sup> = 0.999).



Supplementary Fig. 14 Determination of the molar extinction coefficient of 3a. The absorbance of 3a at 326 nm was determined with the fixed concentration of 3a at 30, 50, 60, 70 and 80  $\mu$ M. The molar extinction coefficient of 3a ( $\epsilon_{3a} = 121 \text{ M}^{-1} \text{ cm}^{-1}$ ) was estimated from the slope of the fitting curve (R<sup>2</sup> = 0.906).



**Supplementary Fig. 15** Kinetic plots of Cu<sup>2+</sup>·ATP catalyzed D-A reactions with dienophile **1a** at the concentrations of **a** 20  $\mu$ M, **b** 30  $\mu$ M, and **c** 50  $\mu$ M. Reaction conditions: **1a** (20, 30, 50  $\mu$ M), **2** (5 mM), ATP (250  $\mu$ M), Cu(OTf)<sub>2</sub> (50  $\mu$ M), MES buffer (2000  $\mu$ L, 20 mM, pH 5.5), 4 °C.



Supplementary Fig. 16 Kinetic plots of Cu(OTf)<sub>2</sub> catalyzed D-A reactions with dienophile 1a at the concentrations of a 20  $\mu$ M, b 30  $\mu$ M, and c 50  $\mu$ M. Reaction conditions: 1a (20, 30, 50  $\mu$ M), 2 (5 mM), Cu(OTf)<sub>2</sub> (50  $\mu$ M), MES buffer (2000  $\mu$ L, 20 mM, pH 5.5), 4 °C.



**Supplementary Fig. 17** Kinetic plots of ATP catalyzed D-A reactions with dienophile **1a** at the concentrations of **a** 20 μM, **b** 30 μM, and **c** 50 μM. Reaction conditions: **1a** (20, 30, 50 μM), **2** (5 mM), ATP (250 μM), MES buffer (2000 μL, 20 mM, pH 5.5), 4 °C.



**Supplementary Fig. 18** Kinetic plots of the D-A reactions in the absence of catalysts with dienophile **1a** at the concentrations of **a** 20 μM, **b** 30 μM, and **c** 50 μM. Reaction conditions: **1a** (20, 30, 50 μM), **2** (5 mM), MES buffer (2000 μL, 20 mM, pH 5.5), 4 °C.

#### **Supplementary Methods**

**General materials.** ATP, GTP, UTP, CTP, dATP, ADP, AMP and adenosine were purchased from Sangon (Shanghai, China). The metal salts, achiral ligands and buffers were purchased from Energy Chemical and J&K Scientific Ltd. The chemicals were used without further purification unless otherwise stated. Water used was distilled and deionized using a Milli-Q A10 water purification system. Azachalcones (**1a-h**) and the racemates (**3a-h**) were prepared followed the literature.<sup>30</sup>

Circular dichroism (CD) spectra were measured on a Chirascan circular dichroism spectrometer (Applied Photophysics Ltd, UK). The CD spectra were performed using a quartz cell (1 mm optical path length), an instrument scanning speed of 100 nm min<sup>-1</sup> and were accumulated by taking the average of three scans made from 200 to 320 nm at 4 °C. UV/Vis spectra were measured on Agilent Cary 3500 or Mettler-Toledo UV5Bio spectrophotometer in a sealed quartz cell with a path length of 1.0 cm. NMR titration spectra (<sup>1</sup>H, <sup>31</sup>P NMR) of ATP with varied concentration of CuCl<sub>2</sub> were performed on a Bruker AV 600 MHz spectrometer. To a solution of ATP (120 mM) in D<sub>2</sub>O in a nuclear magnetic tube, a stock solution of CuCl<sub>2</sub> in D<sub>2</sub>O was gradually added to vary the ratio of ATP/Cu<sup>2+</sup> from 1000:1 to 5:1. <sup>1</sup>H NMR spectra and <sup>31</sup>P NMR spectra were collected at each molar ratio of ATP/Cu<sup>2+</sup>. Electron paramagnetic resonance (EPR) experiments were conducted on a Bruker EMX plus 10/12 CW X-band EPR spectrometer under a microwave frequency of 9.43 GHz equipped with a liquid nitrogen cooling system. The EPR spectra were measured at 100 K with a modulation amplitude of 0.3 mT, a modulation frequency of 100 kHz, and a microwave power of 20 mW. A mixture of ATP (final conc. 50 mM) and Cu(OTf)<sub>2</sub> (final conc. 10 mM) was added to a MES buffer (20 mM, pH 5.5) with glycerol (20 v/v%). After thoroughly mixed, the solution was drawn into a capillary tube and sealed followed by placing into an EPR tube. After cooling in liquid nitrogen for 5 min, the EPR spectra were started to collect.

**Determination of the apparent**  $k_b$ . The apparent binding constant ( $k_b$ ) of the Cu<sup>2+</sup>·ATP complex was estimated from the UV titration experiments adopted from the previous reference.<sup>31</sup> To a fixed concentration of Cu(OTf)<sub>2</sub> (5 µM) aqueous solution at room temperature, a stock solution of ATP in H<sub>2</sub>O was titrated to make the final concentration over the range from 0.1 µM to 100 µM. The apparent  $k_b$  for the Cu<sup>2+</sup>·ATP complex was calculated by the following Supplementary Equation 1:

$$[ATP]/\Delta\varepsilon_a = [ATP]/\Delta\varepsilon + 1/(\Delta\varepsilon_a \cdot k_b)$$
(1)

where  $\Delta \varepsilon_a = |\varepsilon_{app} - \varepsilon_f|$ ,  $\Delta \varepsilon = |\varepsilon_b - \varepsilon_f|$ ,  $\varepsilon_{app}$ ,  $\varepsilon_f$  and  $\varepsilon_b$  are the apparent, free and bound extinction coefficients for the complex, respectively. In a plot of [ATP]/ $\Delta \varepsilon_a$  vs. [ATP],  $k_b$  is given by the ratio of the slope to the y intercept.



Supplementary Fig. 19 UV titration of varied concentrations of ATP (0-100 µM) to a fixed concentration of Cu(OTf)<sub>2</sub> (5 µM) in H<sub>2</sub>O.

**Determination of the absolute configuration of 3a.** For the Cu<sup>2+</sup>·ATP-catalyzed D-A reaction of **1a** and **2**, the absolute configurations of the product **3a** were determined in comparison with the reported literature.<sup>22</sup> Hayashi et al. reported that the enantioselective D-A reaction of **1a** and **2** was catalyzed by NB-Pyr and Cu(II) ions, yielding the chiral product **3a** (*Si-endo*) at 78% ee in (1*R*,2*S*,3*S*,4*S*) configuration. The *endo* and *exo* isomers of **3a** were assigned on the HPLC trace using Chiralpak IA column (hexane/*i*-PrOH = 99.5:0.5, 0.5 mL min<sup>-1</sup>, 264 nm) with the order of the **3a** enantiomers of **3a** (*Re-exo*), **3a** (*Si-endo*) and **3a** (*Re-endo*).<sup>22</sup> Using the same HPLC condition and Chiralpak IA column, we analyzed the product **3a** that was obtained from the Diels–Alder reaction of **1a** and **2** catalyzed by Cu<sup>2+</sup>·ATP. The HPLC traces of **3a** in racemic form and chiral form are shown in Supplementary Fig. 21. By comparison to the reference, we determined that the major product **3a** generated by Cu<sup>2+</sup>·ATP was **3a** (*Si-endo*) in (1*R*,2*S*,3*S*,4*S*) configuration.



Supplementary Fig. 20 The absolute configurations of the D-A product 3a for the endo and exo isomers.



Supplementary Fig. 21 HPLC traces of a racemic and b product of 3a from the Diels-Alder reaction catalyzed by Cu<sup>2+</sup>·ATP using Chiralpak IA column. The major product 3a generated by Cu<sup>2+</sup>·ATP is 3a (*Si-endo*) in (1*R*,2*S*,3*S*,4*S*) configuration.

**DFT calculations.** In order to compare the stability among all candidates, the relative electronic energies were listed in Supplementary Figs. 22-25 in which the most stable configuration was set to zero.



**Supplementary Fig. 22** The proposed models of Cu<sup>2+</sup>·ATP. The relative electronic energies are in the levels of B3LYP-D3/6-311G(d,p)~LANL2DZ with a unit of kcal mol<sup>-1</sup>.



**Supplementary Fig. 23** The precursors of the intermediates of **1a**-Cu<sup>2+</sup>·ATP and **2** that yield the corresponding products **3a** in different configurations. The relative electronic energies ( $\Delta E$ ) of the precursors are shown in the table.



Supplementary Fig. 24 The intermediates of 3a-Cu<sup>2+</sup>·ATP and their relative electronic energies ( $\Delta E$ ).



**Reaction Coordinate** 

Supplementary Fig. 25 The relative electronic energy profile of the reaction path for Cu<sup>2+</sup>·ATP-catalyzed Diels-Alder reaction of 1a and 2 that yields 3a (*endo*) in the absolute configuration of 1*R*, 2*S*, 3*S*, 4*S*. Abbreviation: TS, transition state.

**HPLC analysis.** The enantioselectivity of **3a-h** was determined by a chiral HPLC column (250 × 4.6 mm) with hexane and *i*-PrOH as eluents.



<Peak table>

Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	11. 926	1350700	102932	0.000		М	
2	13. 184	1342646	91959	0.000		М	
3	14. 516	18082841	909607	0.000		М	
4	18. 570	18106792	811480	0.000		М	

**b** mV



<Peak table>

Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	11. 991	197582	13485	0.000		М	
2	13. 228	1691906	99167	0.000		М	
3	15. 130	2615605	122022	0.000		М	
4	18. 848	16044452	651347	0.000		М	

**Supplementary Fig. 26 a** HPLC trace of racemic **3a**. **b** HPLC trace of prodcut **3a** that from the Diels-Alder reaction catalyzed by Cu<sup>2+</sup>·ATP for the *exo* isomer of 79% ee and the *endo* isomer of 72% ee. HPLC condition: Daicel Chiralpak ODH column, hexane/*i*-PrOH 98:2, 0.5 mL min<sup>-1</sup>, 254 nm, 25 °C.



Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	11.091	1780210	113474	0.000		М	
2	12. 464	1809308	101959	0.000		М	
3	13. 615	26721126	1184550	0.000		М	
4	17. 352	26727757	1008109	0.000		М	



Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	11. 245	136723	10191	0.000		М	
2	12.802	2111972	126052	0.000		M	
3	14. 554	2652252	126124	0.000		M	
4	18. 406	23358557	885340	0. 000		M	

**Supplementary Fig. 27 a** HPLC trace of racemic **3b**. **b** HPLC trace of product **3b** from the Diels-Alder reaction catalyzed by Cu<sup>2+</sup>. ATP for the *exo* isomer of 88% ee and the *endo* isomer of 80% ee. HPLC condition: Daicel Chiralpak OD column, hexane/*i*-PrOH 98:2, 0.5 mL min<sup>-1</sup>, 254 nm, 25 °C.



Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	30. 370	326071	7867	0.000		М	
2	33. 291	334338	7125	0.000		М	
3	38. 800	6685877	116095	0.000		М	
4	51. 761	6736538	103189	0.000		M	





<Peak table>

Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	30. 189	1834231	40402	0.000		М	
2	33. 035	199267	4213	0.000		М	
3	38. 389	21151294	354664	0.000		М	
4	51. 446	2745824	41898	0.000		М	

**Supplementary Fig. 28 a** HPLC trace of racemic **3c**. **b** HPLC trace of product **3c** from the Diels-Alder reaction catalyzed by Cu<sup>2+</sup>.ATP for the *exo* isomer of 80% ee and the *endo* isomer of 77% ee. HPLC condition: Daicel Chiralpak AD column, hexane/*i*-PrOH 98:2, 0.5 mL min<sup>-1</sup>, 254 nm, 25 °C.



Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	18. 618	1041998	44316	0.000		М	
2	21.097	983607	37988	0.000		M	
3	24. 432	18842320	468209	0.000		М	
4	29. 255	18676896	406218	0.000		М	



Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	18. 689	144639	5613	0.000		M	
2	21. 248	625606	21924	0.000		M	
3	24.970	4390140	118597	0.000		M	
4	20 660	12240227	204212	0.000		N.4	

**Supplementary Fig. 29 a** HPLC trace of racemic **3d**. **b** HPLC trace of product **3d** from the Diels-Alder reaction catalyzed by Cu<sup>2+</sup>.ATP for the *exo* isomer of 62% ee and the *endo* isomer of 50% ee. HPLC condition: Daicel Chiralpak ODH column, hexane/*i*-PrOH 98:2, 0.5 mL min<sup>-1</sup>, 254 nm, 25 °C.



b

Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	16.907	2717591	117947	0. 000		М	
2	19.914	3118196	121946	0. 000		М	
3	22. 185	16864839	561311	0. 000		М	
4	25.847	16970924	494182	0. 000		М	



<Peak table>

Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	17. 248	108089	4005	0.000		М	
2	19. 781	2974872	98890	0.000		М	
3	21. 992	20989034	625065	0.000		М	
4	25. 777	9242805	241024	0.000		М	

**Supplementary Fig. 30 a** HPLC trace of racemic **3e**. **b** HPLC trace of product **3e** from the Diels-Alder reaction catalyzed by Cu<sup>2+</sup>. ATP for the *exo* isomer of 93% ee and the *endo* isomer of 39% ee. HPLC condition: Daicel Chiralpak AD column, hexane/*i*-PrOH 98:2, 0.5 mL min<sup>-1</sup>, 254 nm, 25 °C.



Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	23.411	13518019	434013	0. 000		M	
2	27.479	52426923	1422579	0. 000		M	
3	30.379	13449763	322863	0. 000		M	
4	34.624	52636351	980594	0. 000		M	





<Peak table>

Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	22.378	13188348	430200	0.000		М	
2	26.071	69324282	1934707	0.000		М	
3	28.797	1778547	36686	0.000		М	
4	31.922	6221126	143456	0.000		М	

**Supplementary Fig. 31 a** HPLC trace of racemic **3f**. **b** HPLC trace of product **3f** from the Diels-Alder reaction catalyzed by Cu<sup>2+</sup>·ATP for the *exo* isomer of 76% ee and the *endo* isomer of 84% ee. HPLC condition: Daicel Chiralpak AD column, hexane/*i*-PrOH 90:10, 0.5 mL min<sup>-1</sup>, 254 nm, 25 °C.



Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	11.477	364209	29507	0.000		М	
2	12.874	360856	26032	0.000		М	
3	14.913	8462127	427237	0.000		М	
4	16.283	8474746	446715	0.000		М	



Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	12.004	237771	16378	0.000		М	
2	13. 807	837634	43784	0.000		Μ	
3	16. 022	5279941	218188	0.000		М	
4	17. 872	13678379	566640	0.000		М	

**Supplementary Fig. 32 a** HPLC trace of racemic **3g**. **b** HPLC trace of product **3g** from the Diels-Alder reaction catalyzed by Cu<sup>2+</sup>·ATP for the *exo* isomer of 56% ee and the *endo* isomer of 44% ee. HPLC condition: Daicel Chiralpak ODH column, hexane/*i*·PrOH 98:2, 0.5 mL min<sup>-1</sup>, 254 nm, 25 °C.



Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	12.461	258100	19771	0. 000		M	
2	14. 526	261632	17065	0. 000		M	
3	15. 884	4149071	222725	0. 000		M	
4	18.048	4169960	211255	0. 000		M	



1      12.720      82533      5867      0.000      M        2      15.048      753665      39701      0.000      M        3      16.503      1647025      78644      0.000      M        4      18.903      8191361      331242      0.000      M	Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
2      15.048      753665      39701      0.000      M        3      16.503      1647025      78644      0.000      M        4      18.903      8191361      331242      0.000      M	1	12. 720	82533	5867	0.000		M	
3      16.503      1647025      78644      0.000      M        4      18.903      8191361      331242      0.000      M	2	15. 048	753665	39701	0.000		M	
4 18 903 8191361 331242 0 000 M	3	16. 503	1647025	78644	0.000		M	
	4	18. 903	8191361	331242	0.000		M	

**Supplementary Fig. 33 a** HPLC trace of racemic **3h**. **b** HPLC trace of product **3h** from the Diels-Alder reaction catalyzed by Cu<sup>2+</sup>.ATP for the *exo* isomer of 80% ee and the *endo* isomer of 67% ee. HPLC condition: Daicel Chiralpak ODH column, hexane/*i*-PrOH 98:2, 0.5 mL min<sup>-1</sup>, 254 nm, 25 °C.

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