

Supplementary Information for

**VAV2 SIGNALING PROMOTES REGENERATIVE
PROLIFERATION IN BOTH CUTANEOUS AND HEAD AND NECK
SQUAMOUS CELL CARCINOMA**

by

Lorenzo-Martín *et al.*

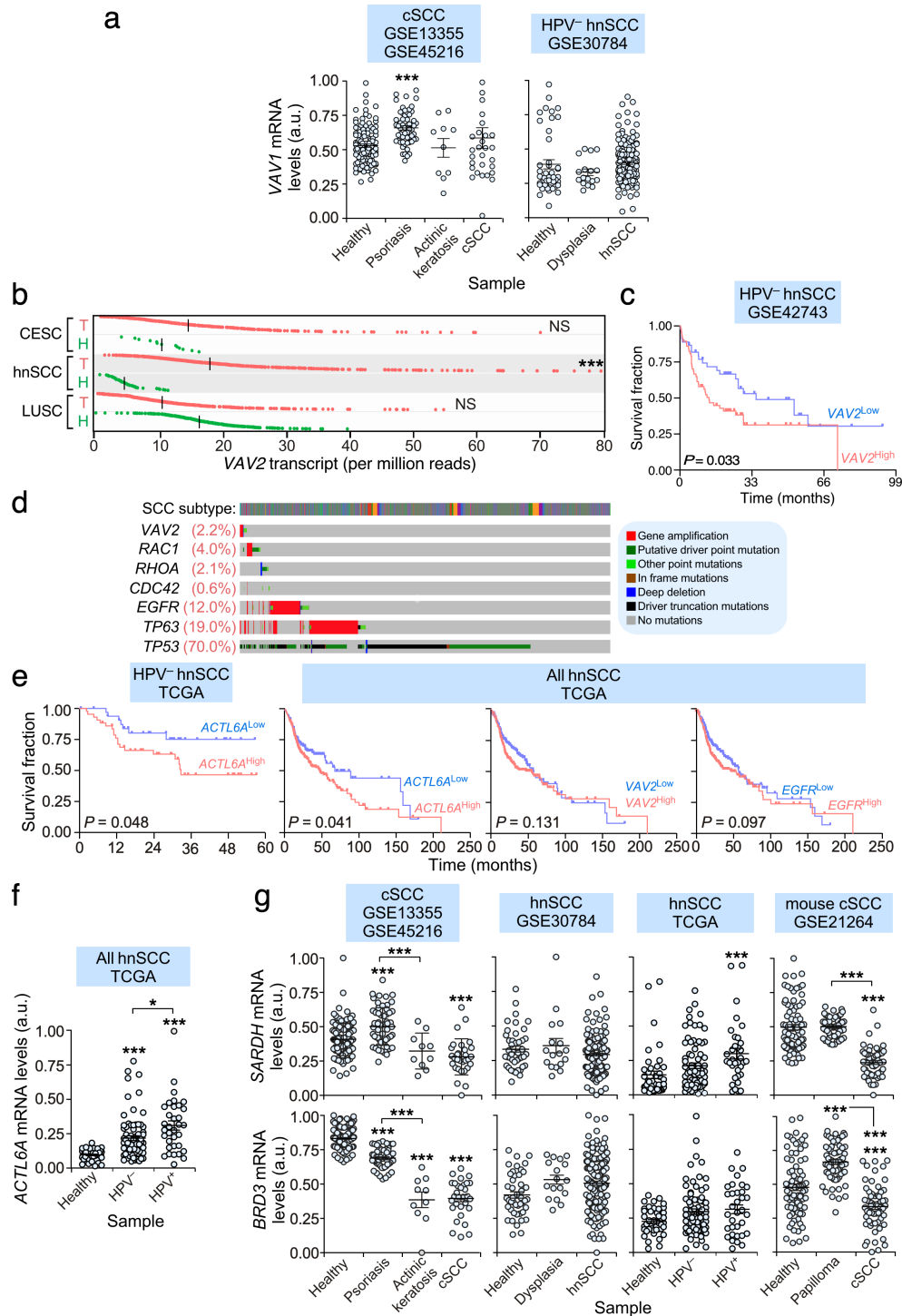
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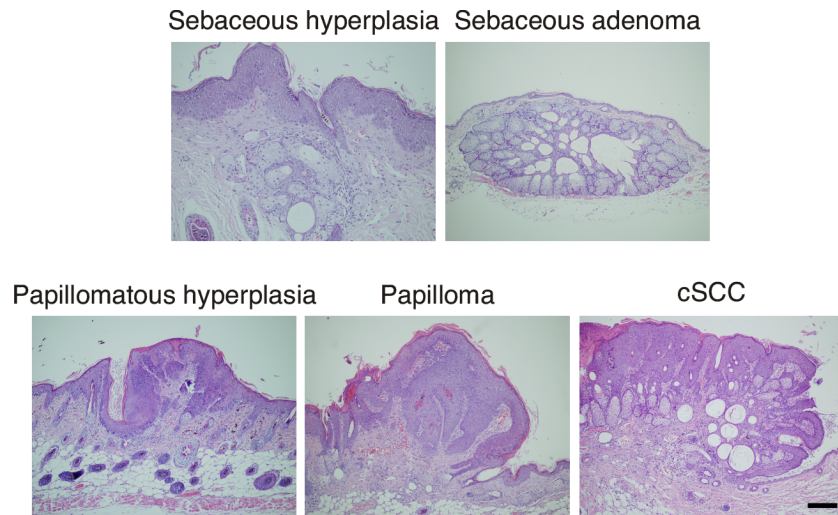


SUPPLEMENTARY FIGURE 1. Increased abundance of VAV2 is associated with poor prognosis in SCC

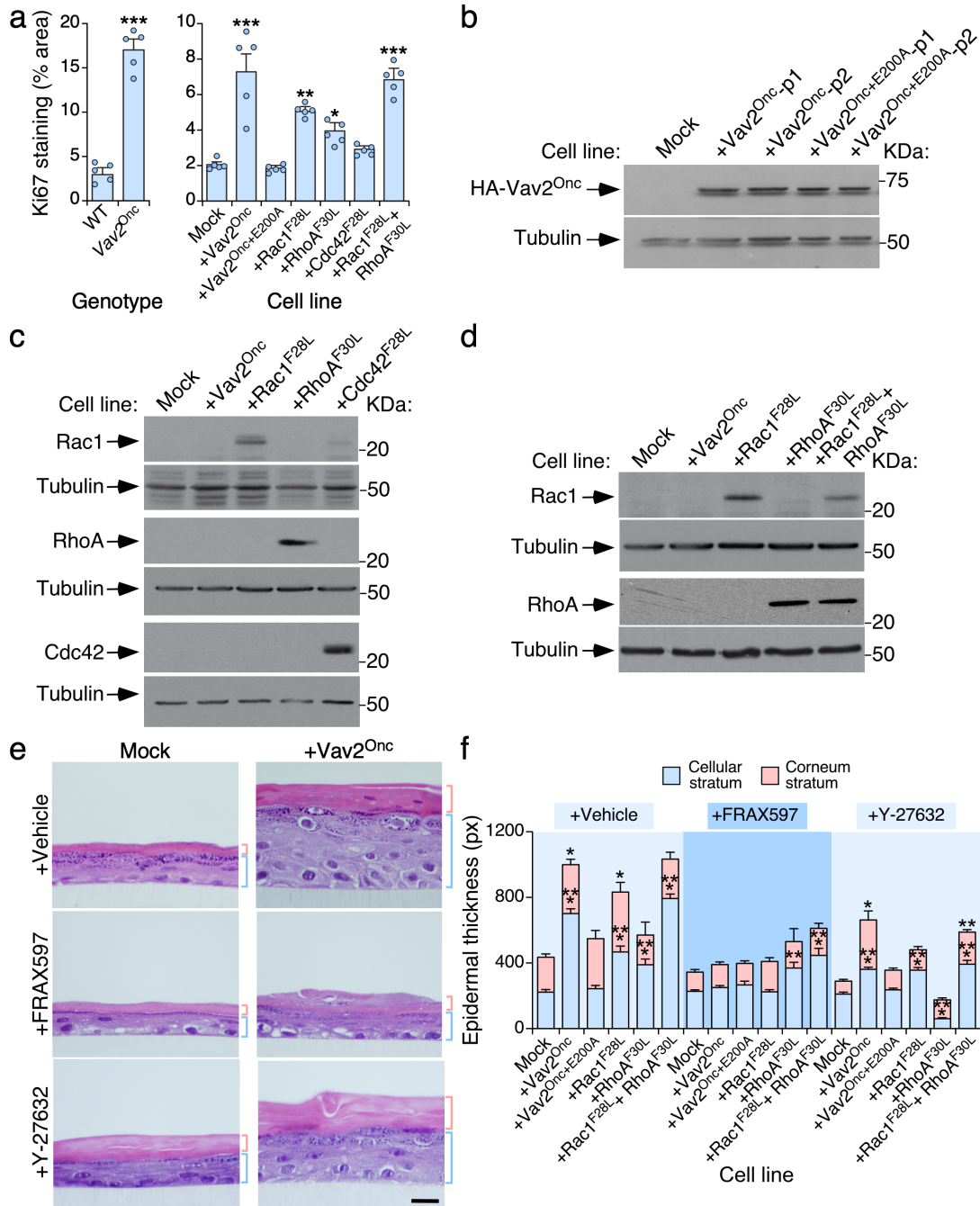
(a) Dot plot showing the normalized expression of the *VAV1* mRNA in indicated hnSCC subtype patients (bottom). The gene expression dataset used for these experiments is shown at the top. ***, $P < 0.0001$ (ANOVA and Tukey's HSD tests, $n = 220$ and 229 patient samples, respectively).

- (b) Dot plot showing the *VAV2* mRNA abundance in healthy ($n = 44$) and tumor samples ($n = 519$) from the indicated cervical (CESC), head and neck, and lung (LUSC) squamous cell carcinoma TCGA datasets (left). ***, $P < 0.0001$. NS, not statistically significant (Student's t -test).
- (c) Survival of the hnSCC patients from the indicated datasets according to the abundance of the *VAV2* mRNA present in tumors. The Mantel-Cox test P value is indicated.
- (d) Genetic alterations found in the indicated genes (left) in human cSCC and hnSCC patients present in the cBioPortal database. The percentage of samples with gene alterations (in %) are indicated on the left in red.
- (e) Survival of the hnSCC patients from indicated datasets according to the abundance of *ACTL6A*, *VAV2* and *EGFR* mRNAs in tumors. The Mantel-Cox test P value is indicated for each transcript.
- (f) Dot plot showing the normalized expression of the *ACTL6A* transcript in indicated hnSCC subtype patients (bottom) according to HPV status. The gene expression dataset used for these experiments is shown at the top *, $P = 0.010$; ***, $P < 0.0001$ (ANOVA and Tukey's HSD tests, $n = 155$ patient samples).
- (g) Dot plots showing the normalized expression of the *SARDH* and *BRD3* mRNAs in indicated tissue samples (bottom) and gene expression datasets (top). ***, $P < 0.0001$ (ANOVA and Tukey's HSD tests, $n = 220, 229, 155$ and 273 patient samples, respectively).

In a, f and g, data represent the mean \pm SEM. Source data for this figure are provided as a Source Data file.



SUPPLEMENTARY FIGURE 2. Examples of the type of tumors detected in WT and *Vav2^{Onc}* mice that have been used to generate the graphs shown in **Figure 3g**. $n = 87$ independent stainings. Scale bar, 200 μm .



SUPPLEMENTARY FIGURE 3 The effect of Vav2^{Onc} in epidermis is both keratinocyte autonomous and catalysis-dependent

(a) Quantification of the Ki67 immunoreactivity obtained in **Figures 4a** (left) and **4c** (right). *, $P = 0.012$; **, $P = 0.001$; ***, $P < 0.0001$ (ANOVA and Dunnett's multiple comparison test, $n = 5$ independent cultures).

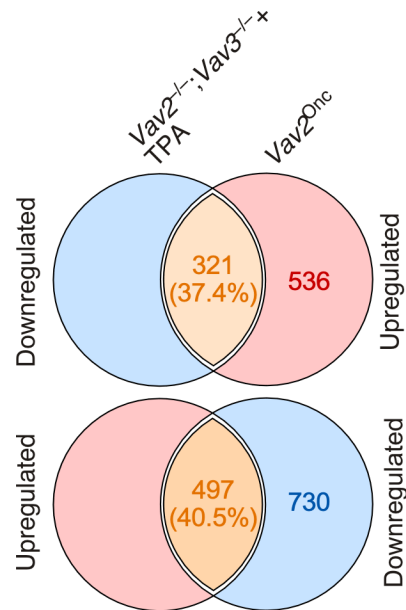
(b, c and d) Immunoblots showing the expression of the indicated proteins in human keratinocytes. Tubulin was used as loading control in all cases ($n = 3$ independent experiments). In B, p1 and p2 refer to two independent pools of cells.

(e) Representative images of hematoxylin-eosin-stained organotypic cultures generated by the indicated cells (top) and under the specified experimental conditions (left). The thickness of the corneum and cellular

epidermal strata is indicated with red and blue brackets, respectively. Scale bar, 10 μm ($n = 3$ independent cultures).

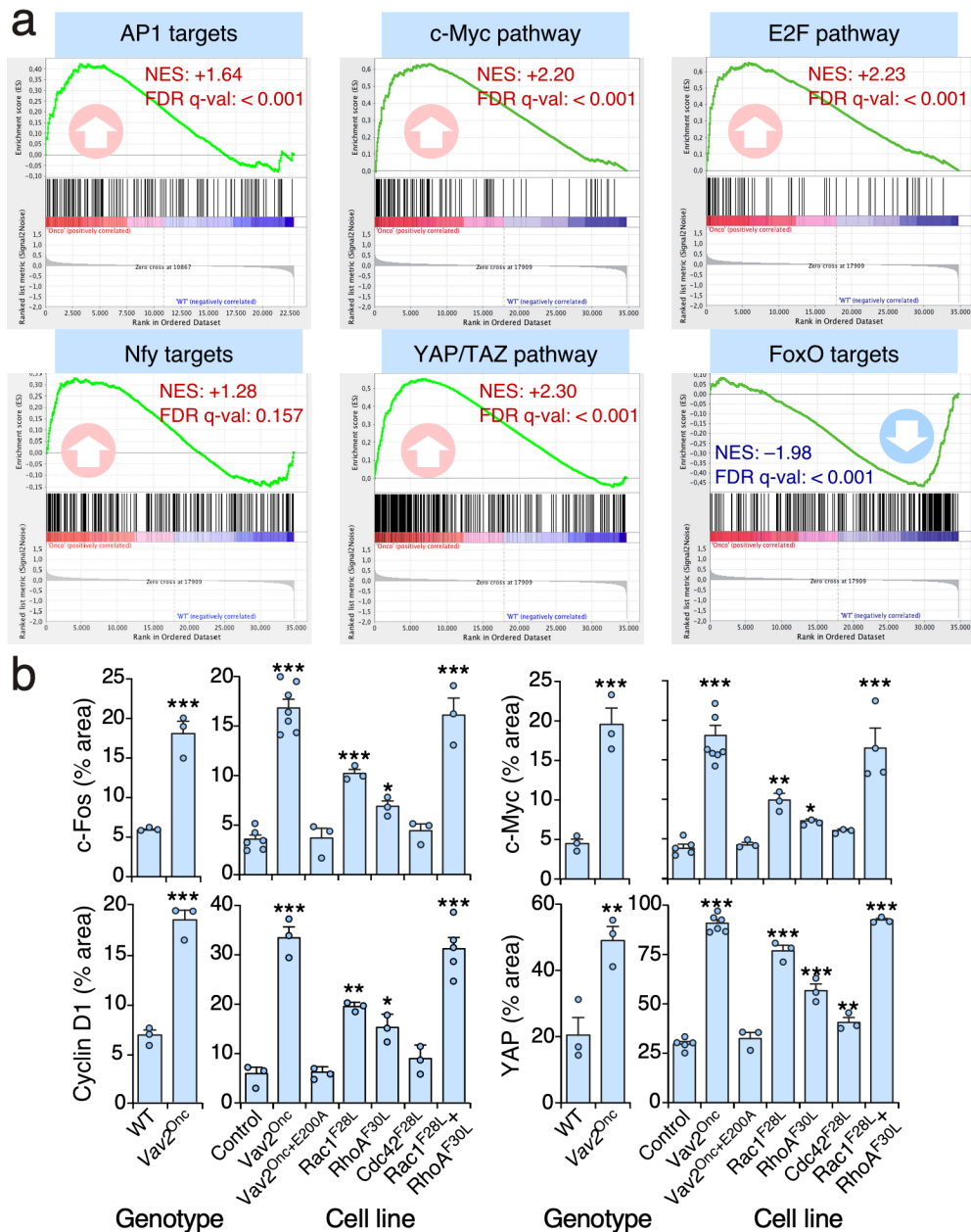
(f) Quantification of the thickness of the indicated skin strata according to the data obtained in panel e. px, pixel. *, $P = 0.041$ (Vav2^{Onc}, vehicle, corneum stratum), 0.025 (Rac1^{F28L}, vehicle, corneum stratum), 0.048 (Vav2^{Onc}, Y-27632, corneum stratum); **, $P = 0.002$ (RhoA^{F30L}, FRAX597, cellular stratum), 0.009 (Rac1^{F28L}+RhoA^{F30L}, FRAX597, corneum stratum); ***, $P < 0.0001$ (all other tests) (ANOVA and Dunnett's multiple comparison test, $n = 3$ independent cultures). In a and f, data are given as the mean \pm SEM.

Source data for this figure are provided as a Source Data file.



SUPPLEMENTARY FIGURE 4. Vav2 controls a protumorigenic stem cell-like program in keratinocytes

Venn diagram comparing the *Vav2^{Onc}*-regulated transcriptome and the *Vav2;Vav3*-dependent fraction of the TPA-stimulated gene expression program in the skin of mice. The number of probesets and level of overlap are indicated.

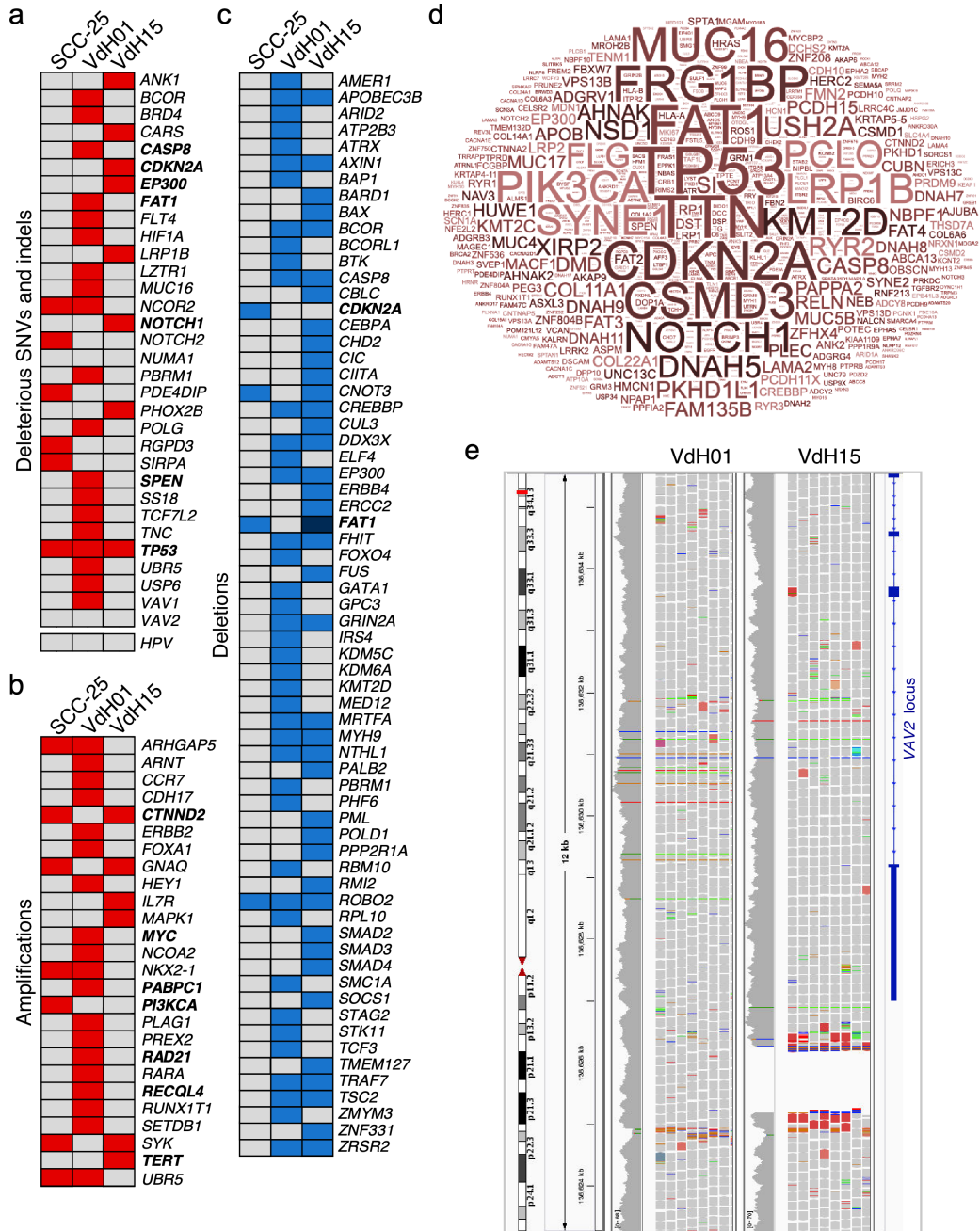


SUPPLEMENTARY FIGURE 5. The *Vav2^{Onc}*-dependent transcriptome is regulated by c-Myc, the YAP/WWTR1/TEAD complex, E2F, and AP1 factors

(a) GSEA of the indicated transcriptional factor-associated gene targets in the *Vav2^{Onc}*-dependent transcriptome. The NES (normalized enrichment score) and FDR (false discovery rate) values are indicated within each graph (GSEA statistical test). Positive and negative enrichments are indicated with upward- and downward-pointing arrows, respectively.

(b) Quantification of the immunoreactivity for the indicated proteins obtained in the experiments shown in Figure 6c,d. Data are given as the mean \pm SEM. *, $P = 0.017$ (RhoA^{F30L} and c-Fos), 0.011 (RhoA^{F30L} and c-Myc), 0.041 (RhoA^{F30L} and Cyclin D1); **, $P = 0.003$ (Rac1^{F28L} and c-Myc), 0.002 (Rac1^{F28L} and Cyclin D1), 0.009 (Cdc42^{F28L} and YAP); ***, $P < 0.0001$ (all other tests) (ANOVA and Dunnett's multiple comparison tests). $n = 3$ independent cultures.

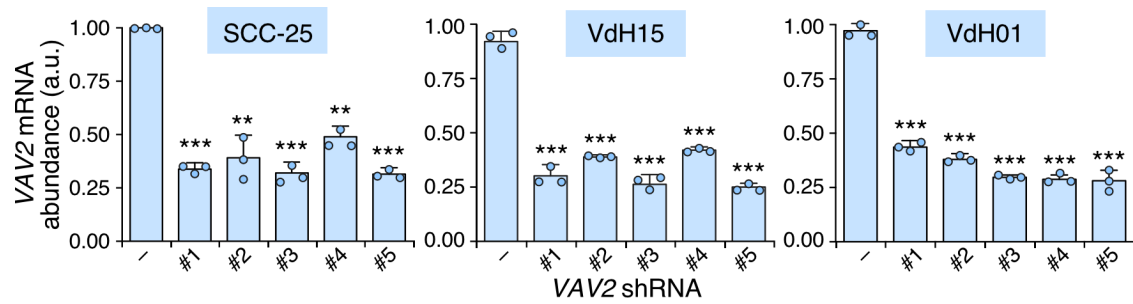
Source data for this figure are provided as a Source Data file.



SUPPLEMENTARY FIGURE 6. Characterization of the main genetic alterations found in VdH01 and VdH15 patient-derived cells.

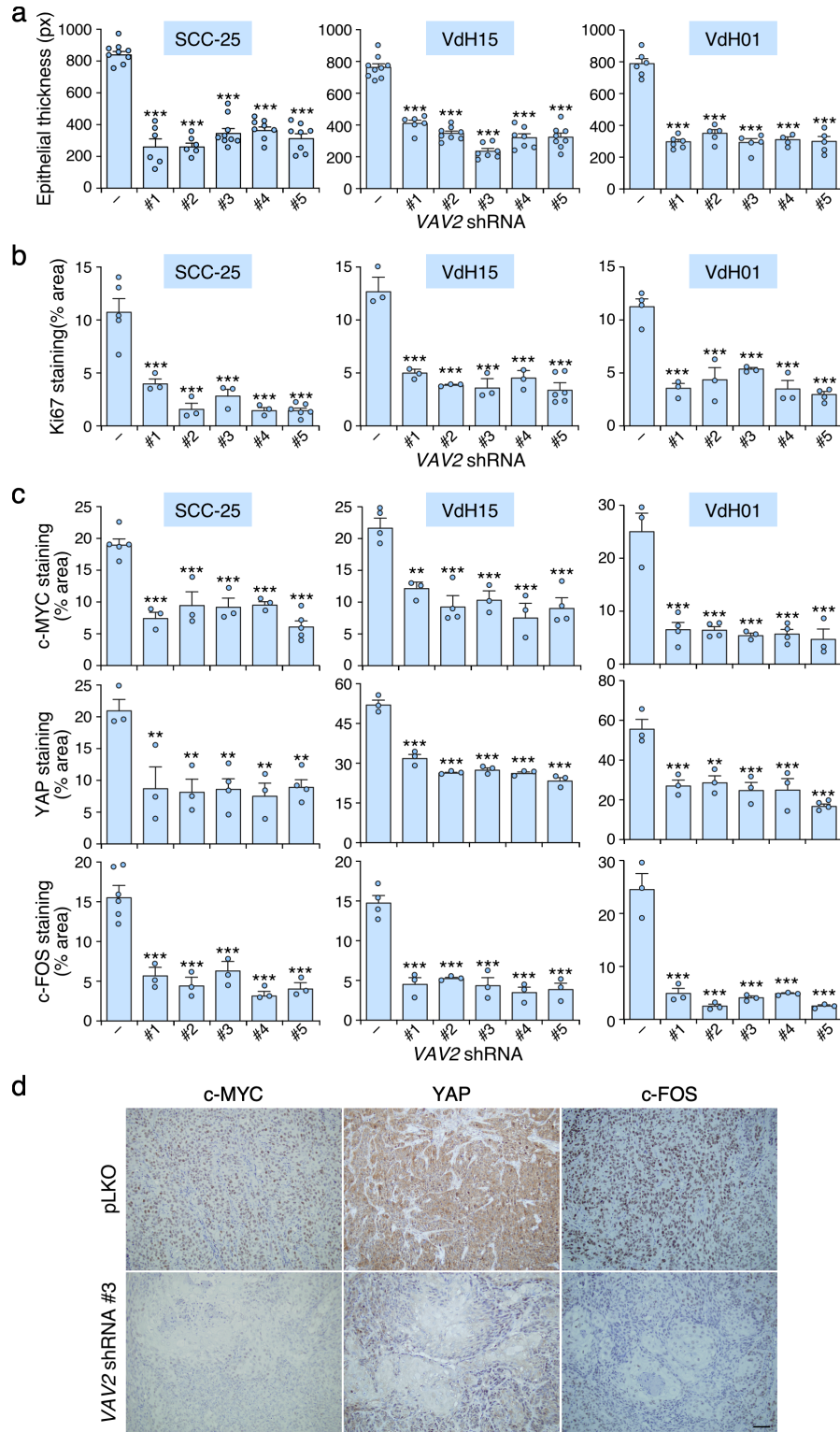
(a to c) Heatmaps showing the deleterious single nucleotide variants (SNVs) and indels (a, red boxes), amplifications (b, red boxes), and deletions (c, blue boxes) identified in the genome of the indicated cells (top). The data for the SCC-25 cell line have obtained from the Ref. #36 cited in the main text (Martín *et al.*, 2014). The genomic lesions that occur in at least 5% of hnSCC patients according to the TCGA dataset are highlighted in bold. In all cases, absence of mutations is depicted as a gray box.

- (d) Wordcloud representing the most frequently mutated genes in hnSCC patient samples according to the TCGA data. The size of each gene symbol is proportional to the mutation frequency of that gene in hnSCC.
- (e) Genomic view of the 3' genomic regions that is downstream of the *VAV2* gene in the indicated hnSCC patient-derived cell lines (top). From left to right, we show the chromosome map, the sequencing coverage, and the alignment of reads.



SUPPLEMENTARY FIGURE 7. VAV2 downmodulation in human SCC patient-derived cells. Level of *VAV2* mRNA knockdown achieved by the indicated *VAV2* shRNAs (bottom) in SCC-25, VdH15, and VdH01 cells. **, $P = 0.009$ (SCC-25 #2), 0.007 (SCC-25 #4); ***, $P < 0.0001$ (all other tests) (ANOVA and Dunnett's multiple comparison test, $n = 3$ independent experiments). Data are given as the mean \pm SEM.

Source data for this figure are provided as a Source Data file.



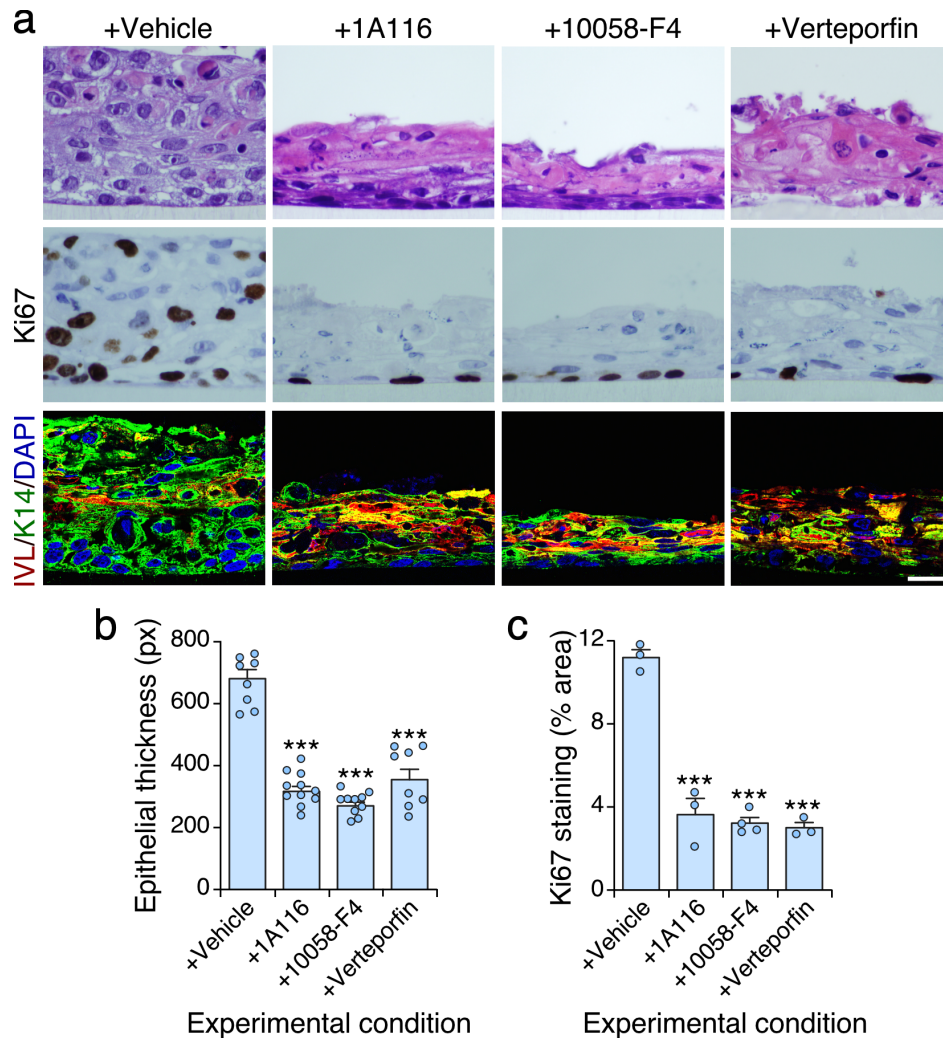
SUPPLEMENTARY FIGURE 8. VAV2 downmodulation abrogates the tumorigenic potential of hnSCC patient-derived cells

(a and b) Quantification of epithelial thickness (a) and Ki67 immunoreactivity (b) in the organotypic cultures displayed in **Figure 9a**. ***, $P < 0.0001$ (two-sided Student's t -test). $n = 5$ (a) and 3 (b) independent cultures. Data represent the mean \pm SEM.

(c) Quantification of the immunoreactivity for the indicated proteins (left) in the specified control and *VAV2* shRNA-knocked down cells (top). The *VAV2* shRNA used (#1 to #5) is indicated at the bottom. **, $P = 0.003$ (VdH15 #1, c-MYC), 0.009 (SCC-25 #1, YAP), 0.007 (SCC-25 #2, YAP), 0.001 (SCC-25 #3, YAP), 0.005 (SCC-25 #4, YAP), 0.001 (SCC-25 #5, YAP), 0.009 (VdH01 #2, YAP); ***, $P < 0.0001$ (all other tests) (ANOVA and Dunnett's multiple comparison tests, $n = 3$ independent cultures). In all panels, data are given as the mean \pm SEM.

(d) Lower magnification of the tumor sections shown in **Figure 9g**. Scale bar, 50 μm ($n = 8$).

Source data for this figure are provided as a Source Data file.



SUPPLEMENTARY FIGURE 9. The impact of the depletion of endogenous VAV2 in oral SCC PDCs is mimicked by the chemical inhibition of endogenous RAC1, c-MYC and YAP/TEAD complexes

(a) Representative images of organotypic cultures of parental VdH15 cells subjected to the indicated experimental conditions (top) upon staining with either hematoxylin-eosin (top panels) or antibodies to Ki67 (middle row of panels), IVL (bottom panels, red color) or K14 (bottom panels, green color). Some sections were counterstained with either hematoxylin (middle row of panels) or DAPI (bottom panels, blue color). Scale bar, 10 μ m ($n = 3$ independent cultures).

(b and c) Quantification the epithelial thickness (b) and Ki67 immunoreactivity (c) obtained in the experiments shown in panel a. ***, $P < 0.0001$ (ANOVA and Dunnett's multiple comparison test, $n = 3$ independent cultures). In b and c, data are given as the mean \pm SEM.

Source data for this figure are provided as a Source Data file.

SUPPLEMENTARY TABLE 1. Transcriptomic datasets used in this study

ACCESSION NUMBER	REFERENCE	SPECIES	SITE	SAMPLES	HEALTHY TISSUE	HPV STATUS	SURVIVAL INFO
GSE21264	PMID: 21244661	Mouse	Skin	273	Yes	Not applicable	No
GSE13355 & GSE45216	PMID: 19169254 & PMID: 24335922	Human	Skin	220	Yes	Not reported	No
GSE30784	PMID: 18669583	Human	Head and neck	229	Yes	Not reported	No
GSE41613	PMID: 23319825	Human	Head and neck	97	No	Negative	Yes
GSE42743	PMID: 23319825	Human	Head and neck	103	Yes	Negative	Yes
TCGA HNSCC	PMID: 25631445	Human	Head and neck	563	Yes	Positive & Negative	Yes
TCGA LUSC	PMID: 22960745	Human	Lung	824	Yes	Not applicable	Yes
TCGA CESC	PMID: 29625050	Human	Cervix	319	Yes	Not applicable	Yes
GSE52651	PMID: 25805135	Human	Epidermis	21	NA	Not applicable	Not applicable
GSE26713	PMID: 21481790	Human	T-ALL	124	Yes	Not applicable	No
GSE30219	PMID: 23698379	Human	Lung	307	Yes	Not applicable	Yes

SUPPLEMENTARY TABLE 2. Genes belonging to the Vav2^{Onc} redundant and nonredundant transcriptomes relative to the Vav2+Vav3-dependent fraction of the TPA-stimulated gene program in the mouse skin

REDUNDANT (Up in Vav2^{Onc})
<p><i>Cenpn, NA, Gas5, Pdap1, Pitrm1, Cdc20, Fam162a, Kpna2, Hmgb3, Cct3, Tmem97, Hist1h4c, Itgb4, Hist1h2bh, Prmt1, Rpl38, Urah, Gnl3, Mcm3, Nop56, Hmgb2, Ftsj3, Ahsa1, Ddx21, Lbr, Mybl2, Dsg3, NA, Mthfd11, Hist1h2be, Cct5, Ppm1g, Hist1h4m, Snrpf, Nt5dc2, Hmgb1, Rps8, Slc7a5, Tk1, Cenpm, Eef1e1, Srm, Rrm2, Nup93, Snord16a, NA, Aurkb, Fbl, Noc2l, Alyref, Rcc1, 2610528A11Rik, Myo10, Stom, Paklip1, Trip13, Erh, Nek2, Hmgb2, Gm3893, Mcm10, Snrpg, Myc, Rpl38, Hmgb1, Srsf9, Gm2173, Gm3696, Hspe1, Snrpd1, Hmgb1, Ccnel, Hmgb1, Nudc, Aurka, Hmgb1, Pla2g4c, Cdca5, Hnrnpa3, 4933409K07Rik, Ppih, Tmem97, Cks2, Rps12, Rplp1, Ndc80, Mki67ip, Myl4, Hist1h2ao, Rpl24, Cdk1, Snora62, Ranbp1, Cdca8, Wdr12, Ldlr, Mlana, Mybbp1a, Hmgb1, Hist1h2br, Myh9, Hist1h4h, Enah, Kif22, Col18a1, Rad51ap1, Sprr1b, Hyal1, Ldlrap1, Cad, Cks1b, Birc5, 9430008C03Rik, Rnu12, Snora74a, H2afx, Nup43, Orcl, Rcl1, Kpna2, Itgb1, Tars, Incenp, NA, Hmgb2, Exosc2, Duox1, Impdh2, Heatr1, Serpinb6c, Gas5, Ass1, Ube2s, Mcoln3, Hist1h2ab, Rpl12, Wfdc12, Hmgn2, Hist1h4b, 9430008C03Rik, Hmgb1, Hmgb1, Ppan, Nop16, Bnc1, Grwd1, Has3, Bcl2l15, Nrp2, Larp4, Snora33, Itga3, Nolc1, 4933409K07Rik, Pmel, Mtl, Scd2, Lrp8, Itpa, Rnu3b1, Myh9, Hirip3, Slc24a5, Ppil1, Slc45a2, Lmnb1, Gm3002, Ccnd1, Snora61, Polal, Gm5416, Cth, Slc20a1, Snora65, Ube2s, Ctps, Fhl2, Col12a1, Timm8a1, Adamts1, E2f7, Ptpz1, S100a6, Serpina9, Hnrnpa3, Pil5, Npm3, Timm8a1, Vaultrc5, Fscn1, Idi1, Erh, Lcn2, Dusp6, Mthfd2, Odc1, Hist2h2ab, Mrto4, Hmgb1, Snord53, Tnfrsf12a, Ccdc86, Junb, Tslp, NA, Fen1, Gjb6, Etv4, Krt35, Bcl3, Tubb6, Gsta4, Beat1, Gjb2, S100a8, S100a9, Odc1, Lipg, Gsr, Hbegf, Hp, Akr1b8, Il33, Ahcy, Dct, Cxcr2, Snord15a, Tyr, Ngf, Rps27a, Ccnblip1, Snord49a, Krt85, Itga5, Socs3, Mrto4, Smim3, Procr, Snora31, Tnc, Slc7a11, Rnu73b, Wif1, Fosl1, Gm5483, Rnu3a, 4933409K07Rik, 5730408K05Rik, Hist1h3g, Clcf1, Fbp1, Defb14, Phlda1, Lamc2, Krt28, Sh2d5, Tubb3, Tyrp1, Saa3, Cdhr1, Cntfr, Pdpn, Saa2, Ms4a4c, Lrat, Areg, 4933409K07Rik, Mmp9, Sirpb1b, Rnu1b1, Nrg1, Slc39a14, Slpi, Krt73, Inhba, Plek, Gm5150, Serpinel, Fam26d, Ifitm1, Timp1, Ptgs2, Trem3, Mmp13, Slc2a3, Ifitm1, Krt31, Epgn, Cyp4f18, Osm, Ifi202b, Krt33a, S100a3, NA, Krtap11-1, Mmp1b, Plaur, Hdc, Krt27, Krtap19-5, Krt71, Upp1, Ccl4, Krt25, Slfn4, Il1b, Prg4, Krtap19-1, Cd14, Spatc1, Nlrp3, Ccl3, Selp, Spp1, Trem1, Krtap15, Clec4d, Il24, Irg1, Cxcl2, Clec4e, Il6</i></p>
REDUNDANT (Down in Vav2^{Onc})
<p><i>Skint9, Skint10, H2-T22, Krt77, Ankrd35, Inmt, Skint11, H2-Q6, Cyp2g1, Sema3d, Clca1, Serpina3b, Alox12e, Skint4, Il22ra2, Serpinb7, Bbox1, Trdn, Il20ra, Clca2, Agbl1, Ucp3, Abca5, Cacna1s, Xirp2, Tcrg-V1, Pkia, Irf4, Scd3, NA, Mettl21c, Mlfl, Mstn, Mmp27, Oas1f, Trgv2, Uprr, H2-Ab1, Myh2, Ampd1, Scn2a1, Myom1, Asb12, Ccr4, Skint7, Flg2, Wnt3, Slc2a12, Ecm2, Skint2, Sbk2, Calm5, Klhl31, Myo18b, Lipn, Ereg, Tmod4, Kcnj2, Myoz1, Filip1l, Scn2a1, 2300005B03Rik, Smyd1, Mrgprb2, Clca3, Chit1, Mybpcl, Lcelm, Fbxo32, Skint3, Cmb1, 9430007A20Rik, Ccdc3, Txlnb, Cyp2b19, Xkrx, Fsd2, Slc5a9, Pdk4, Krt78, Serpinb12, Tmem182, Aloxe3, H19, Casq1, Il20rb, Slurp1, Dab1, Sgcd, Lcel1a2, Zfp943, Ttn, Trim7, Lce6a, Myot, H2-Eb1, Serpina12, Ppp2r3a, Trdn, Klhl38, H2-M2, Clca5, Tmem159, Otop3, Slc15a2, Mill1, Lrrc39, Satb1, Ckmt2, Neb, Ikzf2, Gal3st4, P2ry4, Tgfb2, Cd55, Slc16a14, Adh6a, Exph5, Srl, Lcel1, 9530053A07Rik, Sypl2, Lceli, Osr2, Myom3, Ctnb2, Pcmt1d, Ear2, Lcel1f, Myh1, A930018M24Rik, Ogn, Slc24a3, Pdk2, Lor, Gdgd2, Lcel1d, Pygm, C4b, Sh3rf2, Klhl31, Atp1b1, Lrrc28, Il20rb, Lmod2, Hal, Fndc1, Lipk, Cpxm2, Them5, Adck3, Fmo2, Acvr1c, H2-Aa, Sox6, Rblcl1, B3galt2, Il12rb2, Zbtb16, Cilp, Pou2f3, Mfap4, Lipm, Cobl, Map2, Lcelg, Fam214a, Mettl7a1, Nr1d1, Plekhn1, Thrb, Lmod3, Gm7165, Krt15, Ccl27a, 2310014L17Rik, Slc28a3, Klhl24, Fndc9, Gm11992, Gpld1, Il1f8, Fitm1, Il20rb, Krt10, Aif1l, Rps6ka6, Mrgprb3, Smpd3, Nipal2, Fam189a2, Slitrk6, Il1f6, Gm7609, Tmem229b, Tstd2, Atp1a2, Tmprss11f, Calcb, Nt5c3, Cyp27a1, Ear5, Plb1, Kcnj16, Acot2, Myom2, C6, Cyp2f2, Rora, Gm5105, Nrap, Serpinb11, Ar, Gpr111, Sh3gl2, 2310007B03Rik, Cyp2r1,</i></p>

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NON-REDUNDANT (Up in Vav2^{Onc})

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NON-REDUNDANT (Down in Vav2^{Onc})

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Tmeff2, Rab11fip1, Scn7a, Klk8, Nlrc5, Gypc, Mrgprb1, Man2b1, Sprr1a, Greb1, Plk3, Dok2, Cd209c, Scara3, Ctgf, Ipkb, Camk2n1, Gm11564, H2-M3, Mmp11, Col15a1, Atp6v0d1, Fcgrt, Prkcb, Arrb1, Glrx, Il1a, Hmcn1, Pnp2, H2-Q4, Irgm2, Prex2, Tm6sf1, Mnf1, Pla2g2d, Ifi203, Chac1, Sgce, Gng4, Psmb9, Ctst, Prss23, Abcg1, Tcn2, Irak2, Aoah, Tbxas1, Dcn, Car6, Iigp1b, Csf1r, Ccl19, Ly6d, Serping1, Ndn, Plod2, Wisp2, C3, Plac9a, Fabp4, Reln, Cd209a, Cmkrl1, Hsd17b11, Calcrl, Nid1, Laptm5, Cmtm3, Cybb, Dkk2, Mxra8, Loxl1, Serpina3f, Colec12, Sprr4, Tgm2, Pi16, Fstl1, Cd163, Gbp6, Degs2, Abtb2, Dpysl3, Lox, Gm4951, Loxl2, Krtap5-1, Slc5a3, Spon1, Stard8, Tgtp1, Slc43a3, Ecm1, Itga4, 9930111J21Rik2, Gbp7, Pxdn, Sp100, Irgm1, Blvrb, Igfbp3, Defb1, Mcpt4, Ccl19, Gbp9, Mxd1, Cdh11, Cls, Klk10, Clqtnf1, Vsig4, Tgtp1, Igf1, Plb1, Il1r2, Opct, Cfhr2, Gm5431, Lpgat1, Psmb10, Ddx60, Gm10639, Cd37, Rhoj, Krtap5-4, AI607873, AW551984, Ntrk2, Gfpt2, Gatm, Cfh, Lyz2, Rac2, Ly6g6e, Alox5, Clqa, Svep1, Rassf2, F13a1, Fam26e, Dock2, Dab2, Ndr1, Cd48, Pgm1, Plac9a, Ptplad2, Gbgt1, Lgals3bp, Tlr7, Xist, Krtap1-3, Mrc1, Pld4, Clqc, Tshr, Sfrp2, Anxa3, Gbp2, Krtap4-13, Ms4a8a, Efemp1, Clrb, Gvin1, Emr1, Ggt5, Gpc3, Emp3, Ube2l6, Cyp1b1, Folr2, Slc16a12, Unc93b1, Clqb, Siglec1, Tpsb2, Pla1a, Lbp, Sdc3, Mest, Ednrb, Nrpl, Sfrp4, Mfap5, Osr1, Fcgr2b, Cd68, Selplg, Clra, Krtap9-3, Iigp1, Cfp, Trim30a, Gabra3, Ly86, Lyve1, Krtap4-7, Fcer1g, Cd209e, Ccl21a, Gabrp, Oasl2, Ifi205, Mmrn1, Pf4, Fcgr3, AW112010, Ccl8, Krtap4-2, Sod3, Gsta1, P2ry13, Pinlyp, Krtap2-4, Krtap9-1, Chrll1, Cd209d, Ifit3, Gm12250, Clec10a, Lcp1, Snord118, Nnmt, Ccl11, Krtap4-13, Gm11563, Krtap4-6, Cd38, Krtap2-4, Cp, Alox5ap, Angptl4, Wfdc17, Lep, Plac8, Ccl6, Krtap4-1, Krtap1-5, Gm11567, Ccl9, Gm10228, Mup7, Krtap21-1, Krtap4-16, Ifitm6, Krtap6-2, Krtap6-5, Krtap6-1, Cxcl13, Prr9, Krtap7-1

SUPPLEMENTARY TABLE 3. List of genes belonging to the Vav2^{Onc}-Sig.

GENE SYMBOL	PATTERN IN Vav2 ^{Onc}	PATTERN IN Vav2 ^{-/-} ;Vav3 ^{-/-} + TPA	PATTERN IN cSCC	PATTERN IN ORAL SCC
<i>AREG</i>	up	down	up	up
<i>AURKA</i>	up	down	up	up
<i>AURKB</i>	up	down	up	up
<i>BCAT1</i>	up	down	up	up
<i>BIRC5</i>	up	down	up	up
<i>BNC1</i>	up	down	up	up
<i>CCL4</i>	up	down	up	up
<i>CCT5</i>	up	down	up	up
<i>CDC20</i>	up	down	up	up
<i>CDCA5</i>	up	down	up	up
<i>CDCA8</i>	up	down	up	up
<i>CDK1</i>	up	down	up	up
<i>CENPN</i>	up	down	up	up
<i>CKS1B</i>	up	down	up	up
<i>CKS2</i>	up	down	up	up
<i>CXCL2</i>	up	down	up	up
<i>E2F7</i>	up	down	up	up
<i>ENAH</i>	up	down	up	up
<i>FEN1</i>	up	down	up	up
<i>FOSL1</i>	up	down	up	up
<i>FSCN1</i>	up	down	up	up
<i>HAS3</i>	up	down	up	up
<i>HMGB3</i>	up	down	up	up
<i>IFITM1</i>	up	down	up	up
<i>IL1B</i>	up	down	up	up
<i>IL24</i>	up	down	up	up
<i>INHBA</i>	up	down	up	up
<i>ITGB1</i>	up	down	up	up
<i>KPNA2</i>	up	down	up	up
<i>LAMC2</i>	up	down	up	up
<i>LIPG</i>	up	down	up	up
<i>LRP8</i>	up	down	up	up
<i>MCM10</i>	up	down	up	up
<i>MMP13</i>	up	down	up	up
<i>MMP9</i>	up	down	up	up
<i>MTHFD2</i>	up	down	up	up
<i>MYC</i>	up	down	up	up
<i>NDC80</i>	up	down	up	up
<i>NEK2</i>	up	down	up	up
<i>NOLC1</i>	up	down	up	up
<i>NRG1</i>	up	down	up	up
<i>NRP2</i>	up	down	up	up
<i>ODC1</i>	up	down	up	up
<i>PDPN</i>	up	down	up	up
<i>PII5</i>	up	down	up	up
<i>PLAUR</i>	up	down	up	up
<i>PLEK</i>	up	down	up	up
<i>PTGS2</i>	up	down	up	up
<i>RAD51API</i>	up	down	up	up
<i>RANBP1</i>	up	down	up	up

GENE SYMBOL	PATTERN IN <i>Vav2</i> ^{0nc}	PATTERN IN <i>Vav2</i> ^{-/-} ; <i>Vav3</i> ^{-/-} + TPA	PATTERN IN cSCC	PATTERN IN ORAL SCC
<i>RRM2</i>	up	down	up	up
<i>SERPINE1</i>	up	down	up	up
<i>SH2D5</i>	up	down	up	up
<i>SLC20A1</i>	up	down	up	up
<i>SLC2A3</i>	up	down	up	up
<i>SLC7A11</i>	up	down	up	up
<i>SLC7A5</i>	up	down	up	up
<i>SOCS3</i>	up	down	up	up
<i>SPP1</i>	up	down	up	up
<i>SRM</i>	up	down	up	up
<i>STOM</i>	up	down	up	up
<i>TK1</i>	up	down	up	up
<i>TNC</i>	up	down	up	up
<i>TREM1</i>	up	down	up	up
<i>TRIP13</i>	up	down	up	up
<i>UBE2S</i>	up	down	up	up
<i>ABCA5</i>	down	up	down	down
<i>ABCA6</i>	down	up	down	down
<i>ABCA9</i>	down	up	down	down
<i>ACPP</i>	down	up	down	down
<i>ADSSL1</i>	down	up	down	down
<i>AFF3</i>	down	up	down	down
<i>ALDH1A1</i>	down	up	down	down
<i>ANKRD35</i>	down	up	down	down
<i>ANXA9</i>	down	up	down	down
<i>AR</i>	down	up	down	down
<i>ARHGAP20</i>	down	up	down	down
<i>ATP12A</i>	down	up	down	down
<i>ATP1A2</i>	down	up	down	down
<i>ATP1B1</i>	down	up	down	down
<i>BBOX1</i>	down	up	down	down
<i>CAMK1D</i>	down	up	down	down
<i>CARD14</i>	down	up	down	down
<i>CDO1</i>	down	up	down	down
<i>CDON</i>	down	up	down	down
<i>CGN</i>	down	up	down	down
<i>CILP</i>	down	up	down	down
<i>COBL</i>	down	up	down	down
<i>CUL3</i>	down	up	down	down
<i>DLG2</i>	down	up	down	down
<i>EXPH5</i>	down	up	down	down
<i>FAM189A2</i>	down	up	down	down
<i>FAM3B</i>	down	up	down	down
<i>FMO2</i>	down	up	down	down
<i>FXYD1</i>	down	up	down	down
<i>FYCO1</i>	down	up	down	down
<i>GAN</i>	down	up	down	down
<i>GRAMD1C</i>	down	up	down	down
<i>HMGCS2</i>	down	up	down	down
<i>HSPB6</i>	down	up	down	down
<i>IL18</i>	down	up	down	down
<i>IL20RA</i>	down	up	down	down

GENE SYMBOL	PATTERN IN <i>Vav2</i>^{0nc}	PATTERN IN <i>Vav2</i>^{-/-};<i>Vav3</i>^{-/-} + TPA	PATTERN IN cSCC	PATTERN IN ORAL SCC
<i>INMT</i>	down	up	down	down
<i>ITM2A</i>	down	up	down	down
<i>KRT15</i>	down	up	down	down
<i>LDB3</i>	down	up	down	down
<i>LNXI</i>	down	up	down	down
<i>LRRN4CL</i>	down	up	down	down
<i>MAOB</i>	down	up	down	down
<i>MBOAT2</i>	down	up	down	down
<i>MFAP4</i>	down	up	down	down
<i>MYOZ1</i>	down	up	down	down
<i>NEBL</i>	down	up	down	down
<i>OGN</i>	down	up	down	down
<i>PCOLCE2</i>	down	up	down	down
<i>PKIA</i>	down	up	down	down
<i>PKIB</i>	down	up	down	down
<i>POF1B</i>	down	up	down	down
<i>PYGM</i>	down	up	down	down
<i>RECK</i>	down	up	down	down
<i>RORA</i>	down	up	down	down
<i>SCARA5</i>	down	up	down	down
<i>SCEL</i>	down	up	down	down
<i>SH3RF2</i>	down	up	down	down
<i>SLC16A14</i>	down	up	down	down
<i>SLC24A3</i>	down	up	down	down
<i>SLC2A12</i>	down	up	down	down
<i>SLC6A4</i>	down	up	down	down
<i>SLURP1</i>	down	up	down	down
<i>SOX6</i>	down	up	down	down
<i>SSBP2</i>	down	up	down	down
<i>TCEA3</i>	down	up	down	down
<i>TJP3</i>	down	up	down	down
<i>TPPP</i>	down	up	down	down
<i>TPPP3</i>	down	up	down	down
<i>TRIM7</i>	down	up	down	down

SUPPLEMENTARY TABLE 4. List of genes belonging to the minimal *Vav2*^{Onc} diagnostic gene signature

GENE SYMBOL	PATTERN IN <i>Vav2</i> ^{Onc}	PATTERN IN <i>Vav2</i> ^{-/-} ; <i>Vav3</i> ^{-/-} + TPA	PATTERN IN cSCC	PATTERN IN ORAL SCC
<i>AREG</i>	up	down	up	up
<i>AURKA</i>	up	down	up	up
<i>BCAT1</i>	up	down	up	up
<i>BIRC5</i>	up	down	up	up
<i>CCL4</i>	up	down	up	up
<i>CDC45</i>	up	down	up	up
<i>CDC48</i>	up	down	up	up
<i>CKS1B</i>	up	down	up	up
<i>CXCL2</i>	up	down	up	up
<i>FSCN1</i>	up	down	up	up
<i>HAS3</i>	up	down	up	up
<i>IL24</i>	up	down	up	up
<i>LAMC2</i>	up	down	up	up
<i>MMP9</i>	up	down	up	up
<i>MYC</i>	up	down	up	up
<i>NEK2</i>	up	down	up	up
<i>NOLC1</i>	up	down	up	up
<i>ODC1</i>	up	down	up	up
<i>SERPINE1</i>	up	down	up	up
<i>SLC20A1</i>	up	down	up	up
<i>SLC2A3</i>	up	down	up	up
<i>SLC7A11</i>	up	down	up	up
<i>SLC7A5</i>	up	down	up	up
<i>SOCS3</i>	up	down	up	up
<i>SRM</i>	up	down	up	up
<i>ABCA9</i>	down	up	down	down
<i>ACPP</i>	down	up	down	down
<i>ANXA9</i>	down	up	down	down
<i>ARHGAP20</i>	down	up	down	down
<i>ATP1A2</i>	down	up	down	down
<i>CAMK1D</i>	down	up	down	down
<i>CILP</i>	down	up	down	down
<i>EXPH5</i>	down	up	down	down
<i>FYCO1</i>	down	up	down	down
<i>HSPB6</i>	down	up	down	down
<i>PKIB</i>	down	up	down	down
<i>PYGM</i>	down	up	down	down
<i>SCEL</i>	down	up	down	down
<i>SLC6A4</i>	down	up	down	down
<i>SSBP2</i>	down	up	down	down
<i>TCEA3</i>	down	up	down	down

SUPPLEMENTARY TABLE 5. List of patient samples used in this study

SAMPLE ID	SAMPLE TYPE	SOURCE
B87	Healthy	University Central Hospital of Asturias
B100	Healthy	University Central Hospital of Asturias
B103	Healthy	University Central Hospital of Asturias
B107	Healthy	University Central Hospital of Asturias
B116	Healthy	University Central Hospital of Asturias
B135	Healthy	University Central Hospital of Asturias
B140	Healthy	University Central Hospital of Asturias
B141	Healthy	University Central Hospital of Asturias
B143	Healthy	University Central Hospital of Asturias
B147	Healthy	University Central Hospital of Asturias
B158	Healthy	University Central Hospital of Asturias
B161	Healthy	University Central Hospital of Asturias
B169	Healthy	University Central Hospital of Asturias
B172	Healthy	University Central Hospital of Asturias
B174	Healthy	University Central Hospital of Asturias
B175	Healthy	University Central Hospital of Asturias
B183	Healthy	University Central Hospital of Asturias
T87	hnSCC	University Central Hospital of Asturias
T91	hnSCC	University Central Hospital of Asturias
T92	hnSCC	University Central Hospital of Asturias
T97	hnSCC	University Central Hospital of Asturias
T98	hnSCC	University Central Hospital of Asturias
TL100	hnSCC	University Central Hospital of Asturias
TP100	hnSCC	University Central Hospital of Asturias
T103	hnSCC	University Central Hospital of Asturias
T104	hnSCC	University Central Hospital of Asturias
T106	hnSCC	University Central Hospital of Asturias
T107	hnSCC	University Central Hospital of Asturias
T108	hnSCC	University Central Hospital of Asturias
T109	hnSCC	University Central Hospital of Asturias
T112	hnSCC	University Central Hospital of Asturias
T116	hnSCC	University Central Hospital of Asturias
T119	hnSCC	University Central Hospital of Asturias
T129	hnSCC	University Central Hospital of Asturias
T133	hnSCC	University Central Hospital of Asturias
T135	hnSCC	University Central Hospital of Asturias
T140	hnSCC	University Central Hospital of Asturias
T141	hnSCC	University Central Hospital of Asturias
T143	hnSCC	University Central Hospital of Asturias
T147	hnSCC	University Central Hospital of Asturias
T151	hnSCC	University Central Hospital of Asturias
T155	hnSCC	University Central Hospital of Asturias
T158	hnSCC	University Central Hospital of Asturias
T161	hnSCC	University Central Hospital of Asturias
T168	hnSCC	University Central Hospital of Asturias

SAMPLE ID	SAMPLE TYPE	SOURCE
T169	hnSCC	University Central Hospital of Asturias
T172	hnSCC	University Central Hospital of Asturias
T174	hnSCC	University Central Hospital of Asturias
T175	hnSCC	University Central Hospital of Asturias
T183	hnSCC	University Central Hospital of Asturias
S12	Healthy	Sant Pau i Santa Creu Hospital
S139	Healthy	Sant Pau i Santa Creu Hospital
S463	Healthy	Sant Pau i Santa Creu Hospital
S524	Healthy	Sant Pau i Santa Creu Hospital
S542	Healthy	Sant Pau i Santa Creu Hospital
S548	Healthy	Sant Pau i Santa Creu Hospital
S564	Healthy	Sant Pau i Santa Creu Hospital
S568	Healthy	Sant Pau i Santa Creu Hospital
S743	Healthy	Sant Pau i Santa Creu Hospital
S1039	Healthy	Sant Pau i Santa Creu Hospital
T11	hnSCC	Sant Pau i Santa Creu Hospital
T138	hnSCC	Sant Pau i Santa Creu Hospital
T289	hnSCC	Sant Pau i Santa Creu Hospital
T306	hnSCC	Sant Pau i Santa Creu Hospital
T334	hnSCC	Sant Pau i Santa Creu Hospital
T462	hnSCC	Sant Pau i Santa Creu Hospital
T523	hnSCC	Sant Pau i Santa Creu Hospital
T541	hnSCC	Sant Pau i Santa Creu Hospital
T547	hnSCC	Sant Pau i Santa Creu Hospital
T563	hnSCC	Sant Pau i Santa Creu Hospital
T567	hnSCC	Sant Pau i Santa Creu Hospital
T742	hnSCC	Sant Pau i Santa Creu Hospital
T1038	hnSCC	Sant Pau i Santa Creu Hospital
T27598	hnSCC	University Hospital of Salamanca
T27629	hnSCC	University Hospital of Salamanca
T27767	hnSCC	University Hospital of Salamanca
T27824	hnSCC	University Hospital of Salamanca
T28160	hnSCC	University Hospital of Salamanca
T28252	hnSCC	University Hospital of Salamanca
T28288	hnSCC	University Hospital of Salamanca
T29277	hnSCC	University Hospital of Salamanca
T29331	hnSCC	University Hospital of Salamanca
T29894	hnSCC	University Hospital of Salamanca

SUPPLEMENTARY NOTE 1. Genomic characterization of the hnSCC patient-derived cells used in this study

Using next generation sequencing (see Methods), we found that the VdH01 and VdH015 patient-derived cells are clearly different according to the catalogue of both deleterious single nucleotide (**Supplementary Fig. 6a**) and copy number (**Supplementary Fig. 6b,c**) variations present in them. These disparities are expected, given that each of those cells has been obtained from an independent patient. The two patient-derived cells share, however, alterations that target common loci (*APOBEC3B*, *AXIN1*, *BCOR*, *CASP8*, *CREBBP*, *DDX3X*, *EP300*, *FAT1*, *FHIT*, *GRIN2A*, *MRTFA*, *MYH9*, *NTHL1*, *ROBO2*, *TP53*, *TRAF7*, *TSC2*, *ZRSR29*) (**Supplementary Fig. 6a-c**). Mutations in *FAT1*, *TP53*, and *ROBO2* are also present in the SCC-25 cancer cell line (**Supplementary Fig. 6a-c**). The two patient-derived cells are also HPV⁻ (**Fig. 6a**) and show a gain in the long arm of chromosome 9, a frequent event in hnSCC from oral regions^{1,2}. Many of the genetic alterations found in each of those patient-derived cells have been previously detected at high frequency rates in hnSCC cases (*CASP8*, *CDKN2A*, *EP300*, *FAT1*, *NOTCH1*, *MYC*, *PABPC1*, *RAD21*, *RECQL4*, *TERT*, *SPEN*, *TP53*) (**Supplementary Fig. 6a-d**). In the case of *VAV2*, we did not find any activating single point mutation in any of the two patient-derived cells (**Supplementary Fig. 6a**). The VdH15 cells do bear a small deletion of 1,068 bp in a region downstream of the *VAV2* locus that harbors a putative binding site for the transcriptional factor ZNF263 (**Supplementary Fig. 6e**; coordinates chr9:136625194-136626262), although the functional relevance of this alteration for either *VAV2* expression or hnSCC etiology is unknown. Interestingly, we found a point mutation in *VAV1* (Q122R) in VdH01 patient-derived cells. An analogous mutation (Q122H) has been found before in a cervical SCC case in TCGA studies. These mutations, however, are unlikely to create a catalytic gain-of-function event on the protein according to previous functional and structural studies^{3,4}. The VdH01 cells also show amplification events affecting *PREX2* and *ARHGAP5*, two loci encoding a RAC1 GEF and RHOA GTPase activating protein, respectively. Although paradoxical from a functional point of view, the amplification of the latter gene has been observed before in a number of tumors⁵.

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