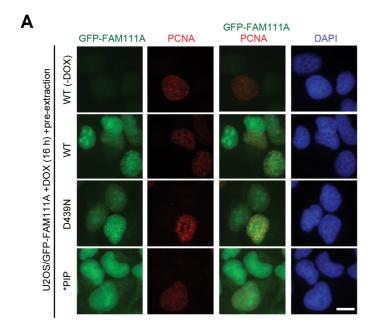
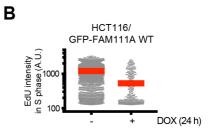
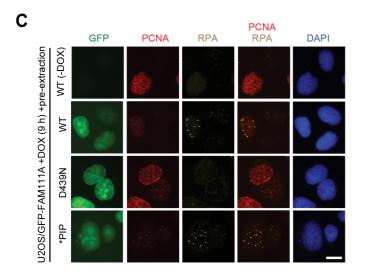
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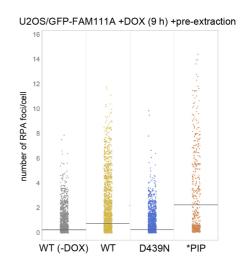
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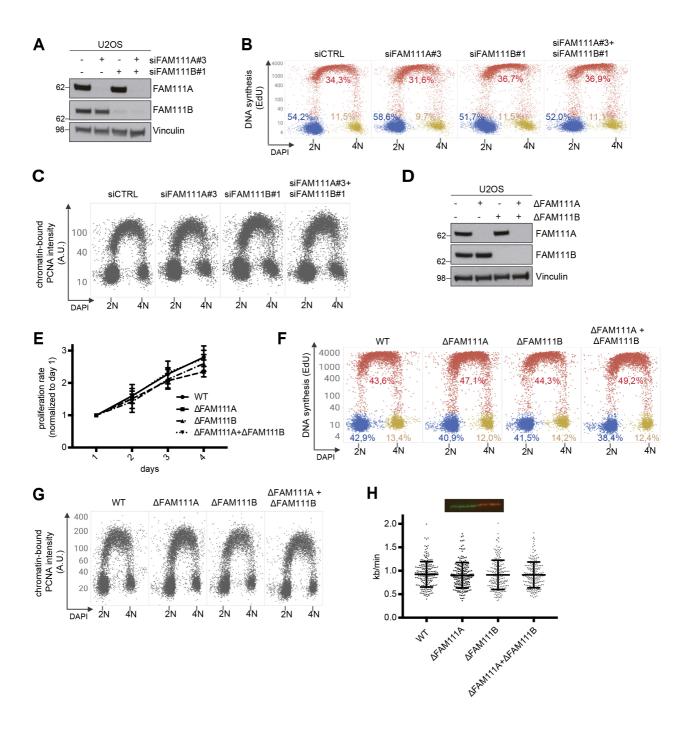


Appendix Figure S1.

Elevated FAM111A proteolytic activity impairs DNA replication and triggers replication stress

A. Representative images of U2OS/GFP-FAM111A cell lines that were treated or not with DOX, fixed and stained with PCNA antibody. Mutation of the PIP box abrogates FAM111A localization to PCNA-positive DNA replication foci. **B.** DNA replication rates in HCT116/GFP-FAM111A WT cells treated or not with DOX, pulse-labeled with EdU and stained with DAPI were analyzed by quantifying EdU signal intensity in S phase cells using QIBC (*n*>500 cells per condition; red bars, mean (A.U., arbitrary units)). **C.** As in (A), except that cells were co-stained with PCNA and RPA2 antibodies. **D.** Quantification of data in (C) (grey bars, average; *n*>2000 cells per condition). Scale bars, 10 μm.

Data (A-D) are representative of three independent experiments with similar outcomes.

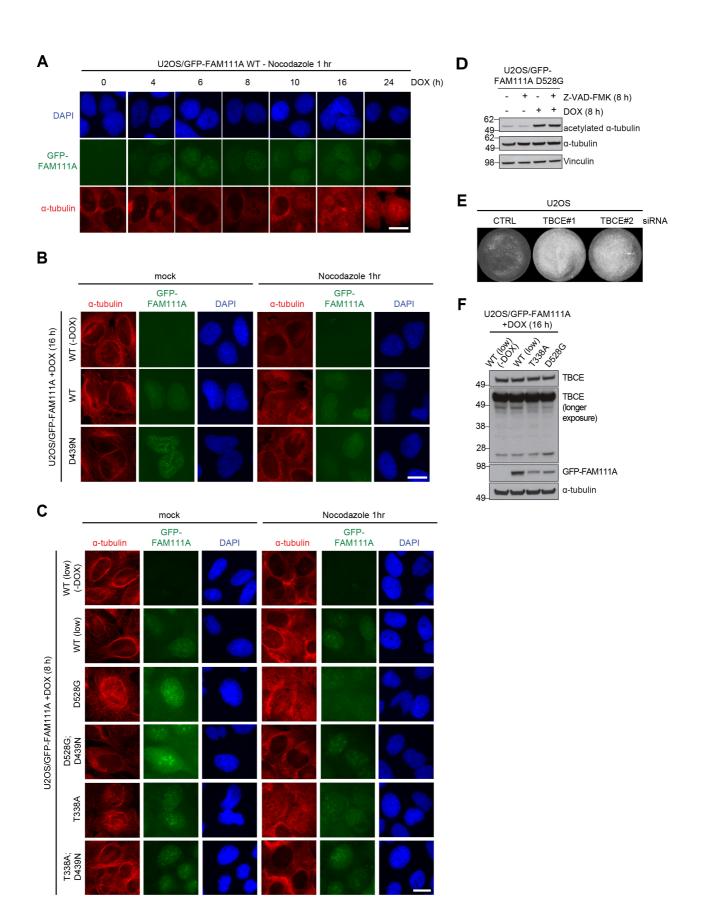


Appendix Figure S2.

FAM111A and FAM111B are dispensable for DNA replication and cell proliferation

A. Immunoblot analysis of U2OS cells transfected with indicated siRNAs. **B.** Cells in (A) were pulse-labeled with EdU, fixed and stained with DAPI. Cells were then subjected to QIBC analysis for quantification of EdU and DAPI signal intensities (*n*>2000 cells per condition). Proportion of cells in G1 (blue), S (red) and G2/M (yellow) phases is indicated. **C.** Cells in (A) were fixed and stained with PCNA antibody and DAPI. Cells were then subjected to QIBC analysis for quantification of PCNA and DAPI signal intensities (*n*>2000 cells per condition). **D.** Immunoblot analysis of U2OS cells and derivative lines with targeted knockout of *FAM111A* and/or *FAM111B*. **E.** Proliferation of cell lines in (D) was determined by quantification of Resazurin incorporation (mean±s.d.; *n*=3 independent experiments). **F.** As in (B), but using U2OS cell lines with targeted knockout of *FAM111A* and/or *FAM111B*. **G.** As in (C), but using U2OS cell lines with targeted knockout of *FAM111A* and/or *FAM111B*. **H.** DNA fiber analysis of indicated U2OS cell lines labeled with CldU (25min; red) followed by IdU (25min; green). Fork speeds were calculated as length of labeled track divided by pulse time (black bars, median±s.d.; at least 200 fibers analysed per condition). A representative fiber image is shown.

Data (A-H) are representative of three independent experiments with similar outcomes.

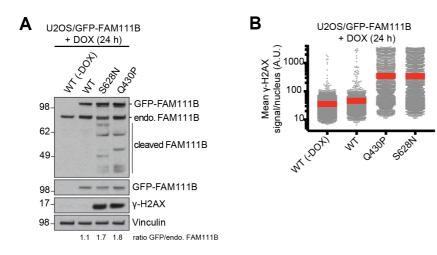


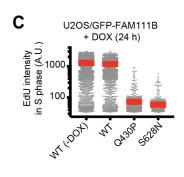
Appendix Figure S3.

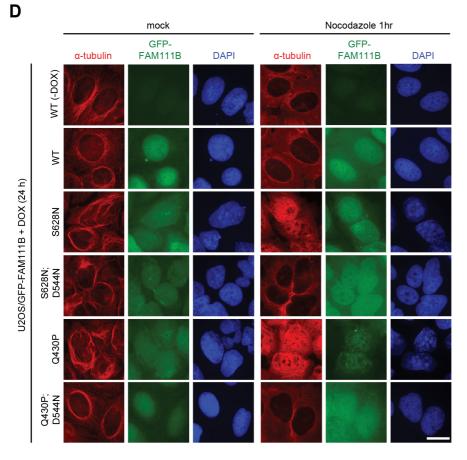
Elevated FAM111A protease activity disrupts microtubule organization

A-C. Representative images of U2OS/GFP-FAM111A cell lines that were treated or not with DOX for the indicated times, then left untreated or exposed to nocodazole for 1 h, and fixed and stained with α -tubulin antibody. Note that cells with elevated FAM111A protease activity display accumulation of nuclear α -tubulin. **D**. Immunoblot analysis of U2OS/GFP-FAM111A D528G cells treated with DOX and Z-VAD-FMK for 8 h, as indicated. **E**. U2OS cells were transfected with the indicated siRNAs, fixed 4 days later and stained with crystal violet. **F**. Immunoblot analysis of U2OS/GFP-FAM111A cell lines that were left untreated or induced with DOX for 16 h. Scale bars, 10 μ m.

Data are representative of three (A-C) and two (D-F) independent experiments with similar outcomes.



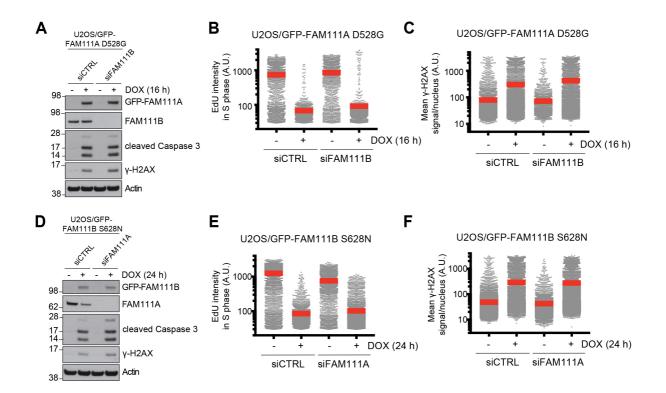


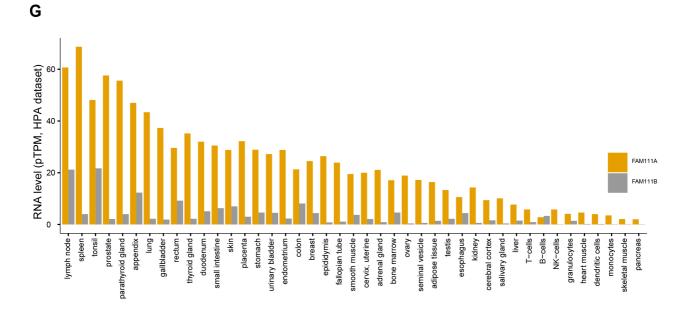


Appendix Figure S4.

Patient-associated FAM111B mutants trigger DNA replication suppression, apoptosis onset and microtubule network disruption in a protease-dependent manner

A. Immunoblot analysis of U2OS/GFP-FAM111B cell lines left untreated or induced with DOX for 24 h to induce expression of WT or disease-associated GFP-FAM111B alleles. **B**. Cells treated as in (A) were fixed, stained with γ-H2AX antibody and DAPI, and γ-H2AX signal intensity was analyzed by QIBC (red bars, mean; n>2000 cells per condition; A.U., arbitrary units). **C**. Cells treated as in (A) were labeled with EdU, fixed and stained with DAPI. EdU signal intensity was analyzed by QIBC (red bars, mean; n>2000 cells per condition). **D**. Representative images of U2OS/GFP-FAM111B cell lines that were treated or not with DOX, then left untreated or exposed to nocodazole for 1 h, and fixed and stained with α-tubulin antibody. Scale bar, 10 μm Data (**A-D**) are representative of three independent experiments with similar outcomes.



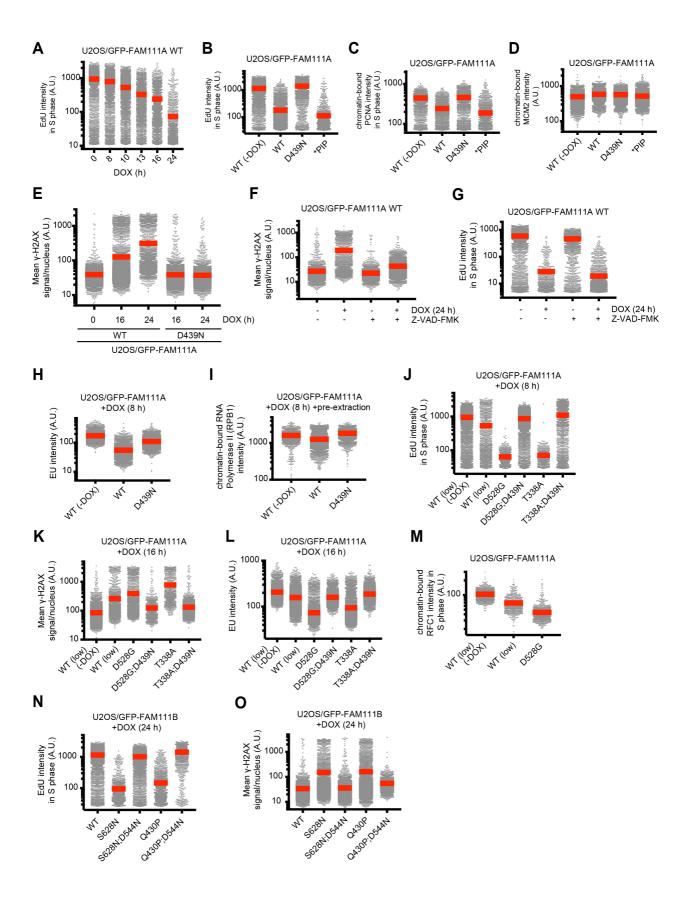


Appendix Figure S5.

FAM111A and FAM111B are differentially expressed in human tissues and impact DNA replication and apoptosis independently of each other

A. Immunoblot analysis of U2OS/GFP-FAM111A D528G cells transfected with control (CTRL) or FAM111B siRNAs, and subsequently left untreated or exposed to DOX. **B**. Cells treated as in (A) were pulse-labeled with EdU, stained with DAPI and analyzed for DAPI and EdU signal intensity using QIBC (n>2000 cells per condition; A.U., arbitrary units). **C**. Cells treated as in (A) were fixed, stained with γ-H2AX antibody and DAPI, and γ-H2AX signal intensity was analyzed by QIBC (red bars, mean; n>2000 cells per condition). **D**. Immunoblot analysis of U2OS/GFP-FAM111B S628N cells transfected with control (CTRL) or FAM111A siRNAs, and subsequently left untreated or exposed to DOX. **E**. Cells treated as in (D) were pulse-labeled with EdU, stained with DAPI and analyzed for DAPI and EdU signal intensity using QIBC (n>2000 cells per condition). **F**. Cells treated as in (D) were fixed, stained with γ-H2AX antibody and DAPI, and γ-H2AX signal intensity was analyzed by QIBC (red bars, mean; n>2000 cells per condition). **G**. Human Protein Atlas RNA-seq data on FAM111A and FAM111B transcript expression levels in 37 human tissues (pTPM, protein transcripts per million).

Data (A-F) are representative of three independent experiments with similar outcomes.



Appendix Figure S6.

Independent replicates of quantitative image-based cytometry (QIBC) experiments A. Biological replicate of Figure 2B (red bars, mean (A.U., arbitrary units); *n*>2000 cells per condition). **B.** Biological replicate of Figure 2D (red bars, mean; *n*>2000 cells per condition). **C.** Biological replicate of Figure 2E (red bars, mean; *n*>2000 cells per condition). **D.** Biological replicates of Figure 2F (red bars, mean; *n*>2000 cells per condition). **E.** Biological replicate of Figure 3C (red bars, mean; *n*>2000 cells per condition). **F.** Biological replicate of Figure EV3B (*n*>2000 cells per condition; red bars, mean). **G.** Biological replicate of Figure EV3E (*n*>1000 cells per condition). **H.** Biological replicate of Figure 3G (red bars, mean; *n*>2000 cells per condition). **J.** Biological replicate of Figure 4D (red bars, mean; *n*>2000 cells per condition). **K.** Biological replicate of Figure 4F (red bars, mean; *n*>2000 cells per condition). **M.** Biological replicate of Figure EV4J (red bars, mean; *n*>2000 cells per condition). **M.** Biological replicate of Figure EV4J (red bars, mean; *n*>2000 cells per condition). **N.** Biological replicate of Figure 5G (red bars, mean; *n*>2000 cells per condition). **O.** Biological replicate of Figure 5K (red bars, mean; *n*>2000 cells per condition).