

# Genomic Sequencing for Newborn Screening: Results of the NC NEXUS Project

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## Summary

Newborn screening (NBS) was established as a public health program in the 1960s and is crucial for facilitating detection of certain medical conditions in which early intervention can prevent serious, life-threatening health problems. Genomic sequencing can potentially expand the screening for rare hereditary disorders, but many questions surround its possible use for this purpose. We examined the use of exome sequencing (ES) for NBS in the North Carolina Newborn Exome Sequencing for Universal Screening (NC NEXUS) project, comparing the yield from ES used in a screening versus a diagnostic context. We enrolled healthy newborns and children with metabolic diseases or hearing loss (106 participants total). ES confirmed the participant's underlying diagnosis in 15 out of 17 (88%) children with metabolic disorders and in 5 out of 28 (~18%) children with hearing loss. We discovered actionable findings in four participants that would not have been detected by standard NBS. A subset of parents was eligible to receive additional information for their child about childhood-onset conditions with low or no clinical actionability, clinically actionable adult-onset conditions, and carrier status for autosomal-recessive conditions. We found pathogenic variants associated with hereditary breast and/or ovarian cancer in two children, a likely pathogenic variant in the gene associated with Lowe syndrome in one child, and an average of 1.8 reportable variants per child for carrier results. These results highlight the benefits and limitations of using genomic sequencing for NBS and the challenges of using such technology in future precision medicine approaches.

## Introduction

Newborn screening was established in the early 1960s, beginning with phenylketonuria<sup>1</sup> (PKU) (MIM: 261600), for which early intervention can prevent permanent intellectual disability. In the 1990s, tandem mass spectrometry (TMS) was employed to efficiently screen neonates for numerous metabolites, which resulted in an increase in the number of genetic conditions screened.<sup>2–4</sup> As of July 2018, The American College of Medical Genetics and Genomics and the United States federal Advisory Committee on Heritable Disorders in Newborns and Children have identified 35 core conditions and 26 secondary conditions to be included on the recommended uniform screening panel (RUSP, recommended by the Secretary of the Department of Health and Human Services), including specific disorders of organic acids, fatty acid oxidation, amino acids, as well as endocrine and hemoglobin disorders.<sup>5–7</sup> Most infants in the United States also receive point-of-

care newborn screening for hearing and critical congenital heart disease.

Advances in genomic sequencing technologies (next-generation sequencing, or NGS), such as exome and genome sequencing (ES or GS, respectively), have emerged as powerful tools for identifying an individual's genomic changes at the DNA base level.<sup>8</sup> These technologies have the potential to dramatically increase the ability to screen newborns for rare hereditary disorders that would not be detected using traditional screening methods. Given this potential, the National Institute of Child Health and Human Development (NICHD) and the National Human Genome Research Institute (NHGRI) sponsored a cohort of studies, the Newborn Sequencing in Genomic Medicine and Public Health (NSIGHT) consortium, to investigate the use of this technology in newborns.<sup>9–12</sup> One of these studies recently reported on the use of NGS in healthy newborns and reported additional findings that would have been missed by existing

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newborn screening modalities.<sup>10</sup> Thus, for some cases, genomic sequencing may clarify the underlying genetic cause of an individual's disease and directly impact a patient's diagnosis, intervention, and/or treatment. In addition, genomic sequencing may also be useful for screening asymptomatic individuals.

In the North Carolina Newborn Exome Sequencing for Universal Screening (NC NEXUS) study, we explored the possible use of ES in the context of newborn screening and studied the responses of parents to this new technology. The study design<sup>13</sup> included two cohorts of infants and children: (1) healthy newborns whose parents were approached for participation in the study prenatally; and (2) infants and children (<5 years of age) already clinically diagnosed with conditions detected with current newborn screening methods (inborn errors of metabolism and hearing loss).

In the current manuscript, we report the molecular analysis results from the cohort of 106 children who were enrolled in the study. Molecular analysts examined ES data using a next-generation sequencing newborn screen (NGS-NBS) panel of 466 genes,<sup>14</sup> while being blinded to the identity of the participant's cohort (well-child, metabolic, or hearing loss). In our analysis strategy, we defined strict criteria for assessing the screen as abnormal or "positive," which allowed us to examine the sensitivity of ES when used as a screening test to detect a genetic alteration in the context of a theoretical newborn screen. For those participants in the metabolic cohort or the hearing loss cohort, we then performed a second, indication-based, diagnostic analysis assessing the ES results for additional genes that were likely associated with the participant's phenotype. In addition, parents were randomized to one of two study arms. In one study arm, parents could elect to learn additional findings from their child's ES. The results of the initial NGS-NBS screening, indication-based analyses, and additional genomic findings are presented in this work.

## Subjects and Methods

### Recruitment and Sample Collection

The study protocol was approved by the University of North Carolina Institutional Review Board and all participants were consented to the study by a certified genetic counselor. The NC NEXUS randomized controlled trial protocol and study design were previously described.<sup>13</sup> As part of that protocol, 17 children previously diagnosed with inborn errors of metabolism and 28 children previously diagnosed with hearing loss were recruited into the study. Additionally, we recruited parents in the prenatal period (pregnancies of 18 weeks or longer) in which the fetus did not have a positive or pending chromosomal abnormality or congenital malformation diagnostic test result. The mothers subsequently gave birth to 61 infants; these babies were classified into the well-child cohort. Participant demographic information is reported in [Table S1](#).

### Decision Aids

As part of the overall study, we previously described a decision aid for parents who would be offered ES for their child.<sup>13,15</sup> This web-

based electronic decision aid was developed to enhance parents' abilities to make decisions about learning genomic information for their child; the aid included both an educational component and a values clarification exercise. The educational component included information about newborn screening, genomic sequencing, and the process of using genomic sequencing to identify genetic variants associated with specific diseases. This educational component was followed by a values clarification exercise in which parents classified five reasons for and five reasons against having genomic sequencing for their child by level of importance.<sup>15-17</sup> After viewing the decision aid, parents met face to face with a genetic counselor for additional clarification before being asked to decide whether to consent to ES for their child.

### Age-Based Semiquantitative Metric

As previously described,<sup>14</sup> we determined actionability of gene-disease pairs using the age-based semiquantitative metric (ASQM). Conditions were placed into one of four categories: category 1, pediatric conditions with high medical actionability; category 2, pediatric conditions with low or no medical actionability; category 3, adult conditions with high medical actionability; and category 4, adult conditions with low or no medical actionability. In total, 822 gene-disease pairs were assessed, enriched for those with pediatric onset of disease and suspected actionability. Category 1 (466 gene-disease pairs) included many core RUSP conditions, as well as other disorders with onset in infancy or in childhood that have treatment, monitoring, and/or medical management that can potentially improve clinical outcomes; these conditions are comparable to those detected by current NBS and therefore all NC NEXUS study participants were eligible for disclosure of abnormal, positive findings.

### Randomization for Additional Findings

Parents were randomized to one of two study arms. One arm was the "control arm" (1/3 of participants) in which parents learned findings from the NGS-NBS panel and, if applicable, indication-based analysis. The second arm was the "decision arm" (2/3 of participants) in which parents learned findings from the NGS-NBS (and indication-based analysis if applicable) and were also asked to decide whether or not they wanted to learn about additional types of genomic findings from up to three categories. These three categories included: childhood-onset conditions with low or no clinical actionability, adult-onset conditions with high actionability, and carrier status for autosomal-recessive disorders.

### Exome Sequencing

The BioSpecimen Processing Facility at the University of North Carolina at Chapel Hill isolated genomic DNA from saliva and oral epithelial cell samples using PureGene chemistry. Exome libraries, including molecular barcoding and exome capture, were prepared using Agilent SureSelect XT kits (Human All Exon V6 probes) according to the manufacturer's low input guidelines. The University of North Carolina High-Throughput Sequencing Facility performed exome sequencing on an Illumina HiSeq2500 with minimum depth of 40× coverage. Raw sequence reads were mapped to the human genome reference GRCh38.p7 using BWA-MEM version 0.7.12,<sup>18</sup> duplicate reads were marked using Picard MarkDuplicates (v.1.130), and variants were called using FreeBayes, v.1.1.0.<sup>19</sup>

## Bioinformatics and Molecular Analysis

Variants were prioritized based on previous classification as pathogenic (P) or likely pathogenic (LP) in a ClinVar submission,<sup>20</sup> minor allele frequency in a reference database,<sup>21,22</sup> and predicted effect of the variant on the protein (frameshift, nonsense, canonical splice-site, missense, synonymous, intronic variants). All rare possibly damaging variants in 466 genes in category 1 were reviewed by a molecular analyst for each of the 106 participants. For the initial NGS-NBS analysis, molecular analysts were blinded to the participant's cohort (metabolic, hearing loss, or well-child). Variants were classified based on ACMG/AMP interpretation guidelines<sup>23,24</sup> using collected evidence from population databases,<sup>21,22</sup> ClinVar,<sup>20</sup> and the primary literature. Pathogenic and/or LP variants that were considered an abnormal "positive screen" (heterozygous P/LP variants in a gene associated with a dominant condition; two or more P/LP variants in a gene associated with a recessive condition) were flagged for further review by a committee of molecular geneticists and physicians, genetic counselors, and researchers. Variants of uncertain significance (VUS) were not returned in the NGS-NBS screen. After completion of the first NGS-NBS analysis, the participant's cohort was revealed to the molecular analyst. If the participant was part of the metabolic cohort or hearing loss cohort, ES data were further analyzed by filtering for variants in a subset of genes within indication-based "diagnostic lists," containing genes associated with the relevant phenotypes. For the indication-based diagnostic analyses, P, LP, and VUS were reported. All variants were confirmed in a duplicate sample using an orthogonal method performed in the UNC Hospitals Molecular Genetics CLIA-certified laboratory. Copy number variant analysis was not conducted due to restrictions based on the study's Investigational Device Exemption approved by the FDA.<sup>13</sup>

For participants who decided to learn some or all of the additional findings categories, ES data were analyzed to identify P/LP variants in 22 genes associated with adult-onset actionable conditions, 234 genes associated with childhood-onset nonactionable conditions, and/or 871 genes associated with carrier findings for autosomal-recessive conditions.

All parents were seen by a genetic counselor and/or clinical geneticist for return of any positive results in diagnostic, NGS-NBS, childhood-onset medically non-actionable, and adult-onset medically actionable categories. Carrier status results were conveyed by a genetic counselor only by phone. Recommendations and options for further testing or evaluation were provided but not part of the study. Parents were given a copy of the CLIA-confirmed result report for their child and were given the option of having the results placed in their child's electronic health record.

## Results

Molecular analysis was completed for 106 participants: 61 in the healthy/well-child cohort, 17 in the metabolic cohort, and 28 in the hearing loss cohort. A total of 43 out of 46 variants detected by ES were confirmed by orthogonal testing in the CLIA-certified laboratory prior to reporting. Additionally, 5 variants that were previously identified by clinical molecular testing were also detected by ES.

### Metabolic Cohort

NGS-NBS analysis was deemed to represent an abnormal positive screen result in 15 of 17 participants (88%) in

the metabolic cohort (Tables 1 and 2). These individuals had a combination of pathogenic and/or likely pathogenic variants in genes implicated in metabolic conditions, that, when blinded to the participants' cohort, were determined to be reportable in a screening setting.

Seven participants previously diagnosed with phenylketonuria (PKU) by newborn screening had pathogenic variants in *PAH* (MIM: 612349); six of them were likely compound heterozygous for two different variants (ranging from missense, canonical splice-site, nonsense, and/or deletion variants). In view of their previously established diagnosis of PKU, we did not routinely perform parental testing to confirm the phase of these *PAH* variants. The seventh participant was likely homozygous for a pathogenic missense variant in *PAH* (c.194T>C [p.Ile65Thr]), although we did not rule out the possibility of a deletion in *trans*.

Seven participants previously diagnosed with medium-chain acyl-coA dehydrogenase (MCAD) deficiency (MIM: 201450) by newborn screening had pathogenic variants in *ACADM* (MIM: 607008); five of these seven were homozygous for a well-known pathogenic missense variant (c.985A>G [p.Lys329Glu]) reported previously in individuals with MCAD deficiency.<sup>25–28</sup> Two of these seven were likely compound heterozygous for two different variants in the *ACADM* gene: c.985A>G (p.Lys329Glu) and c.799G>A (p.Gly267Arg); c.985A>G (p.Lys329Glu) and c.199T>C (p.Tyr67His).

One participant was previously diagnosed with primary carnitine deficiency (MIM: 212140) by newborn screening due to a borderline low free carnitine level (C0) found by TMS. This individual was homozygous for a pathogenic *SLC22A5* (MIM: 603377) missense variant (c.506G>A [p.Arg169Gln]) consistent with his clinical diagnosis (see GeneReviews by El-Hattab in Web Resources).<sup>29–34</sup>

NGS-NBS analysis was deemed normal, or "negative," in 2 of 17 participants (12%) in the metabolic cohort. However, after unblinding, diagnostic analysis revealed suggestive but inconclusive findings (Tables 1 and 2). One participant previously diagnosed with maple syrup urine disease (MSUD [MIM: 248600]) was heterozygous for a pathogenic missense variant in the *BCKDHA* (MIM: 608348) gene (c.1312T>A [p.Tyr438Asn]) associated with classic, autosomal-recessive MSUD.<sup>35–39</sup> Since we did not identify any additional variants in *BCKDHA* that might explain a genetic etiology for this participant's disease, we classified this result as inconclusive. Another participant previously diagnosed with malonyl-CoA decarboxylase deficiency (MIM: 248360) was homozygous for a variant (c.1013T>C [p.Leu338Pro]) in *MLYCD* (MIM: 606761), the gene that encodes the malonyl-CoA decarboxylase enzyme. This variant is present in population databases at low frequency (Genome Aggregation Database total allele frequency: 1 allele/249,584 alleles) and is located in a well-conserved catalytic domain of the encoded protein, with other reported benign variants observed nearby.<sup>20–22,40</sup> The variant was not previously reported in the clinical literature, and many of the other reported pathogenic variants in this

**Table 1. Summary of NGS-NBS and Diagnostic Findings in the NC NEXUS Study**

NC NEXUS Cohort	Total Number of Participants in Cohort	NGS-NBS Abnormal Positive Screen	NGS-NBS Normal Negative Screen	Diagnostic Positive	Diagnostic Inconclusive	Diagnostic Negative
Metabolic	17	15	2	15	2	0
Hearing loss	28	7*	21	5	5	18
Healthy/well-child	61	1*	60	0	0	0
<b>Sum</b>						
Unanticipated findings	106	4	N/A	N/A	N/A	N/A

An abnormal NGS-NBS, or positive screen, was indicated by observing likely pathogenic and/or pathogenic variants in genes associated with pediatric conditions with high medical actionability. A normal NGS-NBS, or negative screen, was defined by the absence of likely pathogenic or pathogenic variants found in these genes. Diagnostic positive indicates the presence of likely pathogenic or pathogenic variants found in gene(s) on the metabolic or hearing loss diagnostic list that are consistent with the participant's disorder. Diagnostic inconclusive indicates an inconclusive result (i.e., a single heterozygous variant found in a gene associated with an autosomal-recessive condition and/or variants of uncertain significance [VUS] in genes on the diagnostic list). A negative diagnostic result indicated that we did not detect any pathogenic or likely pathogenic variants or any VUS on the diagnostic gene lists. Numbers with asterisks include two participants in the hearing loss cohort with an abnormal NGS-NBS due to likely pathogenic variants in *DSC2* or *F11* plus one participant in the healthy/well-child cohort that was determined to have an abnormal NGS-NBS due to a pathogenic variant in *LDLR* associated with familial hypercholesterolemia. NGS-NBS, next-generation sequencing newborn screen.

gene are putative loss of function.<sup>41–44</sup> Importantly, this infant was born prematurely at 23 weeks gestation and required several repeat newborn screens due to inconclusive findings prior to eventual confirmation of malonyl-CoA decarboxylase deficiency at 7 weeks of age. It is worth noting that a ClinVar<sup>20</sup> entry is available for this variant (ClinVar: VCV000432061.2) with a “likely pathogenic” interpretation. Based on clinical testing (not known to the research team at the time of the analysis), we believe this observation to be the same as the NC NEXUS participant described here. The discordance between our interpretation and GeneDx's interpretation is likely due to our more conservative use of evidence codes related to *in silico* predictors and the location of the variant within a catalytic domain. Considering there was no available *in vivo* or *in vitro* functional evidence to confirm a loss-of-function effect for this variant, we continue to interpret this rare novel missense variant as a VUS due to insufficient evidence. In the context of the participant's phenotypic information, the *MLYCD* variant provides a suggestive but not definitive explanation for the genetic cause of the participant's disorder.

### Hearing Loss Cohort

NGS-NBS analysis was deemed to represent an abnormal positive screen result in 5 of 28 participants (18%) in the hearing loss cohort (Tables 1 and 3). These individuals had a combination of pathogenic and/or likely pathogenic variants in genes implicated in hearing loss that, when blinded to cohort, were determined to be reportable in a screening setting. Two participants were each presumed to be compound heterozygous for two variants in an Usher syndrome gene, *USH2A* (MIM: 608400) based on ES data (c.1256G>T [p.Cys419Phe] and c.3686T>G [p.Leu1229Ter]; c.4338\_4339del [p.Cys1447fs] and c.2299del [p.Glu767fs]). In both cases, we subsequently confirmed that the variants were in *trans* by parental testing. A clinical geneticist and genetic counselor discussed the implications of this diagnosis, including the

likelihood of vision loss due to retinitis pigmentosa, with the parents of these children during the in-person return of results visits. In addition, the geneticist and genetic counselor provided information regarding clinical trials for treatment of retinitis pigmentosa to the parents. One participant was homozygous for a 1-bp frameshift deletion (c.35del [p.Gly12fs]) in *GJB2* (MIM: 121011); encoding connexin 26), a common pathogenic variant associated with DFNB1 nonsyndromic deafness (MIM: 220290).<sup>45–48</sup> One participant was compound heterozygous for two pathogenic missense variants (c.626G>T [p.Gly209Val] and c.1151A>G [p.Glu384Gly]) in *SLC26A4* (MIM: 605646), a gene associated with Pendred syndrome (MIM: 274600), an autosomal-recessive form of syndromic hearing loss.<sup>49–53</sup> Finally, one participant was heterozygous for a pathogenic variant (c.5597C>T [p.Thr1866Met]) in *TECTA* (MIM: 602574), a gene associated with autosomal-dominant hearing loss (MIM: 601543).<sup>54–56</sup> Of note, this infant passed her newborn hearing screen, but presented at 1 year of age with unilateral hearing loss that progressed to both ears. She was included in the study after referral by her audiologist, despite not fitting the original enrollment criteria, as her hearing loss was not detected by newborn screening.

In 5 of 28 participants in the hearing loss cohort, NGS-NBS screening analysis was deemed normal, or “negative.” However, after unblinding, diagnostic analysis identified five “inconclusive” results that may provide a possible explanation for their hearing loss (Tables 1 and 3). One participant was heterozygous for the pathogenic c.35del (p.Gly12fs) deletion in *GJB2*. No additional pathogenic variants were detected in *GJB2*, and analysis in the CLIA-certified laboratory for intragenic *GJB6* (MIM: 604418) deletions was negative; however, we cannot rule out the presence of a second pathogenic variant in *GJB2* not detected by ES, such as a partial gene deletion or cryptic splice alteration caused by a non-coding variant.

**Table 2. NGS-NBS and Diagnostic Findings in the NC NEXUS Inborn Errors of Metabolism Cohort**

NC NEXUS Participant	NGS-NBS Result	Diagnostic (Dx) Result	Gene(s)	Disease Association with Gene (Inheritance)	Variant and (Predicted Protein Change)	Zygoty in NC NEXUS Participant
M001	abnormal	positive	<i>ACADM</i>	MCAD deficiency (AR)	c.799G>A (p.Gly267Arg); c.985A>G (p.Lys329Glu)	heterozygous for both variants
M002	abnormal	positive	<i>PAH</i>	PKU (AR)	c.1315+1G>A (p.?); c.782G>A (p.Arg261Gln)	heterozygous for both variants
M003	abnormal	positive	<i>PAH,OTC</i>	PKU (AR), OTC deficiency (XL)	<i>PAH</i> c.1222C>T (p.Arg408Trp) and c.896T>G (p.Phe299Cys); <i>OTC</i> c.1061T>G (p.Phe354Cys)	heterozygous for <i>PAH</i> and <i>OTC</i> variants
M004	abnormal	positive	<i>PAH</i>	PKU (AR)	c.1315+1G>A (p.?); c.284_286del (p.Ile95del)	heterozygous for both variants
M005	abnormal	positive	<i>PAH</i>	PKU (AR)	c.194T>C (p.Ile65Thr); c.814G>T (p.Gly272*)	heterozygous for both variants
M006	abnormal	positive	<i>ACADM</i>	MCAD deficiency (AR)	c.985A>G (p.Lys329Glu)	homozygous
M007	abnormal	positive	<i>PAH</i>	PKU (AR)	c.1315+1G>A (p.?); c.842C>T (p.Pro281Leu)	heterozygous for both variants
M008	abnormal	positive	<i>ACADM</i>	MCAD deficiency (AR)	c.985A>G (p.Lys329Glu)	homozygous
M009	normal	inconclusive (VUS)	<i>MLYCD</i>	malonyl-coA decarboxylase deficiency (AR)	c.1013T>C (p.Leu338Pro)	homozygous
M010	abnormal	positive	<i>PAH</i>	PKU (AR)	c.117C>G (p.Phe39Leu); c.842C>T (p.Pro281Leu)	heterozygous for both variants
M011	abnormal	positive	<i>PAH</i>	PKU (AR)	c.194T>C (p.Ile65Thr)	homozygous
M012	abnormal	positive	<i>ACADM</i>	MCAD deficiency (AR)	c.985A>G (p.Lys329Glu)	homozygous
M013	abnormal	positive	<i>ACADM</i>	MCAD deficiency (AR)	c.985A>G (p.Lys329Glu)	homozygous
M014	abnormal	positive	<i>SLC22A5</i>	renal carnitine transport deficiency (AR)	c.506G>A (p.Arg169Gln)	homozygous
M015	normal	inconclusive (heterozygous variant)	<i>BCKDHA</i>	MSUD type 1A (AR)	c.1312T>A (p.Tyr438Asn)	heterozygous
M016	abnormal	positive	<i>ACADM</i>	MCAD deficiency (AR)	c.985A>G (p.Lys329Glu)	homozygous
M017	abnormal	positive	<i>ACADM</i>	MCAD deficiency (AR)	c.985A>G (p.Lys329Glu); c.199T>C (p.Tyr67His)	heterozygous for both variants

The NC NEXUS participant column includes “M” (indicating inborn errors of metabolism cohort) followed by the participant number. Each row in the table represents one NC NEXUS participant. An abnormal NGS-NBS result was indicated by a likely pathogenic or pathogenic variant found in the newborn screen gene list. A positive diagnostic result was indicated by a likely pathogenic or pathogenic variant found in a gene on the inborn errors of metabolism diagnostic list. An inconclusive diagnostic result was indicated by any variant of uncertain significance (VUS) finding or a single heterozygous variant found in a gene associated with autosomal-recessive inborn errors of metabolism. A negative diagnostic result indicated we did not detect any pathogenic or likely pathogenic variants or any VUS on the inborn errors of metabolism gene list. Gene symbols are italicized. NGS-NBS, next-generation sequencing newborn screen; AR, autosomal-recessive pattern of inheritance; XL, X-linked pattern of inheritance; PKU, phenylketonuria; MCAD deficiency, medium-chain acyl-coA dehydrogenase deficiency; MSUD, maple syrup urine disease.

**Table 3. Participants in the NC NEXUS Hearing Loss Cohort with Positive or Inconclusive Findings**

NC NEXUS Participant	NGS-NBS Result	Diagnostic (Dx) Result	Gene(s)	Disease Association with Gene (Inheritance)	Variant and (Predicted Protein Change)	Zygoty in NC NEXUS Participant
HL003	abnormal	positive	<i>SLC26A4</i>	Pendred syndrome (AR)	c.626G>T (p.Gly209Val); c.1151A>G (p.Glu384Gly)	heterozygous for both variants
HL004	normal	inconclusive VUS	<i>MARVELD2 MYO7A</i>	deafness 49 (AR); deafness 2 (AR); Usher type 1B (AR)	<i>MARVELD2</i> c.1183–1G>A (p.?); <i>MYO7A</i> c.5824G>A (p.Gly1942Arg)	heterozygous for <i>MARVELD2</i> variant; homozygous for <i>MYO7A</i> variant
HL010	abnormal	positive	<i>USH2A</i>	Usher type 2A (AR)	c.1256G>T (p.Cys419Phe); c.3686T>G (p.Leu1229*)	heterozygous for both variants
HL013	abnormal	negative	<i>F11</i>	factor XI deficiency (AR)	c.1489C>T (p.Arg497*); c.1608G>C (p.Lys536Asn)	heterozygous for both variants
HL014	abnormal	positive	<i>GJB2</i>	deafness 1A (AR)	c.35del (p.Gly12fs)	homozygous
HL016	abnormal	positive	<i>USH2A</i>	Usher type 2A (AR)	c.4338_4339del (p.Cys1447fs); c.2299del (p.Glu767fs)	heterozygous for both variants
HL017	abnormal	positive	<i>TECTA</i>	deafness 8/12 (AD)	c.5597C>T (p.Thr1866Met)	heterozygous
HL021	abnormal	negative	<i>DSC2</i>	arrhythmogenic right ventricular dysplasia (ARVD11) (AD)	c.631–2A>G (p.?)	heterozygous
HL022	normal	inconclusive VUS	<i>TMPRSS3</i>	nonsyndromic hearing loss and deafness (AR)	c.208del (p.His70fs); c.1151T>A (p.Met384Lys)	heterozygous for both variants
HL025	normal	inconclusive (heterozygous variant)	<i>GJB2</i>	deafness (AR)	c.35del (p.Gly12fs)	heterozygous
HL026	normal	inconclusive VUS	<i>LOXHD1; CEMIP; MYH14</i>	hearing loss (AR or AD)	<i>LOXHD1</i> c.2914G>A (p.Glu972Lys); <i>LOXHD1</i> c.3161C>T (p.Thr1054Met); <i>CEMIP/KIAA1199</i> c.58C>T (p.Leu20Phe); <i>MYH14</i> c.5942G>C (p.Gly1981Ala)	heterozygous for all variants
HL027	normal	inconclusive VUS	<i>SOX10</i>	Waardenburg syndrome (AD)	c.482G>A (p.Arg161His)	heterozygous

The NC NEXUS participant column includes “HL” (indicating hearing loss cohort) followed by the participant number. Each row in the table represents one NC NEXUS participant. An abnormal NGS-NBS result was indicated by a likely pathogenic or pathogenic variant found in the newborn screen gene list. A positive diagnostic result was indicated by a likely pathogenic or pathogenic variant found in a gene on the hearing loss list. An inconclusive diagnostic result was indicated by any variant of uncertain significance (VUS) finding or a heterozygous variant found in a gene associated with an autosomal-recessive form of hearing loss. A negative diagnostic result indicated we did not detect any pathogenic or likely pathogenic variants or any VUS on the hearing loss gene list. Gene symbols are italicized. NGS-NBS, next-generation sequencing newborn screen; AR, autosomal-recessive pattern of inheritance; AD, autosomal-dominant pattern of inheritance.

<b>Table 4. Other Actionable Findings in the NC NEXUS NGS-NBS</b>					
<b>NC NEXUS Participant</b>	<b>NGS-NBS Result</b>	<b>Gene</b>	<b>Disease Association with Gene (Inheritance)</b>	<b>Variant and (Predicted Protein Change)</b>	<b>Zygoty in NC NEXUS Participant</b>
M003	abnormal	<i>OTC</i>	OTC deficiency (XL)	c.1061T>G (p.Phe354Cys)	heterozygous
NB012	abnormal	<i>LDLR</i>	familial hypercholesterolemia (AD or AR)	c.502G>A (p.Asp168Asn)	heterozygous
HL013	abnormal	<i>F11</i>	factor XI deficiency (AR)	c.1489C>T (p.Arg497*), c.1608G>C (p.Lys536Asn)	heterozygous for both variants
HL021	abnormal	<i>DSC2</i>	arrhythmogenic right ventricular dysplasia (ARVD1) (AD)	c.631-2A>G (p.?)	heterozygous

The NC NEXUS participant column includes "M" (indicating inborn errors of metabolism cohort), "HL" (indicating hearing loss cohort) or "NB" (indicating well-child cohort) followed by the participant number. Each row in the table represents one NC NEXUS participant. Other actionable findings were defined by pathogenic or likely pathogenic variants found in the NGS-NBS that are associated with a disorder not previously known in the NC NEXUS participant. Gene symbols are italicized. NGS-NBS, next-generation sequencing newborn screen; XL, X-linked inheritance pattern; AR, autosomal-recessive pattern of inheritance; AD, autosomal-dominant pattern of inheritance.

Other "inconclusive" findings were reported that did not provide definitive diagnostic results. One participant was heterozygous for four different missense VUSs in genes associated with hearing loss: two variants in *LOXHD1* (MIM: 613072) (c.2914G>A [p.Glu972Lys] and c.3161C>T [p.Thr1054Met]), one variant in *CEMIP*, also known as *KIAA1199* (MIM: 608366) (c.58C>T [p.Leu20Phe]), and one variant in *MYH14* (MIM: 608568) (c.5942G>C [p.Gly1981Ala]). One participant was heterozygous for a missense VUS in *SOX10* (MIM: 602229) (c.482G>A [p.Arg161His]), a gene associated with autosomal-dominant Waardenburg syndrome<sup>57</sup> (MIM: 611584, 613266). One participant was homozygous for a missense VUS in *MYO7A* (MIM: 276903) (c.5824G>A [p.Gly1942Arg]), a gene associated with autosomal-recessive syndromic and non-syndromic hearing loss<sup>58,59</sup> (MIM: 276900, 600060). One participant was compound heterozygous for one likely pathogenic variant and one VUS in *TMPRSS3* (MIM: 605511): c.208del (p.His70fs) and c.1151T>A (p.Met384Lys); parental testing indicated that each variant was in *trans*.

We did not identify any P or LP variants on the diagnostic gene lists for 18 out of 28 (64%) participants in the hearing loss cohort; therefore, diagnostic analyses were deemed "negative" for identifying a genetic cause of hearing loss in these individuals (Table 1). Interestingly, one participant referred by audiologists to the hearing loss cohort had a rare syndromic diagnosis, Warsaw breakage syndrome (MIM: 613398), which is characterized by severe microcephaly and intellectual disability, growth restriction, and sensorineural hearing loss due to cochlear hypoplasia (see GeneReviews by Alkhunaisi et al. in [Web Resources](#)).<sup>60</sup> Clinical quad exome sequencing of the similarly affected sibling, this participant, and parents revealed a homozygous pathogenic variant in exon 13 of the *DDX11* (MIM: 601150) gene (GenBank: NM\_030653.4; c.1403dup [p.Ser469fs]) in the participant and his sibling. This *DDX11* c.1403dup (p.Ser469fs) variant was present in the NC NEXUS research ES data but was not examined because *DDX11* was not included in either the NGS-NBS list or the hearing loss diagnostic list. If additional phenotypic information had been available at the time of molecular analysis, it is likely this variant would have been recognized as having diagnostic significance.

#### NGS-NBS Conditions

An abnormal, positive screen NGS-NBS result was also defined as a finding that predicted a childhood-onset actionable condition in any individual in the well-child cohort, or a condition unrelated to the indicated diagnosis of a member of one of the affected cohorts. NGS-NBS analysis was deemed abnormal, or positive in 4 of 106 participants (Table 4). One participant in the well-child cohort was heterozygous for a missense *LDLR* (MIM: 606945) variant (c.502G>A [p.Asp168Asn]) previously reported in autosomal-dominant familial hypercholesterolemia<sup>61-67</sup> (MIM: 143890). After disclosure of this finding, the parents

stated that they were aware of a family history of hypercholesterolemia.

One female participant in the metabolic cohort, diagnosed with PKU, was a carrier of a missense variant (c.1061T>G [p.Phe354Cys]) in the ornithine transcarbamylase (*OTC* [MIM: 300461]) gene. This variant was previously reported in mild *OTC* deficiency<sup>68,69</sup> (MIM: 311250), an X-linked disease with variable expressivity in carrier females. Although this participant was diagnosed with PKU by the standard newborn screen at birth, the *OTC* deficiency was an unexpected finding arising from the ES analysis. Her ammonia levels are normal, and the variant was also found in her unaffected grandfather. Her younger male sibling was prenatally diagnosed with the variant, which enabled early measurement of ammonia level immediately after delivery and appropriate ongoing health supervision. So far, he has been asymptomatic. One hearing loss cohort participant was heterozygous for a canonical splice-site variant in *DSC2* (MIM: 125645) (c.631-2A>G [p.?]). This variant has been reported previously in individuals with autosomal-dominant arrhythmogenic right ventricular dysplasia<sup>70</sup> (MIM: 610476). This child has had a normal echo and it was recommended that the child be followed by a cardiologist. Another hearing loss cohort participant had two variants in *F11* (MIM: 264900) (nonsense variant c.1489C>T [p.Arg497Ter] and missense variant c.1608G>C [p.Lys536Asn]). Both variants have been reported previously in individuals with autosomal-recessive factor XI deficiency<sup>71-73</sup> (MIM: 612416). This child has a history of frequent episodes of epistaxis (nosebleeds), one requiring cauterization. It was recommended that she be evaluated by a hematologist, and it was also recommended that precautions are taken prior to any surgical procedures.

### Additional Genomic Findings

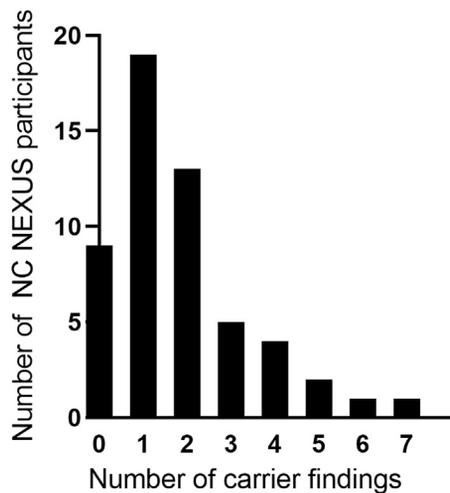
A total of 65 participants were randomized into the experimental decision arm of the study, viewed the decision aid, and attended the study visit with a genetic counselor. More than 90% of these parents chose to learn at least one category of additional genomic information: 47 (72.3%) requested all three categories of additional information, 7 requested additional findings for both adult-onset actionable conditions and carrier status, 2 requested additional findings for the adult-onset actionable category only, 1 requested carrier status only, 1 requested findings for the adult-onset actionable and childhood-onset nonactionable conditions categories, and 1 requested the childhood-onset nonactionable category only. The parents of 6 participants (9.2%) elected not to receive any of the additional information categories. Details about the factors associated with parental decision making will be reported in a separate manuscript.

Two participants had additional findings in the adult-onset actionable category (Table S2). A participant in the metabolic cohort (M007) was heterozygous for a pathogenic deletion *RAD51C* (MIM: 602774) variant

(c.905-2\_905-1del [p.?]), which is predicted to alter a splice site and has been reported previously in autosomal-dominant familial ovarian cancer<sup>74,75</sup> (MIM: 613399). This participant had a known family history of breast cancer but no known family history of ovarian cancer. A participant in the well-child cohort (NB040) was heterozygous for a nonsense c.7480C>T (p.Arg2494Ter) variant in *BRCA2* (MIM: 600185) that is a known pathogenic variant associated with autosomal-dominant familial breast and ovarian cancer<sup>76-83</sup> (MIM: 612555) This participant's parents reported a maternal family history of breast and pancreatic cancer. The child's mother was referred to adult genetics.

One participant in the hearing loss cohort (HL015) had an additional finding for the childhood-onset nonactionable conditions category (Table S2). This participant was hemizygous for a likely pathogenic nonsense variant (c.741G>A [p.Trp247Ter]) in *OCRL* (MIM: 300535). This gene is associated with Lowe syndrome<sup>84</sup> (MIM: 309000), and further correlation with clinical findings indicated that the participant has multiple features consistent with Lowe syndrome, including congenital cataracts with severe vision loss, intellectual disability, and renal impairment. Indeed, we subsequently learned that the patient had been seen in clinical genetics and that clinical exome sequencing was considered but not performed due to concerns about cost. Although hearing loss is not a prominent feature of Lowe syndrome, there is at least one previous case report<sup>85</sup> involving an individual with Lowe syndrome who also had hearing loss, suggesting that the finding may provide a comprehensive diagnostic result for this participant's symptoms. The mother of this patient is a carrier of the variant and cascade testing of other family members was negative.

The number of carrier findings in children whose parents requested this category ranged from 0 to 7 variants, with an average of 1.8 reportable carrier findings per individual (Figure 1, Table S2). A total of 100 out of 109 (92%) potential carrier findings detected by ES were confirmed by orthogonal testing in a CLIA-certified laboratory and reported. The nine variants that failed to be confirmed by orthogonal testing had low read depth for both alleles in the ES data. We observed several recurrent carrier findings; the most common were variants in the genes *HFE* (MIM: 613609), *GALT* (MIM:606999), *SERPINA1* (MIM:107400), and *RBM8A* (MIM: 605313) (Figure 2). A total of 11 participants carried the *HFE* c.187C>G (p.His63Asp) variant, while 4 carried the *HFE* c.845G>A (p.Cys282Tyr) variant, both associated with hereditary hemochromatosis (see GeneReviews by Barton and Edwards in Web Resources)<sup>86</sup> (MIM: 235200). Four participants each carried the *SERPINA1* S or Z alleles (c.863A>T [p.Glu288Val] and c.1096G>A [p.Glu366Lys]), respectively) associated with alpha-1-antitrypsin deficiency (see GeneReviews by Stoller et al. in Web Resources) (MIM: 613490). Five participants were carriers for variants in *GALT* that are associated with the Duarte 2 allele<sup>87-89</sup> (including one missense variant and three intronic variants), while



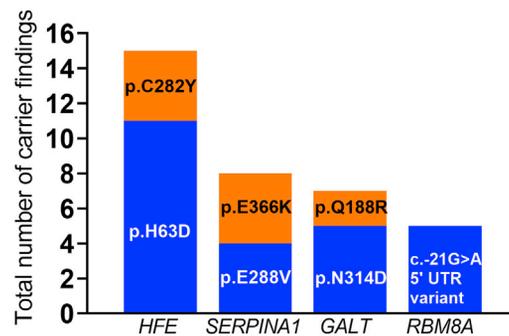
**Figure 1. Number of Carrier Findings for NC NEXUS Participants that Were Randomized into the Experimental Group and Elected to Receive Carrier Findings**

Number of carrier findings is represented on the x axis and the number of NC NEXUS participants is represented on the y axis.

two participants were carriers for a variant associated with classic galactosemia<sup>90,91</sup> (MIM: 230400), c.563A>G (p.Gln188Arg). Five participants carried a hypomorphic variant located in the 5' untranslated region (UTR) in *RBM8A* (c.-21G>A [p.?]), associated with thrombocytopenia-absent radius (TAR) syndrome<sup>92,93</sup> (MIM: 274000). Additionally, we identified three carriers of pathogenic *CFTR* (MIM: 602421) variants associated with cystic fibrosis<sup>94-97</sup> (MIM: 219700); two participants carried the *CFTR* c.1521\_1523del (p.Phe508del) founder variant and one participant carried the *CFTR* c.1519\_1521del (p.Ile507del) variant (Table S2). All variants reported in the NC NEXUS study are listed in Table S3.

## Discussion

The potential use of genome-scale sequencing to screen healthy individuals for monogenic disorders—before symptoms manifest—promises the tantalizing possibility of ameliorating these conditions, but also raises numerous challenges. Among the barriers to the use of genomic sequencing in any screening endeavor is the performance of this technology and our ability to interpret variants in the absence of phenotypic indications. Other critical issues include the kinds of genomic information that should be offered, how parental informed consent can be obtained, and how the overall process and the potential results will impact families. To address these questions, the NC NEXUS study enrolled a cohort of 106 children: 17 with inborn errors of metabolism, 28 with hearing loss, and 61 healthy newborns. We explored the analytical performance of ES in the context of newborn screening, both from a “screening” standpoint as well as in an “indication-based” interpretation mode.



**Figure 2. Variants in *HFE*, *SERPINA1*, *GALT*, and *RBM8A* Were the Most Frequently Observed Carrier Findings**

Variants/predicted protein changes and genes in which they are located are represented on the x axis. Total number of carrier findings in the NC NEXUS study is represented on the y axis.

## Analytic Performance

In order to assess how well ES would perform for predictive purposes, molecular analysts were initially blinded to the cohort status of the individual, enabling us to simulate the conditions of a true screening test, in which the individual is assumed to be at population risk. We did not perform trio analyses, which can be helpful for identifying *de novo* and compound heterozygous variants, although we did conduct follow-up parental sequencing whenever possible where compound heterozygosity was suspected. From the standpoint of implementation workflows, it is reasonable to attempt to obtain trio samples in order to maximize the diagnostic yield of clinical diagnostic sequencing; however, comprehensive trio sample collection would be impractical within current NBS practices.

Screening is unlike indication-based diagnostic testing, in which the analyst can utilize phenotypic information to judge which variants are most likely to be relevant, and where the disclosure of inconclusive results (variants of uncertain significance, single heterozygous variants in genes associated with recessive conditions, etc.) can still be informative and allow for more detailed clinical follow-up. However, in a population at low risk for any given monogenic disorder, disclosure of genomic findings with lower certainty will inevitably lead to an extremely high false positive rate, and we therefore set relatively stringent thresholds for the results to qualify as an abnormal positive screen. This decision reflects the inevitable balance between increasing sensitivity to maximize case finding versus establishing stringent thresholds to reduce false positives. As expected, a small number of participants (4 out of the total 106) had abnormal positive screen NGS-NBS results (beyond those that would be expected for those in the affected cohorts), but the results have implications for their health supervision. These findings exemplify the expansion of the possible range of conditions that could be identified in newborns and thereby influence future health care and screening to ultimately improve clinical outcomes. These findings had implications for other family members, but longitudinal follow-

up of a larger number of screened individuals will be required to assess outcomes, clinical utility, and the economic value of NGS-NBS. Among the affected cohorts, the screening analysis predicted the presence of inborn errors of metabolism in 15 out of 17 individuals and hearing loss in 5 out of 28 individuals. Thus, at a population level, it is unlikely that genomic sequencing could replace current screening based on biochemical measurements or audiology testing.

Rather than being used purely in a screening mode of analysis, genomic information could also be used in conjunction with phenotypic information (such as a metabolite levels) to perform a “secondary” or “indication-based” analysis that may improve the sensitivity and specificity of newborn screening for inborn errors of metabolism. In our study, after determining the “screening” results for a given individual, the analyst was unblinded to the cohort that the individual was from, and a more sensitive diagnostic analysis was conducted. The yield of sequencing improved somewhat in the context of indication-based analysis: the two remaining individuals in the metabolic cohort and several members of the hearing loss cohort had highly suggestive but “inconclusive” results (e.g., a single heterozygous pathogenic variant, a homozygous missense VUS in a gene associated with an autosomal-recessive disease). In the case of the participant with malonyl-CoA decarboxylase deficiency, this phenotype information was valuable for detecting a possible explanation for the child’s deficiency in the indication-based analysis (homozygous *MLYCD* c.1013T>C [p.Leu338Pro] variant). Genomic analysis may also improve the specificity of newborn screening. For example, low total carnitine levels found by TMS may be indicative of primary carnitine deficiency, a condition that can result in encephalopathy, cardiomyopathy, and hypoglycemia; however, carnitine levels may also be influenced by other factors, such as prematurity and/or drug interactions (see GeneReviews by El-Hattab in [Web Resources](#)).<sup>33,34</sup> Detecting pathogenic variants in *SLC22A5* provides an unambiguous molecular diagnosis. These findings suggest that sequencing might be useful as an adjunct to traditional NBS methods, and that with improved detection of variants (such as non-canonical splicing variants, copy number variants, or rearrangements) and more extensive interpretive databases, the positive predictive value of genomic screening may improve.

A potential explanation for the relatively low diagnostic yield of ES for participants in the hearing loss cohort is that numerous factors<sup>98–102</sup> contribute to hearing loss, including non-genetic factors (i.e., environmental, infections, premature birth, etc.) that are estimated to account for approximately 20% of cases of prelingual hearing loss (see GeneReviews by Shearer et al. in [Web Resources](#)). The diagnostic rate in our study likely reflects the genetic heterogeneity of the monogenic forms of hearing loss<sup>103,104</sup> as well as the incomplete knowledge of pathogenic variants. Ascertainment bias also may have led to a larger frac-

tion of the hearing loss cohort having complex syndromic conditions that led the referring providers to consider exome sequencing as a useful diagnostic test. Other hearing loss patients may already have had panel testing, thus depleting the enrolled study population of the more commonly identified nonsyndromic hearing loss conditions. In addition, certain genetic forms of hearing loss are caused by copy number variants or mitochondrial DNA variants (see GeneReviews by Shearer et al. in [Web Resources](#)),<sup>105</sup> which were not assessed in this study. Due to the heterogeneity of the etiologies of hearing loss, it seems clear that phenotypic screening (at birth and during childhood) will remain necessary, even if detection of the genetic mechanisms improves.

On the other hand, many types of hearing loss that would be detectable with genomic sequencing (for example, hearing loss due to pathogenic or likely pathogenic variants in *TECTA* as shown in one NC NEXUS subject) do not have congenital onset and would therefore be missed at birth. The pathogenic *TECTA* variant finding demonstrates the potential for NGS-NBS to detect hearing loss risk in children who are not identified through traditional newborn hearing screening. The identification of a genetic risk for hearing loss in this setting could be followed up with more aggressive audiology evaluations during infancy and early childhood in order to facilitate interventions and reduce the chance of language impairment. In other cases, hearing loss is associated with a broader syndromic condition, for example the NC NEXUS subjects with Warsaw breakage syndrome or Lowe syndrome. These examples raise intriguing points regarding the implementation of genomic screening in newborns, where testing in the context of a specific phenotype can be a powerful diagnostic tool enabling families and clinicians to better manage an individual’s health condition. However, it is unclear whether routine genomic screening of all healthy newborns for such conditions would be useful in a broader context without clinical interventions to ameliorate symptoms; in such cases, parental input will be critical.

### Additional Genomic Findings

One question raised by the prospect of genomic sequencing in newborns is how parents will be engaged in the process of informed decision making. A subsequent manuscript will describe in detail how parents used an online decision aid, factors associated with decisions about whether or not to learn additional genomic findings, and parents’ reactions to learning the results of these additional analyses. From a molecular analysis standpoint, carrier findings reported in this study were similar to other studies.<sup>10,106–109</sup> Interestingly, many of the variants we observed are milder or hypomorphic variants; in order to cause disease, these alleles must be in *trans* with a more severe variant in the same gene (e.g., the *RBM8A* c.–21G>A [p.?] variant and the *HFE* c.187C>G [p.His63Asp] variant). These hypomorphic variants complicate the analysis and the reporting of carrier findings. In addition, identifying

carrier status in a newborn is not fully informative with regards to carrier status in the parents or siblings without further analysis, which decreases the clinical utility of reporting carrier findings in infants.

Other molecular findings predictive of a current or future clinical diagnosis in the patient, which we categorized in this study as childhood-onset nonactionable and adult-onset actionable conditions, can provide clinically relevant information, yet it is unclear whether this information should be sought routinely in the course of newborn screening. In the context of an otherwise healthy newborn, this information could potentially allow parents and providers to prepare for future eventualities, to avoid a “diagnostic odyssey,” or even to alert other family members that they could also have a disease-causing variant that may be immediately relevant to their health, as in the case of hereditary cancer predisposition. However, the long-term impact of this information on clinical outcomes and family psychosocial dynamics is unknown. We also recognize that the screening of healthy adults for medically actionable conditions such as cancer predisposition is becoming increasingly available, thereby reducing the imperative to discover these findings in an infant.

### Conclusions and Future Directions

Clinical applications of genomic sequencing technologies offer great opportunities in both diagnostic testing and screening. In the context of NBS, sequencing-based approaches cannot fully replace biochemical or phenotypic screens because of etiologic heterogeneity and the challenges of variant interpretation. However, sequencing approaches have the advantage of being able to identify virtually any condition with a known genetic cause. Thus, augmentation of newborn screening with some form of genomic sequencing seems inevitable. In the near term, sequencing of genes already associated with conditions that are screened for in traditional NBS may help to improve clinical sensitivity and specificity. Expanding the current NBS panel to include other actionable conditions detectable only by sequencing could further enhance the public health benefits of NBS, if interpretive challenges can be overcome in order to balance case detection versus false positives. Providing additional genomic information beyond the most actionable conditions, while potentially of interest to many parents, may increase the complexity of informed consent and thereby serve to distract from the primary health benefits of NGS-NBS. There will be significant costs, ethical considerations, and implementation challenges involved in conducting genome-scale sequencing in healthy newborns. These challenges may limit broad application across the entire population, thus failing to deliver on the promise of the Human Genome Project. We should therefore continue to explore innovative approaches to NGS-NBS that will enable the most comprehensive adoption in the general population, improve outcomes to the greatest extent

possible, and maximize societal benefits in a cost-effective manner.

### Data and Code Availability

The variants discussed in the paper were submitted to ClinVar and variant accession numbers can be found in Table S3. [ClinicalTrials.gov](https://clinicaltrials.gov/ct2/show/study/NCT02826694) Identifier: NCT02826694.

### Supplemental Data

Supplemental Data can be found online at <https://doi.org/10.1016/j.ajhg.2020.08.001>.

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### Declaration of Interests

D.B.B. is involved in an unrelated research study that receives contributed equipment and reagents from Asuragen and an unrelated research study that receives partial funding from Sarepta Therapeutics. B.C.P. is an investigator on a different research study that receives in-kind support (reagents and sequencing consumables) from Illumina (San Diego, CA). K.E.W. is President and Chair of the Executive Committee and Board of Directors for the Association for Molecular Pathology, a member of the Advisory Committee for the US FDA Medical Devices Molecular and Clinical Genetics Devices Panel, and a member of the Consultant Advisory Panel for BlueCross BlueShield of North Carolina.

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### Web Resources

ClinGen Allele Registry, [http://reg.clinicalgenome.org/redmine/projects/registry/genboree\\_registry/landing](http://reg.clinicalgenome.org/redmine/projects/registry/genboree_registry/landing)

ClinGen Variant Pathogenicity Curation Interface, <https://curation.clinicalgenome.org/>

ClinVar, <https://www.ncbi.nlm.nih.gov/clinvar/submitters/506663>

ExAC Browser, <http://exac.broadinstitute.org/>

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gnomAD Browser, <https://gnomad.broadinstitute.org/>

Mutalyzer, <https://mutalyzer.nl/index>

OMIM, <https://www.omim.org/>

UCSC Genome Browser, <https://genome.ucsc.edu>

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## Supplemental Data

### Genomic Sequencing for Newborn Screening:

#### Results of the NC NEXUS Project

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Self-reported race and/or ethnicity	Percentage
White	66%
Black/African American	10%
Asian	5%
Asian and White	3%
White and Black/African American	3%
White and Native American	3%
White and other (unknown, or not reported, or Latino)	3%
Latina/Latino and Hispanic or Latino	2%
Not reported	2%
Adopted from China	1%
Asian and other	1%
Black/African American and Pacific Islander	1%
Self-reported completed education level	Percentage
Graduate degree (both parents)	26%
Graduate degree (one parent) and four years college (one parent)	20%
Four years college (one parent) and partial or two years college (one parent)	14%
Four years college (both parents)	8%
Graduate degree (one parent) and partial or two years college (one parent)	4%
Two or four years college (one parent) and high school/GED (one parent)	4%
High school or GED (both parents)	4%
High school or GED (mother reported only)	4%
Two years college (one parent) and partial college (one parent)	3%
Partial college (one parent) and high school/GED (one parent)	3%
Partial, two or 4 years of college (mother reported only)	3%
Graduate degree (mother reported only)	2%
Graduate degree (one parent) and high school/GED (one parent)	2%

Four years college (one parent) and completed trade school (one parent)	1%
Partial college (both parents)	1%
High school/GED (one parent) and less than high school (one parent)	1%

**Table S1. Parents' self-reported demographics in the NC NEXUS study.** The demographics were provided by both parents in the study, with the exception of nine individuals in the education level category (indicated by mother reported only). One participant in the NC NEXUS study was adopted from China (as indicated in the table); therefore, their adoptive parents did not report their self-identified race and/or ethnicity information. N=106 NC NEXUS study participants.

NC NEXUS participant	Childhood onset nonactionable category	Adult onset actionable category	Carrier list category	Gene (s)	Disease association with gene (disease inheritance)	Variant and (predicted protein change)	Zygoty in NC NEXUS participant
M001	negative	negative	positive for four variants ( <i>HFE</i> , <i>CFTR</i> , <i>GUSB</i> , <i>AGK</i> ; see gene column)	<i>HFE</i> , <i>CFTR</i> , <i>GUSB</i> , <i>AGK</i>	hemochromatosis (AR); cystic fibrosis (AR); mucopolysaccharidosis VII (AR); Sengers syndrome (AR)	<i>HFE</i> c.845G>A p.(Cys282Tyr); <i>CFTR</i> c.1521_1523del p.(Phe508del); <i>GUSB</i> c.1069C>T p.(Arg357*); <i>AGK</i> c.523_524del p.(Ile175fs)	heterozygous for all four variants listed in variant column
M002	not requested	negative	not requested	N/A	N/A	N/A	N/A
M003	negative	negative	positive for seven variants ( <i>SIL1</i> , <i>CYP21A2</i> , <i>SLC26A4</i> , <i>SERPINA1</i> , <i>ACADM</i> , <i>HPS3</i> , <i>RNASEH2B</i> )	<i>SIL1</i> , <i>CYP21A2</i> , <i>SLC26A4</i> , <i>SERPINA1</i> , <i>ACADM</i> , <i>HPS3</i> , <i>RNASEH2B</i>	Marinesco-Sjogren syndrome (AR); 21-hydroxylase deficiency (AR); Pendred syndrome (AR); Alpha-1 antitrypsin deficiency (AR); Hermansky-Pudlak syndrome (AR), Aicardi-Goutières syndrome (AR)	<i>SIL1</i> c.645+2T>C p.(?); <i>CYP21A2</i> c.1360C>T p.(Pro454Ser); <i>SLC26A4</i> c.2015G>A p.(Gly672Glu); <i>SERPINA1</i> c.1096G>A p.(Glu366Lys); <i>ACADM</i> c.985A>G p.(Lys329Glu); <i>HPS3</i> c.1682_1683del p.(Cys561fs); <i>RNASEH2B</i> c.2T>C p.(Met1Thr)	heterozygous for all seven variants listed in variant column
M005	not requested	negative	positive- <i>RBM8A</i> and <i>HFE</i> variants	<i>RBM8A</i> , <i>HFE</i>	thrombocytopenia-absent radius (TAR) syndrome (AR); hemochromatosis (AR)	<i>RBM8A</i> n.500C>T c.-21G>A p.(?); <i>HFE</i> c.187C>G p.(His63Asp)	heterozygous for both variants listed in variant column
M006	negative	negative	not requested	N/A	N/A	N/A	N/A
M007	negative	positive for <i>RAD51C</i> variant	positive for five variants ( <i>RAD51C</i> , <i>SMARCAL1</i> , <i>TYR</i> , <i>SERPINA1</i> , <i>AARS2</i> )	<i>RAD51C</i> , <i>SMARCAL1</i> , <i>TYR</i> , <i>SERPINA1</i> , <i>AARS2</i>	ovarian cancer (AD); Fanconi anemia (AR); Schimke immunosseous dysplasia (AR); Alpha 1 antitrypsin deficiency (AR); combined oxidative phosphorylation deficiency (AR)	<i>RAD51C</i> c.905-2_905-1del p.(?); <i>SMARCAL1</i> c.2542G>T p.(Glu848*); <i>TYR</i> c.1217C>T p.(Pro406Leu); <i>SERPINA1</i> c.1096G>A p.(Glu366Lys); <i>AARS2</i> c.2599-1G>A p.(?)	heterozygous for all five variants in variant column
M008	negative	negative	positive- <i>SERPINA1</i> variant	<i>SERPINA1</i>	alpha-1 antitrypsin deficiency (AR)	c.863A>T p.(Glu288Val)	homozygous

M009	negative	negative	positive- <i>G6PD</i> variants	<i>G6PD</i>	hemolytic anemia (XL)	c.292G>A p.(Val98Met); c.466A>G p.(Asn156Asp)	heterozygous for both <i>G6PD</i> variants
M011	negative	negative	positive- <i>RPE65</i> and <i>IDUA</i> variants	<i>RPE65</i> , <i>IDUA</i>	retinitis pigmentosa (AR); Leber congenital amaurosis (AR); mucopolysaccharidosis type I (AR)	<i>RPE65</i> c.271C>T p.(Arg91Trp); <i>IDUA</i> c.208C>T p.(Gln70*)	heterozygous for both variants in variant column
M013	negative	negative	positive for variants in <i>HFE</i> , <i>GALT</i> , <i>VWF</i> , <i>KIAA0586</i> , <i>ATP13A2</i>	<i>HFE</i> , <i>GALT</i> , <i>VWF</i> , <i>KIAA0586</i> , <i>ATP13A2</i>	hemochromatosis (AR); galactosemia (AR); von Willebrand disease type 2N (AR or AD); Joubert syndrome and/or short rib thoracic dysplasia with polydactyly (AR); Kufor-Rakeb syndrome and/or spastic paraplegia (AR)	<i>HFE</i> c.845G>A p.(Cys282Tyr); <i>GALT</i> c.378- 27G>C, c.507+62G>A, c.508-24G>A, c.940A>G p.(Asn314Asp); <i>VWF</i> c.2561G>A p.(Arg854Gln); <i>KIAA0586</i> c.428del p.(Arg143fs); <i>ATP13A2</i> c.2135_2136del p.(Val712fs)	heterozygous for all variants listed in variant column
M014	negative	negative	positive- <i>WRN</i> variant	<i>WRN</i>	Werner syndrome (AR)	c.561A>G p.(Lys187fs)	heterozygous
M015	not requested	not requested	positive- <i>HFE</i> variant	<i>HFE</i>	hemochromatosis (AR)	c.187C>G p.(His63Asp)	heterozygous
M017	negative	negative	positive for variants in <i>HFE</i> , <i>PEX1</i> , <i>GALT</i>	<i>HFE</i> , <i>PEX1</i> , <i>GALT</i>	hemochromatosis (AR); Zellweger spectrum disorder (AR); galactosemia (AR)	<i>HFE</i> c.187C>G, p.(His63Asp); <i>PEX1</i> c.2614C>T p.(Arg872*); <i>GALT</i> c.378- 27G>C, c.507+62G>A, c.508-24G>A, c.940A>G p.(Asn314Asp)	heterozygous for the <i>HFE</i> variant and <i>PEX1</i> variant; homozygous for <i>GALT</i> variants
HL001	negative	negative	positive- <i>HFE</i> and <i>PHYH</i> variants	<i>HFE</i> , <i>PHYH</i>	hemochromatosis (AR); Refsum disease (AR)	<i>HFE</i> c.187C>G p.(His63Asp); <i>PHYH</i> c.823C>T p.(Arg275Trp)	heterozygous for both variants in variant column
HL002	negative	negative	positive- <i>GNE</i> variant	<i>GNE</i>	Nonaka myopathy (AR)	c.1985C>T p.(Ala662Val)	heterozygous
HL004	not requested	negative	positive- <i>HFE</i> variant	<i>HFE</i>	hemochromatosis (AR)	c.187C>G p.(His63Asp)	heterozygous
HL006	negative	negative	positive- <i>RBM8A</i> and <i>MMAB</i> variants	<i>RBM8A</i> , <i>MMAB</i>	thrombocytopenia- absent radius syndrome (AR); methylmalonic acidemia (AR)	<i>RBM8A</i> n.500C>T c.-21G>A p.(?); <i>MMAB</i> c.556C>T p.(Arg186Trp)	heterozygous for both variants in variant column

HL007	negative	negative	positive- <i>GALT</i> and <i>F5</i> variants	<i>GALT, F5</i>	galactosemia (AR); Factor V deficiency (AR)	<i>GALT</i> c.563A>G p.(Gln188Arg); <i>F5</i> c.1601G>A p.(Arg534Gln)	heterozygous for both variants in variant column
HL008	negative	negative	positive- <i>SERPINA1</i> variant	<i>SERPINA1</i>	alpha-1 antitrypsin deficiency (AR)	c.863A>T p.(Glu288Val)	heterozygous
HL015	positive- <i>OCRL</i> variant	negative	negative	<i>OCRL</i>	Lowe syndrome (XL)	c.741G>A p.(Trp247*)	Hemizygous
HL016	negative	negative	positive- <i>MMACHC</i> variant	<i>MMACHC</i>	methylmalonic acidemia w/homocystinuria type cblC (AR)	c.331C>T p.(Arg111*)	heterozygous
HL017	negative	negative	negative	N/A	N/A	N/A	N/A
HL019	negative	negative	negative	N/A	N/A	N/A	N/A
HL022	negative	negative	positive- <i>CYP21A2</i> and <i>APTX</i> variants	<i>CYP21A2,</i> <i>APTX</i>	21-hydroxylase deficiency (AR); ataxia w/oculomotor apraxia (AR)	<i>CYP21A2</i> c.1360C>T p.(Pro454Ser); <i>APTX</i> c.837G>A p.(Trp279*)	heterozygous for both variants in variant column
HL023	negative	negative	positive for variants in <i>IGHMBP2,</i> <i>G6PC,</i> <i>SERPINA1,</i> <i>NR2E3,</i> <i>HFE,</i> <i>GALT</i>	<i>IGHMBP2,</i> <i>G6PC,</i> <i>SERPINA1,</i> <i>NR2E3,</i> <i>HFE,</i> <i>GALT</i>	Charcot-Marie-Tooth disease axonal type 2S (AR); spinal muscular atrophy with respiratory distress (AR); glycogen storage disease type I (AR); alpha-1 antitrypsin deficiency (AR); retinitis pigmentosa (AR); enhanced S-cone syndrome (AR); hemochromatosis (AR); galactosemia (AR)	<i>IGHMBP2</i> c.1488C>A p.(Cys496*); <i>G6PC</i> c.562G>A p.(Gly188Ser); <i>SERPINA1</i> c.1096G>A p.(Glu366Lys); <i>NR2E3</i> c.932G>A p.(Arg311Gln); <i>HFE</i> c.845G>A p.(Cys282Tyr); <i>GALT</i> c.378- 27G>C, c.507+62G>A, c.508-24G>A, c.940A>G p.(Asn314Asp)	heterozygous for all variants listed in variant column
HL025	negative	negative	positive- <i>IDUA</i> and <i>F5</i> variants	<i>IDUA, F5</i>	MPS I (AR); Factor V deficiency (AR)	<i>IDUA</i> c.979G>C p.(Ala327Pro); <i>F5</i> c.1601G>A p.(Arg534Gln)	heterozygous for both variants in variant column
HL027	negative	negative	positive for three variants ( <i>GNRHR,</i> <i>ATP13A2,</i> <i>RBM8A</i> )	<i>GNRHR,</i> <i>ATP13A2,</i> <i>RBM8A</i>	Hypogonadotropic hypogonadism 7 w/or w/o anosmia (AR); Kufor-Rakeb syndrome and spastic paraplegia (both AR); thrombocytopenia absent radius (TAR) syndrome (AR)	<i>GNRHR</i> c.785G>A p.(Arg262Gln); <i>ATP13A2</i> c.558- 1G>T p.(?); <i>RBM8A</i> n.500C>T c.-21G>A p.(?)	heterozygous for all three variants in variant column

HL028	negative	negative	positive- <i>SERPINA1</i> variant	<i>SERPINA1</i>	alpha-1 antitrypsin deficiency (AR)	<i>SERPINA1</i> c.1096G>A p.(Glu366Lys)	heterozygous
NB002	negative	negative	negative	N/A	N/A	N/A	N/A
NB003	negative	negative	positive- <i>HFE</i> variant	<i>HFE</i>	hemochromatosis (AR)	c.187C>G p.(His63Asp)	heterozygous
NB004	negative	negative	positive for 4 variants ( <i>CFTR</i> , <i>POLG</i> , <i>SERPINA1</i> , <i>SLC7A9</i> )	<i>CFTR</i> , <i>POLG</i> , <i>SERPINA1</i> , <i>SLC7A9</i>	cystic fibrosis (AR); <i>POLG</i> -related spectrum disorders (AD or AR); alpha-1 antitrypsin deficiency (AR); cystinuria (AR)	<i>CFTR</i> c.1521_1523del p.(Phe508del); <i>POLG</i> c.1399G>A p.(Ala467Thr); <i>SERPINA1</i> c.863A>T p.(Glu288Val); <i>SLC7A9</i> c.544G>A p.(Ala182Thr)	heterozygous for all four variants in variant column
NB005	negative	negative	negative	N/A	N/A	N/A	N/A
NB007	not requested	negative	positive- <i>HFE</i> variant	<i>HFE</i>	hemochromatosis (AR)	c.187C>G p.(His63Asp)	heterozygous
NB008	negative	negative	positive- <i>ERCC6</i> and <i>G6PD</i> variants	<i>ERCC6</i> , <i>G6PD</i>	Cockayne syndrome (AR); hemolytic anemia (XL)	<i>ERCC6</i> c.3952_3953del p.(Arg1318fs); <i>G6PD</i> c.292G>A p.(Val98Met); c.466A>G p.(Asn156Asp)	heterozygous for all variants in variant column
NB011	negative	negative	negative	N/A	N/A	N/A	N/A
NB012	negative	negative	negative	N/A	N/A	N/A	N/A
NB013	negative	negative	positive for four variants ( <i>ARMC4</i> , <i>PFKM</i> , <i>PMM2</i> , <i>CEP290</i> )	<i>ARMC4</i> , <i>PFKM</i> , <i>PMM2</i> , <i>CEP290</i>	Primary ciliary dyskinesia (AR); glycogen storage disease type VII (AR); congenital disorder of glycosylation, type Ia (AR); range of disorders, including Senior-Loken or Joubert syndrome (AR)	<i>ARMC4</i> c.2219G>A p.(Trp740*); <i>PFKM</i> c.496C>T p.(Arg166*); <i>PMM2</i> c.422G>A p.(Arg141His); <i>CEP290</i> c.4437+1G>A p.(?)	heterozygous for all four variants in variant column
NB015	negative	negative	positive for 4 variants in <i>GALT</i>	<i>GALT</i>	galactosemia (AR)	c.378-27G>C, c.507+62G>A, c.508-24G>A, c.940A>G p.(Asn314Asp). (These variants are associated with the Duarte 2 allele).	heterozygous for all variants in variant column
NB017	negative	negative	positive- <i>DYNC2H1</i> variant	<i>DYNC2H1</i>	short-rib thoracic dysplasia (AR)	c.8283del p.(Phe2761fs)	heterozygous
NB018	negative	negative	positive- <i>HFE</i> variant	<i>HFE</i>	hemochromatosis (AR)	c.845G>A p.(Cys282Tyr)	heterozygous

NB020	negative	negative	positive-GAA and CDK5RAP2 variants	GAA, CDK5RAP2	glycogen storage disease type II (AR); primary microcephaly (AR)	GAA c.-32-13T>G p.(?); CDKRAP2 c.4114C>T p.(Arg1372*)	heterozygous for both variants in variant column
NB025	negative	negative	positive-HFE variant	HFE	hemochromatosis (AR)	c.187C>G p.(His63Asp)	heterozygous
NB028	not requested	negative	not requested	N/A	N/A	N/A	N/A
NB031	negative	negative	positive-MCCC1 variant	MCCC1	3-methylcrotonyl-coA carboxylase (3-MCC) deficiency (AR)	c.1483C>T p.(Gln495*)	heterozygous
NB033	negative	negative	positive for four variants (SERPINA1, DARS2, MCPH1, HFE)	SERPINA1, DARS2, MCPH1, HFE	alpha-1 antitrypsin deficiency (AR); leukoencephalopathy with brain stem and spinal cord involvement and lactate elevation (AR); primary microcephaly (AR); hemochromatosis (AR)	SERPINA1 c.863A>T p.(Glu288Val); DARS2 c.493-1G>C p.(?); MCPH1 c.2214+2T>C p.(?); HFE c.187C>G p.(His63Asp)	heterozygous for all four variants in variant column
NB037	negative	negative	positive-USH2A and DOCK8 variants	USH2A, DOCK8	Usher syndrome (AR); hyper-IgE recurrent infection syndrome (AR)	USH2A c.6937G>T p.(Gly2313Cys); DOCK8 c.54-1G>T p.(?)	heterozygous for both variants in variant column
NB040	negative	positive, BRCA2 variant	positive-ASL and DNAH11 variants	BRCA2 (adult onset category), ASL (carrier), DNAH11 (carrier)	hereditary breast and ovarian cancer (AD); argininosuccinic aciduria (AR); primary ciliary dyskinesia (AR)	BRCA2 c.7480C>T p.(Arg2494*); ASL c.544C>T p.(Arg182*); DNAH11 c.6983+1G>A p.(?)	heterozygous for all variants in variant column
NB041	not requested	negative	positive-GALT variant	GALT	classical galactosemia (AR)	c.563A>G p.(Gln188Arg)	heterozygous
NB042	negative	not requested	not requested	N/A	N/A	N/A	N/A
NB043	negative	negative	positive for three variants (BBS1, VWF, CD36)	BBS1, VWF, CD36	Bardet-Biedl syndrome (AR); von Willebrand disease (AD or AR); platelet glycoprotein IV deficiency (AR)	BBS1 c.1169T>G p.(Met390Arg); VWF c.2561G>A p.(Arg854Gln); CD36 c.787_808del p.(Val263fs)	heterozygous for all three variants in variant column
NB044	negative	negative	negative	N/A	N/A	N/A	N/A
NB047	negative	negative	positive for three variants (RBM8A, HFE, SERPINA1)	RBM8A, HFE, SERPINA1	TAR syndrome (AR); hemochromatosis (AR); alpha-1 antitrypsin deficiency (AR)	RBM8A n.500C>T c.-21G>A p.(?); HFE c.187C>G p.(His63Asp); SERPINA1 c.187C>T p.(Arg63Cys)	heterozygous for RBM8A variant and SERPINA1 variant; homozygous for HFE

NB050	negative	negative	positive- <i>F7</i> and <i>HFE</i> variants	<i>F7, HFE</i>	Factor VII deficiency (AR); hemochromatosis (AR)	<i>F7</i> c.1151C>T p.(Thr384Met); <i>HFE</i> c.187C>G p.(His63Asp)	heterozygous for both variants in variant column
NB052	negative	negative	positive- <i>MPL</i> variant	<i>MPL</i>	congenital amegakaryocytic thrombocytopenia (AR)	c.1192del p.(Trp398fs)	heterozygous
NB053	not requested	negative	negative	N/A	N/A	N/A	N/A
NB055	negative	negative	positive for variants in <i>GALT</i> , <i>ATP7B</i> and <i>CFTR</i>	<i>GALT, ATP7B, CFTR</i>	galactosemia (AR); Wilson disease (AR); cystic fibrosis (AR)	<i>GALT</i> c.378-27G>C, c.507+62G>A, c.508-24G>A, c.940A>G p.(Asn314Asp); <i>ATP7B</i> c.1934T>G p.(Met645Arg); <i>CFTR</i> c.1519_1521del p.(Ile507del)	heterozygous for all variants in variant column
NB058	not requested	negative	positive- <i>GJB2</i> variant	<i>GJB2</i>	nonsyndromic hearing loss and deafness (AR)	c.101T>C p.(Met34Thr)	heterozygous
NB060	negative	negative	positive- <i>MYO3A</i> and <i>RBM8A</i> variants	<i>MYO3A, RBM8A</i>	sensorineural hearing loss/deafness (AR); TAR syndrome (AR)	<i>MYO3A</i> c.4681C>T p.(Arg1561*); <i>RBM8A</i> n.500C>T c.-21G>A p.(?)	heterozygous for both variants in variant column

**Table S2: Additional findings in the NC NEXUS cohort.**

The NC NEXUS participant column includes “M” (indicating inborn errors of metabolism cohort), “HL” (indicating hearing loss cohort) or “NB” (indicating healthy/well-child cohort) followed by the participant number. Each row in the table represents one NC NEXUS participant. “Positive” indicates the presence of likely pathogenic or pathogenic variants in genes in an additional findings category (childhood-onset nonactionable, adult-onset actionable, and carrier status). “Negative” indicates the absence of likely pathogenic or pathogenic variants found in genes in the additional findings categories. The “Gene” column indicates the gene(s) in which the additional finding variants were found (if requested). The “Variant and (predicted protein change)” column indicates the coding change in the DNA reference sequence and predicted change of the protein in parentheses. The “Zygoty in NC NEXUS participant” column indicates the zygoty for each variant found in additional findings for each participant (if requested). AR= autosomal recessive pattern of inheritance; N/A= not applicable; XL= X-linked pattern of inheritance; MPS I= mucopolysaccharidosis type I.

Gene	Refseq chromosome reference, Refseq transcript, variant	Chromosome coordinates of variant (hg38)	Canonical allele identifier from ClinGen Allele Registry	Accession number of variants in ClinVar
AARS2	NC_000006.12(NM_020745.3):c.2559-1G>A	chr6:44301465	CA364336148	VCV000873458
ACADM	NC_000001.11(NM_000016.5):c.199T>C	chr1:75732724	CA252836	VCV000003597
ACADM	NC_000001.11(NM_000016.5):c.799G>A	chr1:75749509	CA252824	VCV000003588
ACADM	NC_000001.11(NM_000016.5):c.985A>G	chr1:75761161	CA252821	VCV000003586
AGK	NC_000007.14(NM_018238.3):c.523_524del	chr7:141621734-141621735	CA834306559	VCV000873475
APTX	NC_000009.12(NM_001195248.1):c.837G>A	chr9:32974495	CA253152	VCV000004431
ARMC4	NC_000010.11(NM_018076.4):c.2219G>A	chr10:27936759	CA5453275	VCV000242859
ASL	NC_000007.14(NM_000048.3):c.544C>T	chr7:66086763	CA220291	VCV000092362
ATP13A2	NC_000001.11(NM_022089.3):c.558-1G>T	chr1:17002374	CA338260662	VCV000873473
ATP13A2	NC_000001.11(NM_022089.3):c.2135_2136del	chr1:16991849-16991850	CA915940655	VCV000873463
ATP7B	NC_000013.11(NM_000053.3):c.1934T>G	chr13:51961849	CA252899	VCV000003862
BBS1	NC_000011.10(NM_024649.4):c.1169T>G	chr11:66526181	CA223760	VCV000012143
BCKDHA	NC_000019.10(NM_000709.4):c.1312T>A	chr19:41424582	CA115507	VCV000100009
BRCA2	NC_000013.11(NM_000059.3):c.7480C>T	chr13:32356472	CA025108	VCV000038099
CD36	NC_000007.14(NM_001001547.2):c.787_808del	chr7:80669991-80670012	CA4315410	VCV000873470
CDK5RAP2	NC_000009.12(NM_001011649.2):c.4114C>T	chr9:120419851	CA5213660	VCV000873462
CEP290	NC_000012.12(NM_025114.3):c.4437+1G>A	chr12:88086038	CA6711940	VCV000285948
CEMIP/KIAA1199	NC_000015.10(NM_018689.2):c.58C>T	chr15:80873937	CA7692526	N/A
CFTR	NC_000007.14(NM_000492.3):c.1519_1521del	chr7:117559590-117559592	CA340628	VCV000007106
CFTR	NC_000007.14(NM_000492.3):c.1521_1523del	chr7:117559592-117559594	CA118639	VCV000007105
CYP21A2	NC_000006.12(NM_000500.7):c.1360C>T	chr6:32041006	CA341186	VCV000012159
DARS2	NC_000001.11(NM_018122.4):c.493-1G>C	chr1:173833375	CA1250153	VCV000873471
DNAH11	NC_000007.14(NM_001277115.1):c.6983+1G>A	chr7:21711861	CA4181054	VCV000873476
DOCK8	NC_000009.12(NM_203447.3):c.54-1G>T	chr9:271626	CA4957100	VCV000265359
DSC2	NC_000018.10(NM_024422.6):c.631-2A>G	chr18:31087815	CA022880	VCV000016850
DYNC2H1	NC_000011.10(NM_001080463.1):c.8283del	chr11:103203748	CA601231441	VCV000873461
ERCC6	NC_000010.11(NM_000124.3):c.3952_3953del	chr10:49461382-49461383	CA274713	VCV000190171
F5	NC_000001.11(NM_000130.4):c.1601G>A	chr1:169549811	CA114378	VCV000000642
F7	NC_000013.11(NM_000131.4):c.1151C>T	chr13:113118758	CA7060229	VCV000420160
F11	NC_000004.12(NM_000128.3):c.1489C>T	chr4:186286423	CA3164011	VCV000280137
F11	NC_000004.12(NM_000128.3):c.1608G>C	chr4:186287715	CA3164056	VCV000873465
G6PC	NC_000017.11(NM_000151.3):c.562G>A	chr17:42909418	CA321440	VCV000214465
G6PD	NC_000023.11(NM_000402.4):c.292G>A	chrX:154536002	CA090913	VCV000037123
G6PD	NC_000023.11(NM_000402.4):c.466A>G	chrX:154535277	CA120939	VCV000100055
GAA	NC_000017.11(NM_000152.4):c.-32-13T>G	chr17:80104542	CA116606	VCV000004027
GALT	NC_000009.12(NM_000155.3):c.940A>G	chr9:34649445	CA116380	VCV000003613
GALT	NC_000009.12(NM_000155.3):c.563A>G	chr9:34648170	CA312566	VCV000003614
GJB2	NC_000013.11(NM_004004.5):c.35del	chr13:20189552	CA127023	VCV000017004
GJB2	NC_000013.11(NM_004004.5):c.101T>C	chr13:20189481	CA172206	VCV000017000
GNE	NC_000009.12(NM_001128227.2):c.1985C>T	chr9:36218224	CA253717	VCV000006035
GNRHR	NC_000004.12(NM_000406.2):c.785G>A	chr4:67740682	CA130198	VCV000016024
GUSB	NC_000007.14(NM_000181.3):c.1069C>T	chr7:65974701	CA199717	VCV000000908
HFE	NC_000006.12(NM_000410.3):c.187C>G	chr6:26090951	CA113797	VCV000000010
HFE	NC_000006.12(NM_000410.3):c.845G>A	chr6:26092913	CA113795	VCV000000009

<i>HPS3</i>	NC_000003.12(NM_032383.4):c.1682_1683del	chr3:149157522-149157523	CA2660145	VCV000662551
<i>IDUA</i>	NC_000004.12(NM_000203.4):c.208C>T	chr4:987858	CA204563	VCV000011909
<i>IDUA</i>	NC_000004.12(NM_000203.4):c.979G>C	chr4:1002275	CA234128	VCV000167190
<i>IGHMBP2</i>	NC_000011.10(NM_002180.2):c.1488C>A	chr11:68933864	CA6153662	VCV000234316
<i>KIAA0586</i>	NC_000014.9(NM_001244189.1):c.428del	chr14:58432439	CA204046	VCV000204593
<i>LDLR</i>	NC_000019.10(NM_000527.4):c.502G>A	chr19:11105408	CA023709	VCV000183136
<i>LOXHD1</i>	NC_000018.10(NM_144612.6):c.2914G>A	chr18:46560230	CA8952596	VCV000228822
<i>LOXHD1</i>	NC_000018.10(NM_144612.6):c.3161C>T	chr18:46559503	CA8952552	VCV000873467
<i>MARVELD2</i>	NC_000005.10(NM_001038603.3):c.1183-1G>A	chr5:69432526	CA3294206	VCV000001195
<i>MCCC1</i>	NC_000003.12(NM_020166.4):c.1483C>T	chr3:183037329	CA355320552	VCV000873469
<i>MCPH1</i>	NC_000008.11(NM_024596.4):c.2214+2T>C	chr8:6499931	CA4610864	VCV000873472
<i>MLYCD</i>	NC_000016.10(NM_012213.3):c.1013T>C	chr16:83915020	CA8197484	VCV000432061
<i>MMAB</i>	NC_000012.12(NM_052845.3):c.556C>T	chr12:109561068	CA312714	VCV000003095
<i>MMACHC</i>	NC_000001.11(NM_015506.2):c.331C>T	chr1:45508266	CA251789	VCV000001424
<i>MPL</i>	NC_000001.11(NM_005373.2):c.1192del	chr1:43346818	CA915940656	VCV000873477
<i>MYH14</i>	NC_000019.10(NM_024729.3):c.5942G>C	chr19:50309744	CA406966312	VCV000873466
<i>MYO3A</i>	NC_000010.11(NM_017433.4):c.4681C>T	chr10:26203058	CA5445544	VCV000631635
<i>MYO7A</i>	NC_000011.10(NM_000260.4):c.5824G>A	chr11:77207370	CA132400	VCV000043299
<i>NR2E3</i>	NC_000015.10(NM_014249.3):c.932G>A	chr15:71813573	CA117575	VCV000005532
<i>OCRL</i>	NC_000023.11(NM_000276.3):c.741G>A	chrX:129560568	CA414551955	VCV000873459
<i>OTC</i>	NC_000023.11(NM_000531.6):c.1061T>G	chrX:38421078	CA224450	VCV000097104
<i>PAH</i>	NC_000012.12(NM_000277.2):c.117C>G	chr12:102912842	CA251537	VCV000000605
<i>PAH</i>	NC_000012.12(NM_000277.2):c.194T>C	chr12:102894893	CA251544	VCV000000636
<i>PAH</i>	NC_000012.12(NM_000277.2):c.284_286del	chr12:102894801-102894803	CA251535	VCV000000604
<i>PAH</i>	NC_000012.12(NM_000277.2):c.782G>A	chr12:102852875	CA251528	VCV000000582
<i>PAH</i>	NC_000012.12(NM_000277.2):c.814G>T	chr12:102852843	CA251532	VCV000000596
<i>PAH</i>	NC_000012.12(NM_000277.2):c.842C>T	chr12:102852815	CA220589	VCV000000589
<i>PAH</i>	NC_000012.12(NM_000277.2):c.896T>G	chr12:102851703	CA251541	VCV000000613
<i>PAH</i>	NC_000012.12(NM_000277.2):c.1222C>T	chr12:102840493	CA251523	VCV000000577
<i>PAH</i>	NC_000012.12(NM_000277.2):c.1315+1G>A	chr12:102840399	CA251522	VCV000000576
<i>PEX1</i>	NC_000007.14(NM_000466.2):c.2614C>T	chr7:92499808	CA4341072	VCV000265395
<i>PFKM</i>	NC_000012.12(NM_001166686.1):c.496C>T	chr12:48132913	CA114806	VCV000001158
<i>PHYH</i>	NC_000010.11(NM_006214.3):c.823C>T	chr10:13283695	CA118904	VCV000007580
<i>PMM2</i>	NC_000016.10(NM_000303.2):c.422G>A	chr16:8811153	CA220621	VCV000007706
<i>POLG</i>	NC_000015.10(NM_002693.2):c.1399G>A	chr15:89327201	CA123140	VCV000013496
<i>RAD51C</i>	NC_000017.11(NM_058216.2):c.905-2_905-1del	chr17:58724038-58724039	CA166374	VCV000141768
<i>RBM8A</i>	NC_000001.11(NM_005105.5):c.-21G>A	chr1:145927447	CA249745	VCV000030464
<i>RNASEH2B</i>	NC_000013.11(NM_024570.3):c.2T>C	chr13:50910078	CA388258350	VCV000661979
<i>RPE65</i>	NC_000001.11(NM_000329.2):c.271C>T	chr1:68444858	CA226531	VCV000013115
<i>SERPINA1</i>	NC_000014.9(NM_000295.4):c.187C>T	chr14:94383051	CA325650	VCV000017974
<i>SERPINA1</i>	NC_000014.9(NM_000295.4):c.863A>T	chr14:94380925	CA127679	VCV000017969
<i>SERPINA1</i>	NC_000014.9(NM_000295.4):c.1096G>A	chr14:94378610	CA127676	VCV000017967
<i>SIL1</i>	NC_000005.10(NM_022464.4):c.645+2T>C	chr5:139026799	CA252368	VCV000002624
<i>SLC22A5</i>	NC_000005.10(NM_003060.4):c.506G>A	chr5:132384155	CA312938	VCV000006421
<i>SLC26A4</i>	NC_000007.14(NM_000441.1):c.2015G>A	chr7:107702038	CA261423	VCV000043532
<i>SLC26A4</i>	NC_000007.14(NM_000441.1):c.626G>T	chr7:107674970	CA253304	VCV000004821
<i>SLC26A4</i>	NC_000007.14(NM_000441.1):c.1151A>G	chr7:107690125	CA261398	VCV000004820
<i>SLC7A9</i>	NC_000019.10(NM_014270.4):c.544G>A	chr19:32862521	CA211245	VCV000005782
<i>SMARCAL1</i>	NC_000002.12(NM_014140.3):c.2542G>T	chr2:216478216	CA253017	VCV000004171
<i>SOX10</i>	NC_000022.11(NM_006941.3):c.482G>A	chr22:37978082	CA10228669	VCV000873468
<i>TECTA</i>	NC_000011.10(NM_005422.2):c.5597C>T	chr11:121168064	CA6327783	VCV000236058
<i>TMPRSS3</i>	NC_000021.9(NM_024022.2):c.208del	chr21:42389043	CA273243	VCV000165492
<i>TMPRSS3</i>	NC_000021.9(NM_024022.2):c.1151T>A	chr21:42376584	CA10042329	VCV000873474
<i>TYR</i>	NC_000011.10(NM_000372.4):c.1217C>T	chr11:89284805	CA227519	VCV000003777
<i>USH2A</i>	NC_000001.11(NM_206933.2):c.1256G>T	chr1:216324240	CA252239	VCV000002359
<i>USH2A</i>	NC_000001.11(NM_206933.2):c.2299del	chr1:216247095	CA252226	VCV000002351

<i>USH2A</i>	NC_000001.11(NM_206933.2):c.3686T>G	chr1:216199752	CA344867902	VCV000873464
<i>USH2A</i>	NC_000001.11(NM_206933.2):c.4338_4339del	chr1:216190280-216190281	CA252228	VCV000002353
<i>USH2A</i>	NC_000001.11(NM_206933.2):c.6937G>T	chr1:215970645	CA1394996	VCV000384319
<i>VWF</i>	NC_000012.12(NM_000552.4):c.2561G>A	chr12:6034812	CA114139	VCV000000296
<i>WRN</i>	NC_000008.11(NM_000553.5):c.561A>G	chr8:31067089	CA275342	VCV000198099

**Table S3. Summary table of all variants reported in the NC NEXUS study.**

Table contains columns describing: the gene in which each variant was found (represented by gene symbol); Refseq chromosome reference number, Refseq transcript number and variant; chromosome coordinates of each variant (hg38); the canonical allele identifier of each variant in the ClinGen Allele Registry; the variants' accession numbers in the ClinVar database.