

Supplemental Materials

Molecular Biology of the Cell

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Supplementary Figure 1. (A) Wildtype and *ark1Δ prk1-as* cells were grown to log phase in YPD medium at 30°C and 80 μM 1NA-PP1 was added to both strains. Cells were collected at the indicated time intervals and Ypk-pT662 and Ypk1 protein were assayed by western blot. Error bars represent the standard deviation of the mean of three biological replicates. *** indicates a p-value smaller than 0.005; ** indicates a p-value between 0.01 and 0.005 and * indicates a p-value between 0.05 and 0.01 when the indicated strain is compared to the wildtype. (B) Cells of the indicated genotypes were grown to log phase in YPD medium at 30°C and Ypk-pT662 and Ypk1 protein were assayed by western blot.

Supplementary Figure 2. (A) Wildtype and *ypk1-as ypk2Δ* cells were grown to log phase in YPD medium at 25°C and 50 μM 3MOB-PP1 was added to both strains. Cells were collected after one hour and Ark1 protein was detected by western blot. A fast migrating dephosphorylated form of Ark1 is marked with an asterisk. (B) Wildtype cells were grown to log phase in YPD medium at 30°C in the presence or absence of 0.5 μg/ml myriocin. At the indicated time points, cells were collected and Ark1 protein was detected by western blot. Wildtype and *ypk1-as ypk2Δ* from panel A were loaded as a control.

Supplementary Figure 3. (A) Wildtype and *rts1Δ* cells were grown to log phase in YPD medium at 30°C and phosphorylation of Ark1:3xHA and Prk1:6XHA was analyzed by western blot. (B) Wildtype and *rts1Δ* mutant cells grown in YPD medium to early log phase and were then rapidly washed into YPG/E medium at 30°C. Cells were collected at the indicated time intervals and phosphorylation of Ark1:3xHA and Prk1:6XHA was analyzed by western blot. An asterisk indicates a background band. (C) Same as (B), but phosphorylation was assayed by PhosTag western blot. On the left, wildtype and *rts1Δ* cells were directly compared 10 minutes after a shift to YPG/E. (D) Wildtype and *elm1Δ* cells were grown in YPD medium to early log phase and were then rapidly washed into YPG/E medium at 30°C. Cells were collected at the indicated time intervals and phosphorylation of Ark1:3xHA and Prk1:6XHA was analyzed by western blot. An asterisk indicates a background band. (E) Same as D, but phosphorylation was assayed by PhosTag western blot.

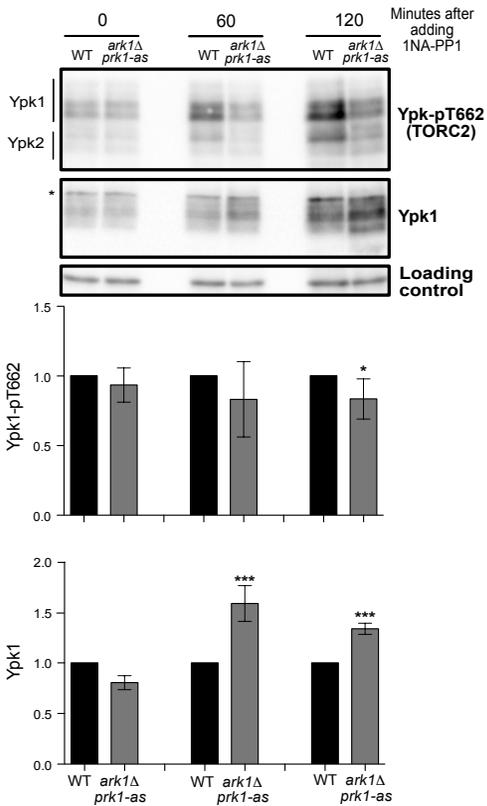
Supplementary Figure 4. (A) A series of 10-fold dilutions of the indicated strains was grown in presence or absence of 15 μM auxin. (B) Wild type and *ent1Δ ent2-AID* cells were grown at 30°C to log phase in YPD medium. 1 mM auxin was added and cells were collected and processed for western blotting with an anti-V5 tag antibody to detect the ent2-AID protein at the indicated time points.

Supplementary Figure 5. (A) Wildtype and *ent1Δ ent2-AID* cells were grown at 30°C to log phase in YPD medium in the presence of 10 μM auxin. As a control, the *ent1Δ ent2-AID* cells were also grown in the presence of the solvent for auxin (ethanol). Cells were collected and Ypk-pT662 and Ypk1 protein were analyzed by western blot. (B) A series of 10-fold dilutions of the indicated strains were grown at 30°C on YPD in the presence or absence of 0.25 μg/ml Myriocin. (C and D) A series of 10-fold dilutions of the indicated strains were grown at 30°C on YPD or YPG/E in the absence or presence 10 μM (C) or 500 μM (D) auxin. (E) A series of 10-fold dilutions of the indicated strains were grown at 30°C on YPD or YPG/E.

Supplementary Figure 6. Protein extracts were made from strains of the indicated genotypes and were used for Western blotting with an anti-Ypk1 antibody. Background bands are marked with an asterisk.

Figure S1

A



B

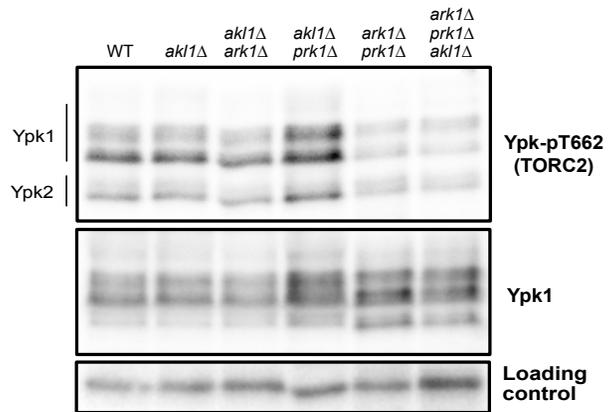
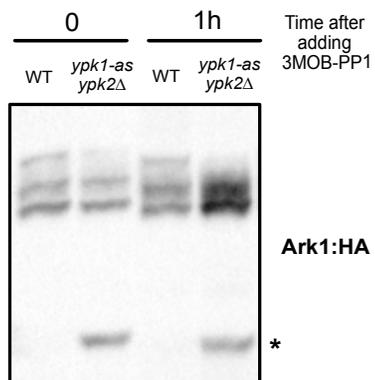


Figure S2

A



B

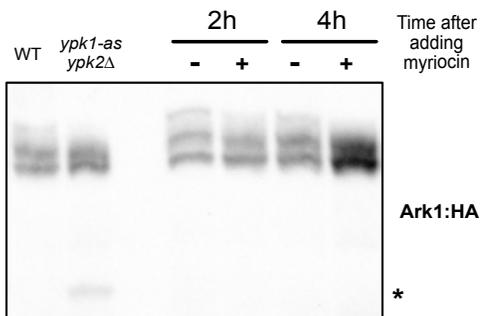
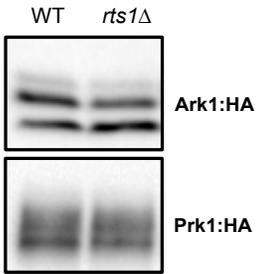
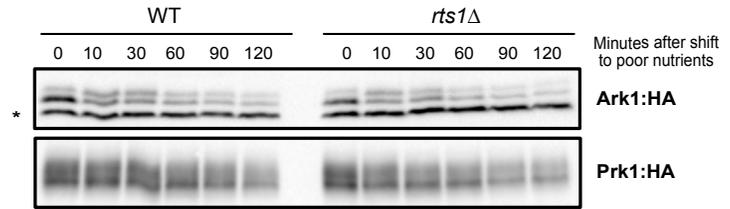


Figure S3

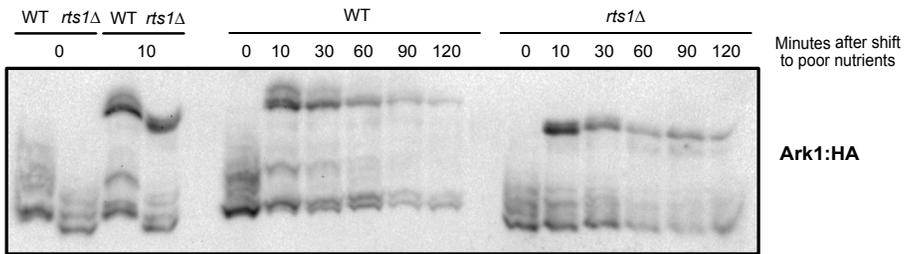
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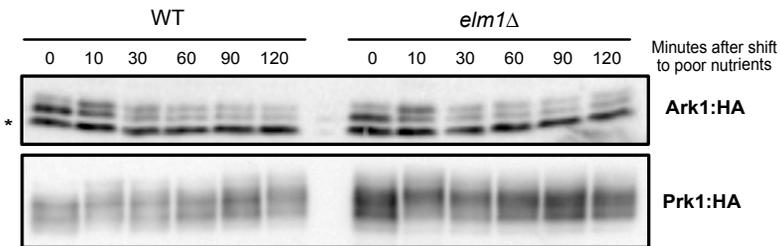
B



C



D



E

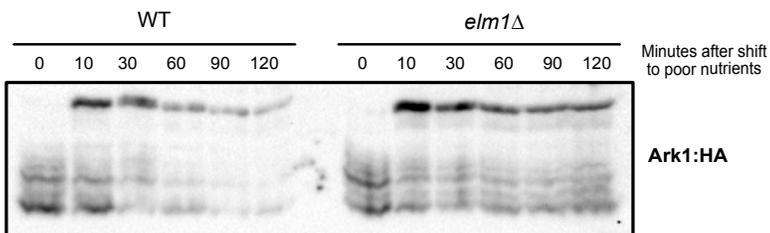
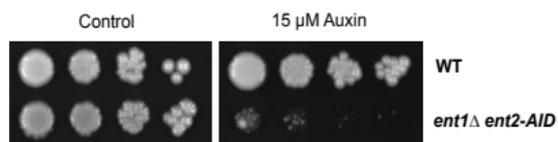


Figure S4

A



B

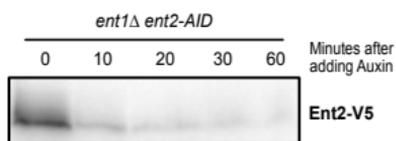
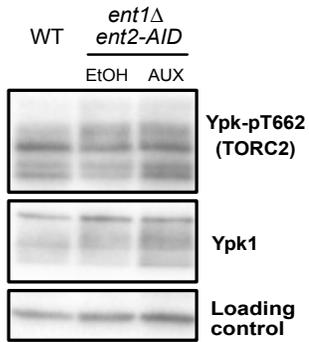
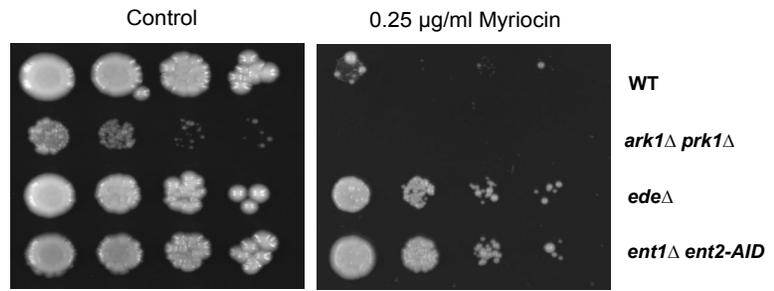


Figure S5

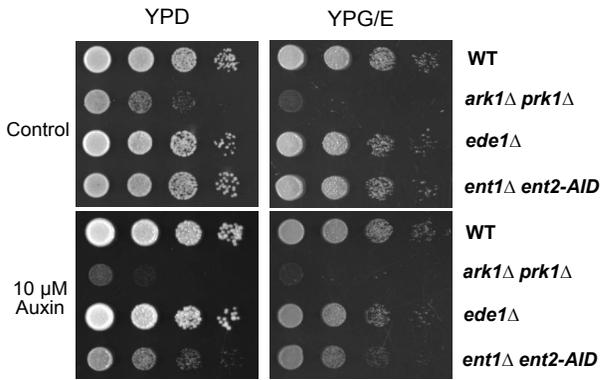
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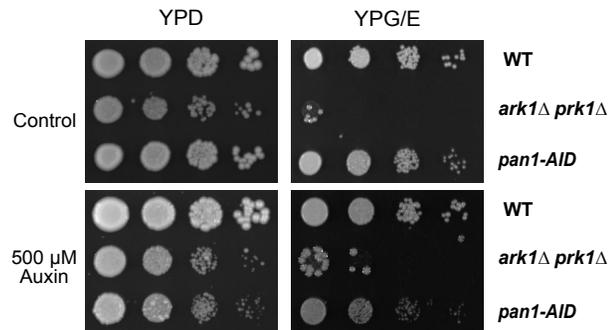
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C



D



E

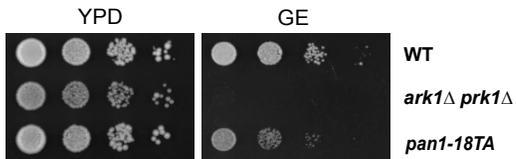


Figure S6

