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Supplementary Information for

Longin R-SNARE is retrieved from the plasma membrane by ANTH domain-containing proteins in *Arabidopsis*

Masaru Fujimoto, Kazuo Ebine, Kohji Nishimura, Nobuhiro Tsutsumi, and Takashi Ueda

Takashi Ueda
Email: tueda@nibb.ac.jp

This PDF file includes:

Figures S1 to S7
Table S1

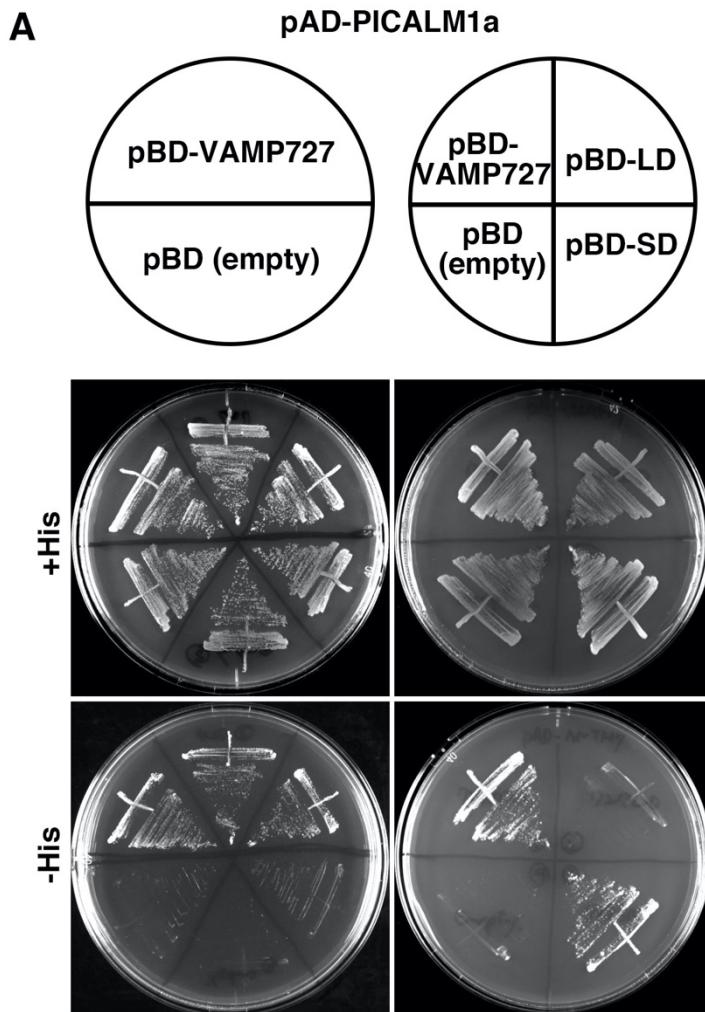


Fig. S1. PICALM1a interacts with the SNARE domain of VAMP727. (A) PICALM1a was expressed as a fusion protein with a transcriptional activation domain (pAD), and the cytoplasmic domain (left) or the longin (LD) or SNARE domain (SD) (right) of VAMP727 was expressed as a fusion protein with the DNA-binding domain (pBD). Interactions between two proteins were tested using the *HIS3* reporter gene.

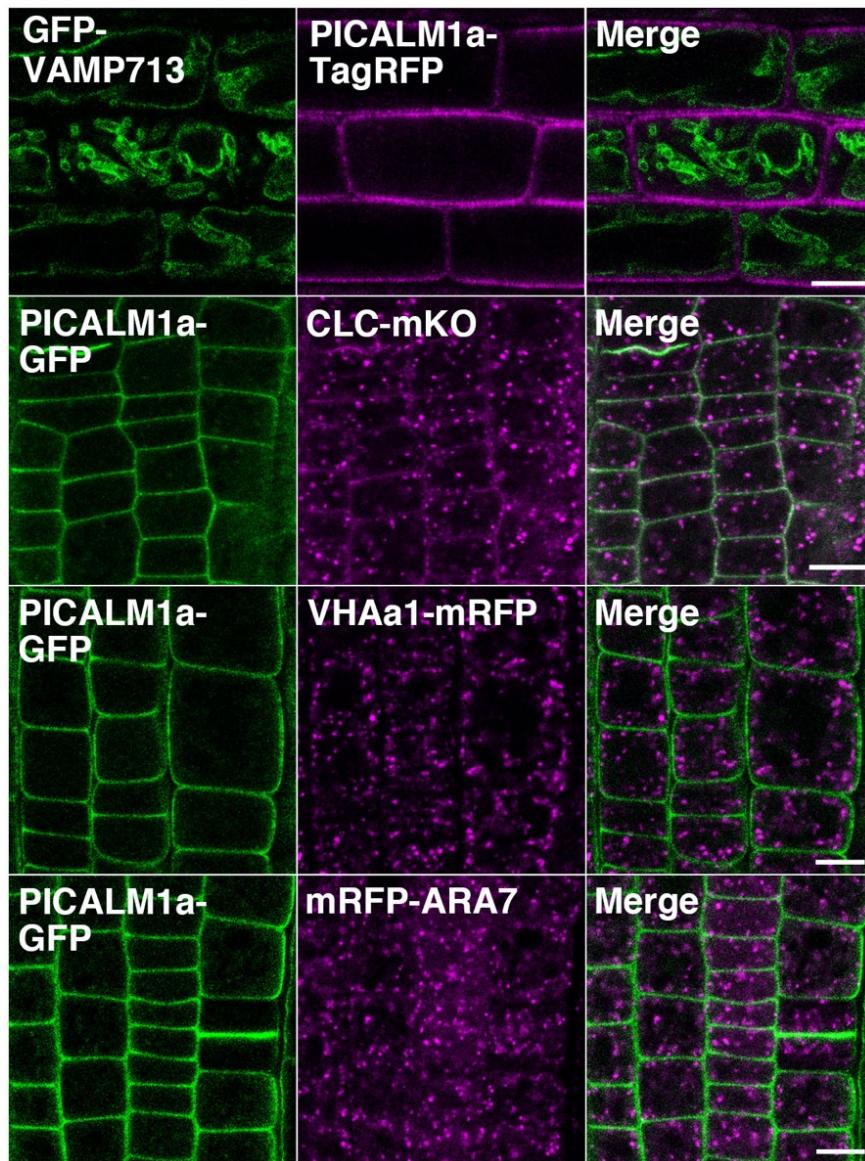


Fig. S2. Comparison of subcellular localization between PICALM1a and organelle markers.

The subcellular localization was compared between fluorescently tagged PICALM1a and GFP-VAMP713 (vacuolar membrane), CLC-mKO, VHAa1-mRFP (TGN), or mRFP-ARA7 (multivesicular bodies) in root epidermal cells of 7-day-old plants by confocal microscopy. Scale bars = 10 μ m.

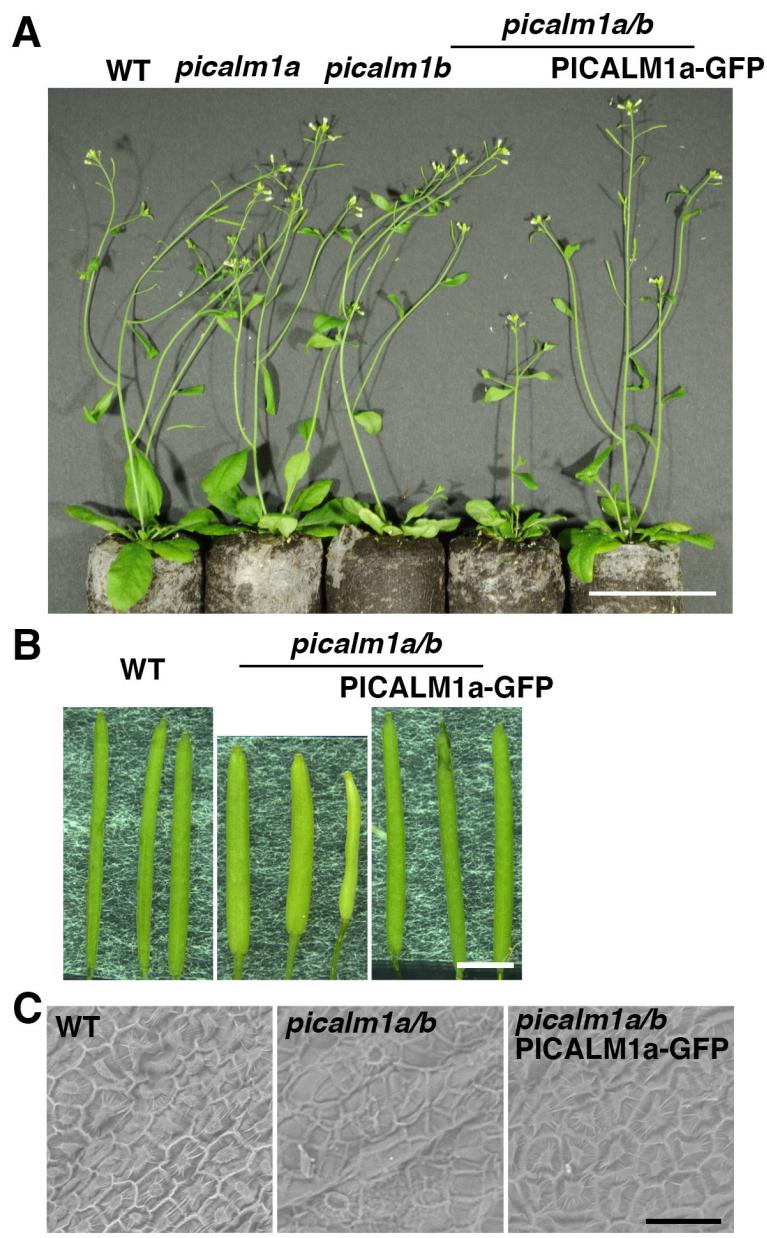


Figure S3. *picalm1* mutants exhibit semi-dwarfism. (A) Forty-day-old *Arabidopsis* plants of wild-type (WT), *picalm1a* and *picalm1b* single mutants, *picalm1a* *picalm1b* (*picalm1a/b*) double mutant, and double mutant transformed with *PICALM1a-GFP*. (B) Siliques collected from the plants with the indicated genotypes. (C) Images of mucilage secretory cells in the seed coat of dry seeds with indicated genotypes. Scale bars = 5 cm (A), 3 mm (B), or 50 μ m (C).

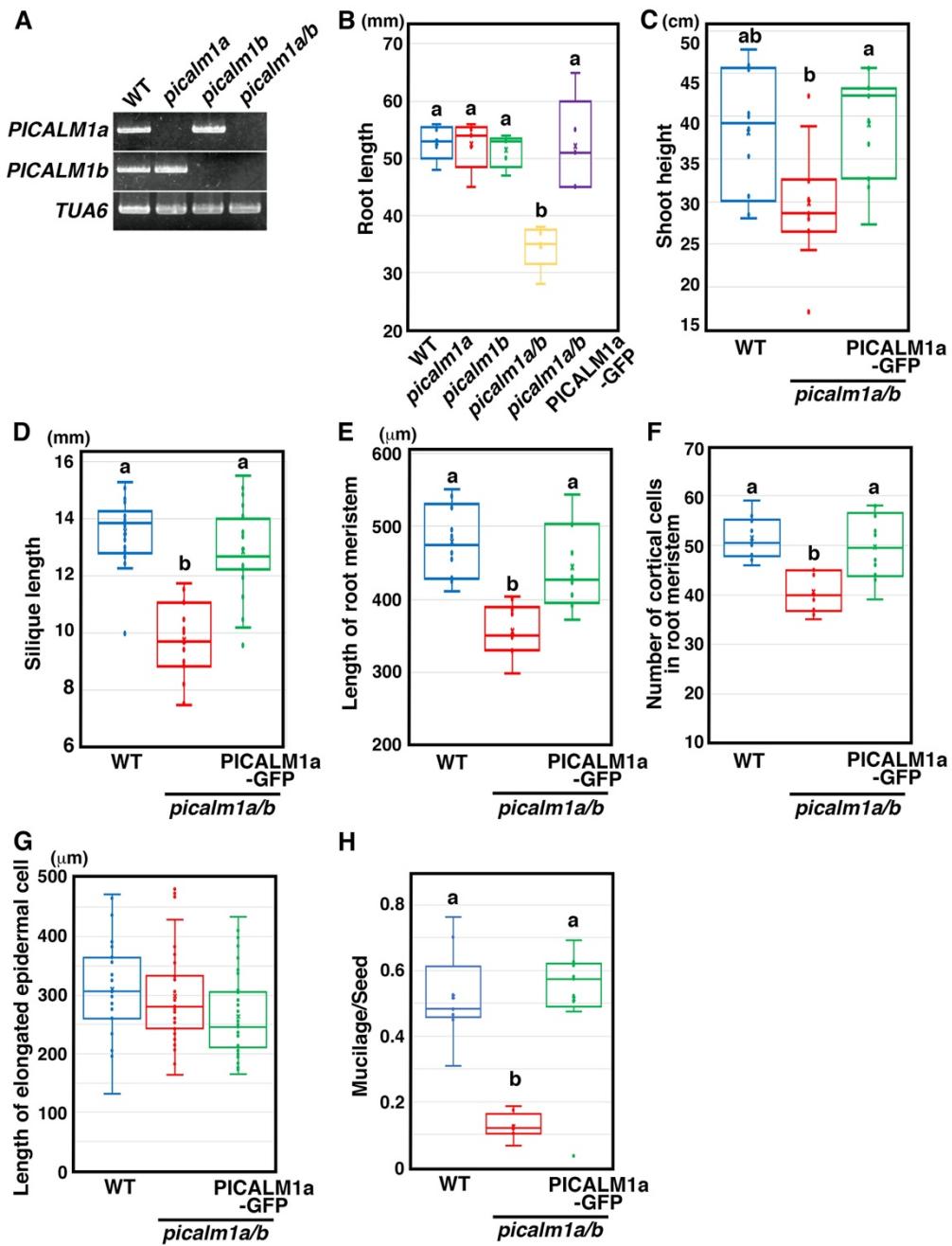


Figure S4. Characterization of *picalm1* mutants. (A) Expression analysis of the *PICALM1* genes by RT-PCR in wild-type and *picalm1* mutant plants. (B) Quantification of root length of 10-day-old *Arabidopsis* seedlings with the indicated genotypes grown on 1/2 MS agar medium. One-way ANOVA followed by Tukey's HSD post-hoc test was used for statistical analyses ($n = 5$ seedlings). (C) Quantification of the height of fully grown plants with the indicated genotypes. One-way ANOVA followed by Steel–Dwass test was used for statistical analyses ($n = 10$ plants for WT and 11 plants for *picalm1a/b* and *picalm1a/b* with *PICALM1a-GFP*). (D) Quantification of the length of siliques collected from *Arabidopsis* plants with the indicated genotypes. One-way

ANOVA followed by Steel–Dwass test was used for statistical analysis (n = 9 siliques for WT and *picalm1a/b* and 12 siliques for *picalm1a/b* with *PICALM1a-GFP*). (E and F) Quantification of the meristem length of roots (E) and number of cortical cells in the root meristem (F) of 6-day-old *Arabidopsis* seedlings with the indicated genotypes grown on 1/2 MS agar medium. One-way ANOVA followed by Steel–Dwass test was used for statistical analysis (n = 10 roots for each genotype). (G) Quantification of the length of fully elongated epidermal cells without a root hair of 6-day-old *Arabidopsis* seedlings with the indicated genotypes grown on 1/2 MS agar medium. Significant difference was not detected among the three groups by the one-way ANOVA. (n = 23 cells for WT, 31 cells for *picalm1a/b*, and 34 cells for *picalm1a/b* with *PICALM1a-GFP* for G) (H) Ratio of the area of mucilage to the seed calculated from images of ruthenium red-stained imbibed seeds with the indicated genotypes. One-way ANOVA followed by Steel–Dwass test was used for statistical analysis (n = 9 seeds for WT, 8 seeds for *picalm1a/b*, and 9 seeds for *picalm1a/b* with *PICALM1a-GFP*).

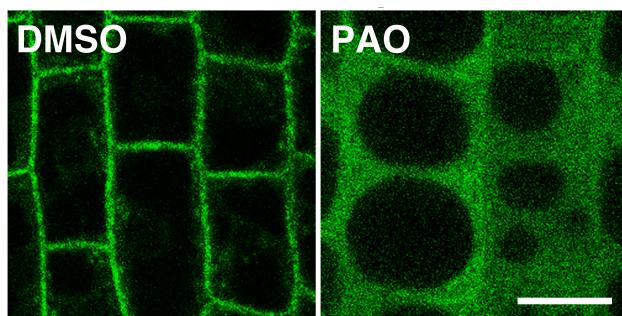


Fig. S5. Effect of phenylarsine oxide (PAO) on PICALM1a-GFP localization. Confocal images of root epidermal cells of 7-day-old *Arabidopsis* plants expressing PICALM1a-GFP treated with DMSO or PAO.

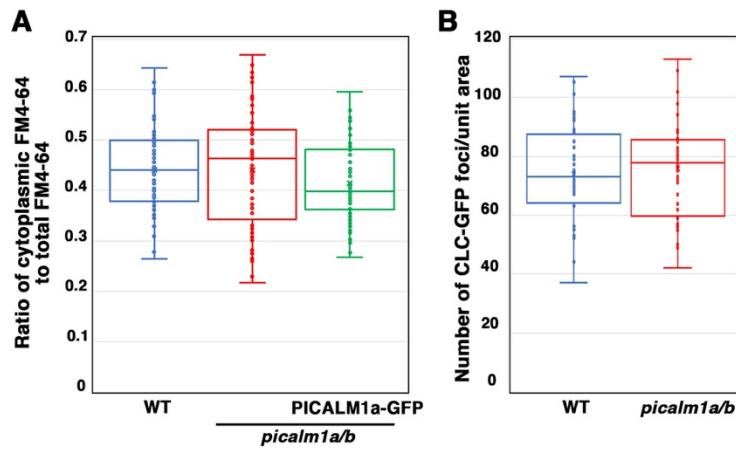


Fig. S6. *picalm1a/b* mutation does not considerably affect general endocytosis. (A)

Quantification of the ratio of internalized to total fluorescence from FM4-64 in root epidermal cells of *Arabidopsis* with the indicated genotypes. n = 51 cells in 5 plants for wild type (WT), 81 cells in 8 plants for *picalm1a/b*, and 66 cells in 6 plants for *picalm1a/b* complemented with PICALM1a-GFP. Significant difference was not detected among the three groups by the one-way ANOVA.

(B) The number of CLC-GFP foci in 100 μm^2 of PM areas in the root epidermal cells of wild-type (WT) and *picalm1a/b* mutant plants observed by VIAFM. n = 44 unit areas in 4 plants for wild type (WT) and 36 unit areas in 4 plants for *picalm1a/b*. Significant difference was not detected using Wilcoxon rank sum test.

Fig 1C, α His

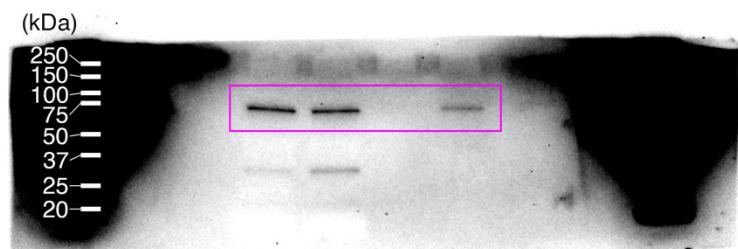


Fig 1C, α GFP



Fig 1D, α CHC

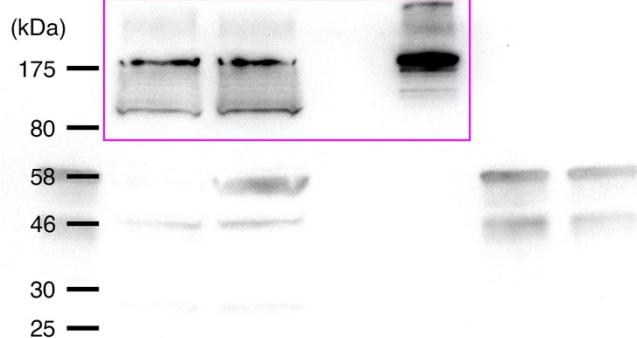


Fig 1D, α GFP

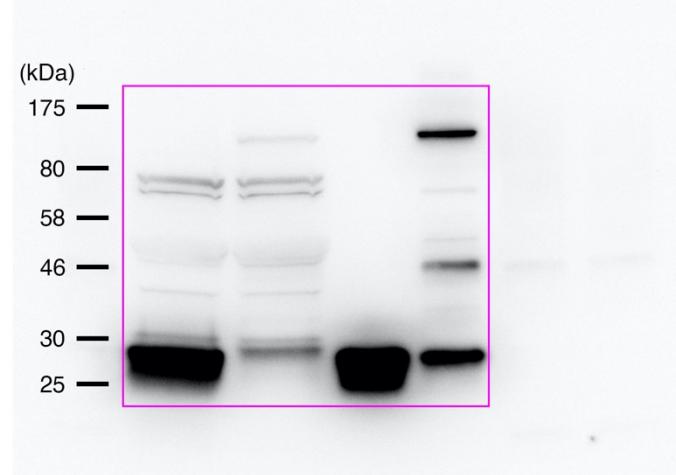


Fig. S7. Full blots.

Table S1. Nucleotide primers used in this study

Genotyping		
PICALM1a	At2g01600-1 for	CTGATCCTCCAACCTCCTCATTTGG
	At2g01600-1 rev	TCAAATGAGGCCAGTGCTACGGA
	LBb1-2	GACCGCTTGCTGCAACTCTCA
Plasmid construction		
PICALM1a genomic / pENTR/D-TOPO	A7g TP fw	caccCAGTGGCCAATTACGTAGTTAGTG
	ANTH7 del stop rv	AATGAGGCCAGTGCTACGGAAC
PICALM1a genomic sequence	A7g s1f	CGGACAGATATTAAAGCCATGAATG
	A7g s2f	GCGAATA CGCCGTACGCTCTC
	A7g s3f	CTCCTTCATTTGTGGGTATATGTG
	A7g s4f	CACA ACTGGACACAGTCGGTG
	A7g s5f	CTTTTCTGACAACAATGGAGGAG
	A7g s6f	CTGATCTCTGTGTTCCAGCTG
	A7g s7f	TGTCTACGGTGCACCAGCTC
PICALM1b genomic / pENTR/D-TOPO	ANTH8 pro TOPO fw	CACCctctccatagactccgacg
	ANTH8 ORF rvG 2	CAAGAAGCCC GTGCTGCCAAGGGTTGCTGTTATG
PICALM1b genomic sequence	A8g sf1	gaattgggcttgcataccatac
	A8g sf2	ctaatcaaaggatataattggacgctag
	A8g sf3	ggaaaagaaatataagaatgagacg
	A8g sf4	GCGATTAGGCCTCGAGCAGATGTTG
	A8g sf5	CACTTATGCGTTGTTCTTGAG
	A8g sf6	cataattacatatatccgctag
	A8g sf7	GCAAAGGATTAGAACTTGCAAGG
	A8g sf8	gtctctgattctcagtattacacacag
	A8g sf9	CAATCCGTTGAGGTCCAAGACCTG
mRFP-CESA2 genomic / pENTR/D-TOPO	CACC-pCESA2 f.	caccGACAGATTGTGAGTGCGAACACACATA
	CESA2 genomic r.	AGCCGTTCGATCAGGTTAAG
	linker-CESA2 f.	GGAGGTAGTGGCGGCAACTGGTGGTCGGCTCAT

	pCESA2-mRFP r. mRFP f. mRFP-linker r.	CTCGGAGGAGGCCATGATGTCTTCTACACCGA ATGGCCTCCTCCGAGGACGT GCCGCCACTACCTCCTCCGGCGCCGGTGGAGTGGCGGC
CESA2 genomic sequence	CESA2 seq1 CESA2 seq2r. CESA2 seq3 CESA2 seq4 CESA2 seq5	ACTCTTACTAGTGCATGCT GTAAACACATACTGAGGTGCAG GGGAACGTCGTGCTATGAA TATGTGTTCATGATGGACC TTTGGTTCTGTCTTGTGA
Y2H Screening		
Library sequence	M5ADLD_scr_amp M3_ins_scr_amp T7_seq_primer	ctattcgatgatgaagatacccccaccaaaccaaaaaaaaagag AGTGAACCTTGCGGGGTTTTCACTATCTACGAT aatacgactcaactatgggc
VAMP721 / pGBK7	VAMP721 fw for pGBK7 VAMP721SN rv for pGBK7	CATGGAGGCCGAATTCATGGCGCAACAATCGTTGATC GCAGGGTCGACGGATCCTCACATGTTCTGAAGGCCACATCTTC
VAMP727 / pGBK7	727 fw for pGBK7 727 SN rv for pGBK7	CATGGAGGCCGAATTCATGAGTCAAAAGGGTTGATATATAGC GCAGGGTCGACGGATCCTCACAGACTCTGTAGGCCACATCTTC
sequence	pGBK7 sf pGBK7 sr pGADT7 sf pGADT7 sr	GAAGAGAGTAGTAACAAAGGTC CGGAATTAGCTTGGCTGCAAG CTATTGATGATGAAGATAACCCAC AGATGGTGCACGATGCACAG
Y2H assay		
VAMP721 / pENTR/D-TOPO	VAMP721 ORF TOPO fw2 VAMP721 SN TOPO fw VAMP721 SN rv AtVAMP721 LD rv	caccATGGCGCAACAATCGTTGATCTACAGTTCTAGCTCGCG caccCATCCTGATGAGATTAGCAAG TCACATGTTCTGAAGGCCACATCTTC TTAACCATGCAATACTGCATATGCTC
VAMP727 / pENTR/D-TOPO	VAMP727 ORF TOPO fw2 727 SN rv 727 SN topo fw 727 LD rv	caccATGAGTCAAAAGGGTTGATATATAGC TCACAGACTCTGTAGGCCACATCTTC caccCATCCAGAAGAGATGAGCAAACCTTC TCAGCTCATACAATACTGCATATGCTC
VAMP713 / pENTR/D-TOPO	TPA713Fw TPA713-TMDRv	caccATGGCGATCATATTGCGTTG TTATGTAAGCTTGACATTCTC
EPSIN1 / pENTR/D-TOPO	At5g11710 topo fw	caccATGGATTTCATGAAGGTCTCGATC

	At5g11710 rv	TCACTGCTTAAAGCCACCAAGATTG
EPSIN2 / pENTR/D-TOPO	AT2G43160 TOPO fw	caccATGAAGAAAGTCTCGGACAAACTG
	AT2G43160 rv	TTACCGGTATCCACCACCATAGGATTGTTG
EPSIN3 / pENTR/D-TOPO	At3g59290 topo fw	caccATGAAGAAAGCATTGGTCAAACCTG
	At3g59290 rv	TTACCGGTAAACCTCCACCATATGG
PICALM5a/ECA2 / pENTR/D-TOPO	At1g03050 topo fw	caccATGGGTTCGAGTAAGTTAAACGAG
	At1g03050 rv	TCAATATTGAGGTGTGAAGAGTAAGGTTG
PICALM5b / pENTR/D-TOPO	At4g02650 topo fw	caccATGGGTTCAAGTAAGCTAAAACGTG
	At4g02650 rv	TCAATAGCGAGGCCTGTAAAGAG
PICALM4a/ECA4 / pENTR/D-TOPO	At2g25430 topo fw	caccATGGCTCCGAGTATTGAAACG
	At2g25430 rv	TCAGTAAGGATTGTTGTAGTAATACCCCG
PICALM6/AP180 / pENTR/D-TOPO	At1g05020 topo fw	caccATGCCGAGCAAGCTAAAAAAGC
	At1g05020 rv	TCAACTCAAGTGCTGGCTATGATC
PICALM3 / pENTR/D-TOPO	At5g35200 topo fw	caccATGTCAGGTGGTGGATC
	At5g35200 rv	TTACATATACCTGGATATGGGTTGAAAC
PICALM2b / pENTR/D-TOPO	At4g25940 topo fw	caccATGCCAACGTTAACAGCTTCC
	At4g25940 rv	CTAGAGGAGCATATGATGATGGTTG
PICALM1a / pENTR/D-TOPO	AT2G01600 TOPO fw	caccATGGGAACGCTACAGTCATGG
	AT2G01600 rv	TCAAATGAGGCCAGTGCTACG
PICALM1b / pENTR/D-TOPO	At1g14910 topo fw	caccATGGGAACGCTACAGTCATGG
	At1g14910 rv	TCACAAGAAGCCCGTGCTGC
PICALM2a / pENTR/D-TOPO	ANTH10_TOPO_fw_3	caccATGGGAACGTTCACGAGCTTCGAAAAGCCTATG
	ANTH10_TOPO_rv_3	CTAGAGAAGCATATGATTAGGACTGTATTGCTGCTGTGA
Sequencing	E1 s1f	CTGGTTCTCAGCTTCATTG
	E1 s2f	AGCTCCTCCGACGGTTGATC
	E2 s1f	GATTACGGAAGCCGGGATGAAG
	E2 s2f	CTCAGAAAGTTGAGACTTCGATG
	E2 s3f	GCTACATCTCAGAGTGGTATTG
	E2 s4f	TCATCAGTCTGGGCAGACAC
	E3 s1f	CCGCTATGGAGACAGAGATG
	E3 s2f	GAATCATCTCCTCAGCAAGTTGAG
	E3 s3f	CACCGAACTTATCCGTCTTCC

E3 s4f	GACACAGCAACCTTATGGTGTG	
E3 s5f	CAAATCCATTGGCGGATATAGGAG	
A1 s1f	TCTCCACGGCTATCGTGGTC	
A1 s2f	GATATGAACGCAATAAAGGCCTAC	
A2 s1f	GGAAGATGATCATCGAGGGAC	
A2 s2f	CAAATCTGAGGAGGGAGGAGGC	
A3 s1f	AGAGATGATTCAGGTCTCCAC	
A3 s2f	GTGTGGCGAGATCGTCTGAG	
A3 s3f	CAGACAGCCGCTTGGTG	
A4 s1f	CAGCACCAACTCAACAACAAG	
A4 s2f	CCACGATTGATACGTCGGAG	
A5 s1f	GACACTCCTGATTGCTCGAG	
A5 s2f	CAGAAGCAGAGAACGCCTGTAG	
A6 s1f	CAAGGATGCTCTCCGATGAAG	
A6 s2f	CCAGCCGCTTATCGAAGAAG	
A7 orf s1	CTTGAGGAACGGCTTGAATGCTTC	
A7 orf s2	GGCCGGATGATGGACTGACTACCG	
A7 orf s3	GAGGGTTAGATACGCTTACACTTAGCAG	
A8 s1f	AAAACTCTGGAACAGTTGCCAG	
A8 s2f	TCCCAGTCTCCTCCTTCAGTAG	
A8 s3f	GCCACAGCTACAGGTTGCAC	
ANTH10_sf1	AGAAGAGCGGCTTGAATGTTATC	
ANTH10_sf2	AGTACATTAAGAACGCGCCTC	
ANTH10_sf3	ACAACAATCCTCGTCCCGTTAG	
RT-PCR		
PICALM1a	ANTH7 RT fw	ATGGGAACGCTACAGTCATGGAG
	ANTH7 RT rv	TCAAATGAGGCCAGTGCTACGG
PICALM1b	ANTH8 RT fw	ATGGGAACGCTACAGTCATGGC
	ANTH8 RT rv	TCACAAGAACGCCGTGCTGC
TUA6	TUA6 RT fw	ATGAGAGAGTGCATTCGATCCAC
	TUA6 RT rv	TTAGTATTCCCTCCTTCATCATCCTCATC