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Hydrolyzable vs. Condensed Wood Tannins for Bio-based **Antioxidant Coatings**

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Figure S1. Comparison of the UV-vis spectra at 2 h of 0.5 mg/mL (a) QT and (b) CT solutions in the presence or absence of the nylon filter.

Sample	mg of tannins adsorbed/mg substrate (after 2 h)		
QT (0.5 mg/mL) in H2O	0.28 ± 0.01		
QT (0.1 mg/mL) in H2O	0.031 ± 0.001		
QT (0.02 mg/mL) in H2O	0.0061 ± 0.0003		
QT (0.1 mg/mL) + laccase in H2O	0.031 ± 0.002		
QT (0.1 mg/mL) + laccase in phosphate buffer pH 6.0	0.021 ± 0.001		
QT (0.1 mg/mL) in carbonate buffer pH 9.0	0.021 ± 0.001		
CT (0.5 mg/mL) in H ₂ O	0.26 ± 0.01		
CT (0.1 mg/mL) in H ₂ O	0.042 ± 0.002		
CT (0.02 mg/mL) in H ₂ O	0.041 ± 0.002		
CT (0.1 mg/mL) + laccase in H2O	0.041 ± 0.002		
CT (0.1 mg/mL) + laccase in phosphate buffer pH 6.0	0.032 ± 0.002		
CT (0.1 mg/mL) in carbonate buffer pH 9.0	0.011 ± 0.001		
1 Reported are the mean \pm SD values of at least three experiments.			

Table S1. Amounts of tannin adhered to the nylon membrane filter under the different coating conditions.^{1.}

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Figure S2. Correlation between the amounts of tannin adhered to the nylon membrane filters and the initial concentrations of (a) QT and (b) CT in water.



Figure S3. Correlation between the percentages of DPPH reduced after 10 min (left) and 2.5 h (right) and the amounts of **(a)** QT and **(b)** CT adhered to the nylon membrane filters.



Figure S4. UV-vis spectra of (**a**) released QT and (**b**) released CT from the nylon filters after washing in ethanol for 2.5 h.

Sample	nmol Trolox/mg filter	nmol Trolox/mg filter
	(after 10 min)	(after 2.5 h)
QT (0.5 mg/mL) in H2O	6.9 ± 0.7^{a}	32 ± 3^{a}
QT (0.1 mg/mL) in H2O	$5.8 \pm 0.6^{\mathrm{a,b}}$	$29 \pm 3^{a,b}$
QT (0.02 mg/mL) in H2O	5.5 ± 0.5^{b}	$25 \pm 2^{b,c}$
QT (0.1 mg/mL) + laccase in H2O	$4.8 \pm 0.5^{\rm b,c}$	$26 \pm 3^{a,b,c}$
QT (0.1 mg/mL) + laccase in phosphate buffer (pH 6.0)	11 ± 1^{d}	$26 \pm 3^{a,b,c}$
QT (0.1 mg/mL) in carbonate buffer (pH 9.0)	5.2 ± 0.5^{b}	$25 \pm 2^{b,c}$
QT (0.1 mg/mL) + FeSO ₄ in H ₂ O	$5.8 \pm 0.6^{\mathrm{a,b}}$	$28 \pm 3^{a,b}$
CT (0.5 mg/mL) in H2O	10 ± 1^{d}	35 ± 4^{a}
CT (0.1 mg/mL) in H2O	9 ± 1^{d}	33 ± 3^{a}
CT (0.02 mg/mL) in H ₂ O	6.1 ±0.7 ^{a,b}	22 ± 2^{c}
CT (0.1 mg/mL) + laccase in H ₂ O	$4.9 \pm 0.5^{\mathrm{b,c}}$	$25 \pm 3^{b,c}$
CT (0.1 mg/mL) + laccase	$4.2 \pm 0.4^{\circ}$	21 ± 2^{c}
in phosphate buffer (pH 6.0)		
CT (0.1 mg/mL)	3.1 ± 0.3^{e}	12 ± 1^{d}
in carbonate buffer (pH 9.0)		
CT (0.1 mg/mL) + FeSO4 in H2O	5.9 ±0.6 ^{a,b}	$26 \pm 3^{a,b,c}$

Table S2. Trolox equivalents determined for the tannin-coated nylon membrane filters in the FRAP assay.^{1.}

Reported are the mean ± SD values of at least three experiments. Values in a column without a common letter are significantly different (P< 0.05).



Figure S5. Correlation between the Fe³⁺-reducing properties of the functionalized substrates after 10 min (left) and 2.5 h (right) and the amounts of (a) QT and (b) CT adhered to the substrate.



Figure S6. UV-vis spectra of (**a**) QT and (**b**) CT solutions in 0.1 M carbonate buffer (pH 9.0), before and after addition of different additives.



Figure S7. UV-vis spectra of (**a**) QT and (**b**) CT solutions in 0.05 M phosphate buffer (pH 6.0) containing laccase, before and after addition of different additives.



Figure S8. UV-vis spectra of (a) QT and (b) CT solutions in water containing FeSO₄, before and after addition of different additives.