

**Supporting Information for:**

**Microfluidic encapsulation of hydrophobic antifouling biocides in calcium-alginate hydrogels for controllable release**

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Supporting information includes:

**Figure S-1.** Introduction of antifouling biocides into paint formulation.

**Figure S-2.** Antifouling mechanism of Irgarol on aquatic plant species.

**Figure S-3.** Scanning electron microscopy (SEM) images of the cross-section of the Ca-alginate hydrogel particles.

**Figure S-4.** ATR-FTIR spectra of sodium alginate (Na-alginate) powder, synthesized Ca-alginate particles, Ca-alginate particles containing Irgarol, and Irgarol powder.

**Figure S-5.** High speed photomicrographs showing the break-off of a Na-alginate droplet containing both the drug and cellulose fibers.

**Figure S-6.** A photograph from the side showing the particles deformation measurement.

**Figure S-7.** A SEM image of the cellulose fibers.

**Figure S-8.** Schematic illustration of the UV-Vis measurement process.

**Figure S-9.** Measurement of UV-Vis absorbance spectra of Irgarol (Sample 1) at different dilution ratios.

**Figure S-10.** Relationship between the UV-Vis absorbance and the drug concentration.

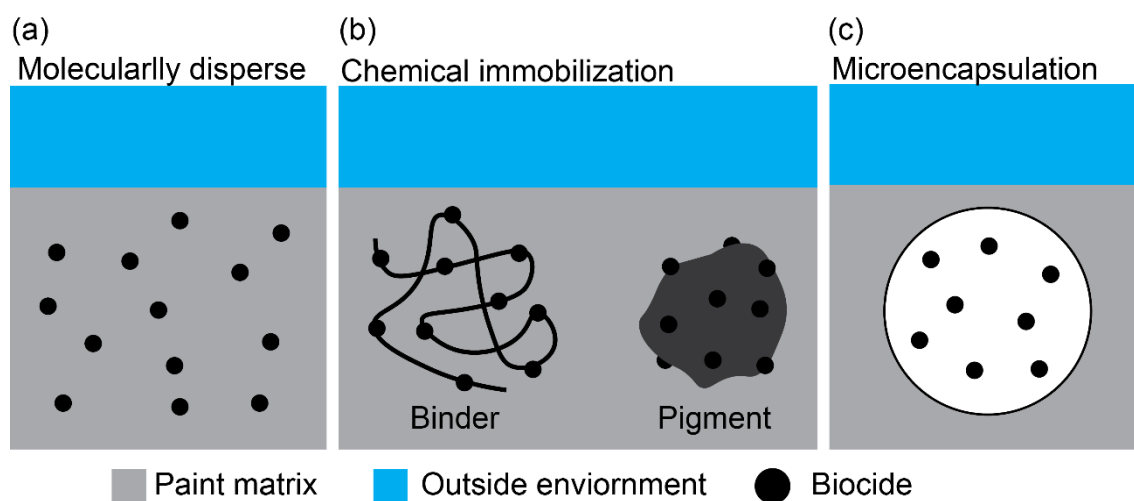
**Table S-1.** Sizes and size distributions of microcapsules for biocides encapsulation in literature.

**Table S-2.** Contents of the disperse phase used in the experiment.

**Table S-3.** The details of the five samples.

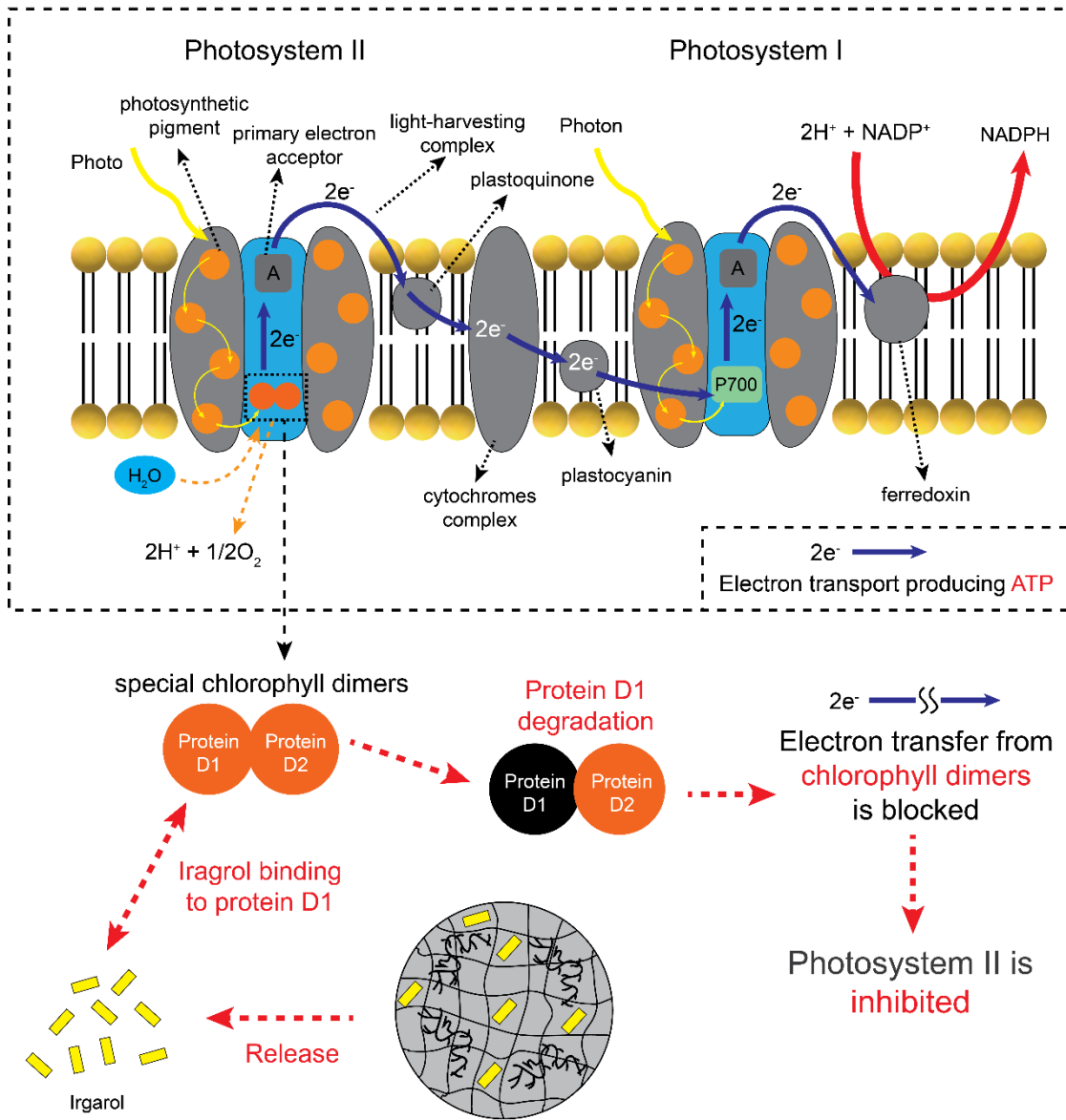
**Table S-4.** Results of the linear fitting.

**Table S-5.** Details of the hydrogel samples for the UV-Vis measurement.

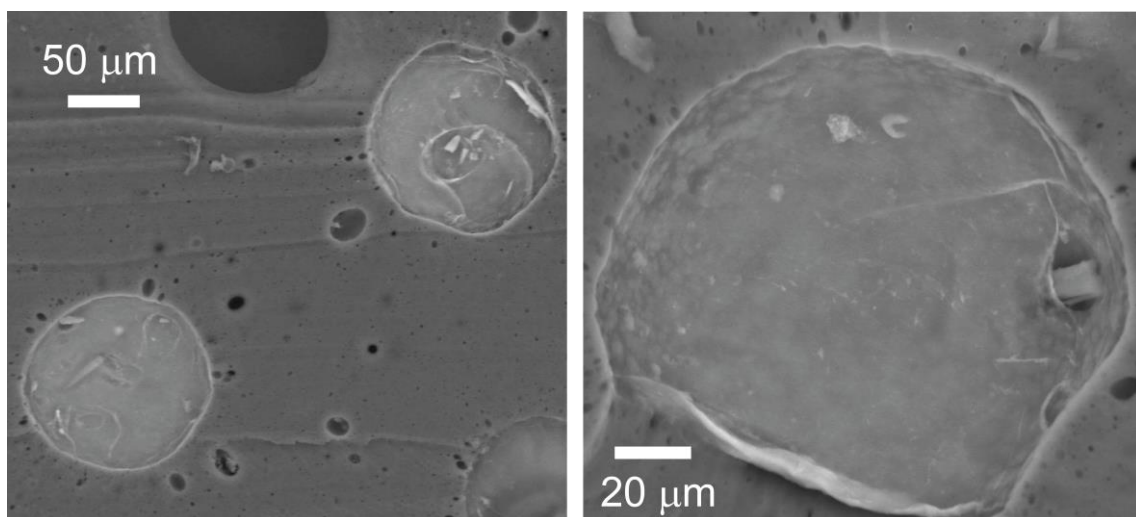


**Figure S-1.** Antifouling biocides can be introduced into paint formulation by (a) molecular dispersion, (b) immobilization to a large component, and (c) encapsulation.

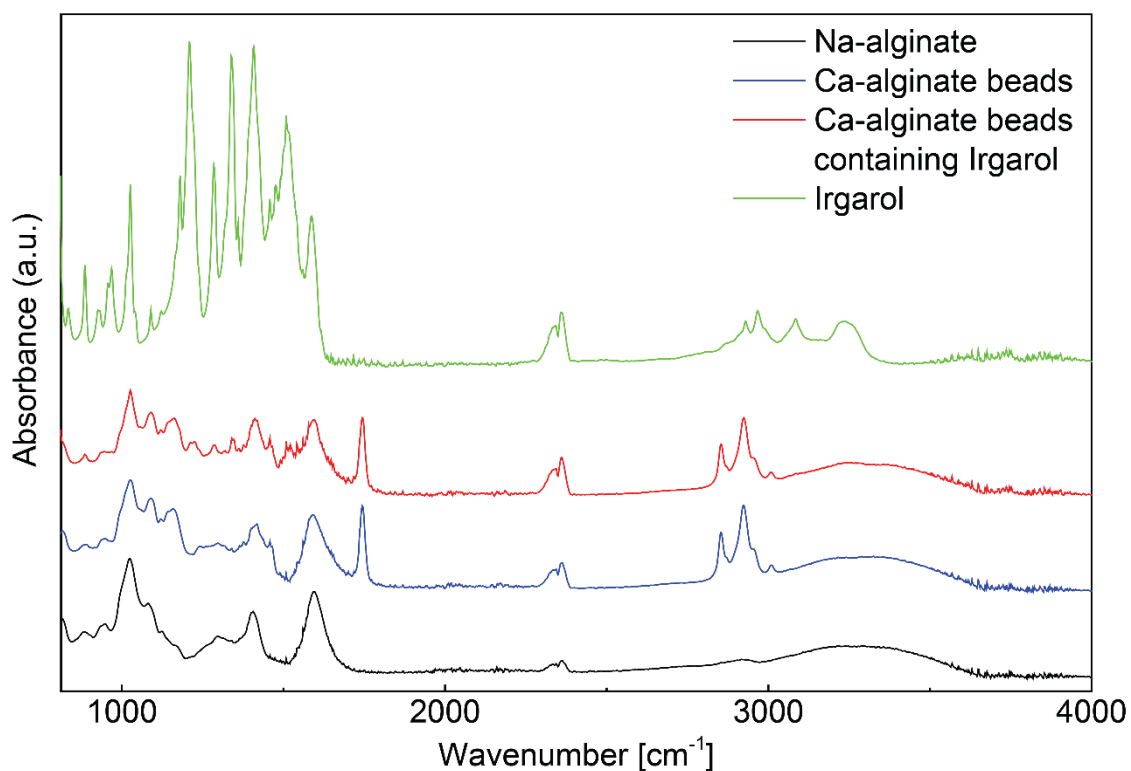
# Photosynthesis process



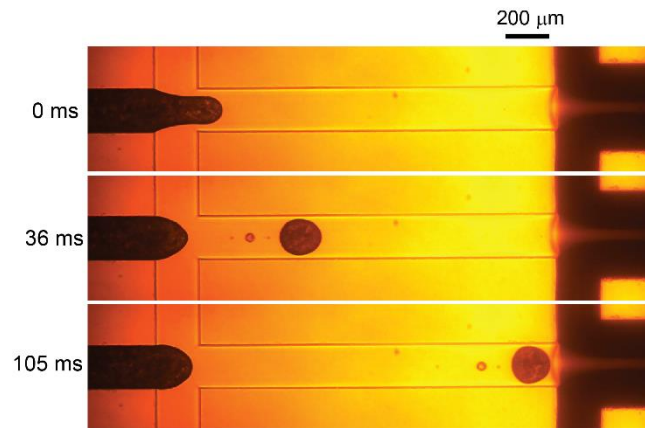
**Figure S-2.** Antifouling mechanism of Irgarol on aquatic plant species.



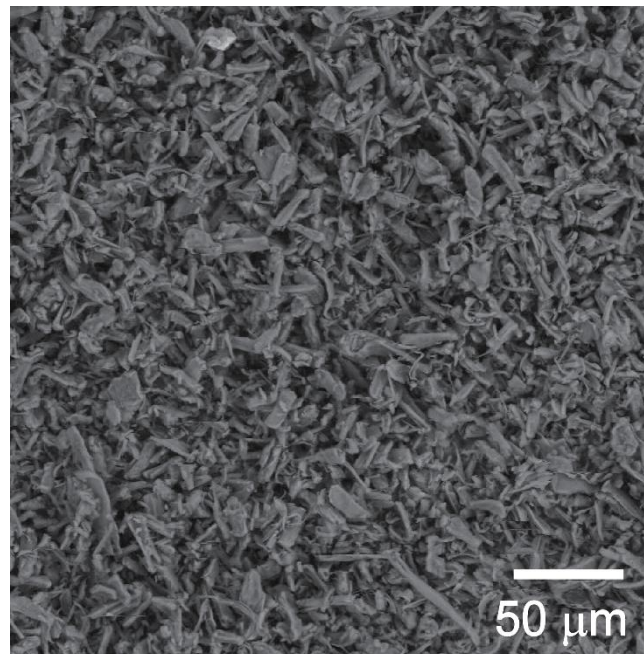
**Figure S-3.** Scanning electron microscopy (SEM) images of the cross-section of the Ca-alginate hydrogel particles.



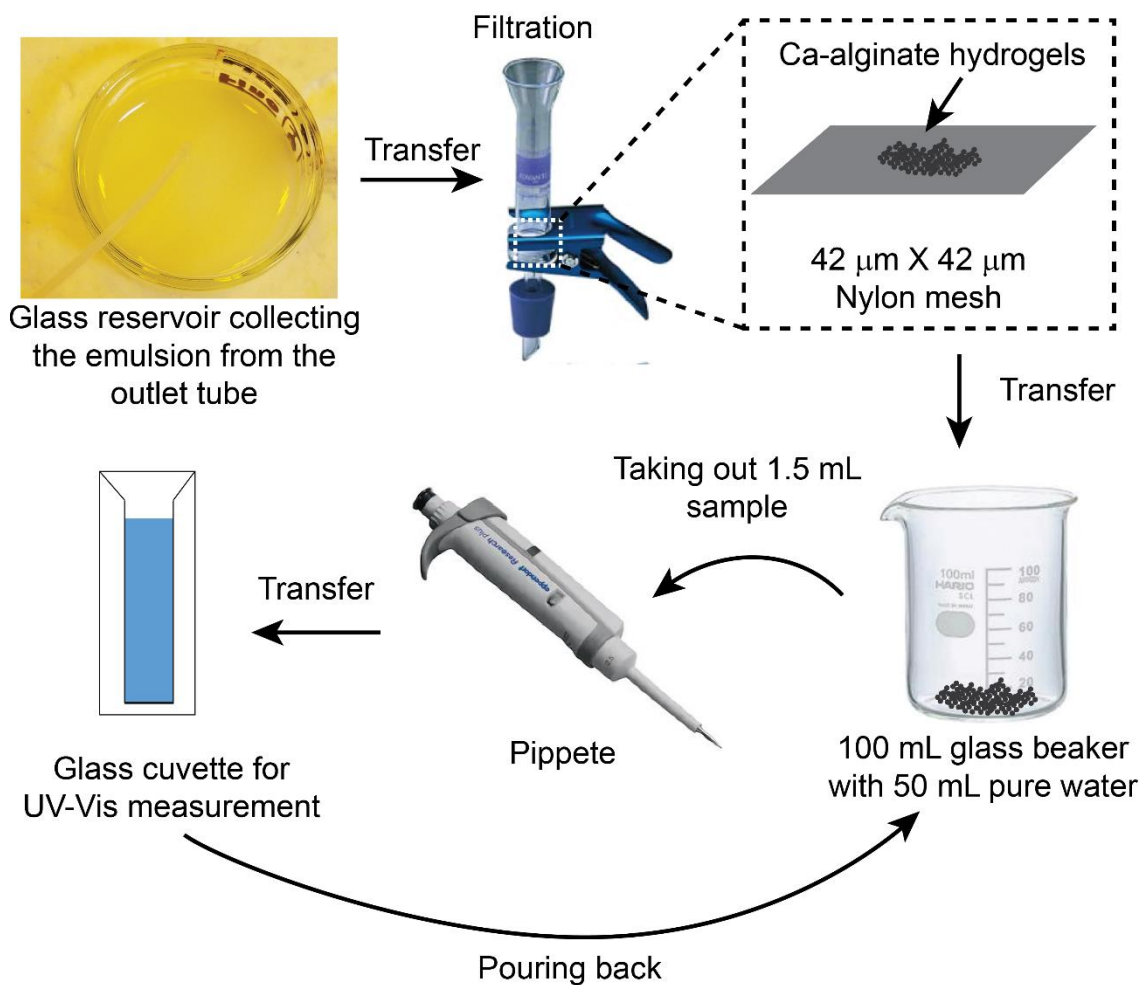
**Figure S-4.** ATR-FTIR spectra of sodium alginate (Na-alginate) powder, synthesized Ca-alginate particles, Ca-alginate particles containing Irgarol, and Irgarol powder. For the spectra of Ca-alginate particles, compared with the Na-alginate powder, the asymmetric stretching peak of  $\text{-COO}^-$  at  $1590\text{ cm}^{-1}$  is gradually replaced by a new absorption peak at  $1624\text{ cm}^{-1}$ , which is caused by the interaction of  $\text{Ca}^{2+}$  with the  $\text{-COO}^-$  groups of Na-alginate <sup>1</sup>. Further, the spectrum of the Ca-alginate beads with Irgarol shows more absorption peaks than the Ca-alginate beads without Irgarol in the wavenumber ranging from  $1552\text{ cm}^{-1}$  to  $1198\text{ cm}^{-1}$ , suggesting the presence of Irgarol. The absorption peaks at  $2925\text{ cm}^{-1}$  and  $2854\text{ cm}^{-1}$  of the hydrogel samples were caused by the residual corn oil <sup>2</sup>.



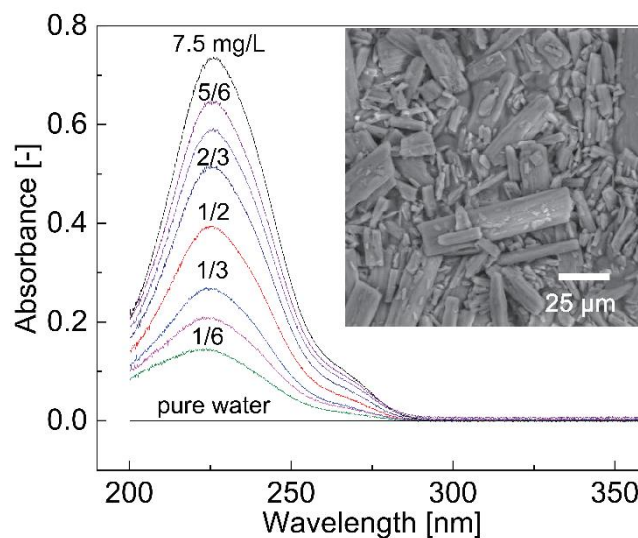
**Figure S-5.** High speed photomicrographs showing the break-off of a Na-alginate droplet containing both the drug and cellulose fibers. Flow rates of the droplet phase, carrier oil phase, and reactant emulsion stream are 0.1 mL/h, 2.0 mL/h ( $= 1.0 \text{ mL/h} \times 2$ ) and 20.0 mL/h ( $= 10.0 \text{ mL/h} \times 2$ ), respectively.



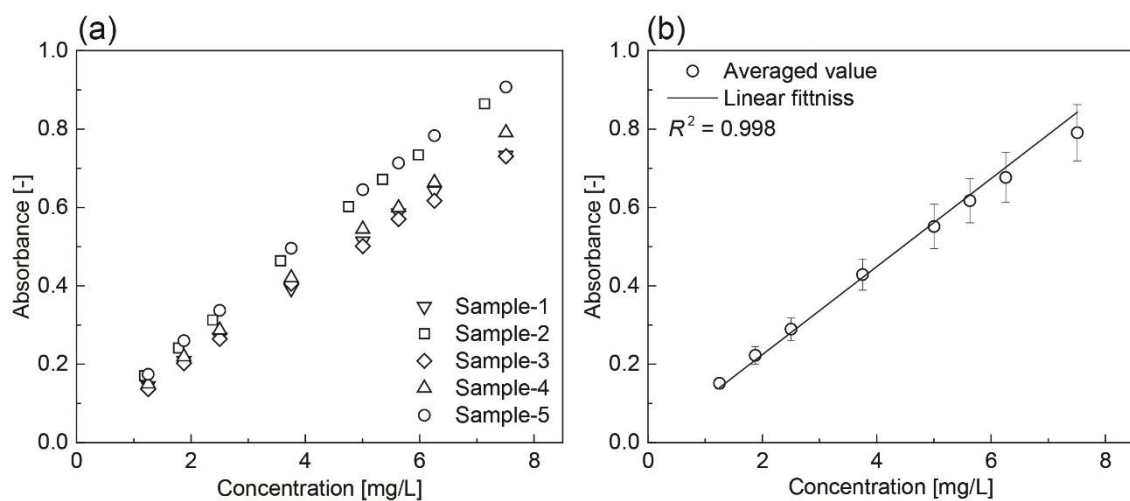
**Figure S-6.** An SEM image of the cellulose fibers.



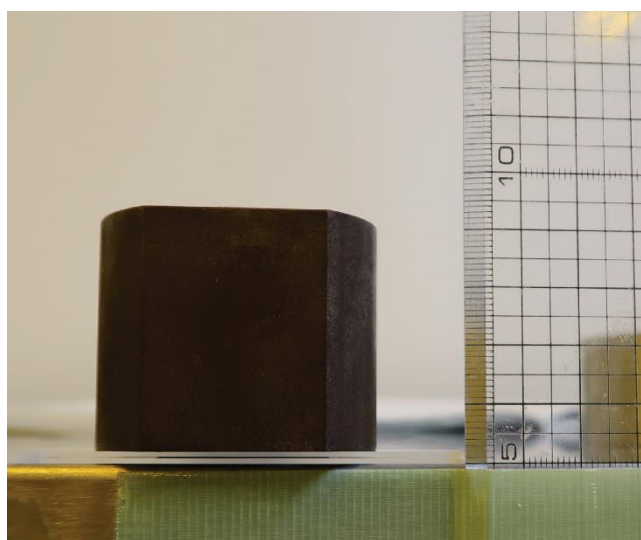
**Figure S-7.** Schematic illustration of the UV-Vis measurement process.



**Figure S-8.** Measurement of UV-Vis absorbance spectra of Irgarol (Sample 1) at different dilution ratios. The inset is an SEM image of Irgarol.



**Figure S-9.** Relationship between the UV-Vis absorbance and the drug concentration. (a) Relationship between the UV-Vis absorbance and the drug concentration of five samples. (b) The average data and linear fitting.



**Figure S-10.** A photograph from the side showing the particles deformation measurement.



**Table S-1.** Sizes and size distributions of microcapsules for biocides encapsulation in literature.

Materials	Biocides	Average diameter	Coefficient variation	Reference
PLA (Poly(L-lactide))	Chlorhexidine	1 $\mu\text{m}$	N.A.	[19]
PLGA and PLGA-PEG	Perfluorooctyl bromide	12–26 $\mu\text{m}$	N.A.	[20]
Melamine formaldehyde	Aldehydes, alcohols, esters	15 $\mu\text{m}$	13%	[21]
PMMA	Medetomidine	12 $\mu\text{m}$	41.6%	[8]
PMMA	Dodecane	2.4 $\mu\text{m}$	19%	[22]
PMMA	IPBC	N.A.	N.A.	[23]
PMMA	Aurantiol	20–200 $\mu\text{m}$	N.A.	[24]

**Table S-2.** Contents of the disperse phase used in the experiment.

	3 wt% Na-alginate solution	Irgarol <sup>®</sup> 1071	Cellulose fiber	Mixing time (30000 rpm)
Disperse #1	20.0 g	0.1 g	-	3 min
Disperse #2	20.0 g	0.05 g	-	3 min
Disperse #3	20.0 g	0.1 g	1.0 g	5 min

**Table S-3.** The details of the five samples.

	Water (mL)	Drug weight (mg)	Concentration (mg/L)
Sample 1	399.6	3.0	7.5
Sample 2	399.4	2.9	7.3
Sample 3	399.5	3.0	7.5
Sample 4	399.4	3.0	7.5
Sample 5	399.6	3.0	7.5

**Table S-4.** Results of the linear fitting.

Intercept		Slope		Statistics
Value	Standard Error	Value	Standard Error	Adj. R-Square
0.02245	0.00363	0.10524	0.00137	0.99841

**Table S-5.** Details of the hydrogel samples for the UV-Vis measurement.

	The type of used disperse phase	Collection time	Encapsulated amount ( $m_e$ )
Type 1	Disperse #1	30 min	0.250 mg
Type 2	Disperse #2	30 min	0.125 mg
Type 3	Disperse #1	60 min	0.500 mg
Type 4	Disperse #2	60 min	0.250 mg
Type 5	Disperse #3	60 min	0.500 mg

## References

- (1) Saarai, A.; Kasparikova, V.; Sedlacek, T.; Saha, P. On the development and characterisation of crosslinked sodium alginate/gelatine hydrogels. *J. Mech. Behav. Biomed.* **2013**, *18*, 152–166.
- (2) Setyaningrum, D. L.; Riyanto, S.; Rohman, A. Analysis of corn and soybean oils in red fruit oil using FTIR spectroscopy in combination with partial least square. *Int. Food Res. J.* **2013**, *20*, 1977–1981.