## **ARRIVE** The ARRIVE Essential 10: author checklist

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These items are the basic minimum to include in a manuscript. Without this information, readers and reviewers cannot assess the reliability of the findings.

| ltem                                   |    | Recommendation   | Section/line<br>number, or reason<br>for not reporting      |
|--|----|--|---|
| Study design                           | 1  | For each experiment, provide brief details of study design including:  | Control and treated group                                   |
|  |    | <ul> <li>The groups being compared, including control groups. If no control group has<br/>been used, the rationale should be stated.</li> </ul>  | group   |
|  |    | b. The experimental unit (e.g. a single animal, litter, or cage of animals).   | litter  |
| Sample size                            | 2  | a. Specify the exact number of experimental units allocated to each group, and the total number in each experiment. Also indicate the total number of animals used.  | 5 per group<br>total 10 mice                                |
|  |    | <ul> <li>Explain how the sample size was decided. Provide details of any a priori sample<br/>size calculation, if done.</li> </ul>   | Based on power<br>analysis                                  |
| Inclusion and<br>exclusion<br>criteria | 3  | a. Describe any criteria used for including and excluding animals (or experimental<br>units) during the experiment, and data points during the analysis. Specify if these<br>criteria were established a priori. If no criteria were set, state this explicitly. | All with successful<br>xenograft are<br>included.           |
|  |    | b. For each experimental group, report any animals, experimental units or data points not included in the analysis and explain why. If there were no exclusions, state so.   | All animals were included.                                  |
|  |    | c. For each analysis, report the exact value of <i>n</i> in each experimental group.   | n=5 in each<br>experimental group                           |
| Randomisation                          | 4  | a. State whether randomisation was used to allocate experimental units to control<br>and treatment groups. If done, provide the method used to generate the<br>randomisation sequence.   | Randomisation 1-10<br>number table used to<br>allocate mice |
|  |    | b. Describe the strategy used to minimise potential confounders such as the order<br>of treatments and measurements, or animal/cage location. If confounders were<br>not controlled, state this explicitly.  | Random order for<br>treatment, and<br>measurements          |
| Blinding                               | 5  | Describe who was aware of the group allocation at the different stages of the experiment (during the allocation, the conduct of the experiment, the outcome assessment, and the data analysis).  | Only technician<br>performed treatment<br>was aware         |
| Outcome<br>measures                    | 6  | a. Clearly define all outcome measures assessed (e.g. cell death, molecular markers, or behavioural changes).  | Tumor size, invasion depth, eNAMPT                          |
|  |    | <ul> <li>b. For hypothesis-testing studies, specify the primary outcome measure, i.e. the<br/>outcome measure that was used to determine the sample size.</li> </ul>   | Tumor invasion depth  |
| Statistical methods                    | 7  | <ul> <li>Provide details of the statistical methods used for each analysis, including<br/>software used.</li> </ul>  | Student t test, Mann-<br>Whitney U test                     |
|  |    | b. Describe any methods used to assess whether the data met the assumptions of<br>the statistical approach, and what was done if the assumptions were not met.   | Q-Q plot, Levene's test                                     |
| Experimental<br>animals                | 8  | <ul> <li>Provide species-appropriate details of the animals used, including species, strain<br/>and substrain, sex, age or developmental stage, and, if relevant, weight.</li> </ul>   | SCID (C.B-Igh-1b/<br>IcrTac-Prkdcscid)                      |
|  |    | b. Provide further relevant information on the provenance of animals, health/immune status, genetic modification status, genotype, and any previous procedures.  | T an B cell immune deficient                                |
| Experimental procedures                | 9  | For each experimental group, including controls, describe the procedures in enough detail to allow others to replicate them, including:  | Intraperitoneal injection of PC3 and                        |
|  |    | a. What was done, how it was done and what was used.   | eNAMPT antibody   |
|  |    | b. When and how often.   | 3x/week, 3 weeks  |
|  |    | c. Where (including detail of any acclimatisation periods).  | Peritoneal space<br>Test PC3 invasion                       |
| Results                                | 10 | d. Why (provide rationale for procedures).<br>For each experiment conducted, including independent replications, report:   | Mean+/-SD and   |
| Results                                | 10 | a. Summary/descriptive statistics for each experimental group, with a measure of   | median  |
|  |    | variability where applicable (e.g. mean and SD, or median and range).<br>b. If applicable, the effect size with a confidence interval.   | 95% confidence<br>interval                                  |