Additional Table 1. Sources and cell culture media for established HNSCC cell lines used in this study.

Cell Line	HPV Status	Tumour Site (if available)	Patient Information (if available)	Growth Medium	Source		
93-VU-147T	Positive	Floor of mouth	Male, T4N2	DMEM/F12	VUMC		
Cal33	Negative	Tongue	Male	DMEM + HI FBS + NEAA	DSMZ		
SCC15	Negative	Tongue	Male	DMEM/F12	ATCC		
PCI6A	Negative	_	_	DMEM/F12	University of Pittsburgh		
SCC61	Negative	_	_	DMEM/F12	Yale		
RF37A	Negative	_	_	DMEM/F12	University of Pittsburgh		
HSC2	Negative	Oral cavity	Male	EMEM	·		
UMSCC47	Positive	Lateral tongue	Male, T3N1M0	DMEM/F12	University of Michigan		
UPCI:SCC154	Positive	Oral cavity	Male, T4N2	DMEM/F12	University of Pittsburah		
JHU029	Negative	Oropharynx	Male, T4N0	DMEM/F12	Johns Hopkins		
RF15B	Negative	_	_	DMEM/F12	University of Pittsburah		
HMS001	Positive	Oropharynx (tonsil)	Male	DMEM/F12	Harvard Medical School		
PCI6B	Negative	Oropharynx	Male, T3N3M0	DMEM/F12	University of Pittsburgh		
D562	Negative	Pharynx	Female	DMEM/F12	ATCC		
SCC25	Negative	Hypopharynx	Male	DMEM/F12	ATCC		
UPCI:SCC090	Positive	Oropharynx (tongue base)	Male, T2N0	DMEM/F12	University of Pittsburgh		
PCI13	Negative	Oral cavity	Male, T4N1M0	DMEM/F12	University of Pittsburgh		
RF22A	Negative	_	_	DMEM/F12	University of Pittsburgh		
JHU006	Negative	_	_	DMEM/F12	Johns Hopkins		
Cal27	Negative	Tongue	Male	DMEM/F12	ATCC		
SCC9	Negative	Tongue	Male	DMEM/F12	ATCC		
JHU011	Negative	Larynx	Male, T3N0	DMEM/F12	Johns Hopkins		
SCC4	Negative	Tongue	Male	DMEM/F12	ATCC		
SCC2	Positive	_	_	DMEM/F12			
FaDu	Negative	Hypopharynx	Male	DMEM/F12	ATCC		

DMEM, Dulbecco's Modified Eagle Medium; EMEM, Eagle's Minimum Essential Medium; NEAA, nor essential amino acids; VUMC, VU University Medical Center Amsterdam; DSMZ, Deutsche Sammlun von Mikroorganismen und Zellkulturen; ATCC, American Type Culture Collection Additional Table 2. Short-tandem repeat (STR) profiling results confirming matching identities of primary tumor, blood, xenograft tumors and cell lines, where available.

Sample	Amelogenin	CSF1PO	D13S317	D16S539	D18S51	D19S433	D21S11	D2S1338	D3S1358	D5S818	D7S820	D8S1179	FGA	TH01	трох	vWA
PDX-C Blood	X,Y	10,12	11,13	11,12			28,32.2			11,11	10,10			6,9.3	8,11	16,16
PDX-C Primary	X,Y	10,12	11,13	11,12			28,32.2			11,11	10,10			6,9.3	8,11	16,16
PDX-C C4	X,Y	10,12	11,13	11,12	11,13	14,14	28,28	17,18	15,15	11,11	10,10	13,13	25,25	6,9.3	8,11	16,17
PDX-C C5	X,Y	10,12	11,13	11,12			28,32.2			11,11	10,10			6,9.3	8,11	16,17
PDX-C B2	X,Y	10,12	11,13	11,12			28,28			11,11	10,10			6,9.3	8,11	16,17
PDX-C B3	X,Y	10,12	11,13	11,12	11,13	14,14	28,28	17,18	15,15	11,11	10,10	13,13	25,25	6,9.3	8,11	16,17
PDX-C B4	X,Y	10,12	11,13	11,12	11,13	14,14	28,32.2	17,18	15,15	11,11	10,10	13,13	25,25	6,9.3	8,11	16,17
PDX-C Cell Line	X,Y	12,12	11,13	11,12	11,13	14,14	28,32.2	17,18	15,15	11,11	10,10	13,13	25,25	6,9.3	8,11	16,17
PDX-E Primary	X,X	12,12	12,12	9,9			30,30,2			12,13	11,11			7,9.3	8,11	17,17
PDX-E C3	X,X	12,12	12,12	9,9	10,18	12,16	30,30,2	19,19	17,17	12,13	11,11	13,13	22,23	7,9.3	11,11	17,17
PDX-E C4	X,X	12,12	12,12	9,9			30,30,2			12,13	11,11			7,9.3	11,11	17,17
PDX-E B2	X,X	12,12	12,12	9,9	10,18	12,16	30,30,2	19,19	17,17	12,13	11,11	13,13	22,23	7,9.3	8,11	17,17
PDX-E B3	X,X	12,12	12,12	9,9			30,30,2			12,13	11,11			7,9.3	8,11	17,17
PDX-E B4	X,X	12,12	12,12	9,9	10,18	12,16	30,30,2	19,19	17,17	12,13	11,11	13,13	22,23	7,9.3	8,11	17,17

## Additional Table 3. Antibodies used in this study.

Antibody	Company	Catalogue Number	Dilution		
pAkt (T308)	CST	4056	1:1000		
pAkt (S473)	CST	9271	1:1000		
Akt (pan)	CST	4685	1:1000		
pERK1/2 (T202/Y204)	CST	4370	1:1000		
ERK1/2	CST	4696	1:1000		
pP90RSK (S380)	CST	11989	1:1000		
P90RSK (pan)	CST	9355	1:1000		
PARP	BD Pharmingen	556494	2ug/mL		
Cleaved PARP	CST	5625	1:1000		
TYRO3	CST	5585	1:1000		
AXL	CST	8661	1:1000		
MER-TK	abcam	ab52968	1/2000		
GAB2	CST	3239	1:1000		
pMEK1 (S298)	CST	9128	1:1000		
MEK1	CST	2352	1:1000		
pS6 (S235/6)	CST	4858	1:1000*		
<b>S</b> 6	CST	2217	1:1000*		
EGFR	CST	4267	1:2000		
MER-TK <sup>+</sup>	abcam	ab52968	1/500		
TYRO3⁺	BETHYL Laboratories Inc.	IHC-00410	1:100-1:500		
α-tubulin	CST	2125	1:1000		

CST, Cell Signaling Technology \* primary antibody incubation: 1hr at room temperature <sup>+</sup> for IHC

Additional Table 4. Clinical features of HNSCC patients used to generate	PDX models of
acquired drug resistance.	

	PDX ID	Gender	Age	T	TNM Stage		Disease	Subsite	HPV	Smoking	Alcohol	Recurrent	
ļ			-			IAI	Site		Status	History	Consumption		
	PDX-A	М	72	T2	N2b	MO	Lip & Oral Cavity	Tongue	nt	Ex-smoker	Non-drinker	No	
	PDX-B	М	60	Т3	N2c	MO	Oropharynx	Base of Tongue	+	Non-smoker	Non-drinker	Yes	
	PDX-C	М	44	T2	N1	MO	Lip & Oral Cavity	Tongue	-	Non-smoker	Non-drinker	No	
	PDX-D	F	87	T2	N2b	MO	Lip & Oral Cavity	Tongue	nt	Non-smoker	Non-drinker	No	
	PDX-E	М	63	T4	N2b	MO	Hypopharynx	Piriform Sinus	-	NA	NA	NA	



Additional Fig. 1. (A) Schematic outlining the derivation of cell line from PDX-C. (B) Phase contrast microscopy image of PDX-C cells. (C) Flow cytometry for cell surface expression of EpCAM (CD326) in PDX-C cells. Over 99% of PDX-C cells were found to be CD326-positive.



Additional Fig. 2. Schematic outlining the development of the alpelisibresistant HNSCC cell lines. Parental cells (Cal33 and 93VU-147T) were treated with increasing doses of alpelisib, beginning with their IC<sub>50</sub> values ( $0.5\mu$ M for Cal33,  $1.7\mu$ M for 93VU-147T), as previously established (20).





93-VU-147T



Additional Fig. 3. Relative abundance of small, medium and large sized colonies for parental and alpelisib-resistant cell lines ((A) Cal33 cells, (B) 93-VU-147T cells) when untreated, and when treated with alpelisib. Colony counts and sizes were analyzed in ImageJ version 1.52a. Briefly, RGB images of wells were converted in binary images and analyzed using the Analyze Particles feature. Colony size cutoffs were set as follows (in pixels): Small 0-100; Medium 101-500; Large  $\geq$  501.

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Cal33





Β.

Additional Fig. 4. (A) & (B) Immunoblot of MER-TK expression in parental and alpelisib-resistant Cal33 and 93-VU-147T cells. Short and long exposures of MER-TK blot are shown. HEK293T cells served as a positive control for MER-TK expression.



Additional Fig. 5. Histological comparison of PDX tissues and their corresponding primary tumors (where available), stained with H&E. Scale bar represents  $50\mu$ M.



Additional Fig. 6. Representative IHC sections showing Ki67 staining PDX tissues treated with the vehicle agent (corn oil) or alpelisib (endpoint either while still responding or treated out to the emergence of resistance). Scale bar represents  $100\mu$ M.



Additional Fig. 7. Representative IHC sections showing AXL staining in PDX-C and PDX-E models. Quantification completed using Fiji software is shown below. ns = not significant, unpaired Student's *t*-test. Scale bars represent  $100\mu$ M.



**Additional Fig. 8.** Immunoblot of TYRO3 and AXL expression in 25 HNSCC cell lines, ordered by sensitivity to alpelisib (IC50 values indicated).