Supplementary Information for

Driver mutations of the adenoma-carcinoma sequence govern the intestinal epithelial global translational capacity.

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Fig. S1.

(A) Representative images of Apc^{-/-} (left panel), Kras^{+/G12D} (middle panel), and Apc^{-/-} Kras^{+/G12D} organoids (right panel) 3 days after passaging. Each panel also shows the method of selection for mutant cells using culture factor withdrawal; E = EGF, N = Noggin, R = R-spondin, scale bar = 250 μ m (B) L-[³⁵S]-Methionine incorporation assay in Apc^{-/-} organoids recombined for several weeks (>2 weeks) assessed at day 4 after passaging (n =2) (C) Flow cytometric quantification of L-azidohomoalanine incorporation represented as relative fluorescence intensity compared to non-recombined controls assessed at day 4 after passaging. (D) L-[35 S]-Methionine incorporation assay in Apc^{-/-} organoids 48 h after recombination, assessed at day 4 after passaging (n = 4). (E) L-[³⁵S]-Methionine incorporation assay in wildtype organoids cultured for 3 days in ENR followed by 24 hour withdrawal of R-spondin (n = 3). (F) Example of PAS and MUC2 staining in Kras^{+/G12D} organoids and quantification of MUC2-positive cells, indicated by black arrows (40x objective). Data presented as number of MUC2-positive cells relative to the number of cells per organoid. (G) Quantitative RT-PCR analysis of Notch target gene Hesl in $Kras^{+/G12D}$ organoids (n = 3). (H) L-[³⁵S]-Methionine incorporation assay in wildtype organoids cultured for 3 days in ENR followed by 24 hour withdrawal of EGF (n = 3). (1) Heatmap of microarray *mRNA* expression data containing a set of genes encoding for small nucleolar *RNAs* in wildtype, $Apc^{-/-}$ and $Apc^{-/-}Kras^{+/G12D}$ organoids (n = 4). (J) Quantitative RT-PCR analysis of the 5' external transcribed spacer of 47S pre-*rRNA* as a by proxy measurement of rRNA synthesis (n = 3). (K) Quantitative RT-PCR analysis of Myc in Apc⁻ ^{/-} and $Apc^{-/-}Kras^{+/G12D}$ organoids (n = 3). (L) Representative immunoblot of MYC levels in Apc^{-/-} and Apc^{-/-}Kras^{+/G12D} organoids (n = 2). (M) Quantification of the number of viable organoids treated for 72 h with 2.5 ng/ml Actinomycin D and presented as relative growth reduction to their respective untreated control (n = 2). (N) Representative microscopic image of organoids treated with 2.5 ng/ml Actinomycin D.

Significance in all figures analyzed by Student *t*-test, *P < 0.05, **P < 0.01, ***P < 0.001.



D









AK (shControl) AKS (sh #2)







Ranking metric scores

— Enrichment profile — Hits





Fig. S2.

(A) Quantitative RT-PCR analysis of Smad4 in wildtype organoids transduced with two shRNAs against Smad4 and scrambled control shRNAs (n = 3). (B) Representative image of generated shSmad4 organoids 3 days after passaging and the method of selection using long-term Noggin withdrawal; E = EGF, N = Noggin, R = R-spondin, scale bar = 250 μ m. (C) $L-[^{35}S]$ -Methionine incorporation assay in wildtype organoids cultured for 3 days in ENR followed by 24 h withdrawal of Noggin (n = 3). Student *t*-test, *P < 0.05. (D) Representative image of generated Apc^{-/-}Kras^{+/G12D} shSmad4 #2 (AKS) organoids and the method of selection showing the regenerative potential under incubation with 10 ng/ml human recombinant TGF β . Scale bar = 250 µm. (E) Gene set enrichment plot comparing AK and AKS organoids against a published gene set of a MCF10A cell line with high EIF4G1 expression. (F) Gene set enrichment plot comparing AK and AKS organoids against a published gene set of 4ebp1/4ebp2 double KO mouse fibroblasts. (G) Quantitative RT-PCR analysis of 4ebp1 and 4ebp2 expression in shControl and shSmad4 organoids. (H) Quantitative RT-PCR analysis of 4ebp1 and 4ebp2 expression in AK and AKS organoids (n = 3).

Significance in all figures analyzed by One-way ANOVA, *P < 0.05, **P < 0.01.



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	WT	A	AK	AKS	AKSP
untreated	6° 5 30 _				
torin1	6		· · · · · · · · · · · · · · · · · · ·		©

AKT2 AKT1

-1

z-score

	WT	A	AK	AKS	AKSP
untreated	A CARLE			8.0	
ribavirin	800 - 100 -			0	

Fig. S3.

(A) Quantitative RT-PCR analysis of Trp53 mRNA expression MSCV-Cre Trp53^{-/-} organoids 48 h after recombination (n = 2). (B) Representative image of $Trp53^{-/-}$ organoids 3 days after passaging, and the method of selection using 5 day incubation with 10 μ M Nutlin-3. Scale bar = 250 μ m. (C) Quantitative RT-PCR analysis of Trp53 in AKS organoids transduced with two different *shRNAs* against *Trp53* (n = 3). (D) Representative image of generated AKSP organoids 3 days after passaging, and the method of selection for Smad4 and Trp53 knockdown showing the regenerative potential during incubation with a combination of 10 ng/ml recombinant TGFB and 10 μ M Nutlin-3. Scale bar = 250 μ m. (E) Heatmap of microarray mRNA expression data containing a set of pro-apoptotic genes in AKSP and AKS organoids (n = 4). (F) L-[³⁵S]-Methionine incorporation assay in wildtype organoids treated for 2 h treatment in the presence of 10 µg/ml cycloheximide (CHM). (G) EdU incorporation assay in organoids treated for 2 h with 10 μ g/ml cycloheximide or 4 h with 1 mM ribavirin. Data presented as percentage of the number of EdU-positive cells to the total cell population (n = 2). (H) Heatmap of microarray mRNA expression data containing genes related to mTORC1 and mTORC1 signaling in organoids of all ACS genotypes (n = 4). (1) Representative experiment of growth in organoids with ACS mutations treated for 48 h with 100 nM rapamycin and presented as quantified growth reduction after treatment relative to untreated control (n = 2). (J) Representative images of organoids with ACS mutations treated for 48 h with 50 nM torin 1. (K) Quantitative RTqPCR analysis of stem cell marker Lgr5 in organoids with ACS mutations treated for 48 h with 50 nM Torin 1 (n = 2). (L) Representative images of organoids with ACS mutations treated for 48 h with 10 µM ribavirin.

Significance in all figures analyzed by One-way ANOVA, *P < 0.05, **P < 0.01, ***P < 0.001.

Gene	Forward primer	Reverse primer	
Lgr5	TGTGTCAAAGCATTTCCAGC	CAGCGTCTTCACCTCCTACC	
Lyz1	ATGGAATGGCTGGCTACTATGG	ACCAGTATCGGCTATTGATCTGA	
Muc2	GAAGCCAGATCCCGAAACCA	GAATCGGTAGACATCGCCGT	
Chgal	GTCTCCAGACACTCAGGGCT	ATGACAAAAGGGGACACCAA	
Alpi	CACAGCTTACCTGGCACTGA	GGTCTCTGACGACAGGGGTA	
Cd44	TCTGCCATCTAGCACTAAGAGC	GTCTGGGTATTGAAAGGTGTAGC	
Sox9	AGGAAGCTGGCAGACCAGTA	TCCACGAAGGGTCTCTTCTC	
Smad4	GGTTGTCTCACCTGGAATTGA	GGCTGTCCTTCAAAGTCGTG	
P53	AAAGGATGCCCATGCTACAG	CCCCACTTTCTTGACCATTG	
Hes1	CCAGCCAGTGTCAACACGA	AATGCCGGGAGCTATCTTTCT	
eIf4ebp1	GGGGACTACAGCACCACTC	CTCATCGCTGGTAGGGCTA	
ETS (47S pre- <i>rRNA</i>)	TTTTGGGGAGGTGGAGAGTC	AGAGAACTCCGGAGCACCAC	

Table 1. List of used RT-qPCR primers

Movie S1.

Cross sectional view of a wildtype small intestinal organoid transduced with scrambled control *shRNA*, three days after passaging. Visible is the central lumen containing cellular debris and normal buds consisting of stem cells and Paneth cells (cells with dark grey vesicles in the crypt bottom).

Movie S2.

Cross sectional view of a small intestinal organoid transduced with *shRNA* against *Smad4*, three days after passaging. Knockdown of *Smad4* results in extensive formation of buds compared to wildtype organoids (movie S1), indicating enrichment for both stem cell and Paneth cell populations.