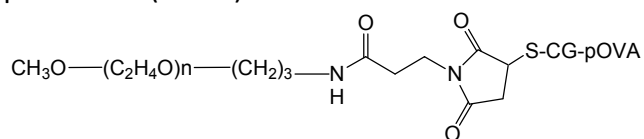


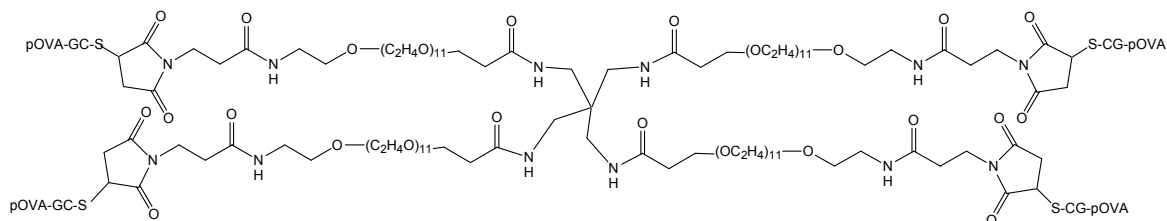
Supplementary Material

pOVA-PEG (linear)

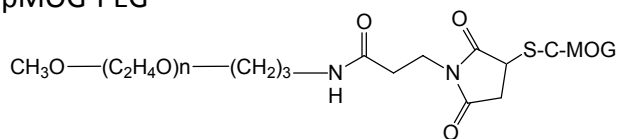


$n = \text{approx. } 450 \text{ or } 900$

pOVA-PEG (tetramer)



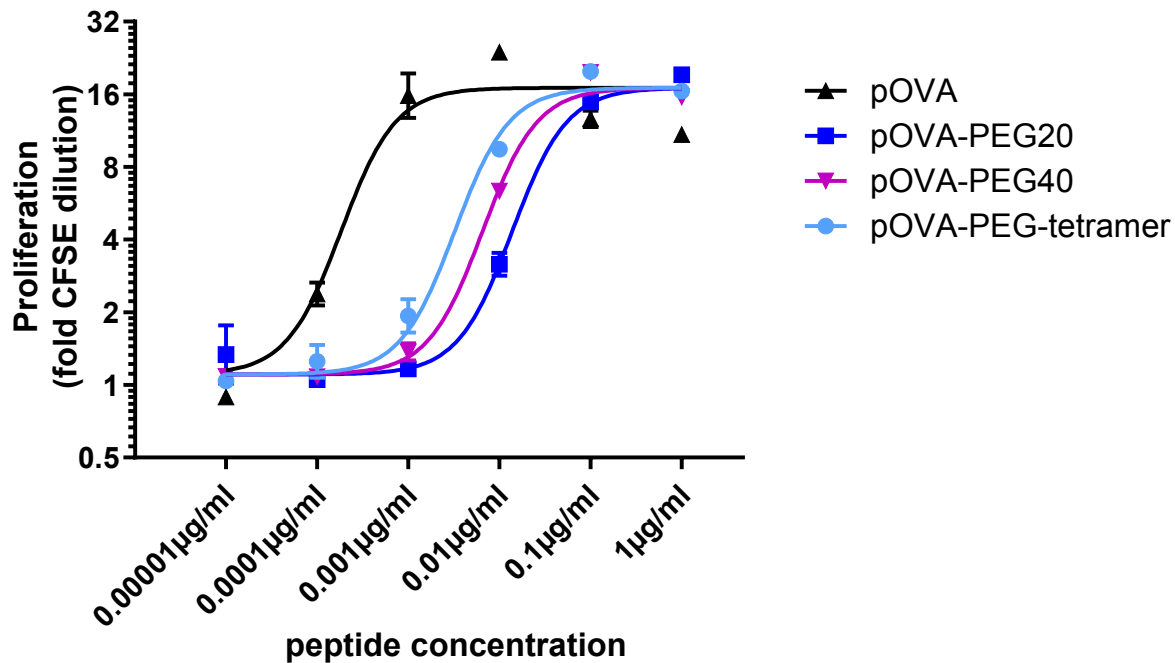
pMOG-PEG



$n = \text{approx. } 450$

Supplementary Figure 1:

Formulas of PEG-peptide conjugates used.



Supplementary Figure 2

Stimulatory capacity of PEGylated peptides *in vitro*: Fitted dose-response curves of peptide and conjugates.

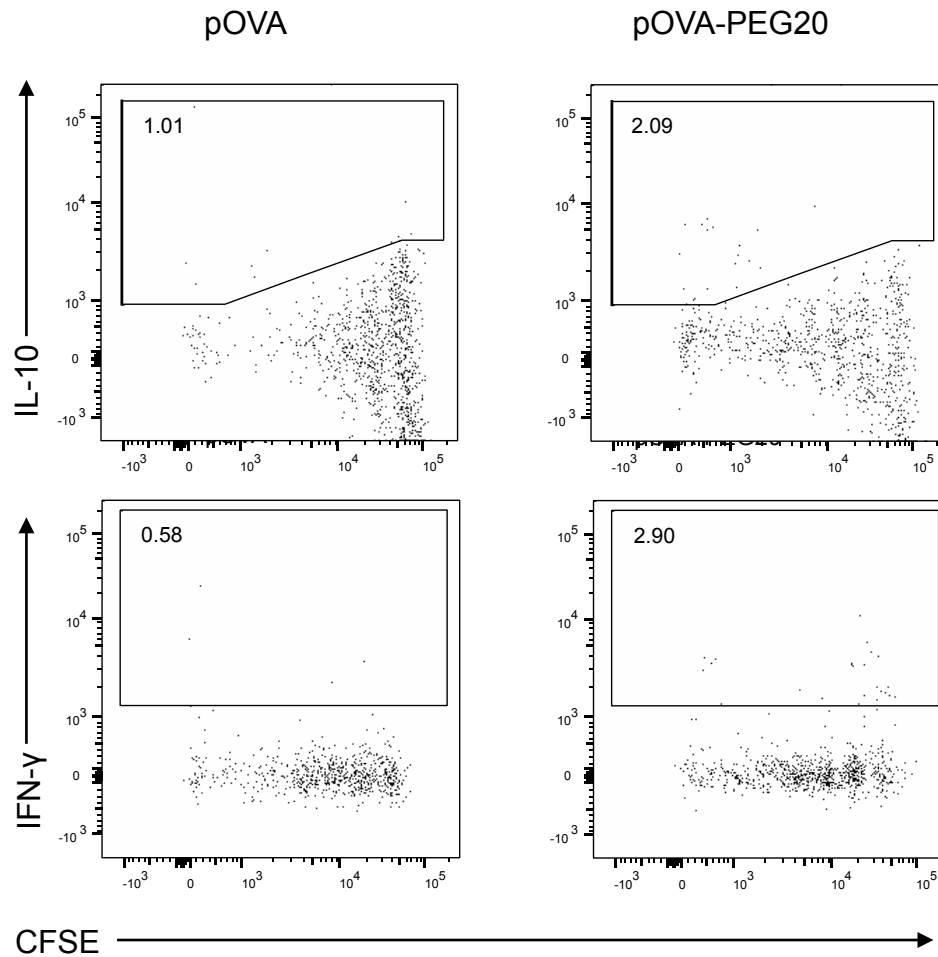
Higher concentrations of PEGylated peptides were required to induce half-maximal proliferation compared to unmodified pOVA with 21 to 91-fold higher effective concentration (EC₅₀).

EC₅₀ pOVA-PEG20: 91.0 (fold EC₅₀ of pOVA); 95% confidence interval (CI): 26.0-156.0;

EC₅₀ pOVA-PEG40: 43.9; CI: 11.4-76.4;

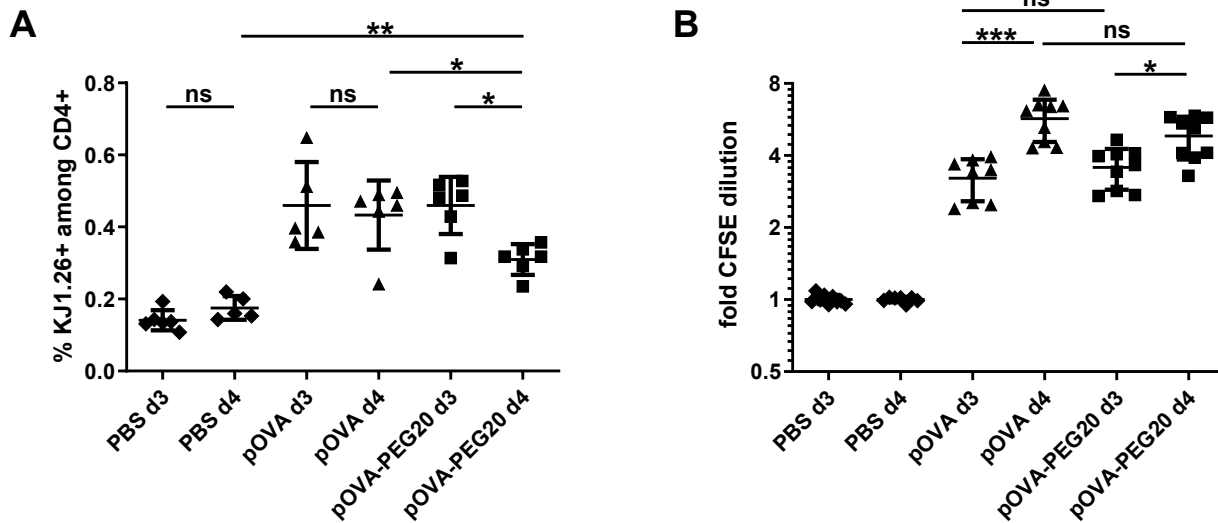
EC₅₀ pOVA-PEG-tetramer: 21.0; CI: 5.4-36.7).

Dose-response analysis was performed using GraphPad Prism version 7. EC₅₀ shift was estimated by global nonlinear fitting of the default Hill model, assuming same minimal and maximal proliferation and variable Hill-slope for each conjugate. The number of cell divisions, that is log₂ of the dilution factor of CFSE, was used as the proliferation output. Statistical differences between the EC₅₀ values of the conjugates were computed applying the sum-of-squares F-Test for the unconstrained model and EC₅₀ shift of 1. All dose-response curves were significantly different from each other with $p < 0.001$.



Supplementary Figure 3

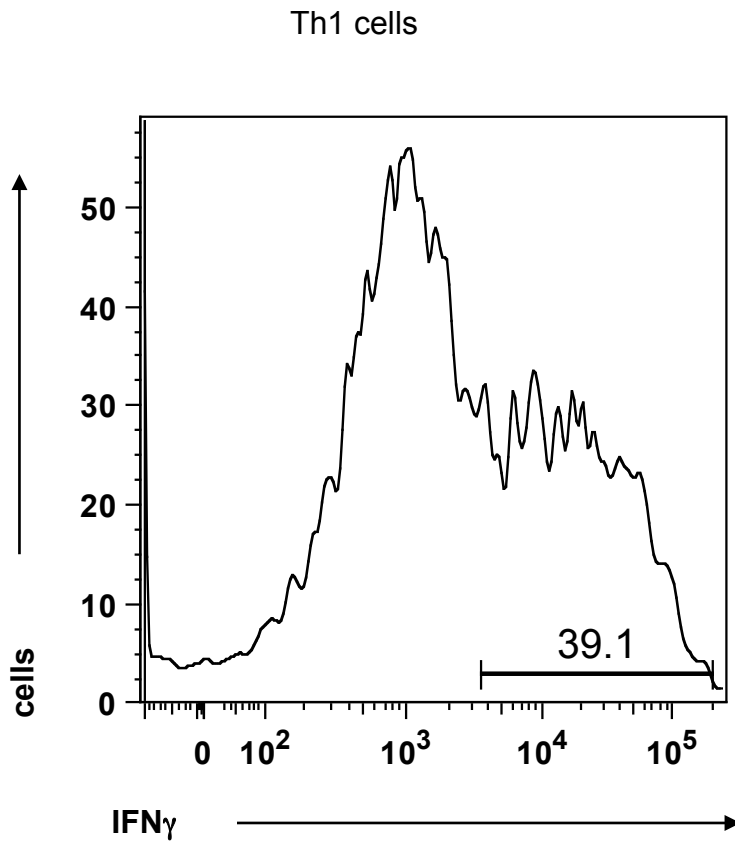
Representative IL10 and IFN γ staining (after restimulation) upon stimulation with 5 μ g pOVA and pOVA-PEG20 *in vivo*; day 7.



Supplementary Figure 4:

Faster decline in the number of OVA-specific CD4⁺ T cells following pOVA-PEG20 after day 3, despite ongoing proliferation.

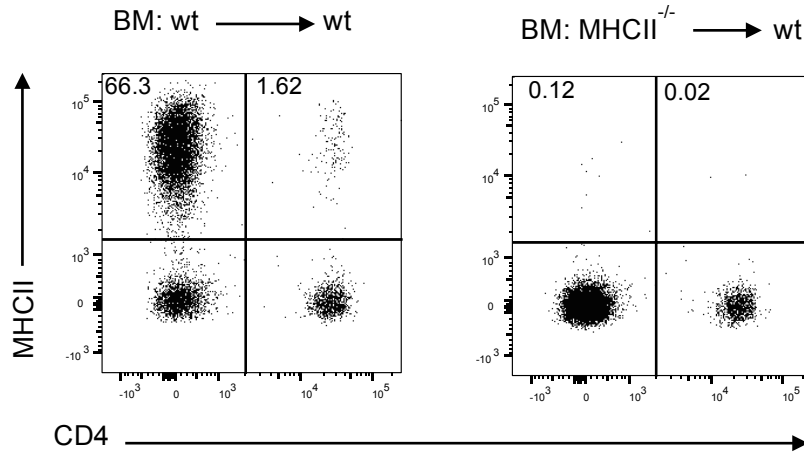
24 h after adoptive transfer of OVA-specific CFSE-labeled CD4⁺ T cells, recipients received *i.v.* PBS (control), 5 μ g pOVA or equimolar amounts of pOVA-PEG20. Splenocytes were isolated on day 3 (d3) and day 4 (d4) and analyzed using flow cytometry. (A) % KJ1.26⁺ cells (OVA-specific T cells) among total CD4⁺ cells. Mean \pm SD of (n = 6). One representative of two independent experiments is shown. (B) Proliferation (mean x-fold CFSE dilution \pm SD) of OVA-specific CD4⁺ T cells. Data from two independent experiments, n = 7-9. Statistical testing was performed using the nonparametric Mann Whitney test and Holm-Bonferroni correction for multiple comparisons.



Supplementary Figure 5:

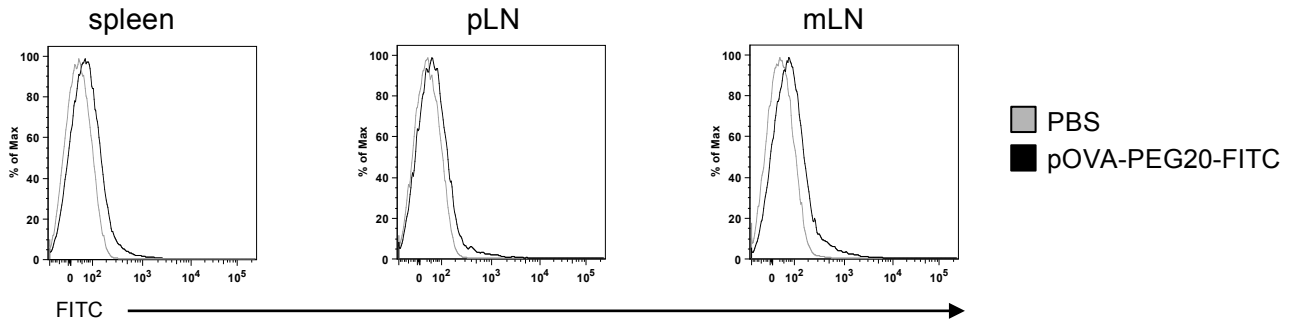
Impact of pOVA-PEG20 vaccination in presence of pre-existing T effector cells.

Representative histogram of IFN- γ expression of *in vitro* generated OVA-specific Th1 cells on day 5 of culture before adoptive transfer.

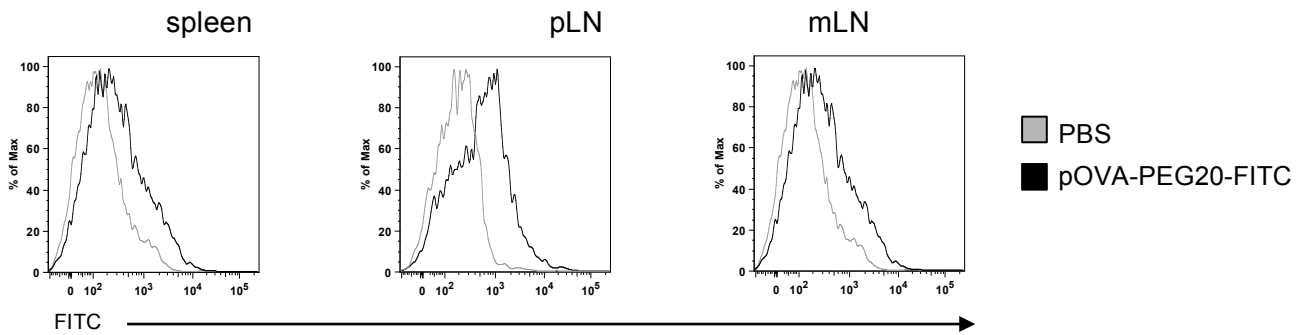
**Supplementary Figure 6:**

Control staining of blood leucocytes for MHCII, 8 weeks after reconstitution of BM-depleted B6 mice with BM from MHCII^{-/-} mice.

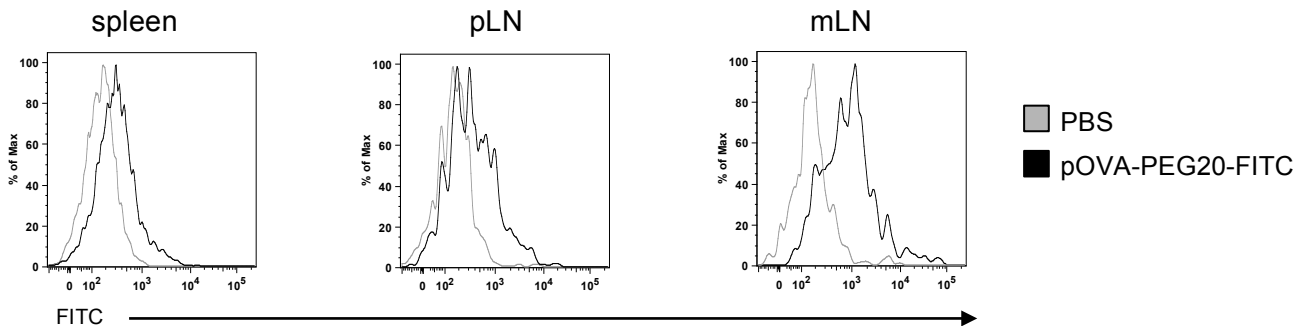
B cells



DCs



macrophages

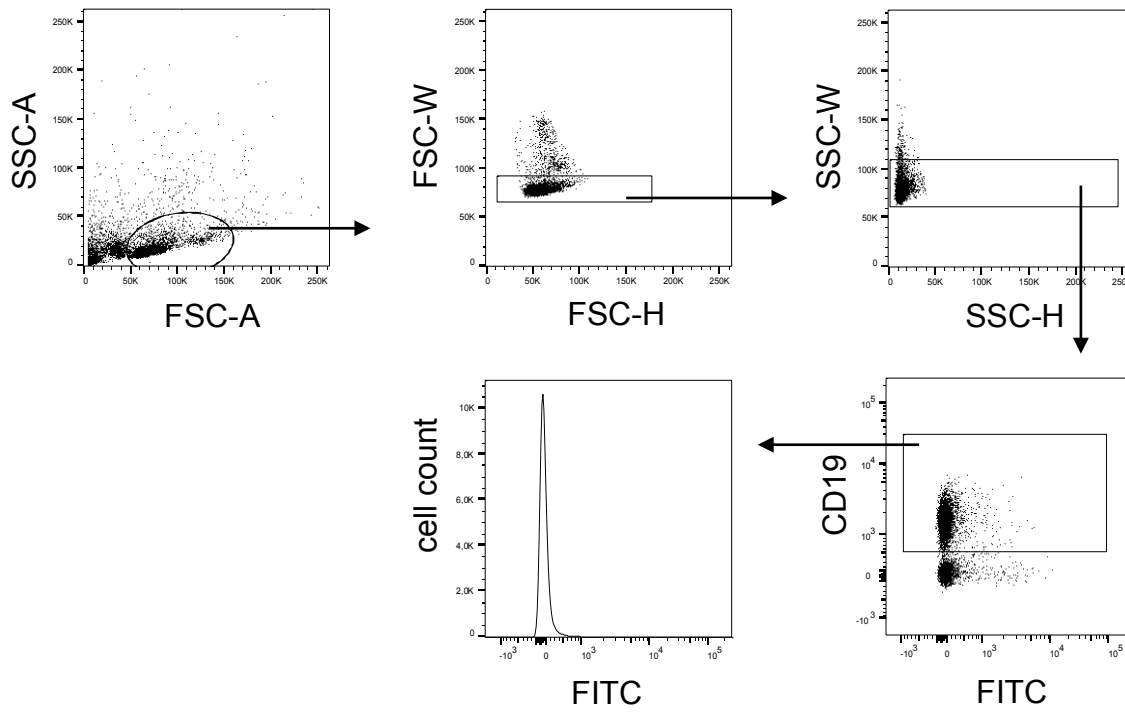


Supplementary Figure 7:

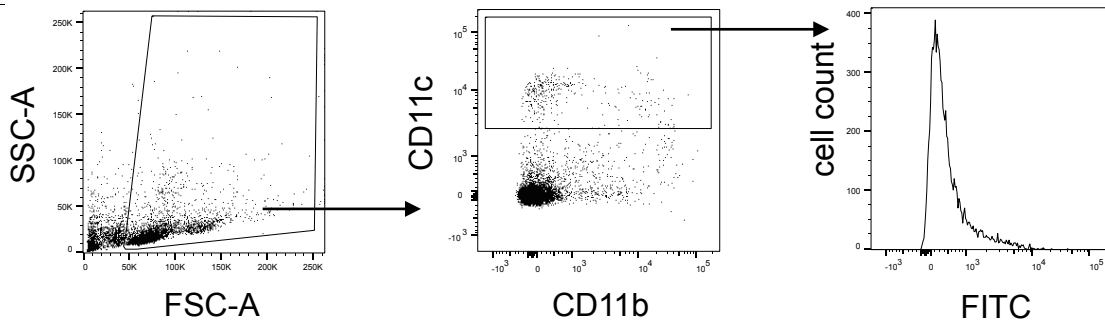
Uptake of pOVA-PEG20 by different hematopoietic APC subsets.

Mice received PBS (grey) or 100 μ g (based on peptide amount) pOVA-PEG20 (black) conjugated to FITC *i.v.*. After one hour mice were sacrificed and secondary lymphoid organs were isolated. Cells were stained with surface markers and analyzed flow cytometry. APC populations were characterized as follows: B cells (CD19⁺), DCs (CD11c⁺) and macrophages (CD11c⁻ CD11b⁺). Representative histograms of FITC-intensity. Representative data from two independent experiments.

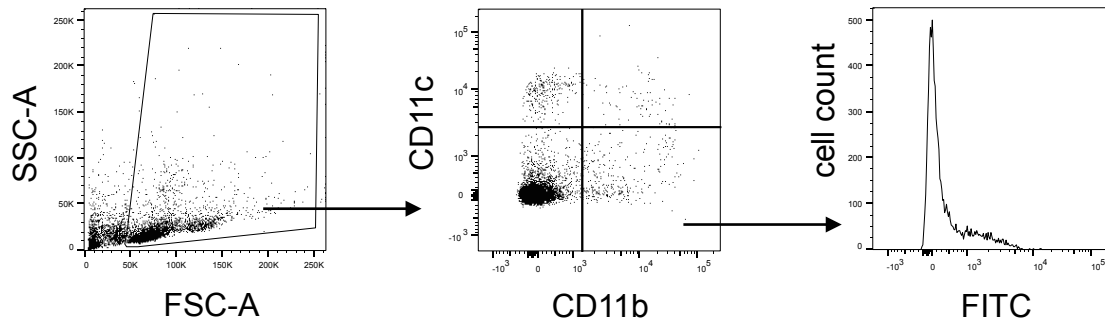
B cells



DCs



Macrophages



Supplementary Figure 8
Gating strategy for the identification of cells taking up pOVA-PEG20-FITC