

PNAS

www.pnas.org

Supplementary Information for

Metformin rescues Parkinson's disease phenotypes caused by hyperactive mitochondria

Danielle E. Mor, Salman Sohrabi, Rachel Kaletsky, William Keyes, Alp Tartici, Vrinda Kalia, Gary W. Miller, and Coleen T. Murphy

Coleen T. Murphy
ctmurphy@princeton.edu

This PDF file includes:

Figures S1 to S5
Legends for Datasets S1 to S2
SI References

Other supplementary materials for this manuscript include the following:

Datasets S1 to S2

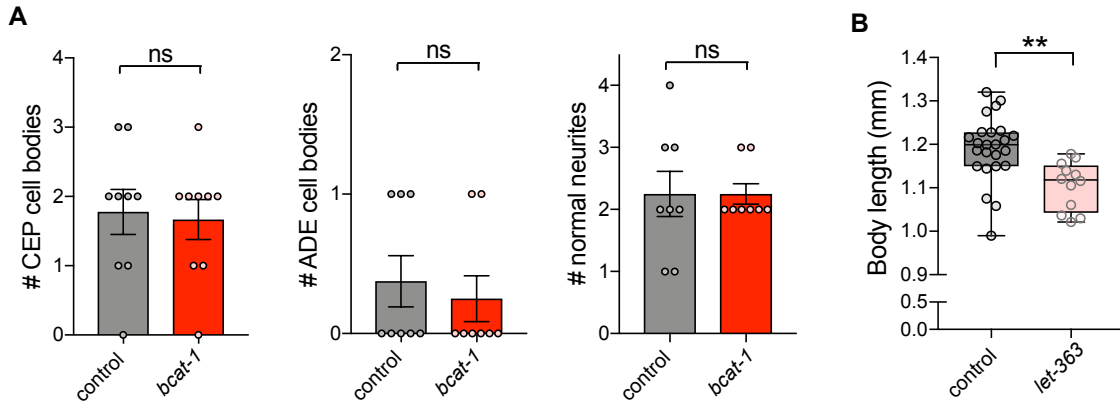


Fig. S1. A, α -synuclein-expressing dopaminergic neurons with or without *bcat-1* knockdown are largely degenerated by day 8. $n=9$ each for control and *bcat-1* CEP, 8 each for control and *bcat-1* ADE and neurites. Data are mean \pm s.e.m. **B**, Whole-life *let-363* RNAi causes a growth defect in TU3311 worms, measured on day 1 of adulthood. $n=24$ for control, 12 for *let-363*. Two-tailed *t*-tests. ns, not significant. ** $P<0.01$. Box-plots show minimum, 25th percentile, median, 75th percentile, maximum.

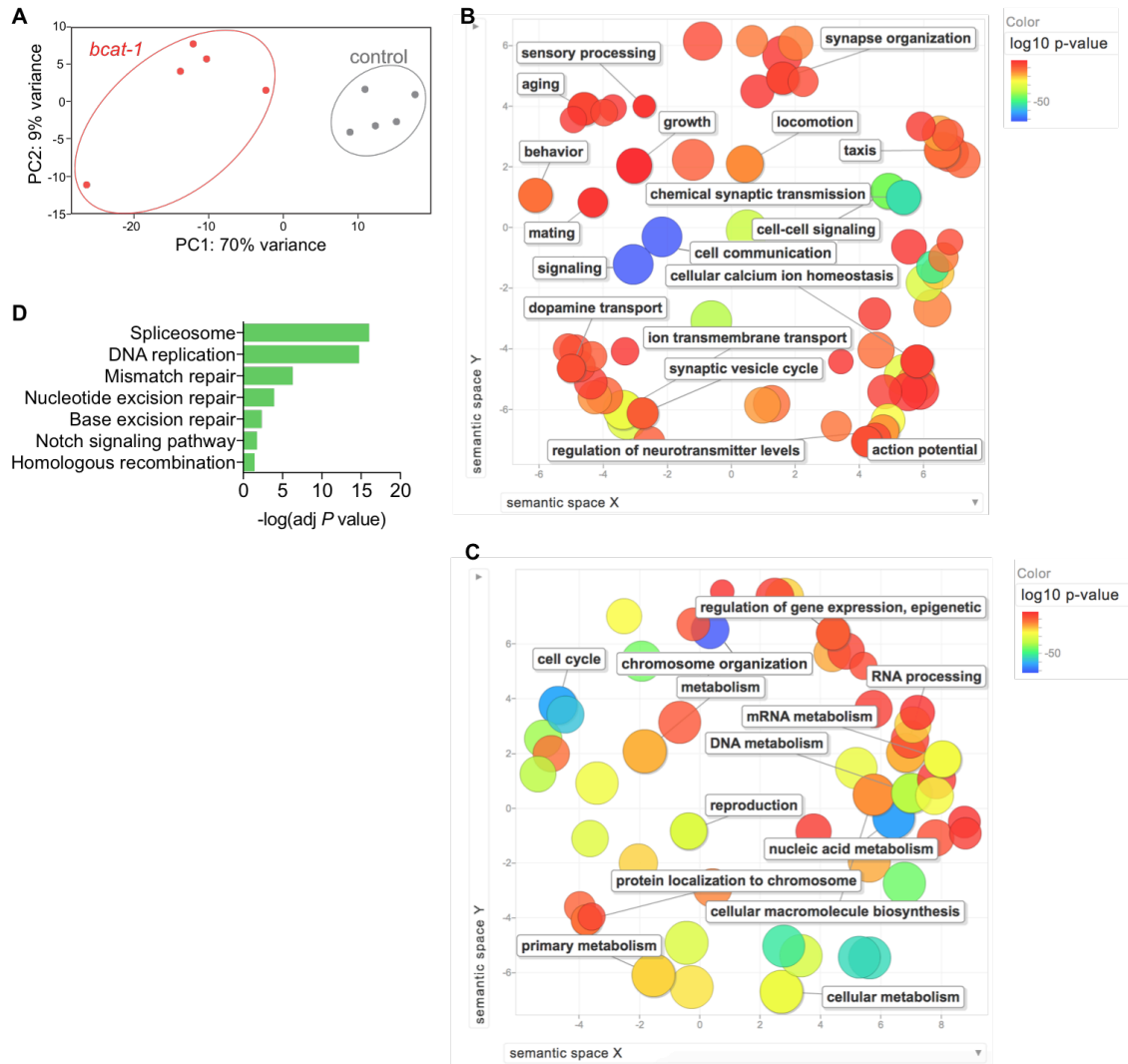


Fig. S2. A, PCA plot of neuronal transcriptomes from worms with *bcat-1* knockdown and controls on day 5. **B-C**, Gene Ontology analysis of significantly upregulated (**B**) and downregulated (**C**) genes in *bcat-1(RNAi)* neurons. **D**, KEGG pathway analysis of significantly downregulated genes in *bcat-1(RNAi)* neurons. FDR<0.05. n=5 independent collections/group.

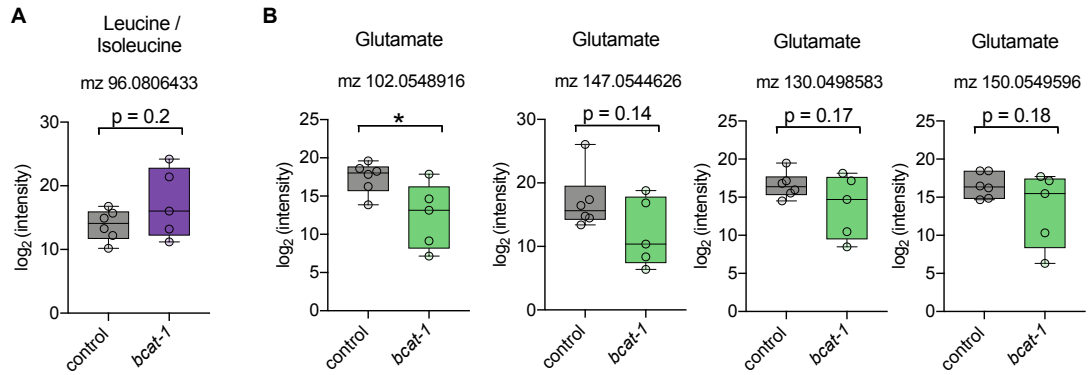


Fig. S3. A-B, Levels of additional features putatively annotated by mummichog as leucine/isoleucine (**A**) or glutamate (**B**) in *bcat-1(RNAi)* worms and controls. n=6 independent collections for control, 5 for *bcat-1*. Two-tailed *t*-tests. **P*<0.05. Box-plots show minimum, 25th percentile, median, 75th percentile, maximum.

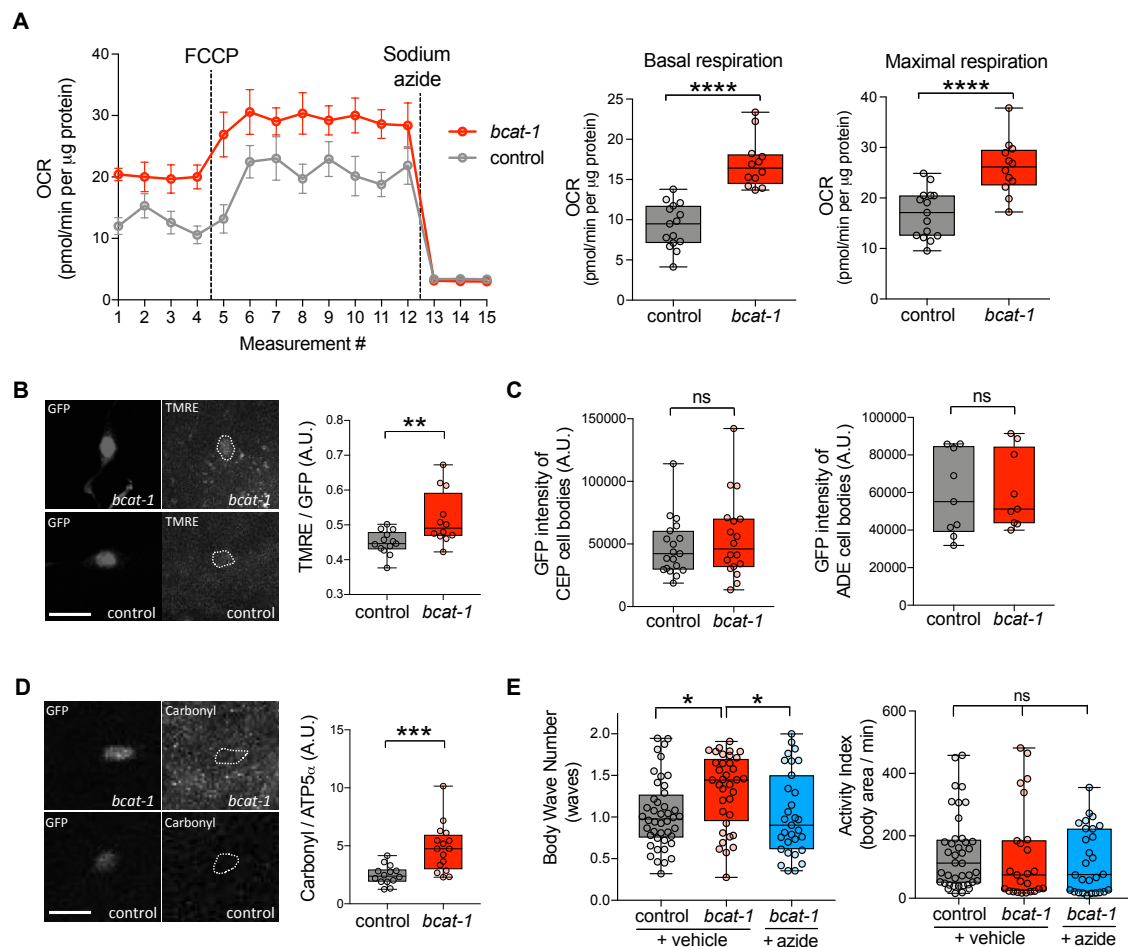


Fig. S4. A, Mitochondrial respiration was increased on day 5 in N2 worms (which are refractory to RNAi in neurons) fed *bcat-1* RNAi. $n=15$ wells totaling 152 worms for control, 12 wells totaling 111 worms for *bcat-1*. Two-tailed t -tests. **B**, Mitochondrial activity was increased on day 5 in ADE α -synuclein-expressing dopaminergic neurons with *bcat-1* knockdown. Scale bar, 10 μ m. $n=13$ worms totaling 13 ADEs for control, 11 worms totaling 12 ADEs for *bcat-1*. Two-tailed t -test. **C**, GFP intensity (used for normalization in each TMRE experiment) was unchanged by *bcat-1* knockdown in α -synuclein-expressing dopaminergic neurons, as measured on day 5. $n=9$ worms totaling 19 CEPs for control, 10 worms totaling 18 CEPs for *bcat-1*, and 9 worms totaling 9 ADEs each for control and *bcat-1*. Two-tailed t -tests. **D**, Protein carbonylation was increased on day 8 in ADE α -synuclein-expressing dopaminergic neurons with *bcat-1* knockdown. Scale bar, 10 μ m. $n=10$ worms totaling 14 ADEs for control, 10 worms totaling 16 ADEs for *bcat-1*. Two-tailed t -test.

E, *bcat-1(RNAi)* worms (strain CF512) treated with 100 μ M sodium azide starting on day 4 had improved body wave number on day 8, while total activity levels remained unchanged. n=28 worms for control, 41 for *bcat-1*+vehicle, 30 for *bcat-1*+azide. One-way ANOVA with Tukey's post-hoc. A.U., arbitrary units. ns, not significant. * P <0.05, ** P <0.01, *** P <0.001, **** P < 0.0001. Box-plots show minimum, 25th percentile, median, 75th percentile, maximum.

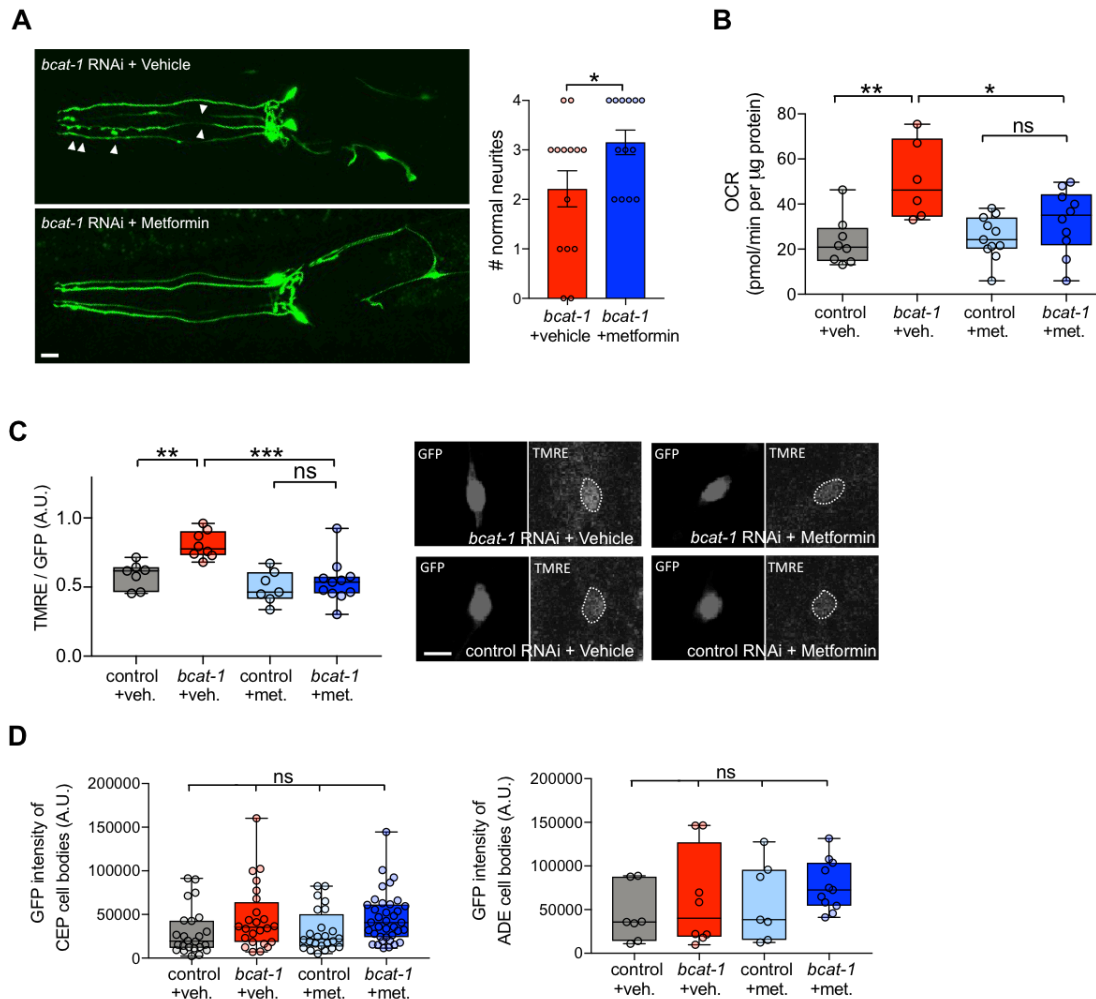


Fig. S5. A, Metformin reduced degenerated morphologies (blebbing and fragmentation, arrowheads) in α -synuclein-expressing dopaminergic neurites with *bcat-1* knockdown, as measured on day 6, following only two days of treatment. Scale bar, 10 μ m. n=14 for vehicle, 13 for metformin. Two-tailed *t*-test. Data are mean \pm s.e.m. **B**, Metformin reduced maximal mitochondrial respiration in *bcat-1(RNAi)* worms (strain CF512) on day 8. n=8 wells totaling 119 worms for control vehicle, 6 wells totaling 50 worms for *bcat-1* vehicle, 11 wells totaling 139 worms for control metformin, 10 wells totaling 123 worms for *bcat-1* metformin. Two-way ANOVA with Tukey's post-hoc. **C**, Metformin reduced mitochondrial activity in α -synuclein-expressing ADE dopaminergic neurons with *bcat-1* knockdown, as measured on day 6. Scale bar, 5 μ m. n=7 worms totaling 7 ADEs for control vehicle, 7 worms totaling 8 ADEs for *bcat-1* vehicle, 7 worms

totaling 7 ADEs for control metformin, 10 worms totaling 11 ADEs for *bcat-1* metformin. Two-way ANOVA with Tukey's post-hoc. **D**, GFP intensity (used for normalization in each TMRE experiment) was unchanged by either *bcat-1* knockdown or metformin treatment in α -synuclein-expressing dopaminergic neurons, as measured on day 6. n=8 worms totaling 24 CEPs for control vehicle, 7 worms totaling 24 CEPs for *bcat-1* vehicle, 9 worms totaling 24 CEPs for control metformin, 10 worms totaling 37 CEPs for *bcat-1* metformin, 7 worms totaling 7 ADEs for control vehicle, 7 worms totaling 8 ADEs for *bcat-1* vehicle, 7 worms totaling 7 ADEs for control metformin, 10 worms totaling 11 ADEs for *bcat-1* metformin. Two-way ANOVAs. A.U., arbitrary units. met., metformin. ns, not significant. veh., vehicle. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$. Box-plots show minimum, 25th percentile, median, 75th percentile, maximum.

Dataset S1 (separate file). Differentially-expressed genes in *bcat-1(RNAi)* neurons. Lists of all genes significantly up- or downregulated in *bcat-1(RNAi)* neurons (FDR<0.05), as well as subsets of each that were previously identified as neuronally-expressed (1).

Dataset S2 (separate file). Annotated features significant at $p < 0.05$ from metabolomics analysis of *bcat-1(RNAi)* worms. m.z, mass-to-charge ratio; retention time, retention time off of the column; KEGG.ID, KEGG ID determined by mummichog hosted on metaboanalyst; Annotation, Compound name from KEGG. Level 5 confidence in annotation (2). Adduct, Metabolite adduct annotated; Mass.Diff, Difference in mass of adduct detected and true mass of adduct.

SI References

1. R. Kaletsky *et al.*, Transcriptome Analysis of Adult *Caenorhabditis elegans* Cells Reveals Tissue-specific Gene and Isoform Expression. *PLoS Genet.* **14**, e1007559 (2018).
2. E. L. Schymanski *et al.*, Identifying small molecules via high resolution mass spectrometry: communicating confidence. *Environ. Sci. Technol.* **48**, 2097–8 (2014).