

## Supplementary material

### **The inhibition of NLRP3 inflammasome and IL-6 by *Hibiscus noldeae Baker f.* derived constituents provides a link to its anti-inflammatory therapeutic potentials**

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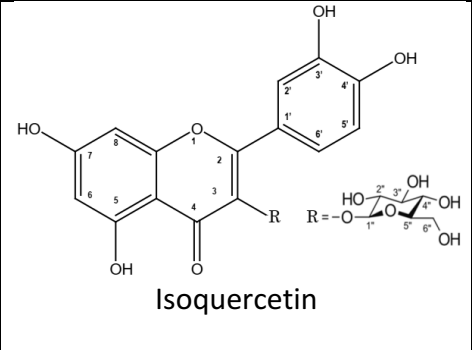
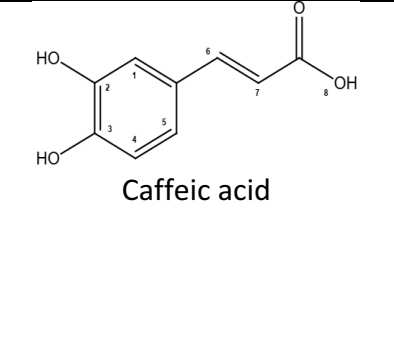
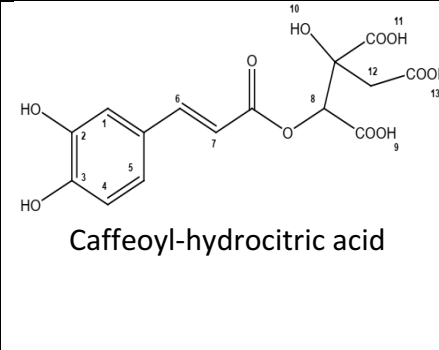
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**Table S.1:** The  $^1\text{H}$  NMR chemical shifts for the 3 purified compounds from *H. noldeae* ( $\text{DMSO-}d_6$ , 500 MHz).

 <p>Isoquercetin</p>		 <p>Caffeic acid</p>		 <p>Caffeoyl-hydrocitric acid</p>	
Chemical shifts (ppm)	Protons	$\delta_{\text{H}}$ (ppm), Chemical shifts (ppm)	Protons	Chemical shifts (ppm)	Protons
12.64 (1H, s)	OH-5	12.15 (1H, s)	H-8	13.3-12.35 (3H, s)	H-9, 11, 13
10.86 (1H, s)	OH-7	9.52 (1H, s)	H-3	9.64 (1H, s)	OH-2
9.72 (1H, s)	OH-3'	9.12 (1H, s)	H-2	9.18 (1H, s)	OH-3
9.15 (1H, s)	OH-4'	7.41 (1H, d)	H-6	7.57 (1H, d)	H-6
7.67 (1H, dd)	H-2'	7.02 (1H, d)	H-1	7.07 (1H, d)	H-1
7.52 (1H, d)	H-6'	6.96 (1H, dd)	H-5	7.03 (1H, dd)	H-5
6.81 (1H, d)	H-5'	6.75 (1H, d)	H-4	6.78 (1H, d)	H-4
6.40 (1H, d)	H-8	6.16 (1H, d)	H-7	6.31 (1H, d)	H-7
6.20 (1H, d)	H-6			5.24 (1H, s)	H-8
5.38 (1H, d)	H-1''			3.17 (2H, s)	H-12
5.12 (1H, s)	OH-3''				
4.84 (1H, s)	OH-4''				
4.42 (2H, s)	OH-6''				
3.65 (1H, s)	H-6''a				
3.57 (1H, m)	H-6''b				

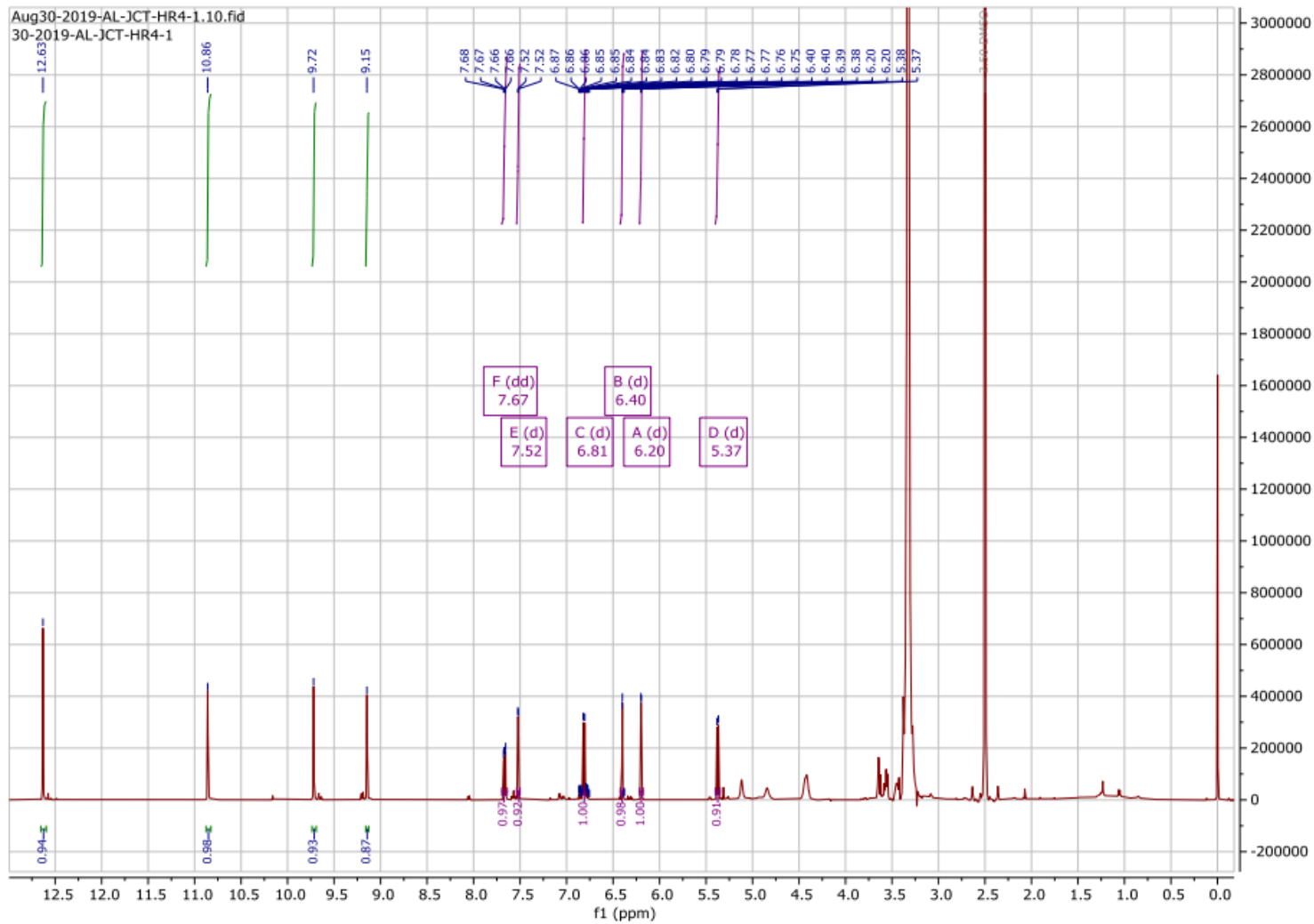


Figure S.1:  $^1\text{H}$ -NMR spectrum of Quercetin-3-O-glucoside (Isoquercetin) ( $\text{DMSO-d}_6$ , 500 MHz)

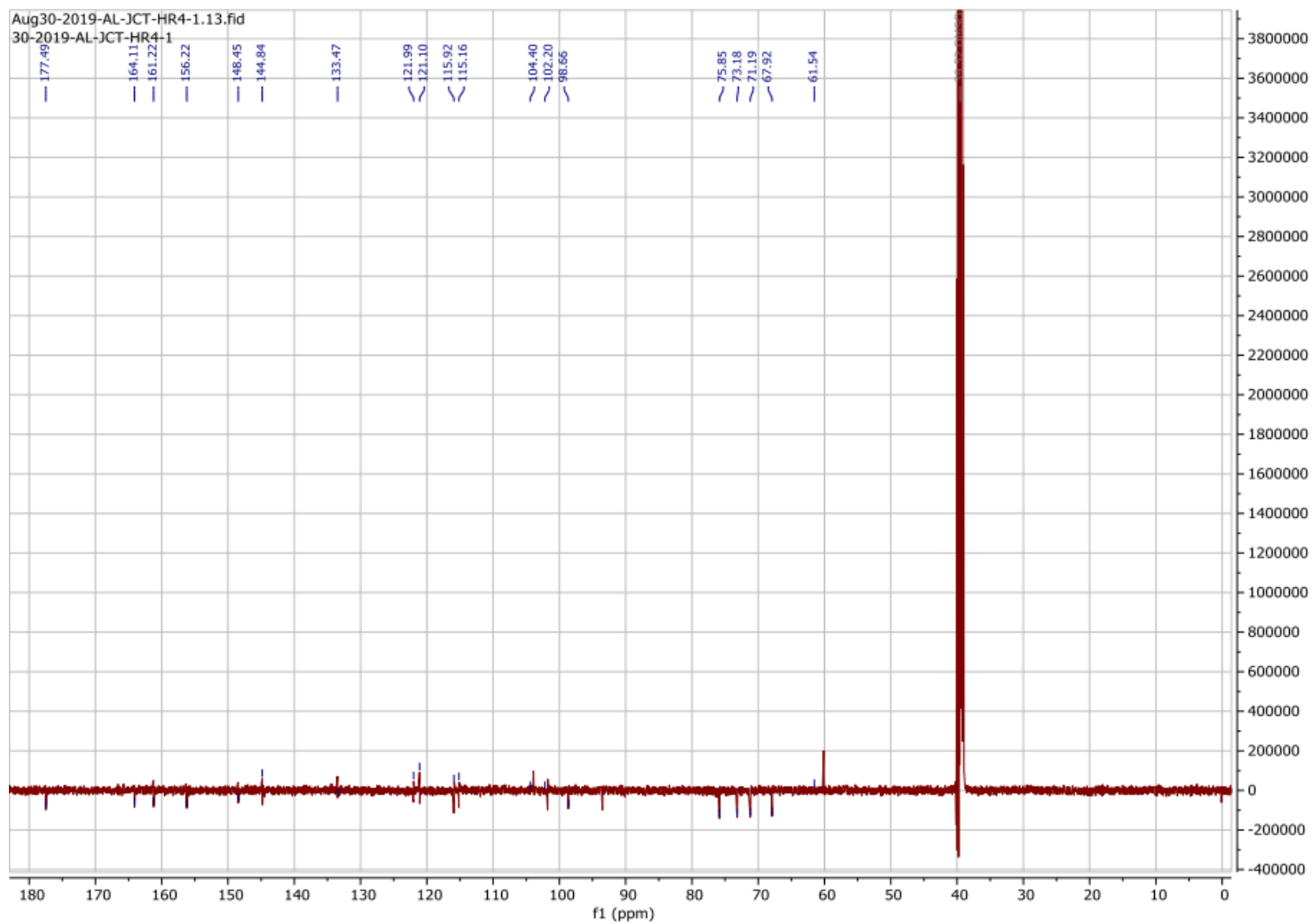


Figure S.2:  $^{13}\text{C}$  spectrum of Quercetin-3-O-glucoside (Isoquercetin) ( $\text{DMSO-d}_6$ , 500 MHz)

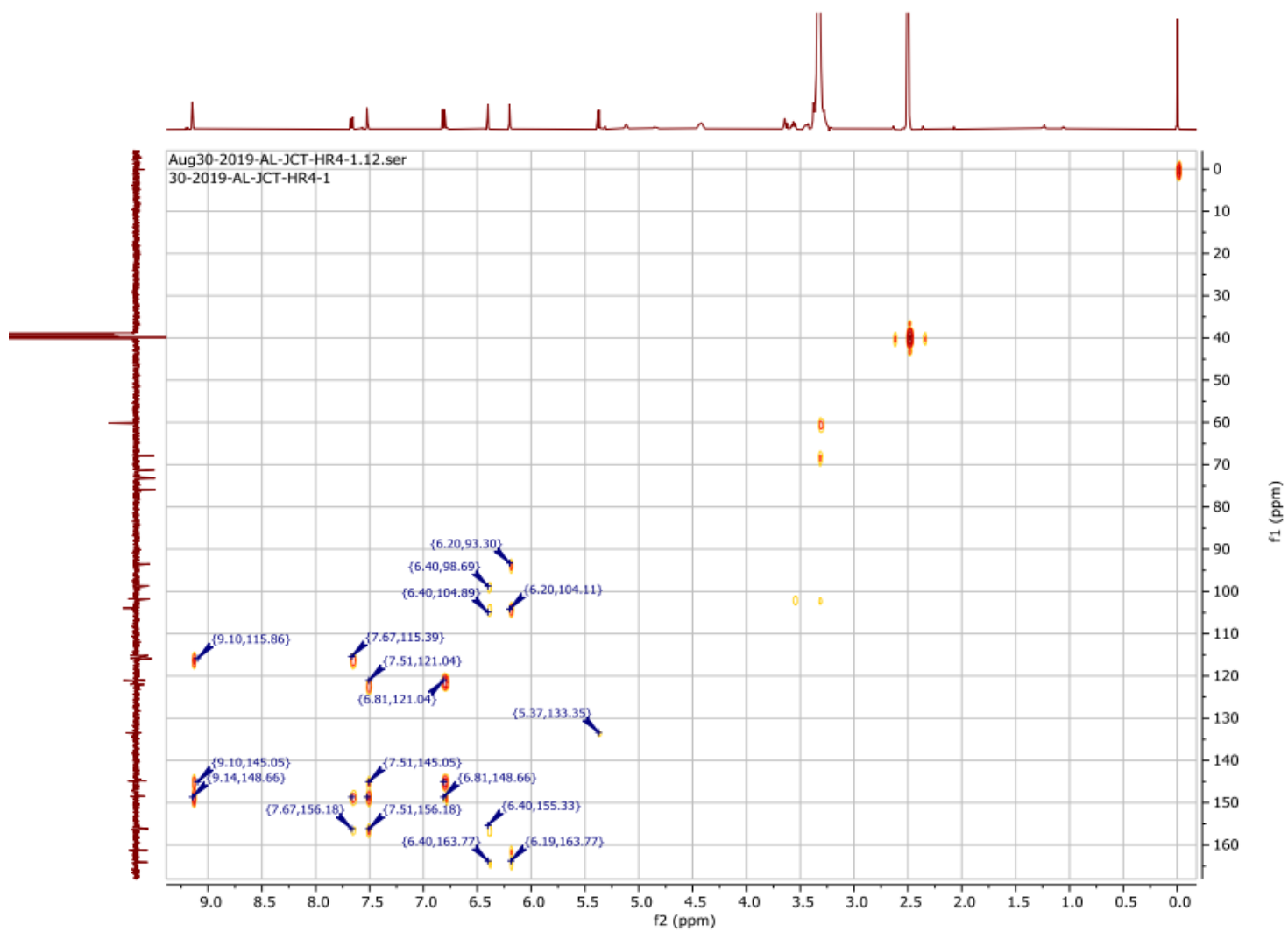
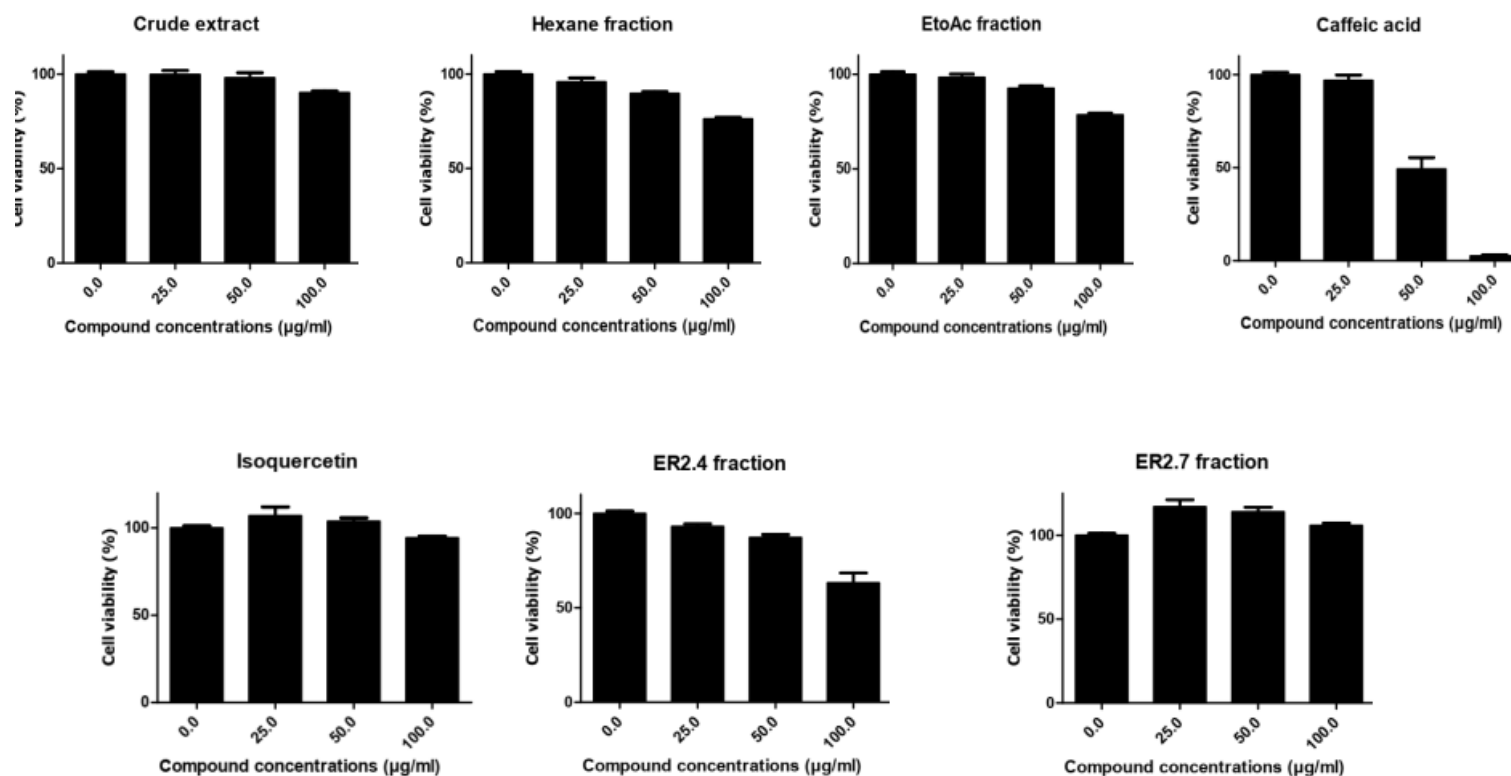
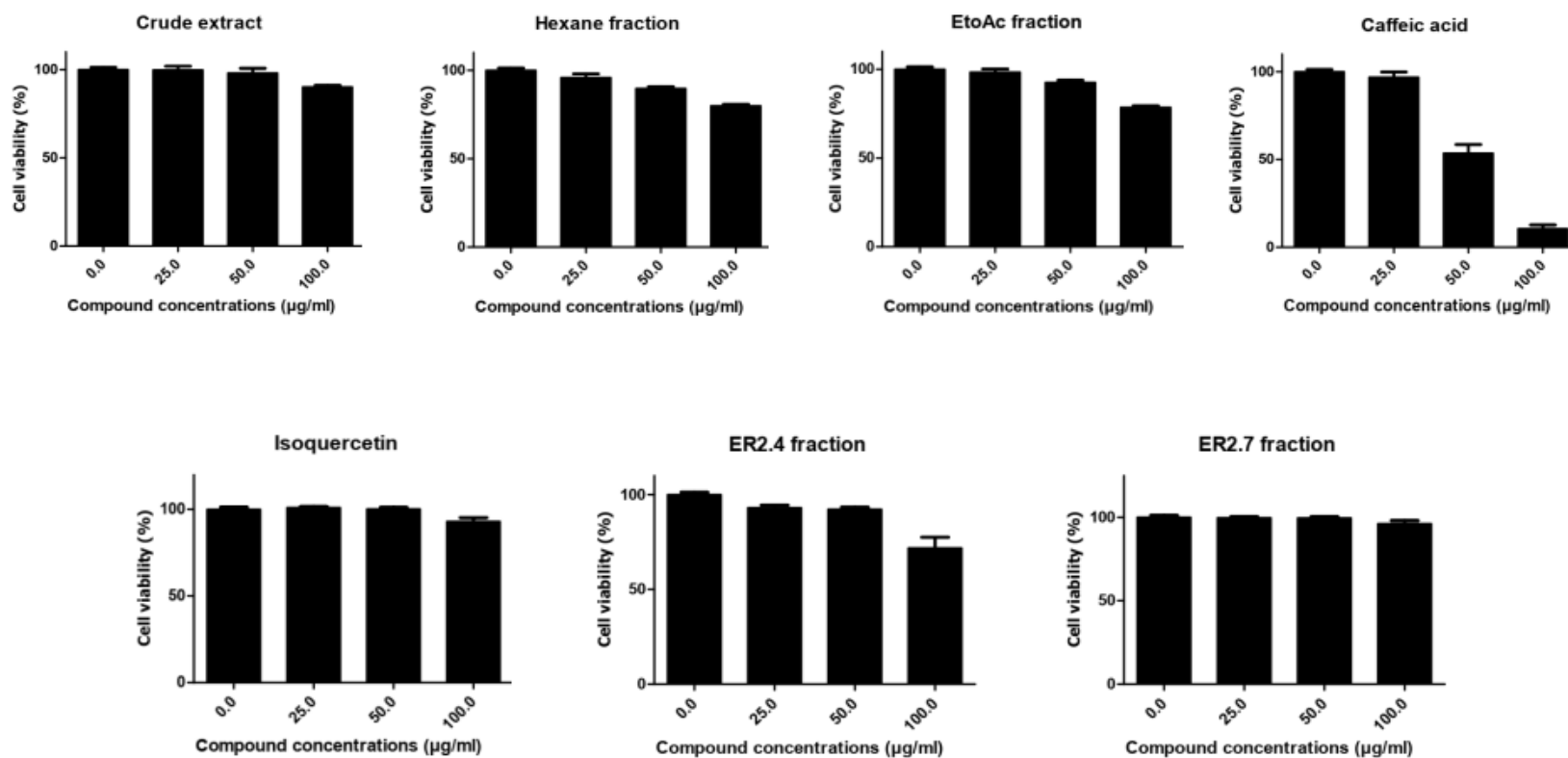


Figure S.3: <sup>1</sup>H, <sup>13</sup>C HMB of Quercetin-3-O-Glucoside (Isoquercetin) (DMSO-d<sub>6</sub>, 500 MHz)



**Figure S.4: Effect of different *H. noldeae*'s constituents on the THP-1 cell viability.** Human leukemia THP-1 monocytes were cultured in RPMI 1640 supplemented with 10% heat-inactivated Fetal Bovine Serum (FBS), 1% glutamine and 1% of penicillin-streptomycin and maintained at 37 °C under 5% CO<sub>2</sub> in a humidified atmosphere. Cells were passaged at least 5 times before any experiment. Cells were then incubated with different compounds at given concentrations for 24 h at 37°C, 5%CO<sub>2</sub> in a humidified atmosphere. The cell viability was then evaluated by celltiter glo luminescent cell viability assay (Promega) according to the manufacturer instructions. Viability (%) = [RLU (treated cells)/ RLU (control cells)] x 100.



**Figure S.5: Effect of different *H. noldeae*'s constituents on the RAW264.7 cell viability.** RAW264.7 murine cells were cultured in DMEM supplemented with 10% heat-inactivated FBS, 1% glutamine, 1% penicillin-streptomycin and 1% Sodium pyruvate and maintained at 37 °C under 5% CO<sub>2</sub> in a humidified atmosphere. Cells were passaged at least 5 times, and allowed to acclimate for 24h, before any experiment. Cells were then incubated with different compounds at given concentrations for 24 h at 37°C, 5%CO<sub>2</sub> in a humidified atmosphere. The cell viability was then evaluated by celltiter-glo luminescent cell viability assay (Promega) according to the manufacturer instructions. Viability (%) = [RLU (treated cells)/ RLU (control cells)] × 100.

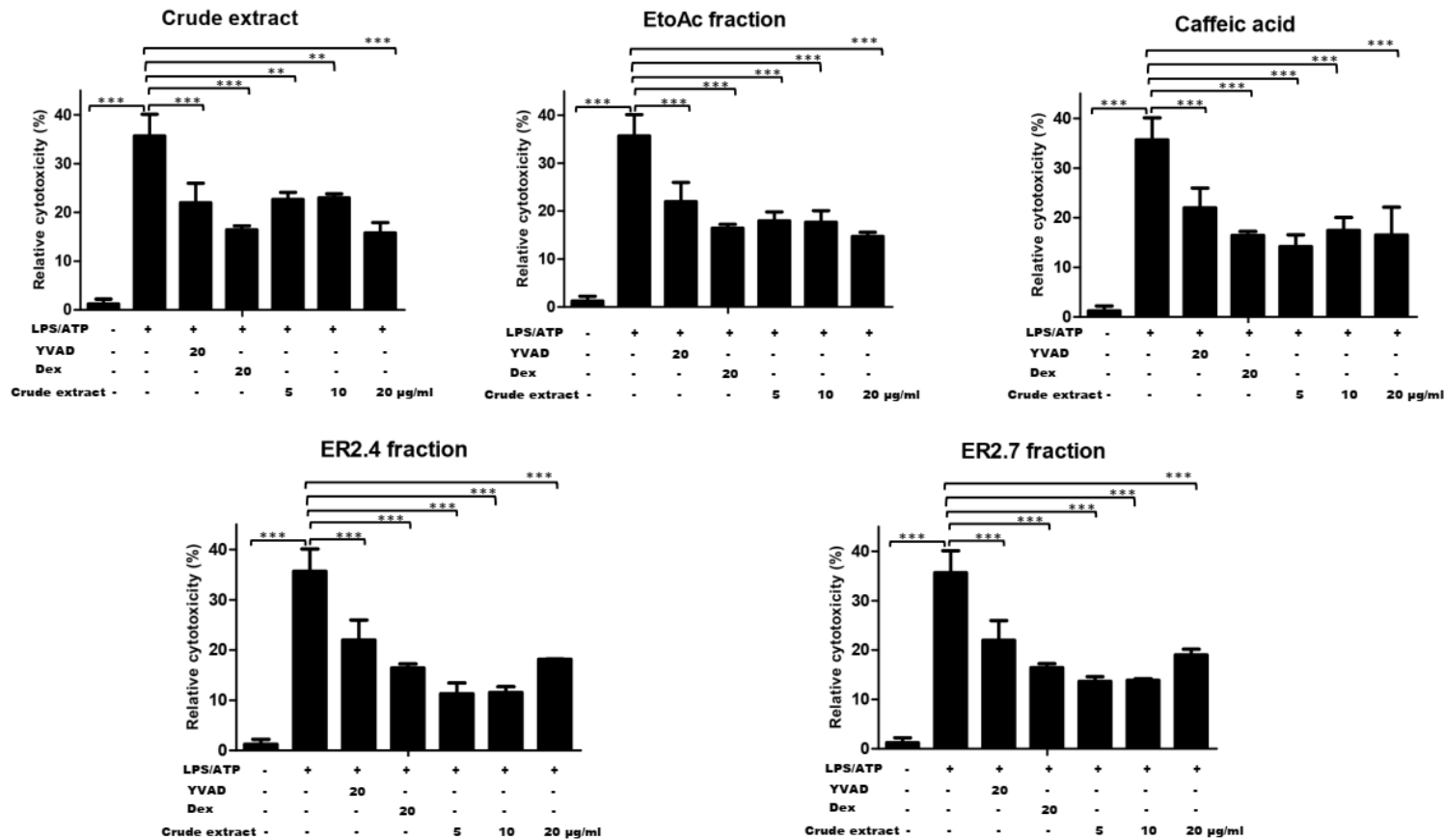


Figure S.6: Crude extract and the ethylacetate fraction as well as the caffeic acid and semi-purified fractions (ER 2.4 and ER 2.7), thereof exhibited significant inhibition of pyroptosis in THP-1 derived macrophages. PMA-differentiated cells were pretreated by corresponding compounds for 1 , and then primed by LPS (1µg/ml) for 3h. Cells were then stimulated by ATP (5mM) for additional 1 h. Samples(5µl) were collected and frozen in LDH Storage Buffer at 1:20 dilution. Samples were thawed, further diluted 5-fold and 50µl of diluted samples were added to 50µl LDH Detection Reagent. Luminescence was recorded after 60 min incubation at room temperature. The % Cytotoxicity = 100 X (Experimental LDH Release – Medium Background)/(Maximum LDH Release Control-Medium Background).