

SUPPORTING INFORMATION

Targeted amino acid substitutions in a *Trichoderma* peptaibol confer activity against fungal plant pathogens and protect host tissues from *Botrytis cinerea* infection

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I. Elmaghraby,^{2,3} S. Tundo,² F. Favaron²**

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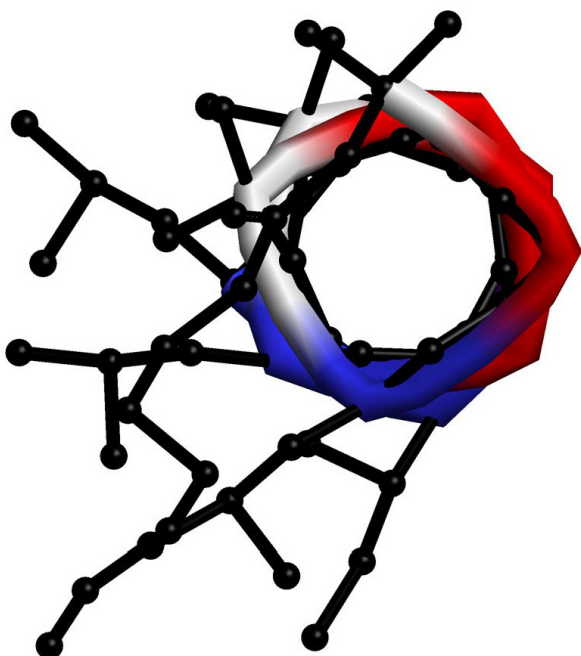
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³*Agricultural Research Center, Central Laboratory of Organic Agriculture, 9, Cairo Univ. St., Giza 12619, Egypt*

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native trichogin GA IV

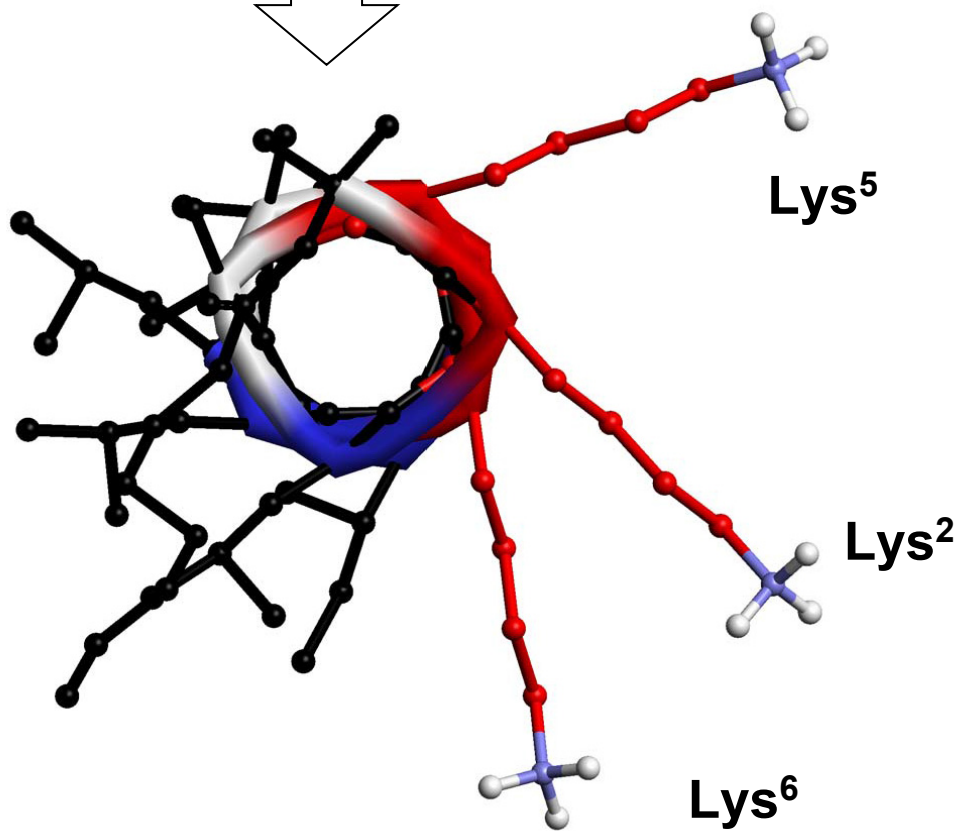
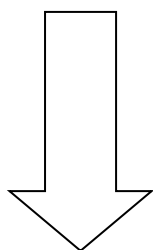


Figure S1. 3D-structure for compounds 1-5 and 7. View along the helical axis.

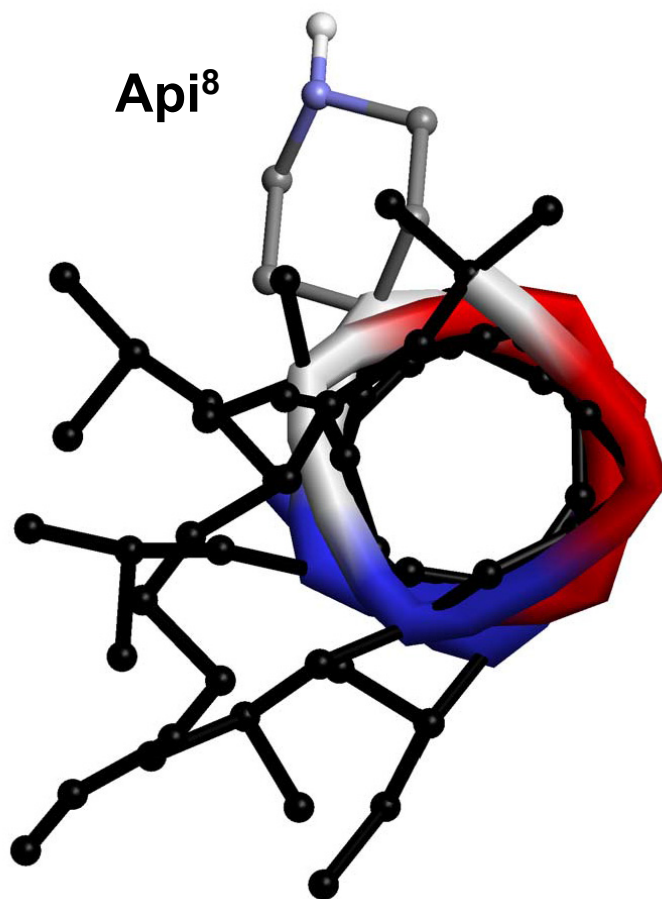


Figure S2. 3D-structure for compound **8**. View along the helical axis.

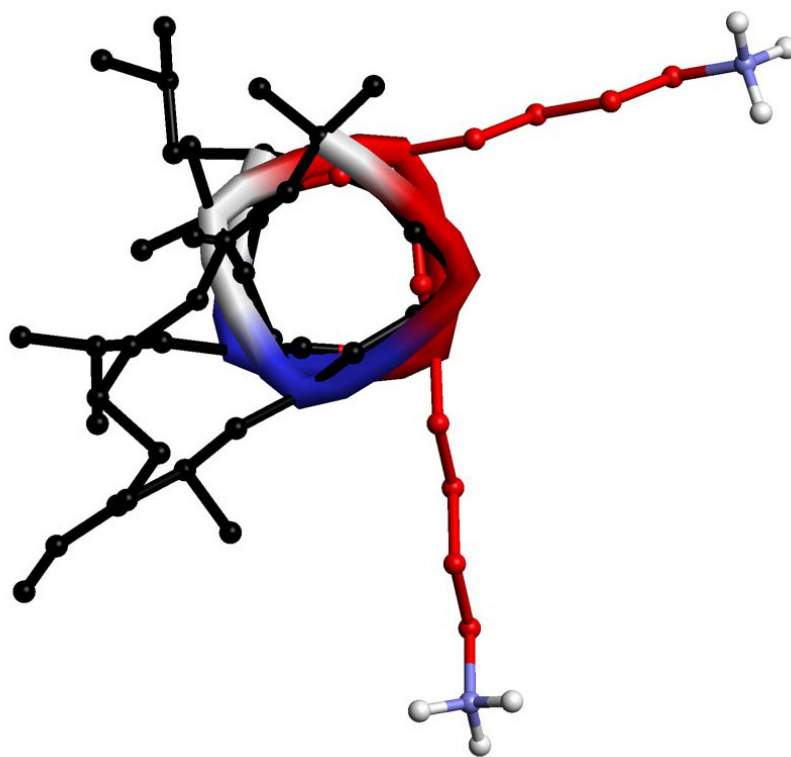


Figure S3. 3D-structure for compounds **4c** and **4c1**. View along the helical axis.

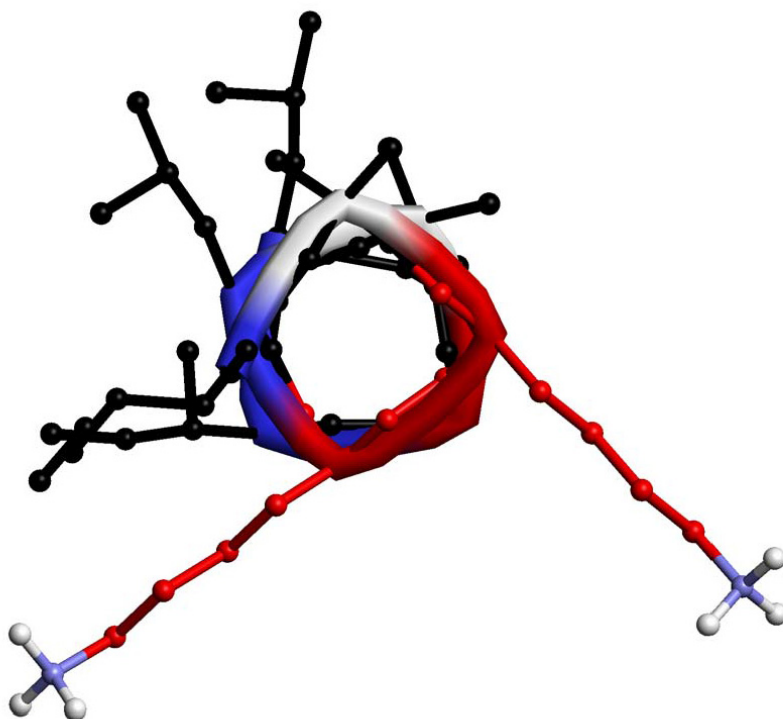


Figure S4. 3D-structure for compound **4c2**. View along the helical axis.



Untreated control

50 μM peptide 4

15 μM peptide 4

Fig. S5. Representative symptoms produced by *Botrytis cinerea* on common bean leaves (Borlotto var. Teggia) untreated or treated with the peptide 4 at 50 μM or 15 μM at 7 days post inoculation (dpi). Leaves were arranged on moist towels in plastic trays and inoculated on two or three points with 10 μL of a conidial suspension containing $2.5 \times 10^6 \text{ mL}^{-1}$ *B. cinerea* conidia in potato dextrose broth (PDB).



Untreated control

50 μ M peptide 4

Fig. S6. Representative symptoms produced by *Botrytis cinerea* on grape berries (cv. Glera) untreated or treated with the peptide 4 at 50 μ M at 4 days post inoculation (dpi). Ripe grape berries were surface-sterilized, rinsed with sterile water, separately arranged on a plastic tray and sprayed with *B. cinerea* conidia at a concentration of 1.85×10^6 mL⁻¹.

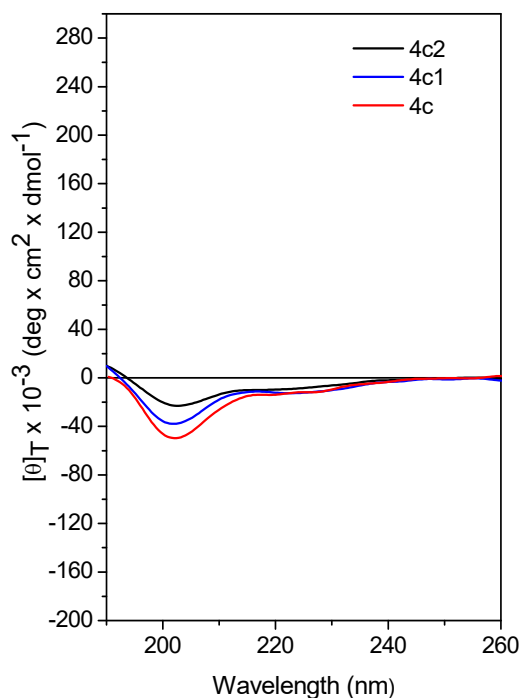


Fig. S7. CD profiles acquired for peptides 4c, 4c1 and 4c2 in aqueous solution (peptide concentration: 10^{-4} M).

Peptide Characterization (HPLC, ESI-HRMS, ¹H NMR)

Table S1. Sequences of the synthesized trichogin analogs.

n°	peptide sequence ^a
1	<i>n</i> Oct-Aib-Lys ² -Leu-Aib-Lys ⁵ -Gly-Leu-Aib-Gly-Ile-Lol
2	<i>n</i> Oct-Aib-Lys ² -Leu-Aib-Gly ⁵ -Gly-Leu-Aib-Gly-Ile-Lol
3	<i>n</i> Oct-Aib-Gly ² -Leu-Aib-Lys ⁵ -Gly-Leu-Aib-Gly-Ile-Lol
4	<i>n</i> Oct-Aib-Gly ² -Leu-Aib-Lys-Lys-Leu-Aib-Gly-Ile- Lol
4c	<i>n</i> Oct-Aib - Lys ² -Lys-Leu-Aib-Gly-Ile- Lol
4r	<i>n</i> Oct-Aib-Gly ² -Leu-Aib-Lys ⁵ -Lys-Leu-Aib-Gly-Ile-Leu-NH ₂
4c1	<i>n</i> Oct-Aib-Gly ² -Leu-Aib-Lys ⁵ -Lys-Leu - Leu-NH ₂
4c2	<i>n</i> Oct-Aib - Lys ² -Lys-Leu-Aib-Gly-Ile- Leu-NH ₂
5	<i>n</i> Oct-Aib-Gly ² -Leu-Aib-Gly ⁵ -Lys-Leu-Aib-Gly-Ile- Lol
6	<i>n</i> Oct-Aib-Gly ² -Leu-Aib-Lys ⁵ -Aib-Leu-Aib-Gly-Ile- Lol
7	<i>n</i> Oct-Aib-Lys ² -Leu-Aib-Gly ⁵ -Lys-Leu-Aib-Gly-Ile- Lol
8	<i>n</i> Oct-Aib-Gly ² -Leu-Aib-Gly ⁵ -Gly-Leu-Api ⁸ -Gly-Ile-Lol

^a *n*Oct, *n*-octanoyl; Aib, α-aminoisobutyric acid; Lol, leucinol; Api, 4-aminopiperidine-4-carboxylic acid.

Abbreviations:

ESI, Electron Spray Ionization

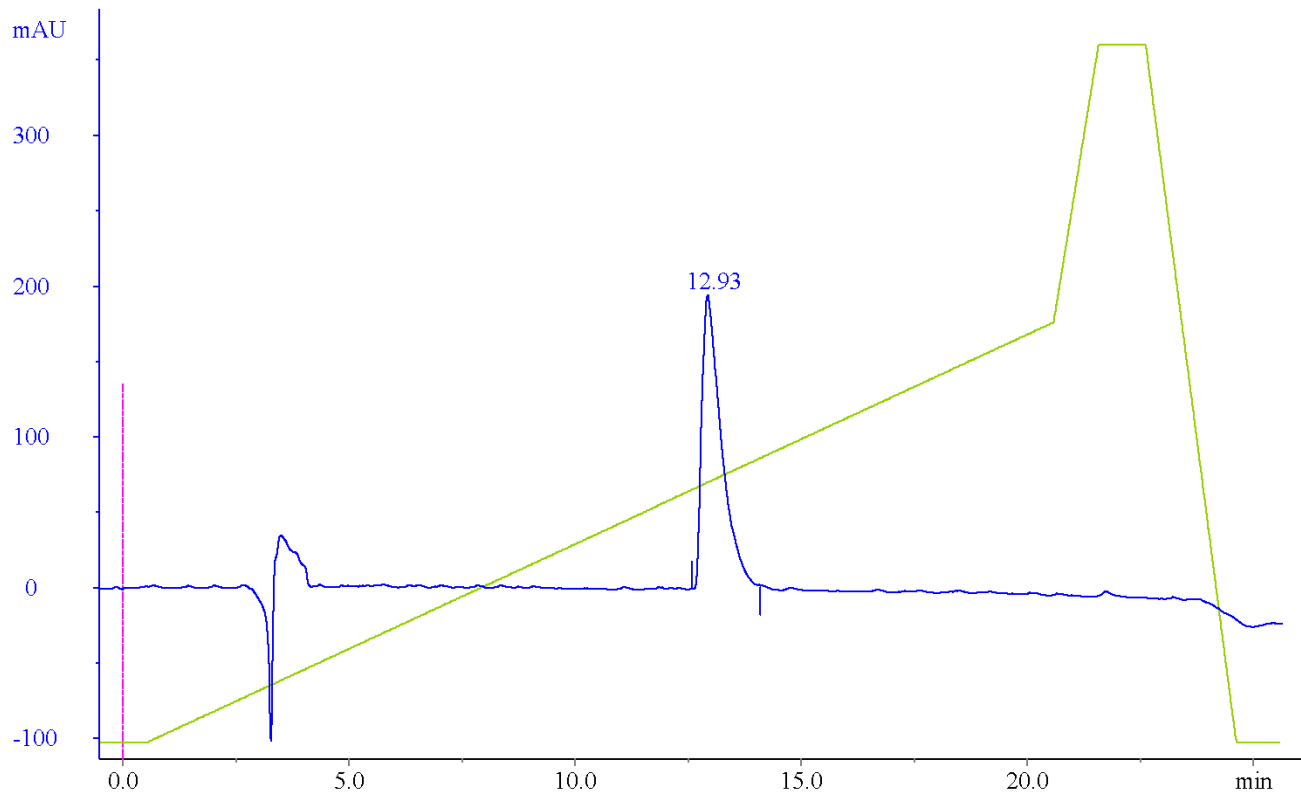
HRMS, High Resolution Mass Spectrometry

Eluants for HPLC:

A: H₂O/CH₃CN 9:1 + 0.05% TFA

B: CH₃CN/H₂O 9:1 + 0.05% TFA

peptide **1** (**K25**)

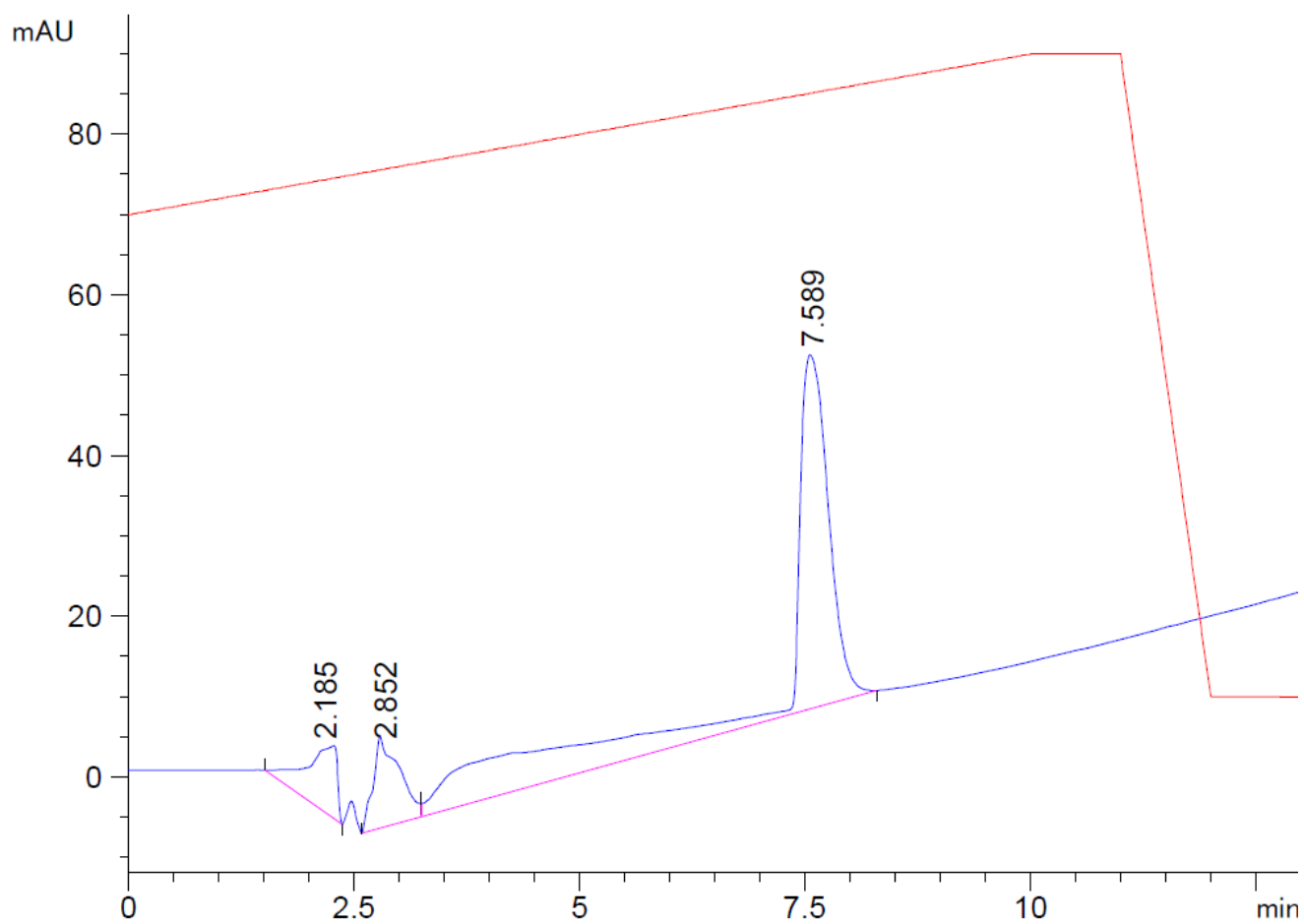


HPLC profile obtained for peptide **1** (gradient: 50→80%B [CH₃CN/H₂O 9:1 + 0.05% TFA] in 20 minutes; flow rate: 1mL/min; C₁₈ Phenomenex 300Å) Retention time (R_t) = 12.93 min. Detection Wavelength: 216 nm.



ESI-HRMS for peptide **1**: $[M+H]^+$ calcd. 1208.8632; $[M+H]^+$ found: 1208.8896 (*monoisotopic*), $[M+2H]^{2+}$ found: 604.9448.

peptide 2 (K2)

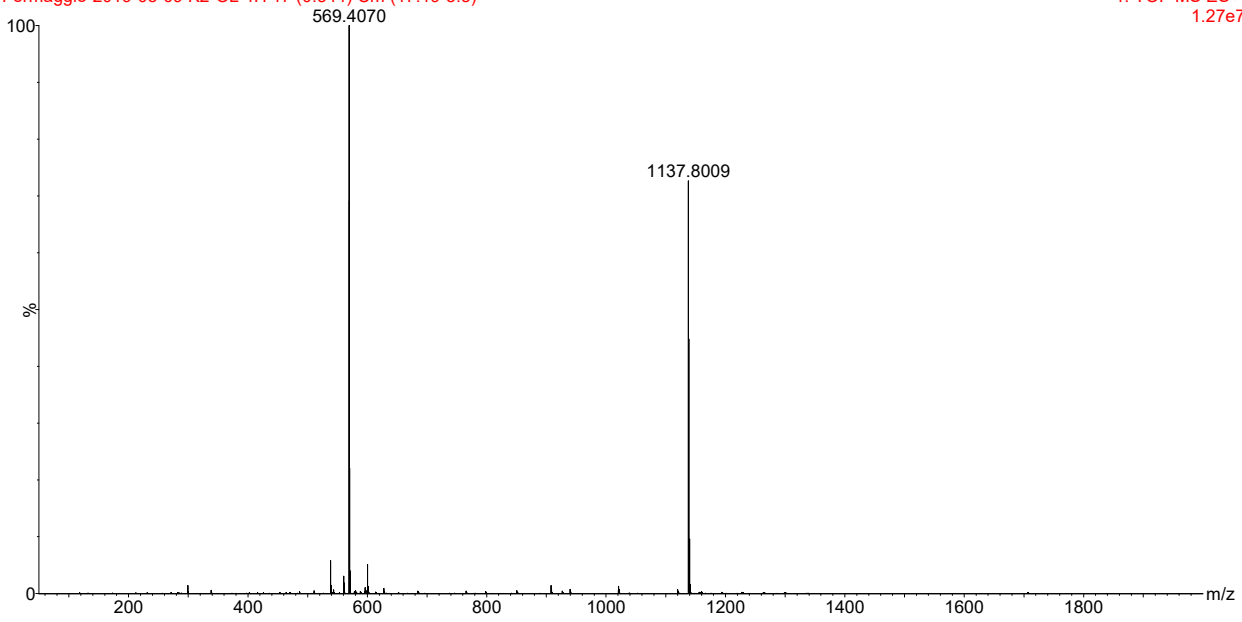


HPLC profile obtained for peptide 2 (gradient: 70→90%B [B: CH₃CN/H₂O 9:1 + 0.05% TFA] in 10 minutes; flow rate: 1mL/min; C₁₈ Phenomenex 100Å) Retention time (R_t) = 7.59 min. Detection Wavelength: 226 nm.

2KV - Dil 1-100 in acqua

Formaggio-2019-05-09 K2-OL 4H 17 (0.344) Cm (17:19-3:5)

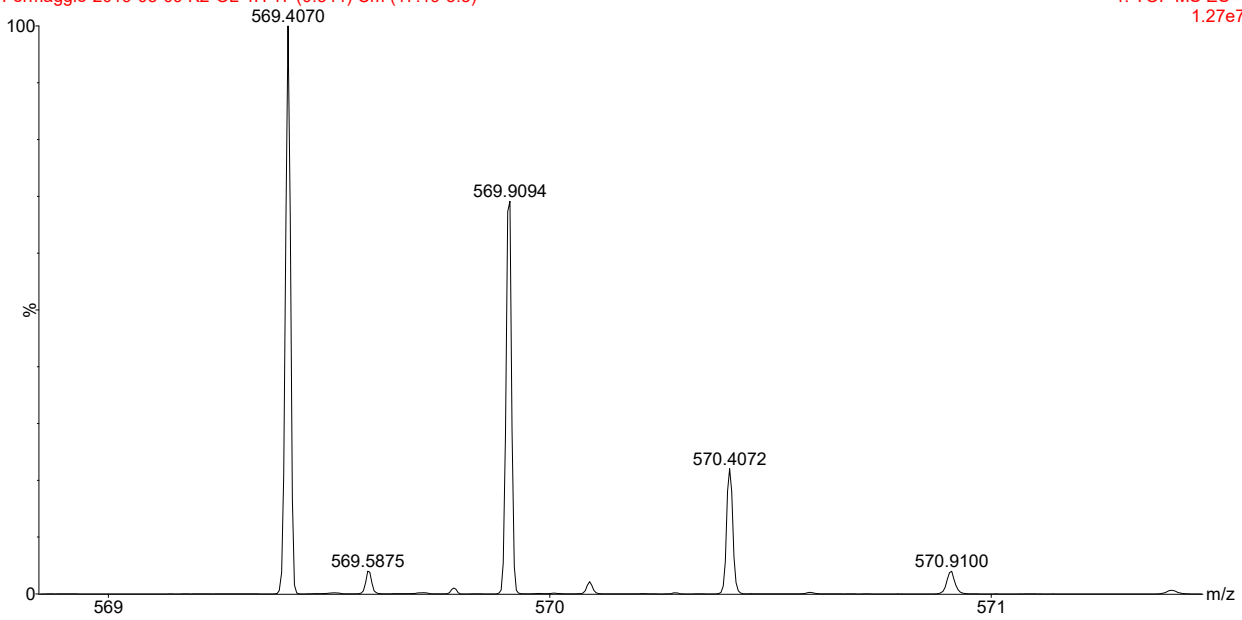
1: TOF MS ES+
1.27e7



2KV - Dil 1-100 in acqua

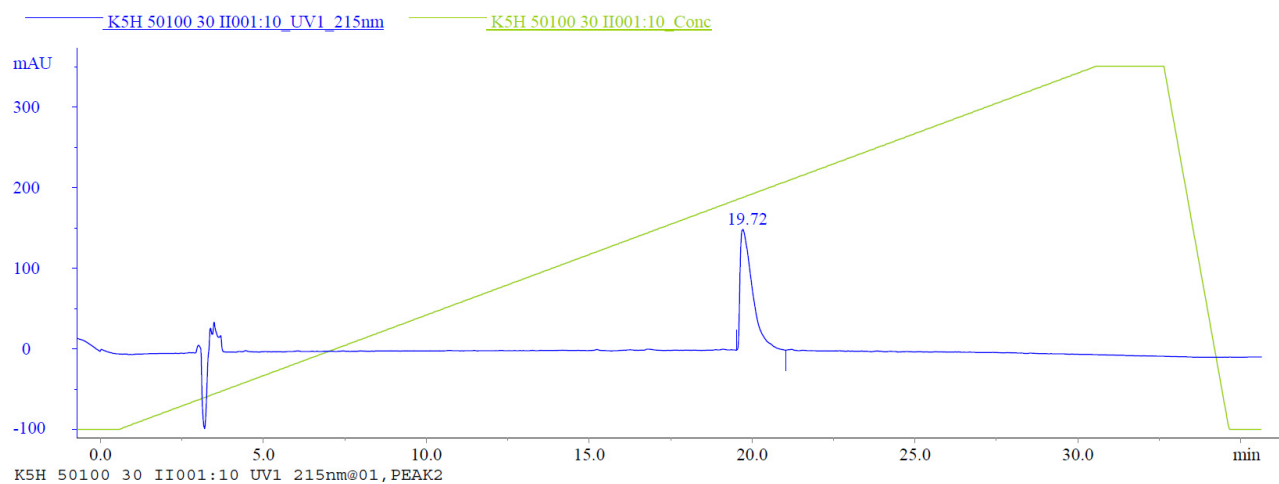
Formaggio-2019-05-09 K2-OL 4H 17 (0.344) Cm (17:19-3:5)

1: TOF MS ES+
1.27e7



ESI-HRMS for peptide 2: $[M+H]^+$ calcd. 1137.7997; $[M+H]^+$ found: 1137.8009 (*monoisotopic*), $[M+2H]^{2+}$ found: 569.4070.

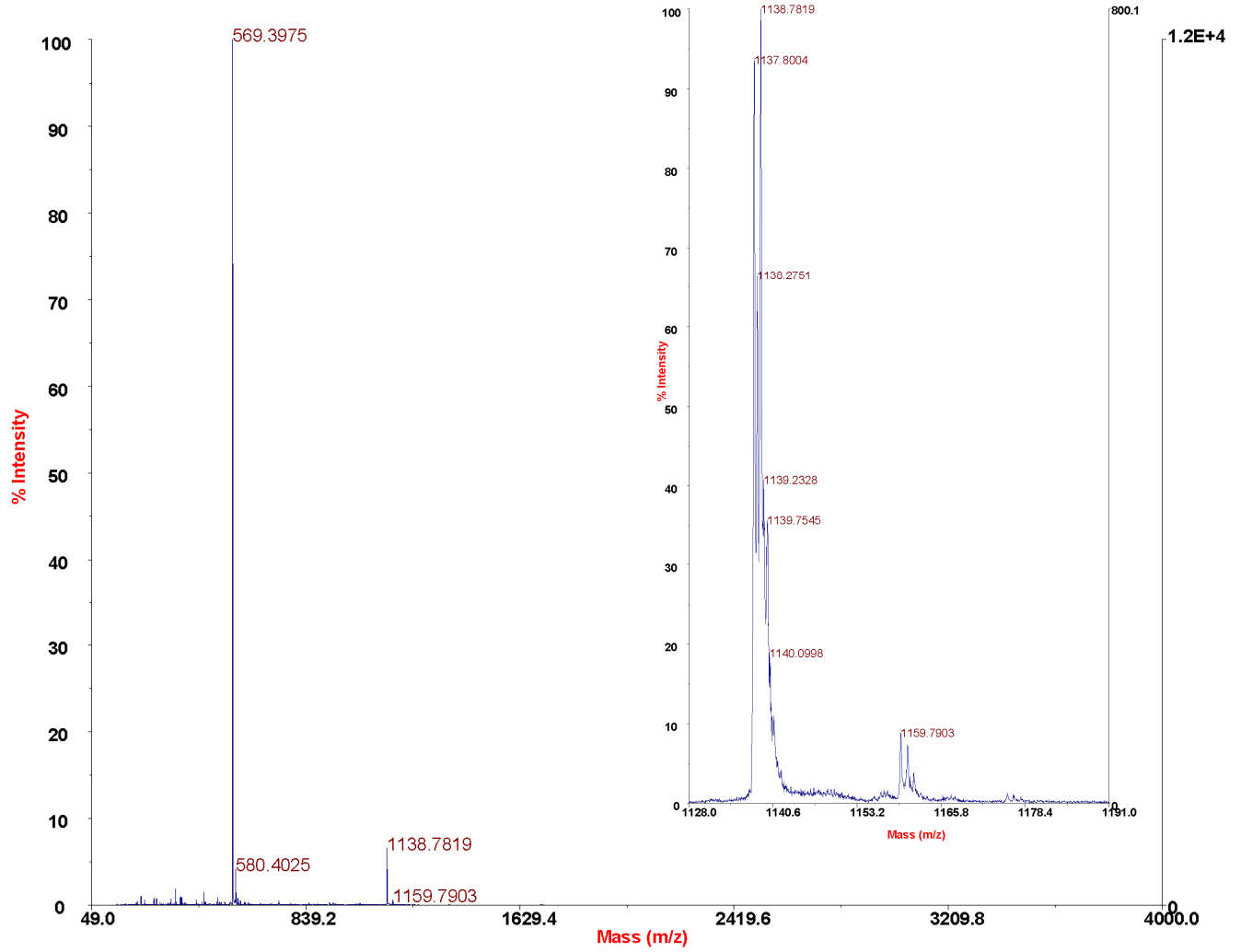
peptide 3 (K5)



HPLC profile obtained for peptide 3 (gradient: 50→100%B [B: CH₃CN/H₂O 9:1 + 0.05% TFA] in 30 minutes; flow rate: 1mL/min; C₁₈ Phenomenex 300Å) Retention time (R_t) = 19.72 min. Detection Wavelength: 215 nm.

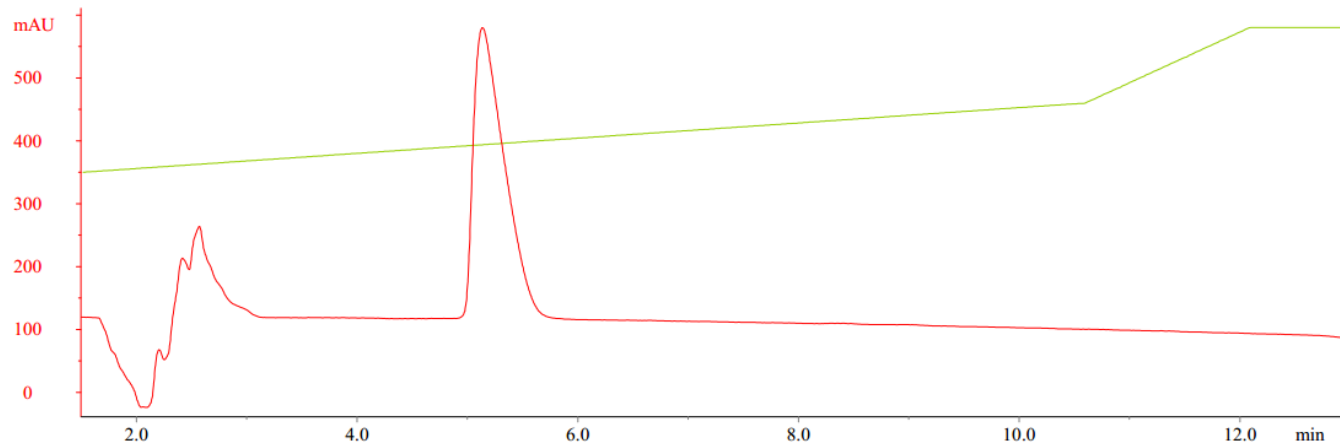
Applied Biosystems Mariner System 5220

Mariner Spec /8:11 (T/0.62:0.90) ASC[BP = 569.4, 12193]



ESI-HRMS for peptide 3: $[M+H]^+$ calcd. 1137.7997; $[M+H]^+$ found: 1137.8004 (monoisotopic), $[M+Na]^+$ found: 1159.7903 (monoisotopic), $[M+2H]^{2+}$ found: 569.3975.

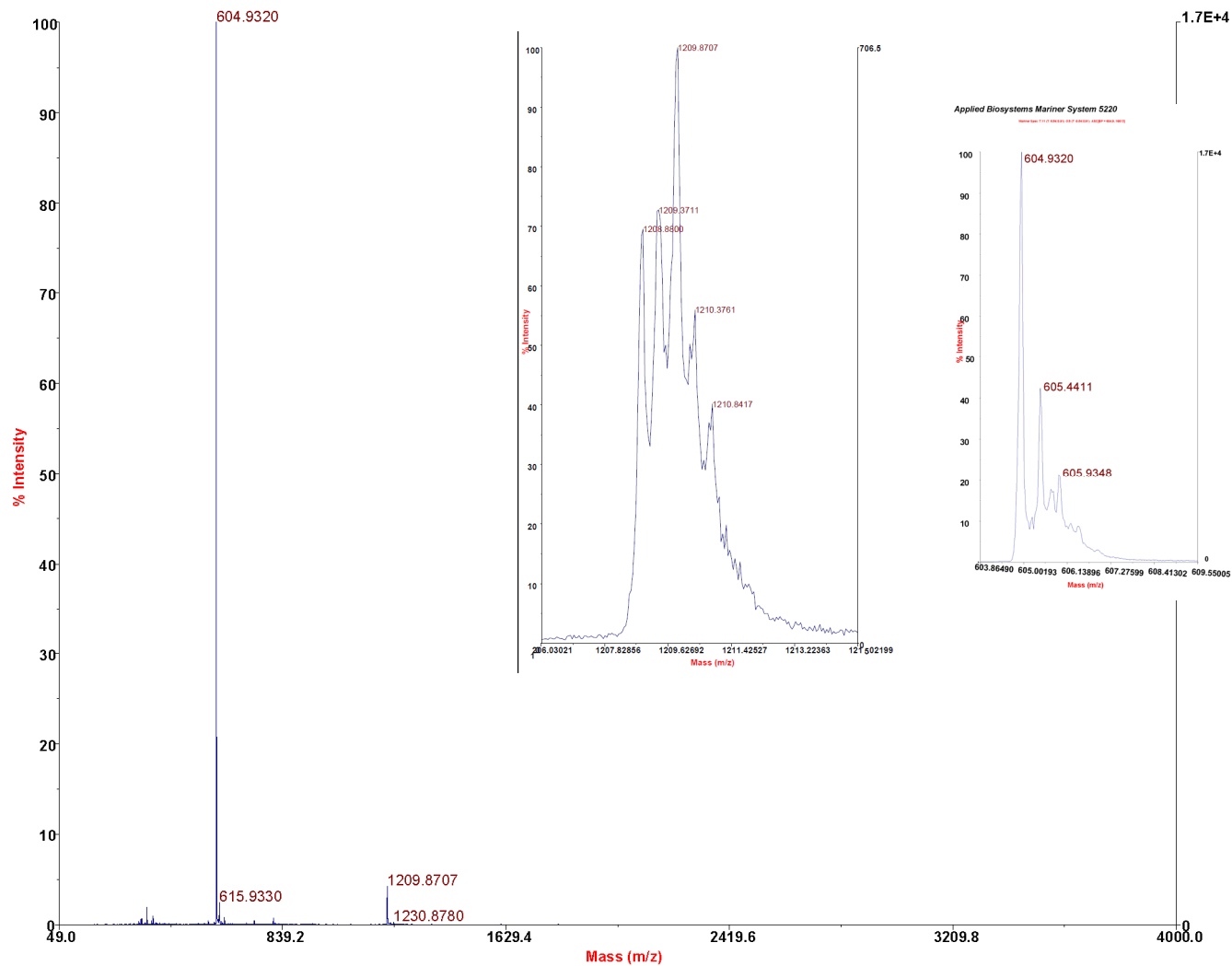
peptide 4 (K56)



HPLC profile obtained for peptide 4 (gradient: 60→80%B [B: CH₃CN/H₂O 9:1 + 0.05% TFA] in 10 minutes; flow rate: 1mL/min; C₁₈ Agilent Zorbax RX-C18) Retention time (R_t) = 5.14 min. Detection Wavelength: 226 nm.

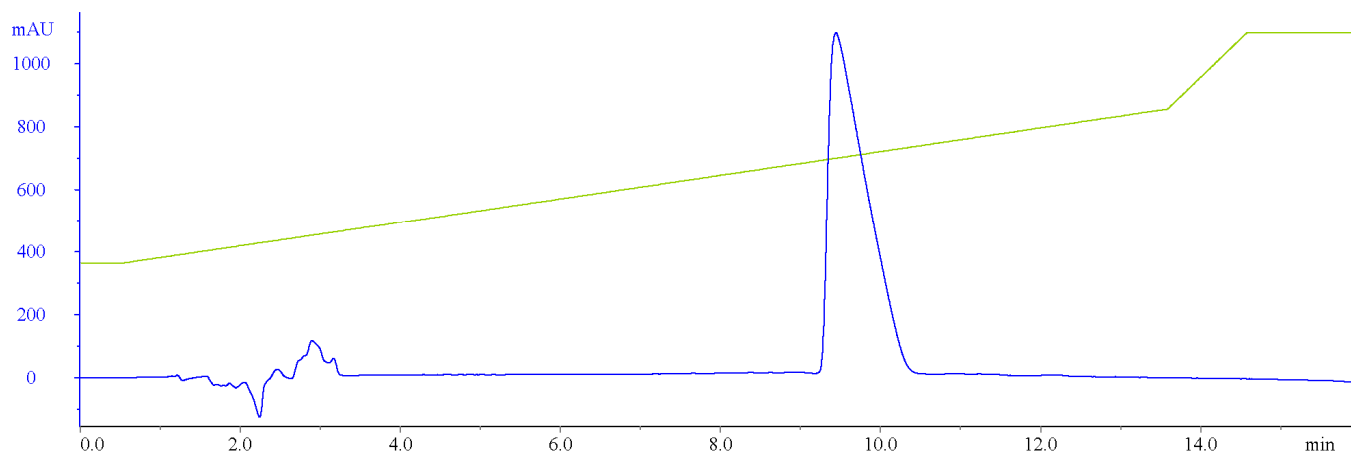
Applied Biosystems Mariner System 5220

Mariner Spec /7:11 (T /0.54:0.91) -3:5 (T -0.54:0.91) ASC[BP = 604.9, 16613]



ESI-HRMS for peptide 4: $[M+H]^+$ calcd. 1208.8732; $[M+H]^+$ found: 1208.8800 (*monoisotopic*), $[M+Na]^+$ found: 1230.8780 (*monoisotopic*), $[M+2H]^{2+}$ found: 604.9320.

peptide **4r** (K56-NH2)

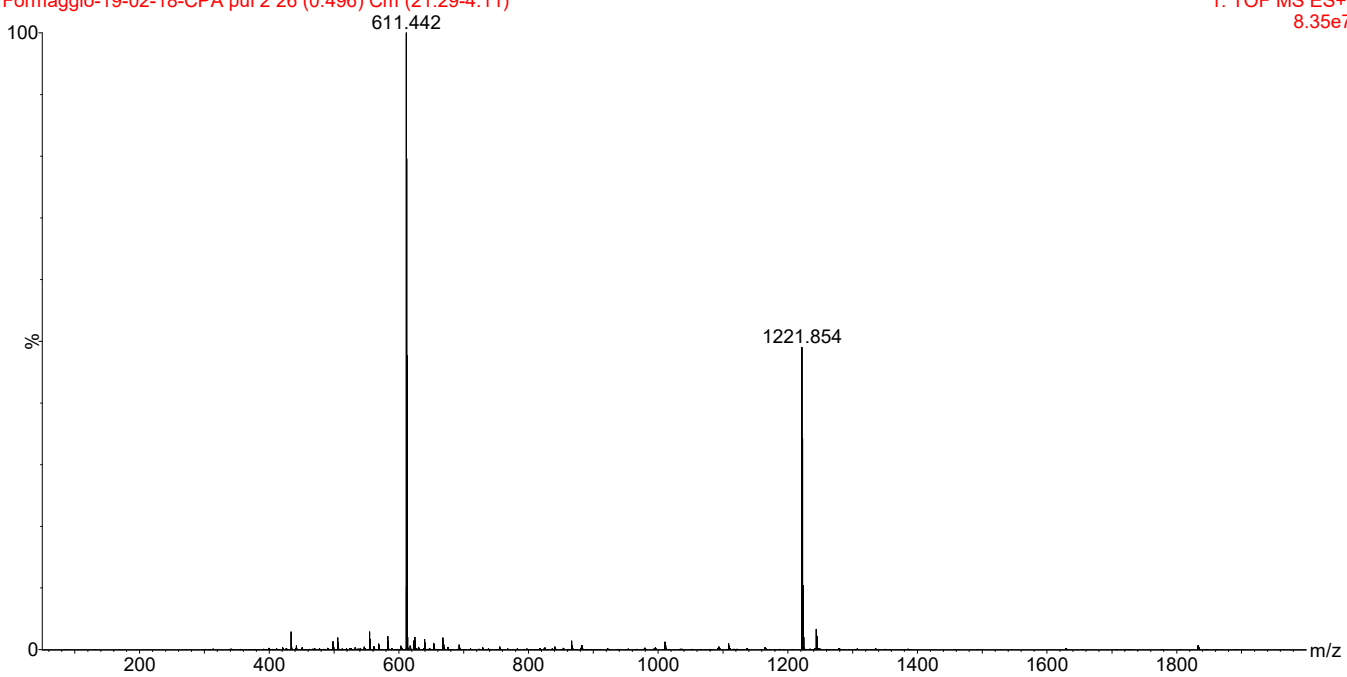


HPLC profile obtained for peptide **4r** (gradient: 40→80%B [CH₃CN/H₂O 9:1 + 0.05% TFA] in 15 minutes; flow rate: 1mL/min; C₁₈ Phenomenex 300Å) Retention time (R_t) = 9.44 min. Detection Wavelength: 208 nm.

1.5 KV

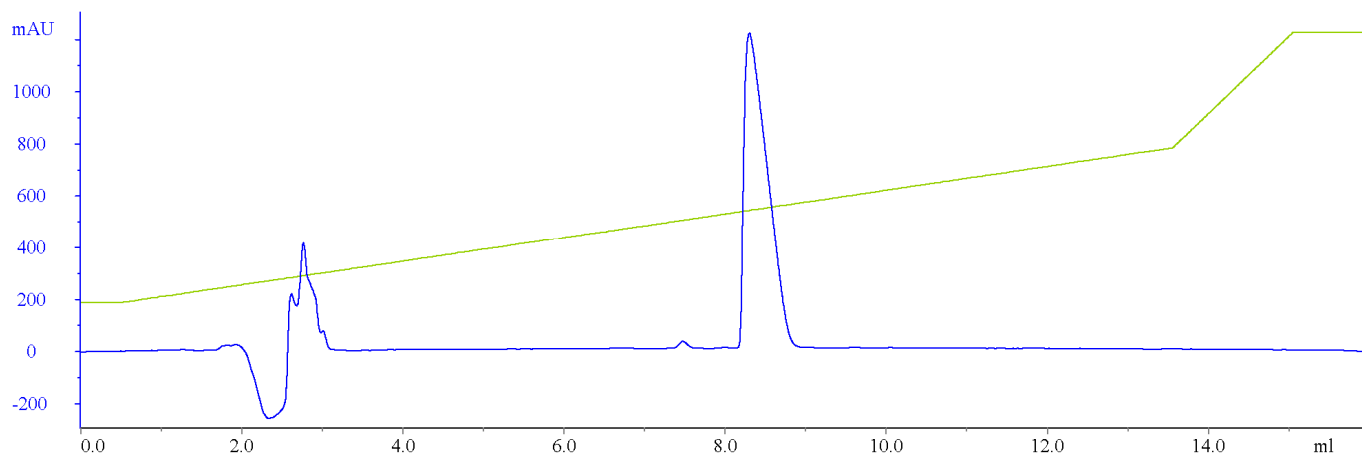
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1: TOF MS ES+
8.35e7

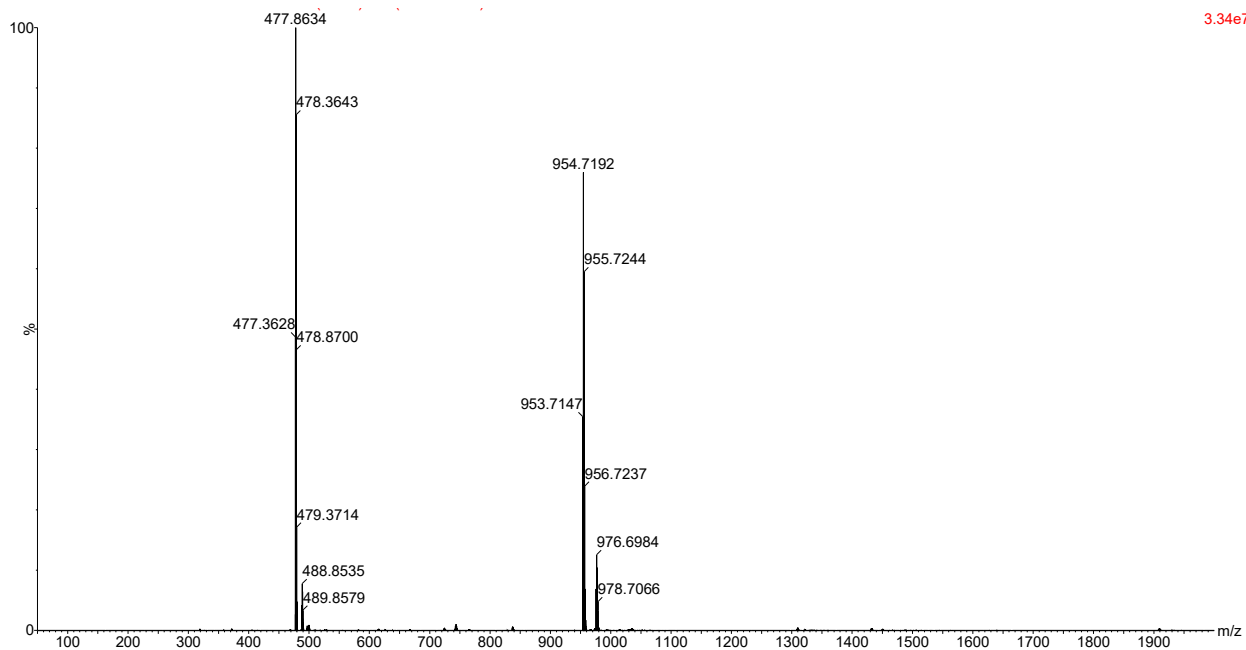


ESI-HRMS for peptide **4r**: $[M+H]^+$ _{calcd.} 1221.8584; $[M+H]^+$ _{found:} 1221.8540 (*monoisotopic*), $[M+2H]^{2+}$ _{found:} 611.4420.

peptide 4c

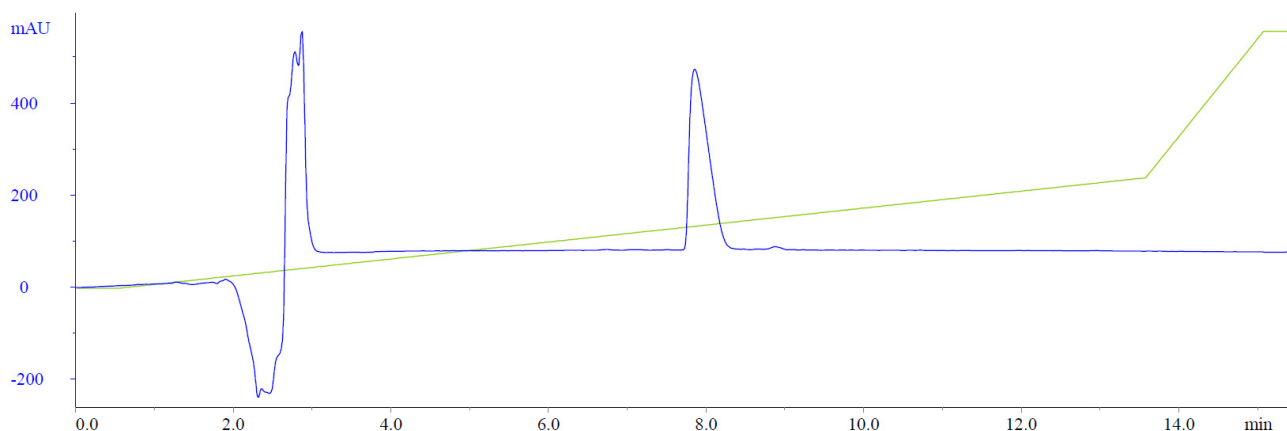


HPLC profile obtained for peptide **4c** (gradient: 30→70%B [CH₃CN/H₂O 9:1 + 0.05% TFA] in 15 minutes; flow rate: 1mL/min; C₁₈ Phenomenex 300Å) Retention time (R_t) = 8.31 min. Detection Wavelength: 208 nm.

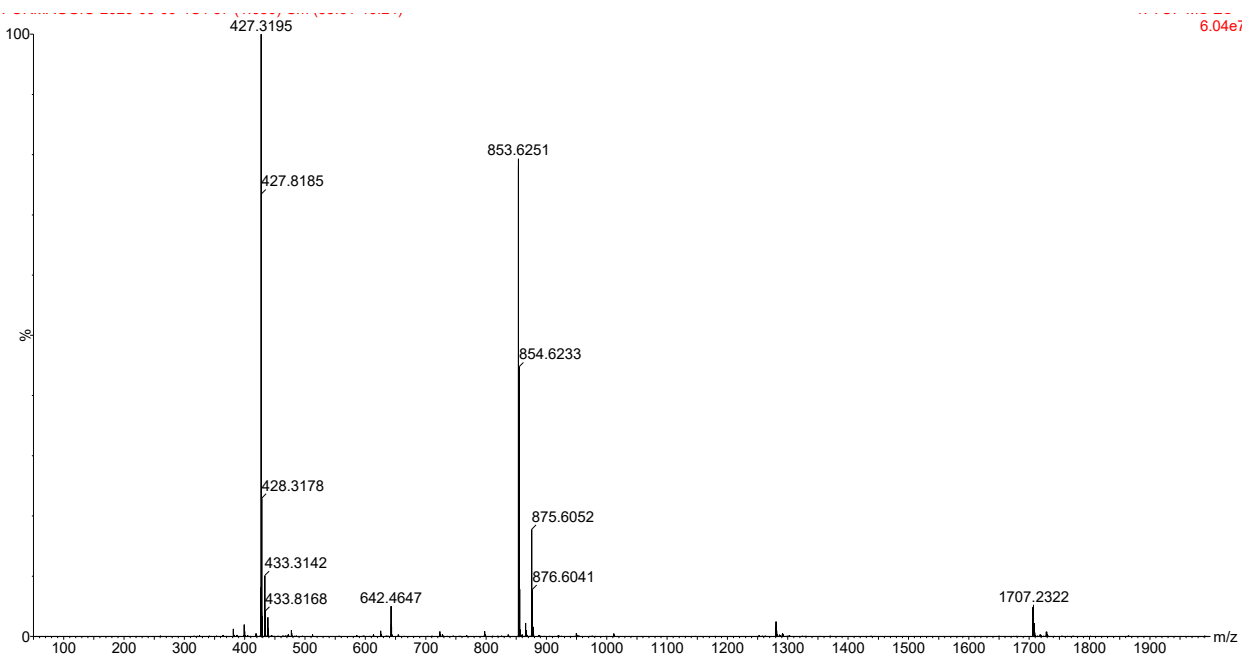


ESI-MS for peptide **4c**: [M+H]⁺_{calcd.} 953.7049; [M+H]⁺_{found:} 953.7147 (*monoisotopic*) [M+Na]⁺_{found:} 975.6951, [M+2H²⁺]_{found:} 477.3628.

peptide **4c1** (Oct-UGLUKKL-NH₂)

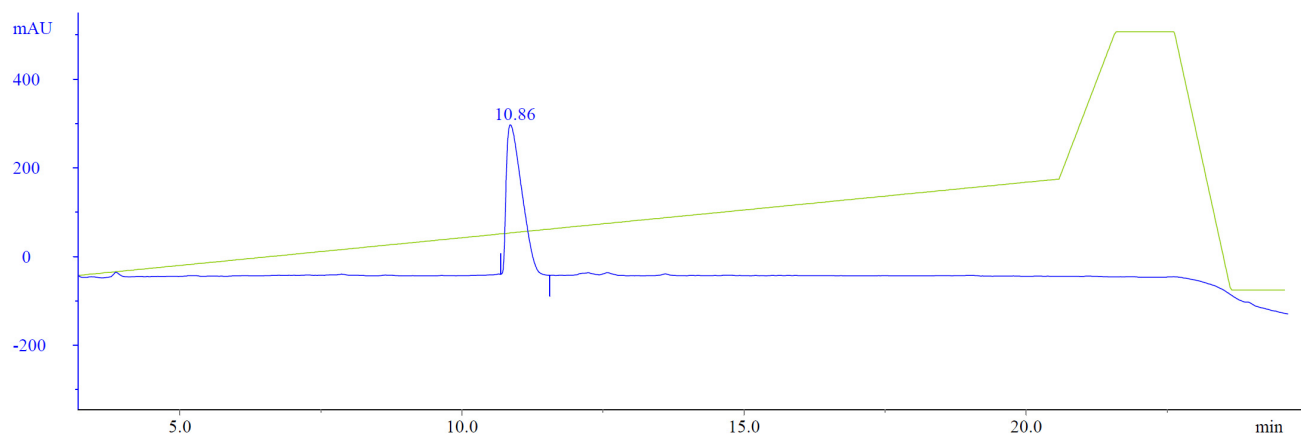


HPLC profile obtained for peptide **4c1** (gradient: 30→60%B [CH₃CN/H₂O 9:1 + 0.05% TFA] in 14 minutes; flow rate: 1 mL/min; C₁₈ Phenomenex 300Å) Retention time (R_t) = 7.85 min. Detection Wavelength: 208 nm.

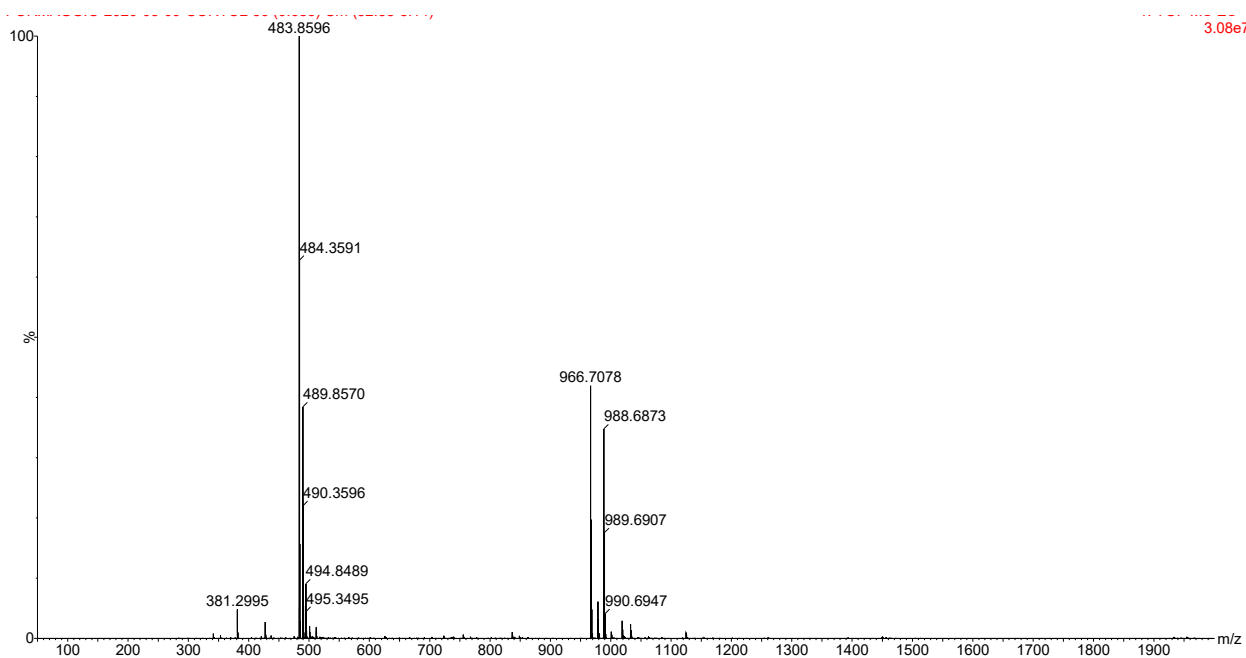


ESI-MS for peptide **4c1**: [M+H]⁺_{calcd.}: 853.6261; [M+H]⁺_{found}: 853.6251, [M+2H²⁺]_{found}: 427.3195. [2M+H]⁺_{found}: 1707.2322.

peptide **4c2** (Oct-AibLLAibGIL-NH₂)

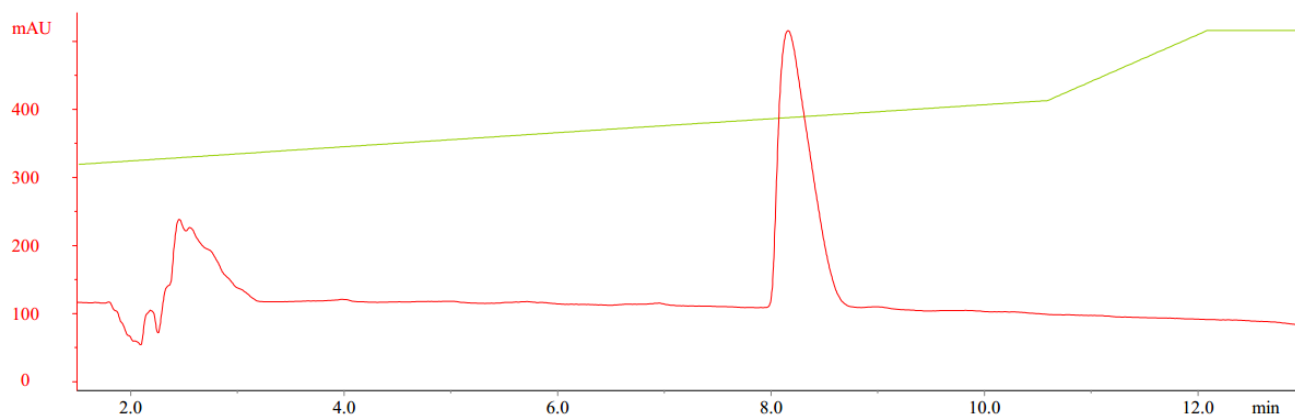


HPLC profile obtained for peptide **4c2** (gradient: 30→60%B [CH₃CN/H₂O 9:1 + 0.05% TFA] in 20 minutes; flow rate: 1mL/min; C₁₈ Phenomenex 300Å) Retention time (R_t) = 10.86 min. Detection Wavelength: 208 nm.



ESI-MS for peptide **4c2**: [M+H]⁺_{calcd.} 966.7001; [M+H]⁺_{found}: 966.7078, [M+Na]⁺_{found}: 988.6873, [M+2H²⁺]_{found}: 483.8596.

peptide 5 (K6)

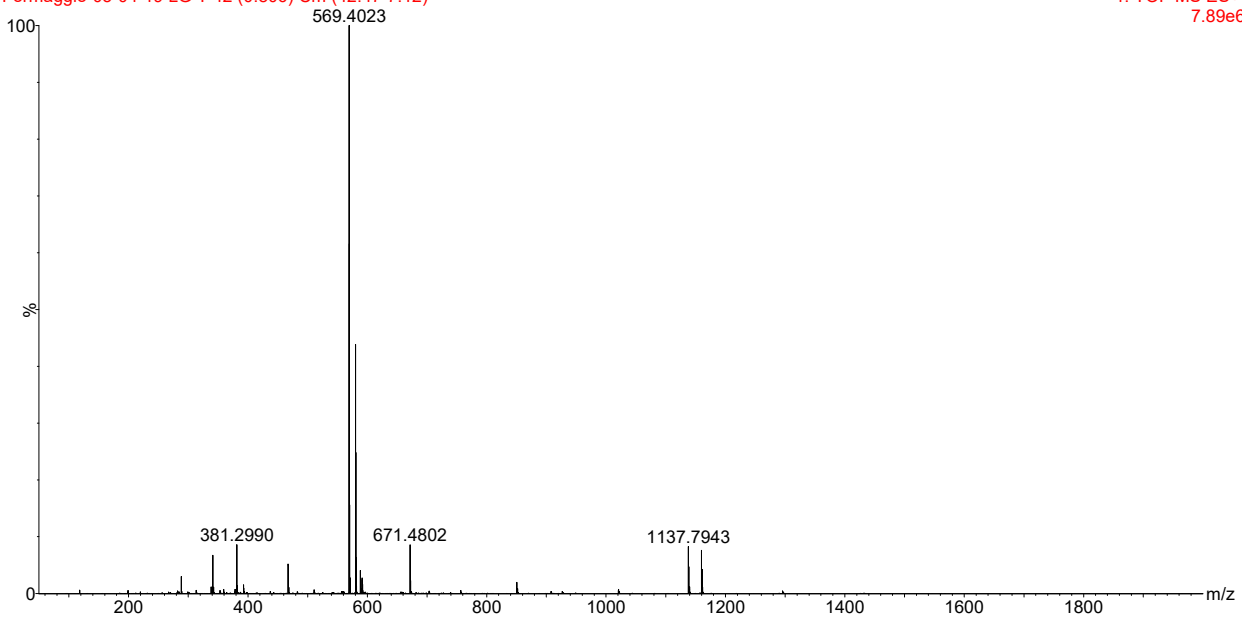


HPLC profile obtained for peptide 5 (gradient: 60→80%B [CH₃CN/H₂O 9:1 + 0.05% TFA] in 10 minutes; flow rate: 1mL/min; C₁₈ Agilent Zorbax RX-C18) R_t = 8.16 min. Detection Wavelength: 216 nm.

Marta

Formaggio-05-04-19 LG T 42 (0.806) Cm (42:47-7:12)

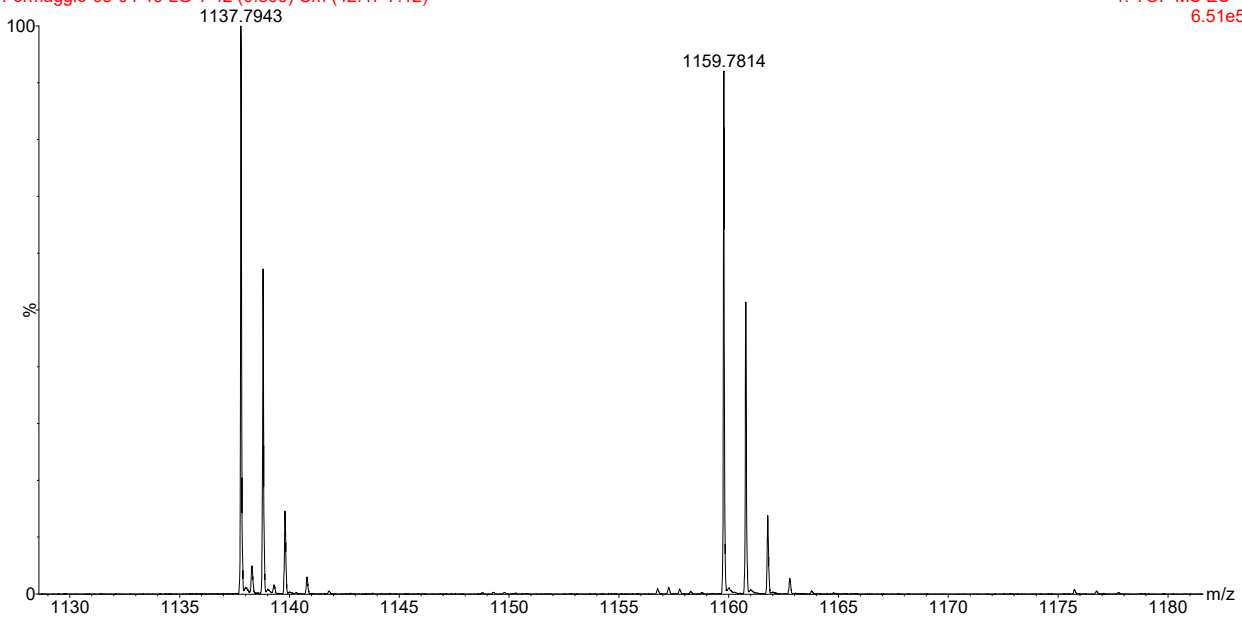
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7.89e6



Marta

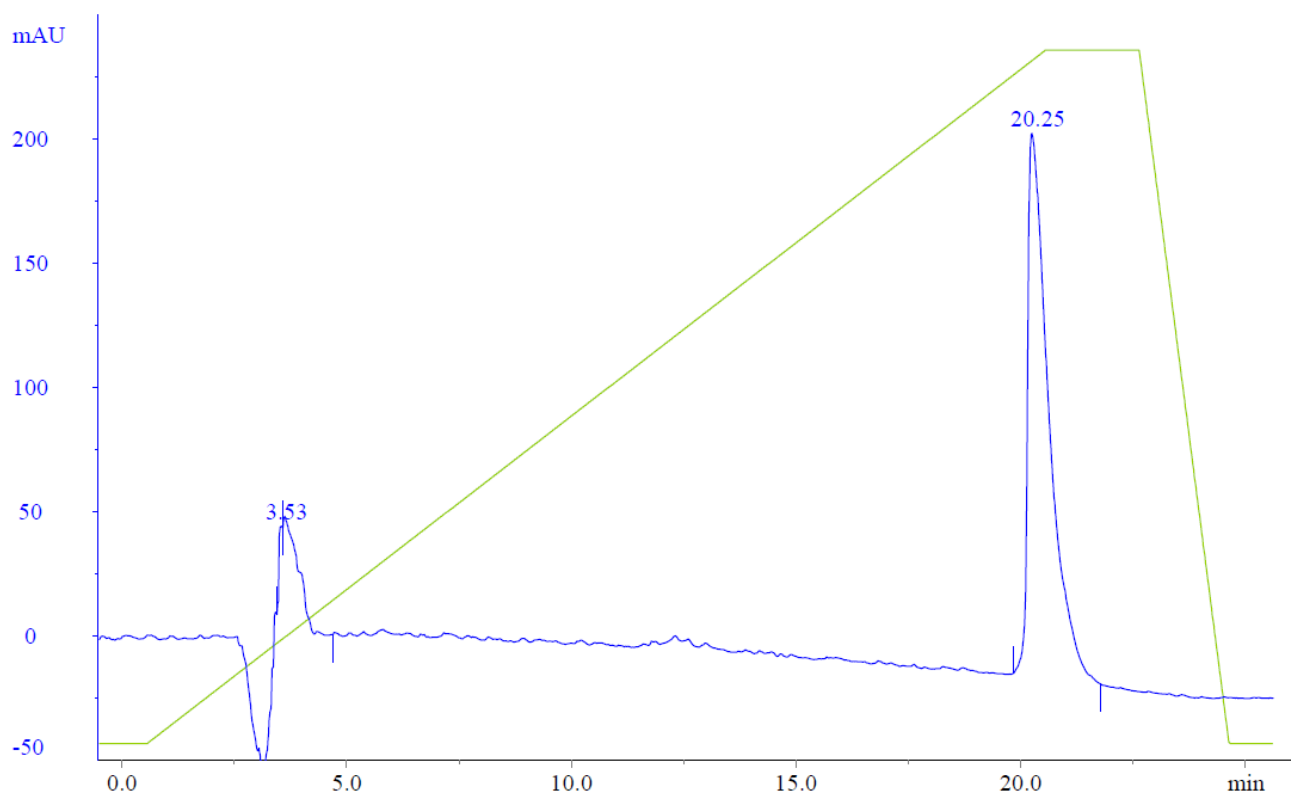
Formaggio-05-04-19 LG T 42 (0.806) Cm (42:47-7:12)

1: TOF MS ES+
6.51e5



ESI-HRMS for peptide **5**: $[M+H]^+$ calcd. 1137.7997; $[M+H]^+$ found: 1137.7943 (*monoisotopic*), $[M+Na]^+$ found: 1159.7814 (*monoisotopic*), $[M+2H]^{2+}$ found: 569.4023. Peaks at 381.2990 and 671.4802 are impurities from samples injected before (the instrument was contaminated).

peptide **6** (K5U6)

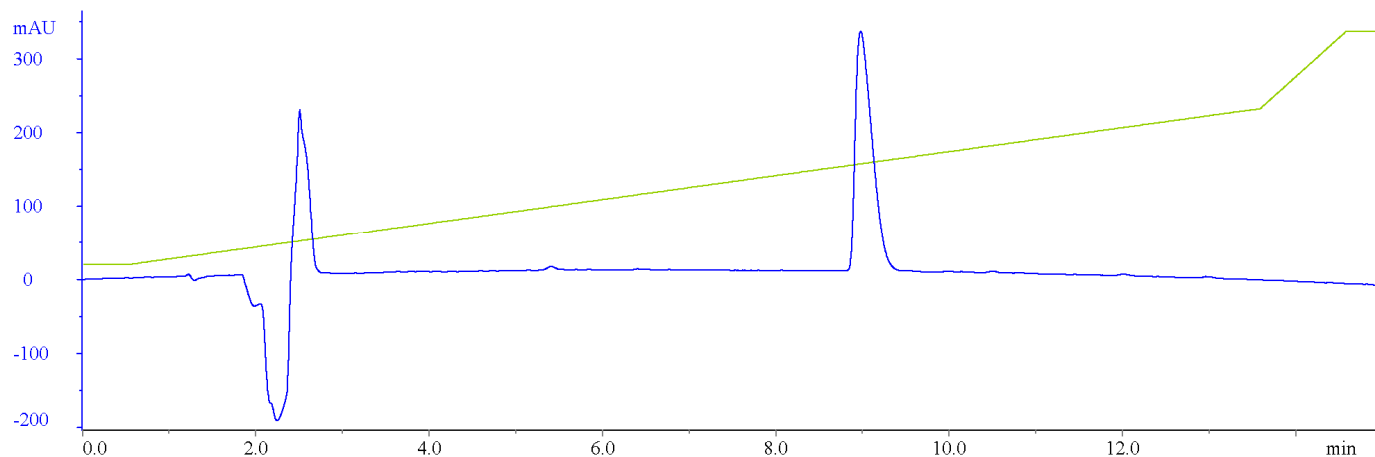


HPLC profile obtained for peptide **6** (gradient: 60→100%B [CH₃CN/H₂O 9:1 + 0.05% TFA] in 20 minutes; flow rate: 1mL/min; C₁₈ Phenomenex 300Å) R_t = 20.25 min. Detection Wavelength: 216 nm.

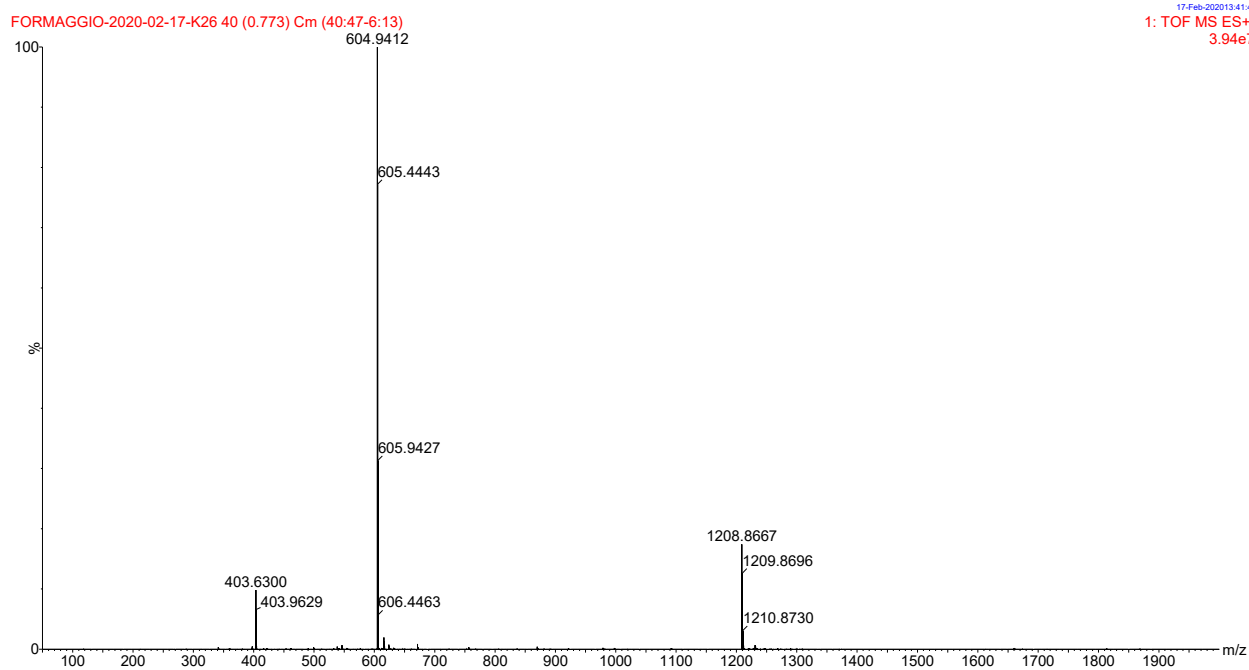


ESI-MS for peptide **6**: $[M+H]_{\text{calcd.}}$ 1165.8210; $[M+H]_{\text{found:}}$ 1165.8271 (*monoisotopic*), $[M+2H^{2+}]_{\text{found:}}$ 583.4162, $[M+H+Na^{2+}]_{\text{found:}}$ 594.4195.

peptide 7 (K26)



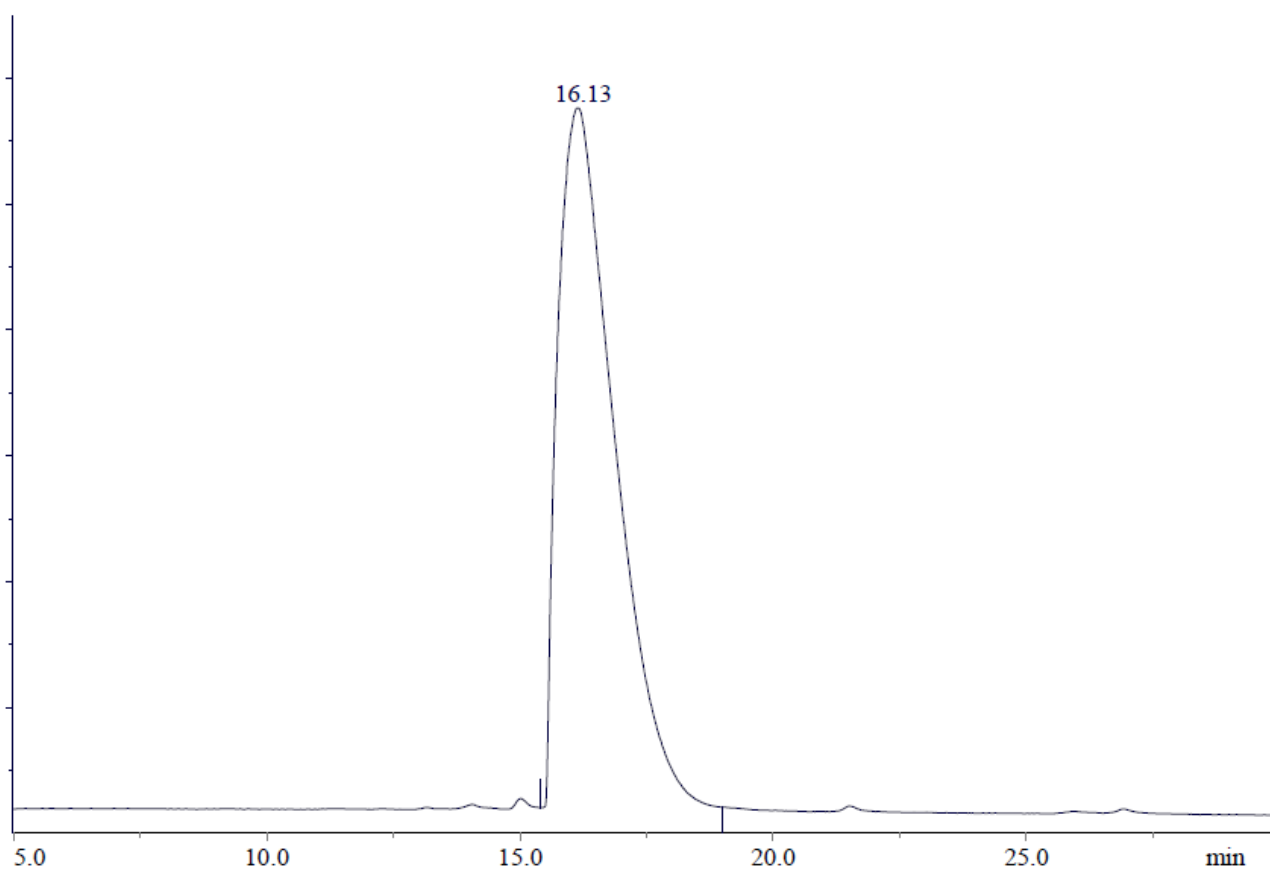
HPLC profile obtained for peptide 7 (gradient: 40→80%B [CH₃CN/H₂O 9:1 + 0.05% TFA] in 15 minutes; flow rate: 1mL/min; C₁₈ Phenomenex 300Å) R_t = 8.97 min. Detection Wavelength: 208 nm.



17-Feb-2020 13:41:49
1: TOF MS ES+
3.94e7

ESI-MS for peptide 7: [M+H]_{calcd.} 1208.8632; [M+H]_{found:} 1208.8667, [M+2H²⁺]_{found:} 604.9412; [M+3H³⁺]_{found:} 403.6300.

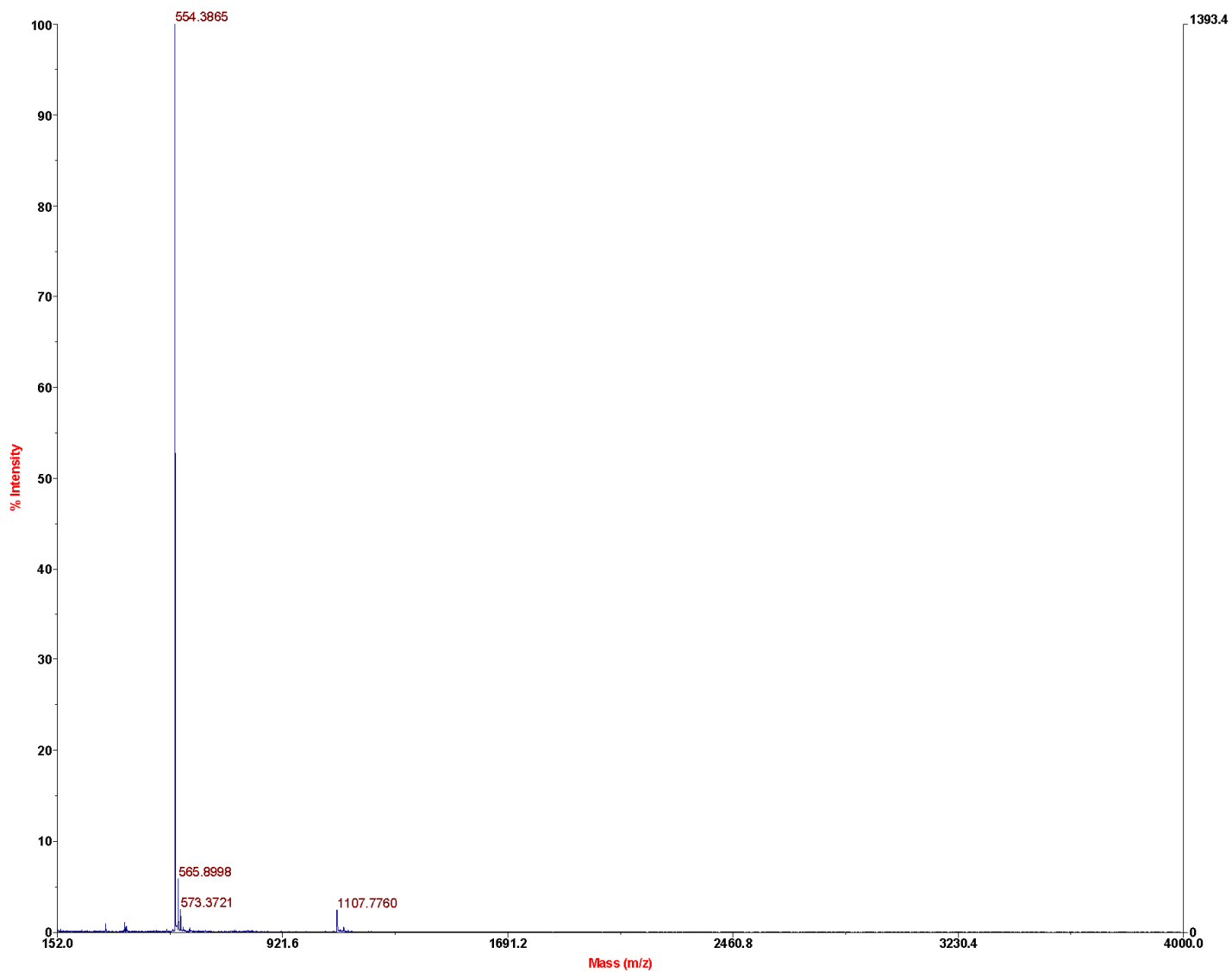
peptide **8** (Api8)



HPLC profile obtained for peptide **8** ([Api⁸]-Tric) (gradient: 45→75% B [CH₃CN/H₂O 9:1 + 0.05% TFA] in 30 minutes; flow rate: 1mL/min; C₁₈ Phenomenex 300Å); R_t = 16.13 min.

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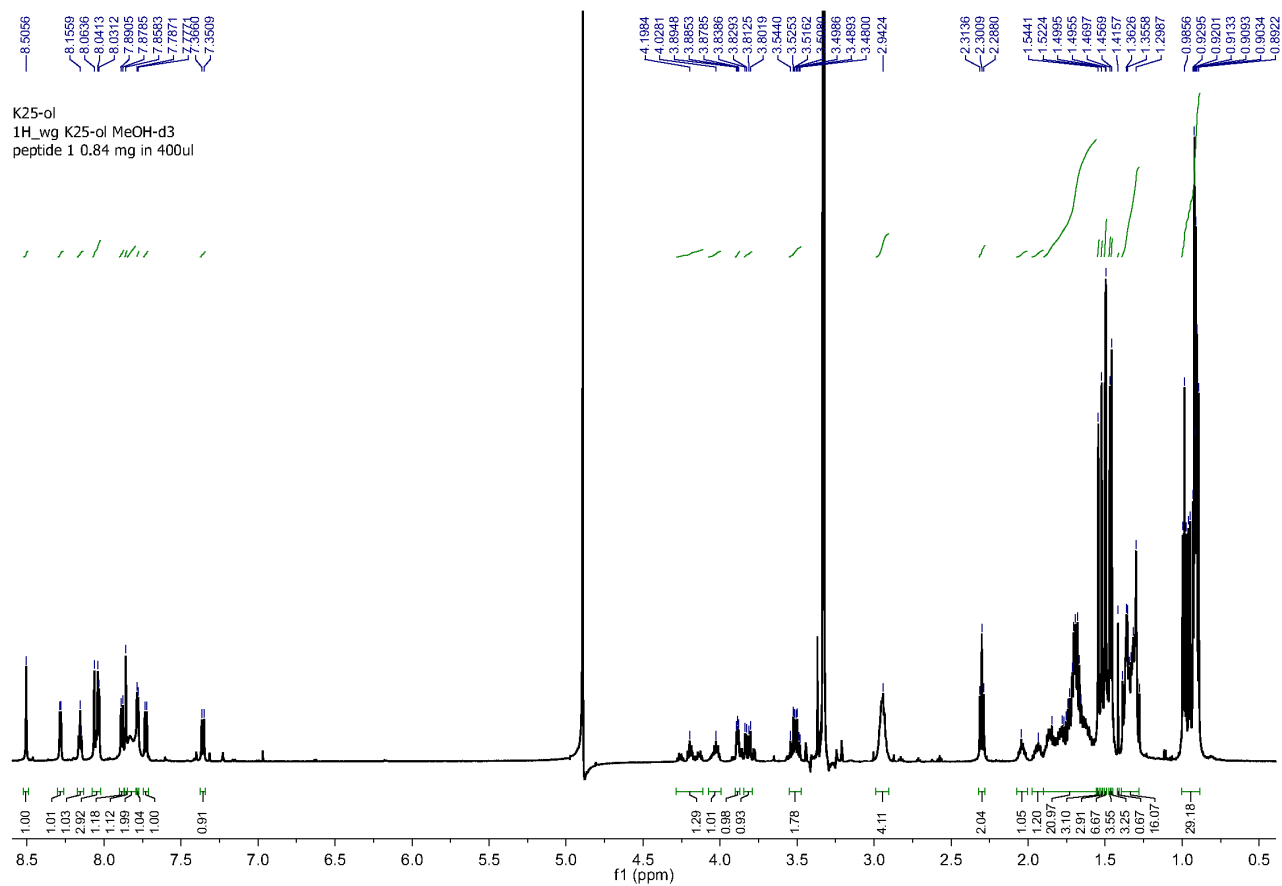
Mariner Spec /7:9 (T /0.53:0.70) -3:5 (T -0.53:0.70) ASC[BP = 554.4, 1393]



ESI-MS for peptide **8**: $[M+H]_{\text{calcd.}}$ 1107.7427; $[M+H]_{\text{found}}$ 1107.7760, $[M+2H^{2+}]_{\text{found}}$ 554.3865.

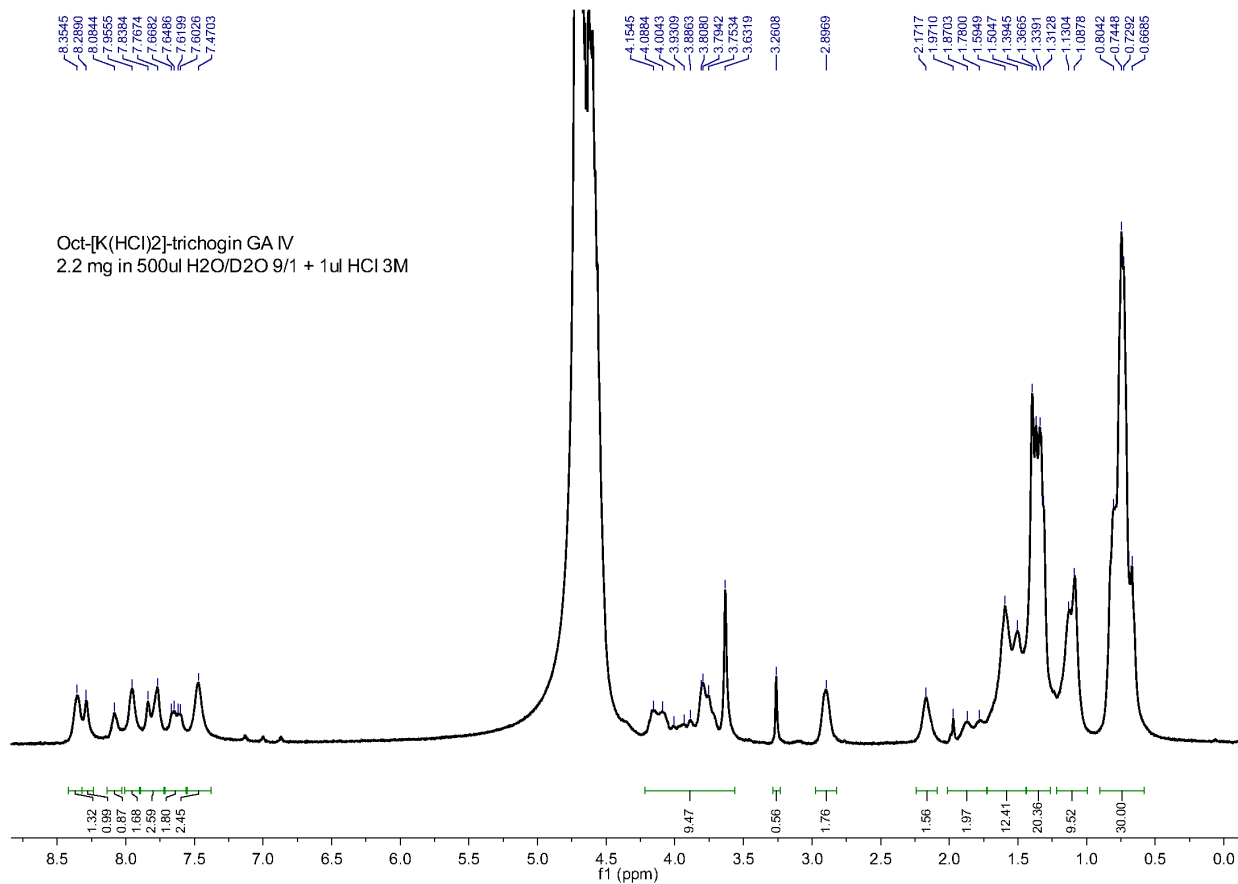
¹H NMR characterizations

peptide 1 (K25)



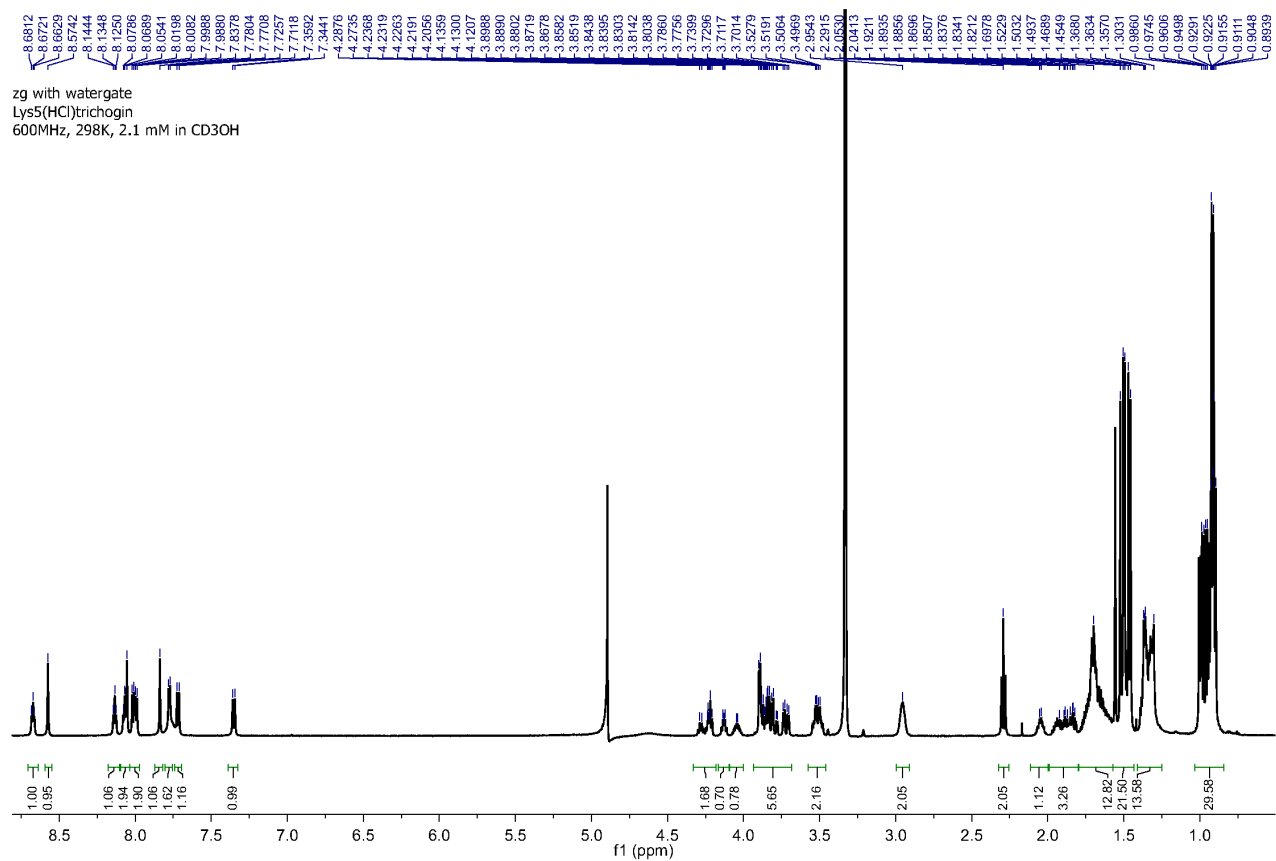
1D ¹H NMR spectrum for peptide 1 in CD₃OH, 298K, 600MHz (peptide concentration ca. 2 mM).

peptide 2 (K2)



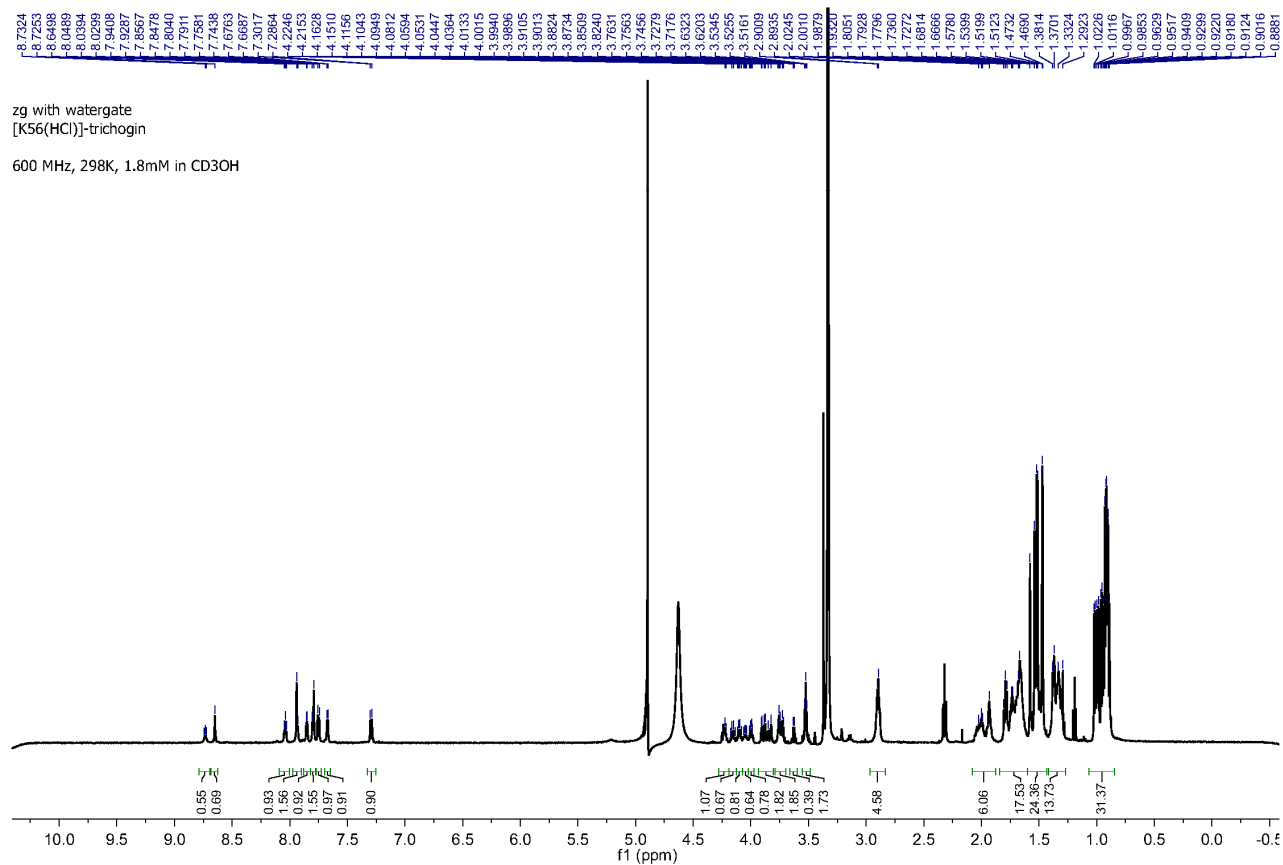
1D ¹H NMR spectrum for peptide 2 in H₂O/D₂O 9:1(+ 1μl HCl 3 M), 298K, 400MHz (peptide concentration ca. 4 mM).

peptide 3 (K5)

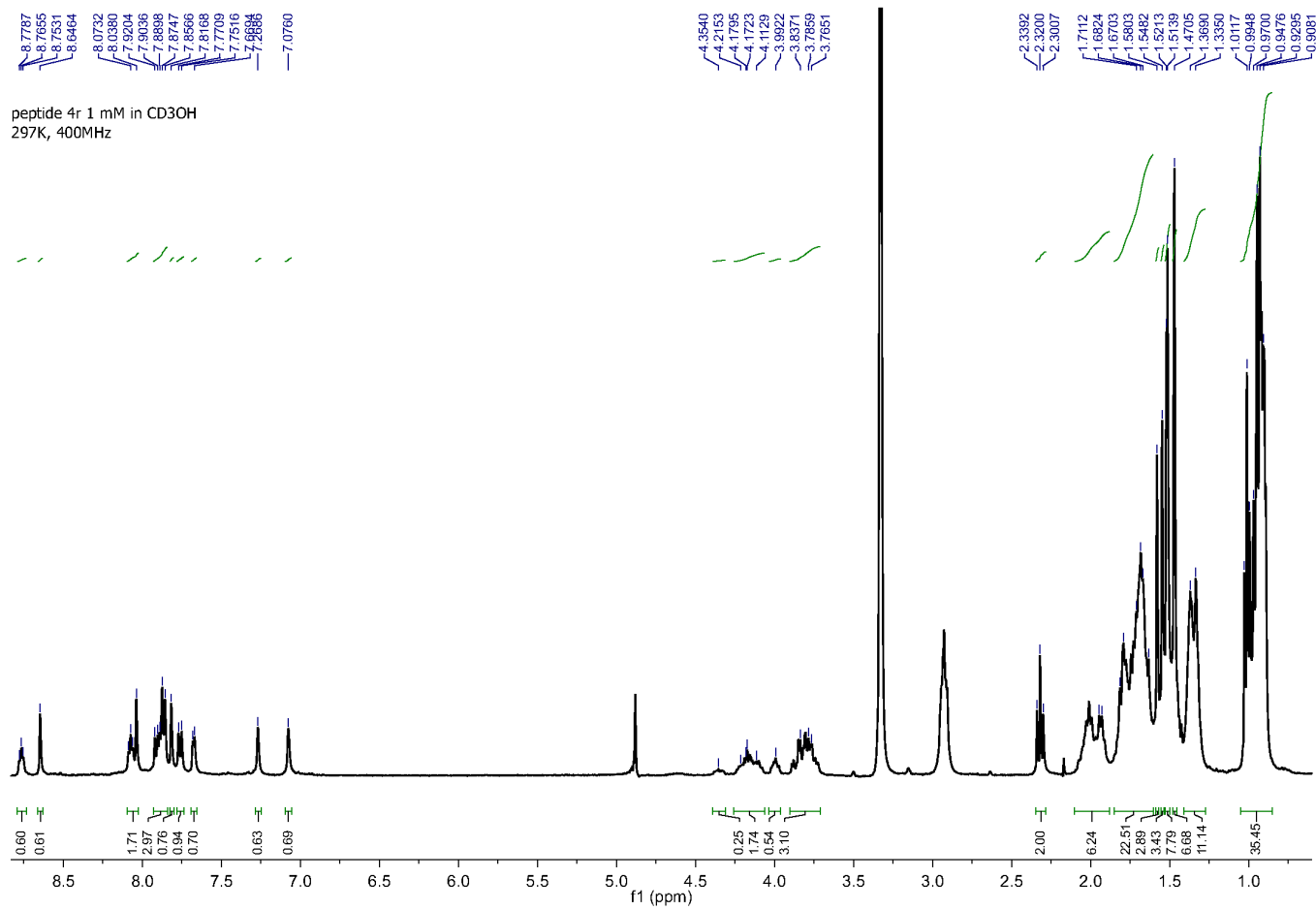


1D ^1H NMR spectrum for peptide 3 in CD_3OH , 298 K, 600MHz (peptide concentration ca. 2.1 mM).

peptide 4 (K56)

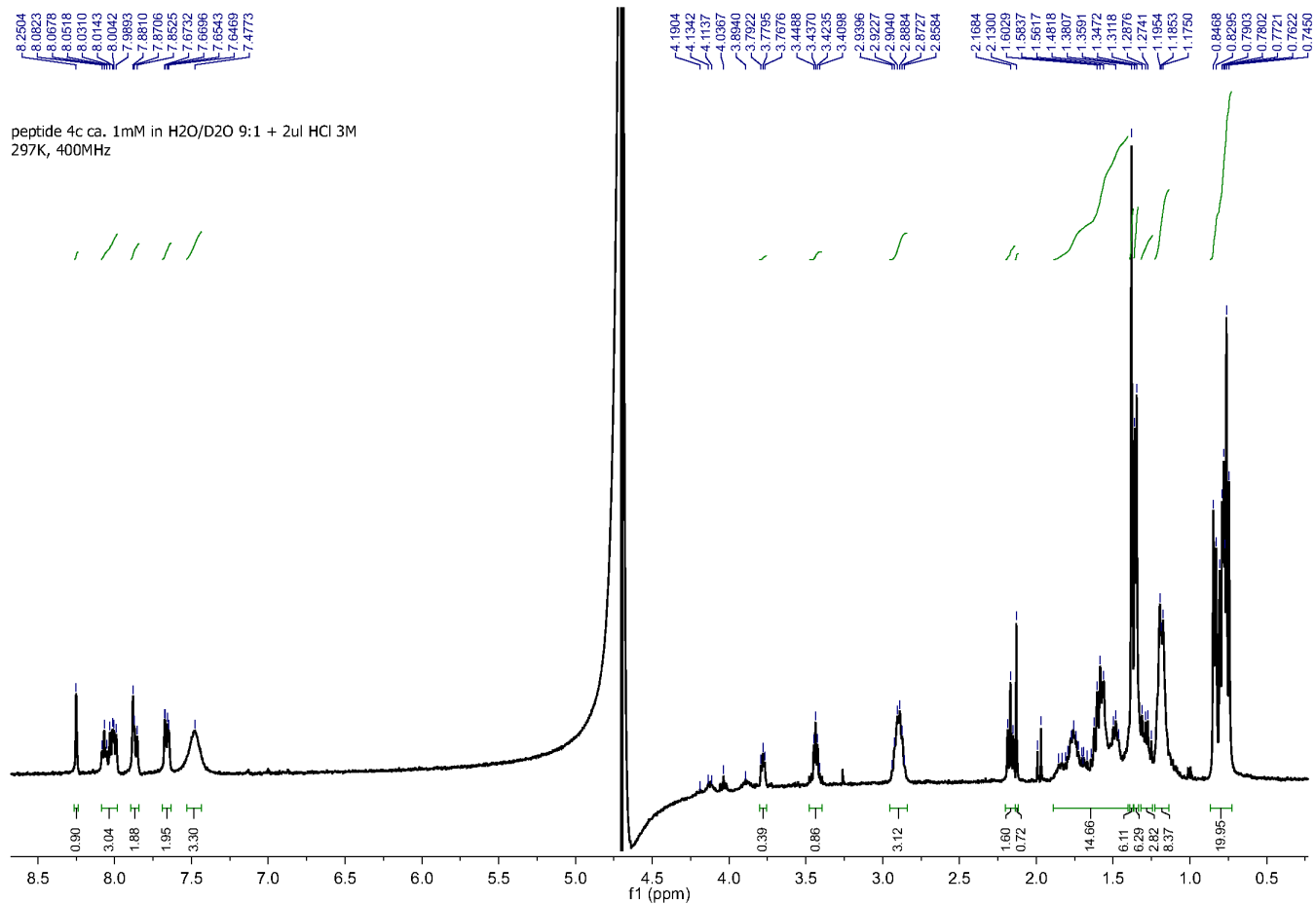


1D ¹H NMR spectrum for peptide 4 in CD₃OH, 298K, 600MHz (peptide concentration ca. 1.8 mM).



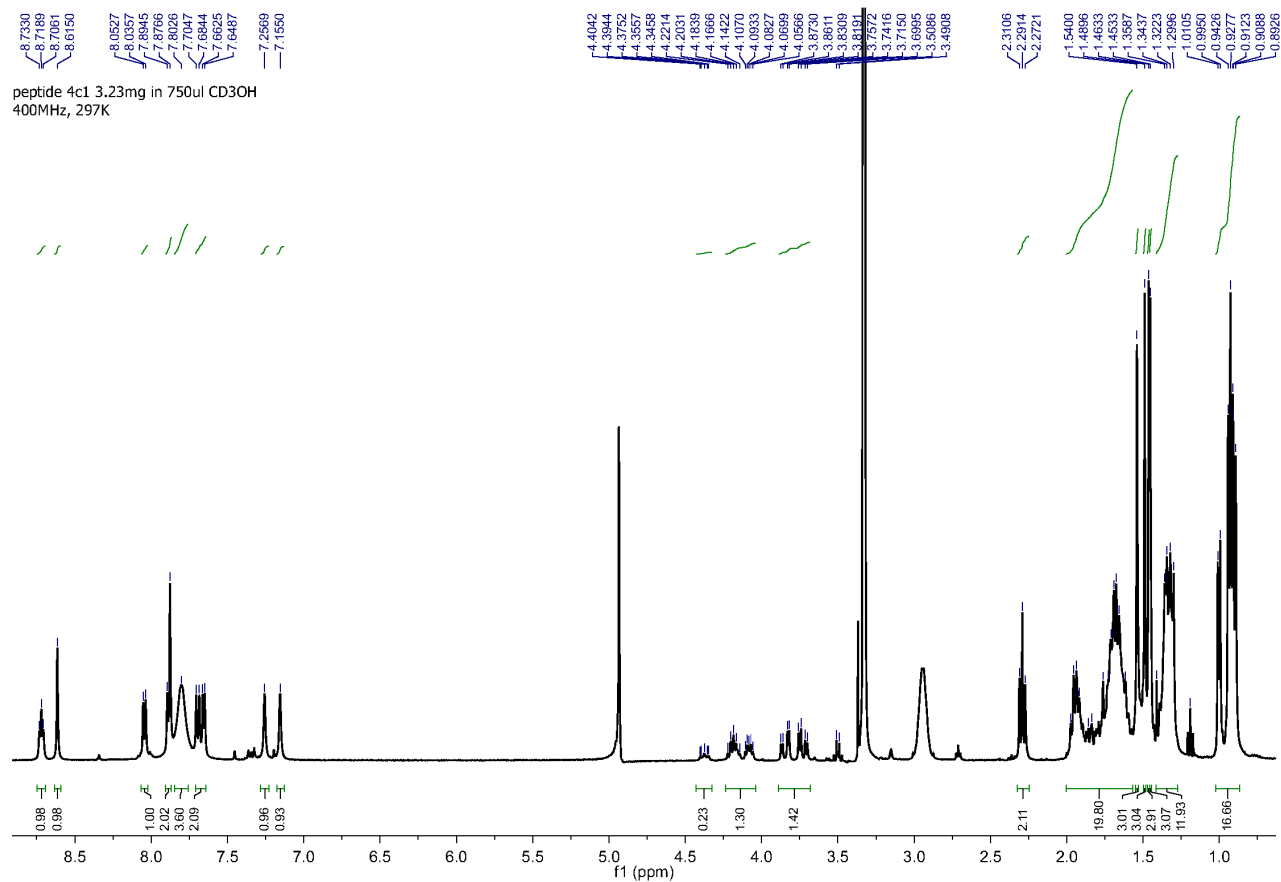
1D ^1H NMR spectrum for peptide **4r** in CD_3OH , 297K, 400MHz (peptide concentration 1 mM).

peptide 4c



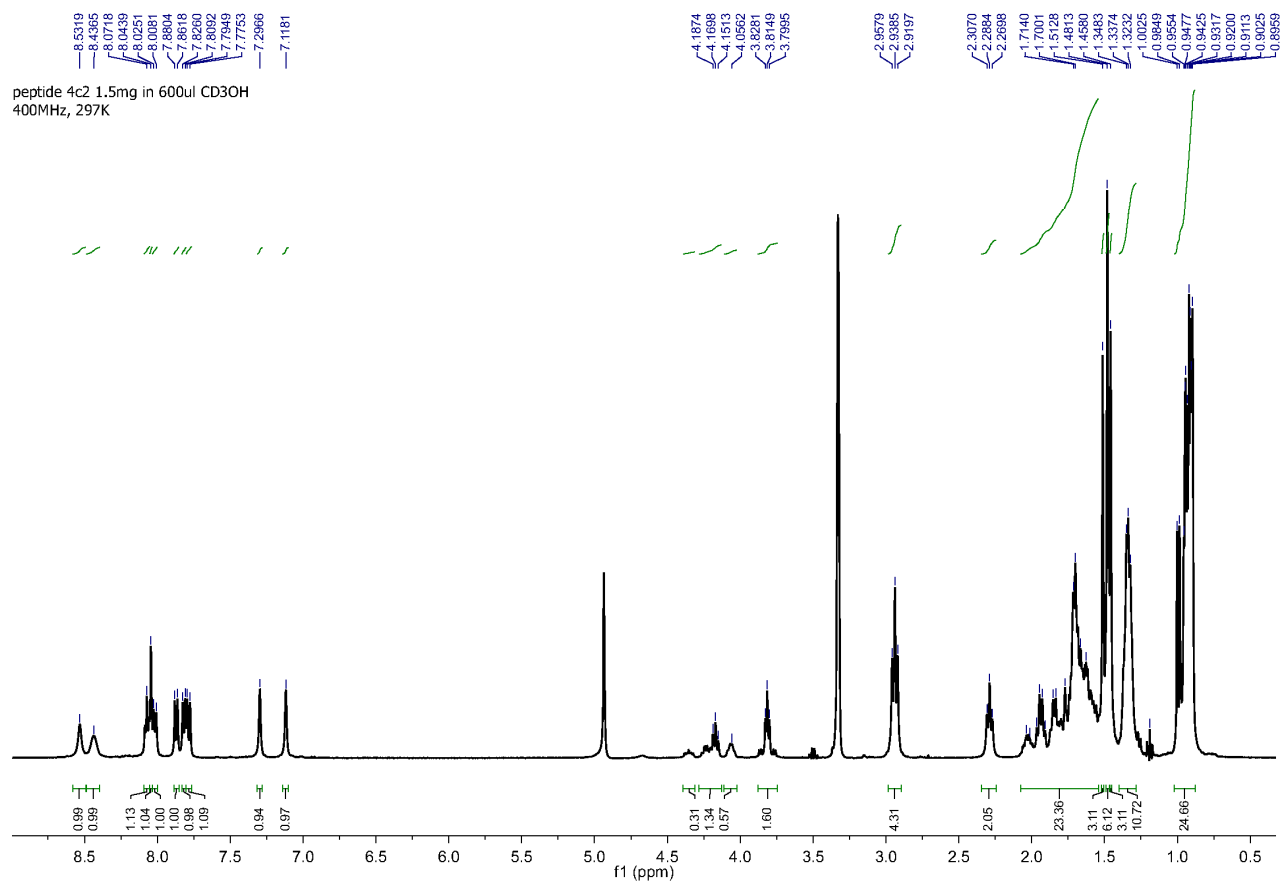
1D ¹H NMR spectrum for peptide **4c** in H₂O/D₂O 9:1 (+2μl HCl 3M in D₂O), 297K, 400MHz (peptide concentration 1 mM).

peptide 4c1



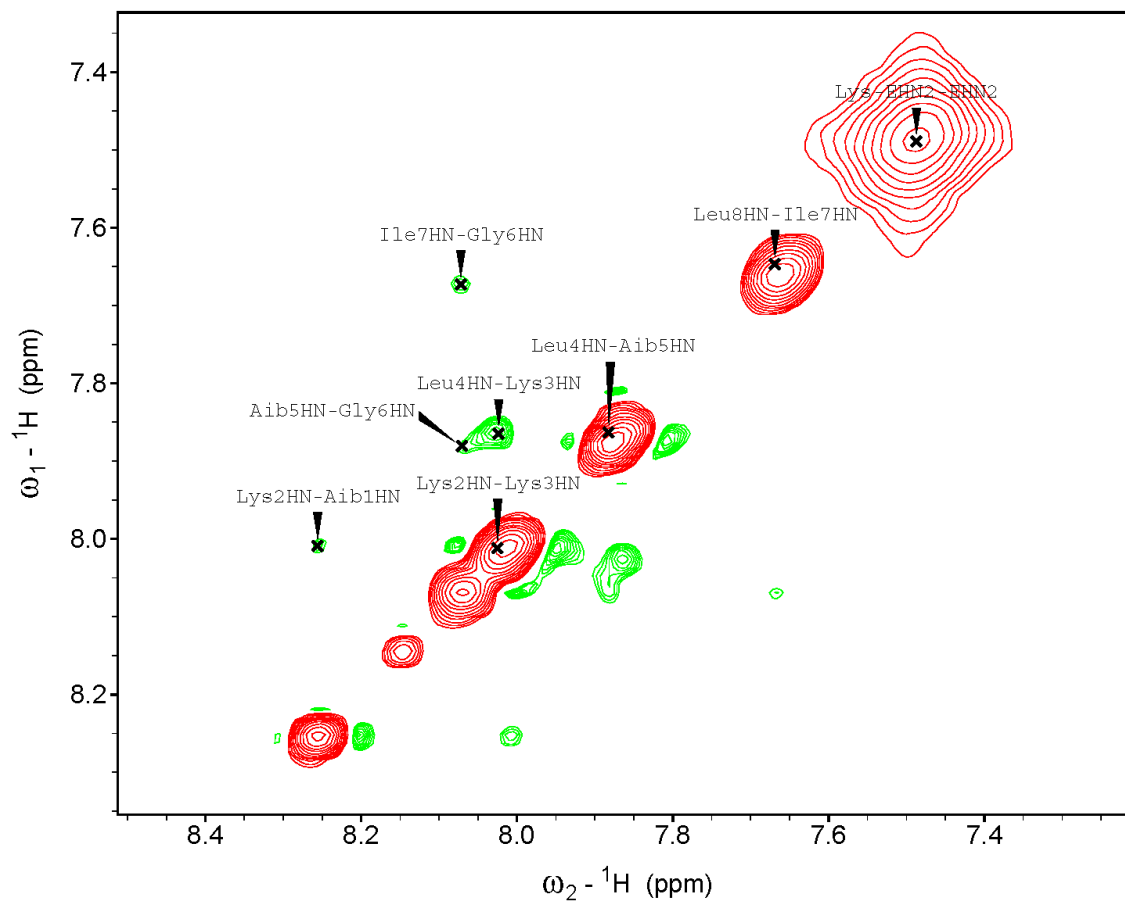
1D ^1H NMR spectrum for peptide **4c1** in CD_3OH , 297K, 400MHz (peptide concentration ca. 5 mM).

peptide **4c2**



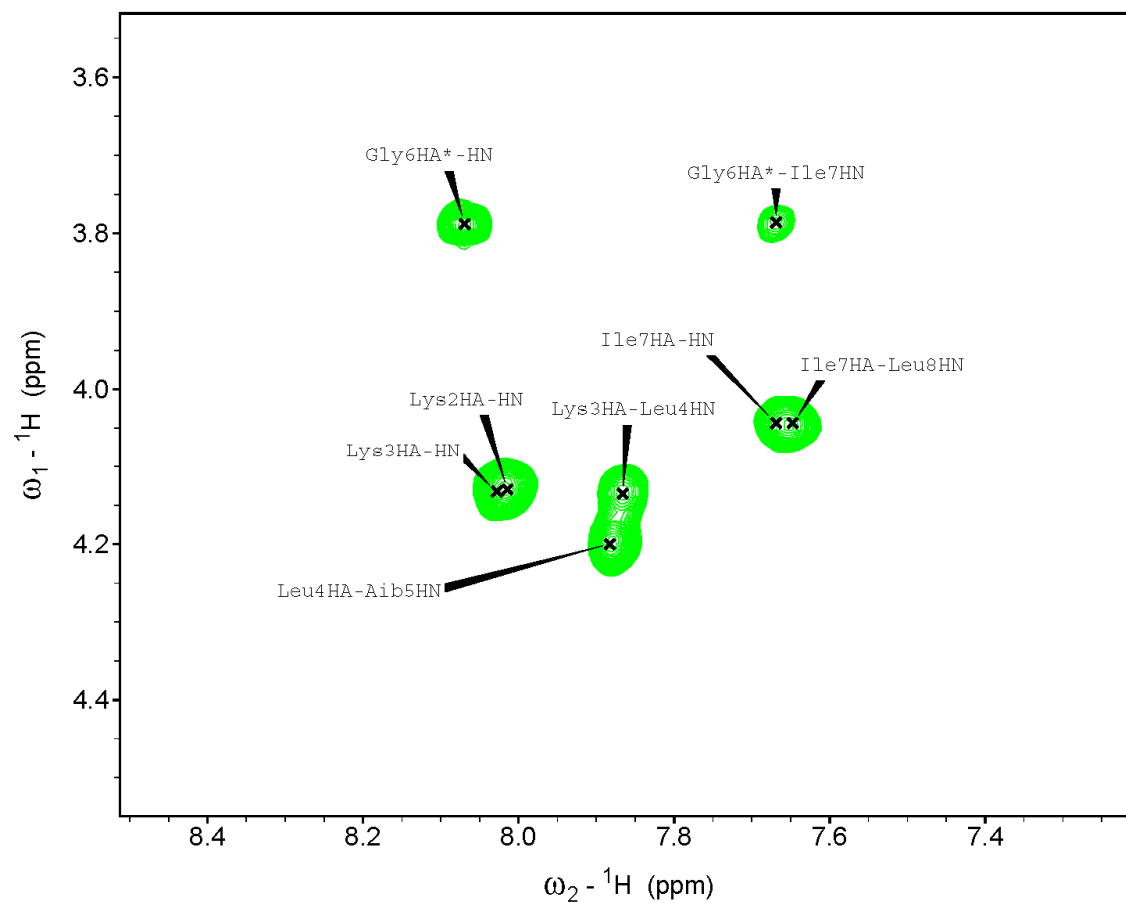
1D ^1H NMR spectrum for peptide **4c2** in CD_3OH , 297K, 400MHz (peptide concentration ca. 2.6 mM).

2D-NMR peptide **4c**



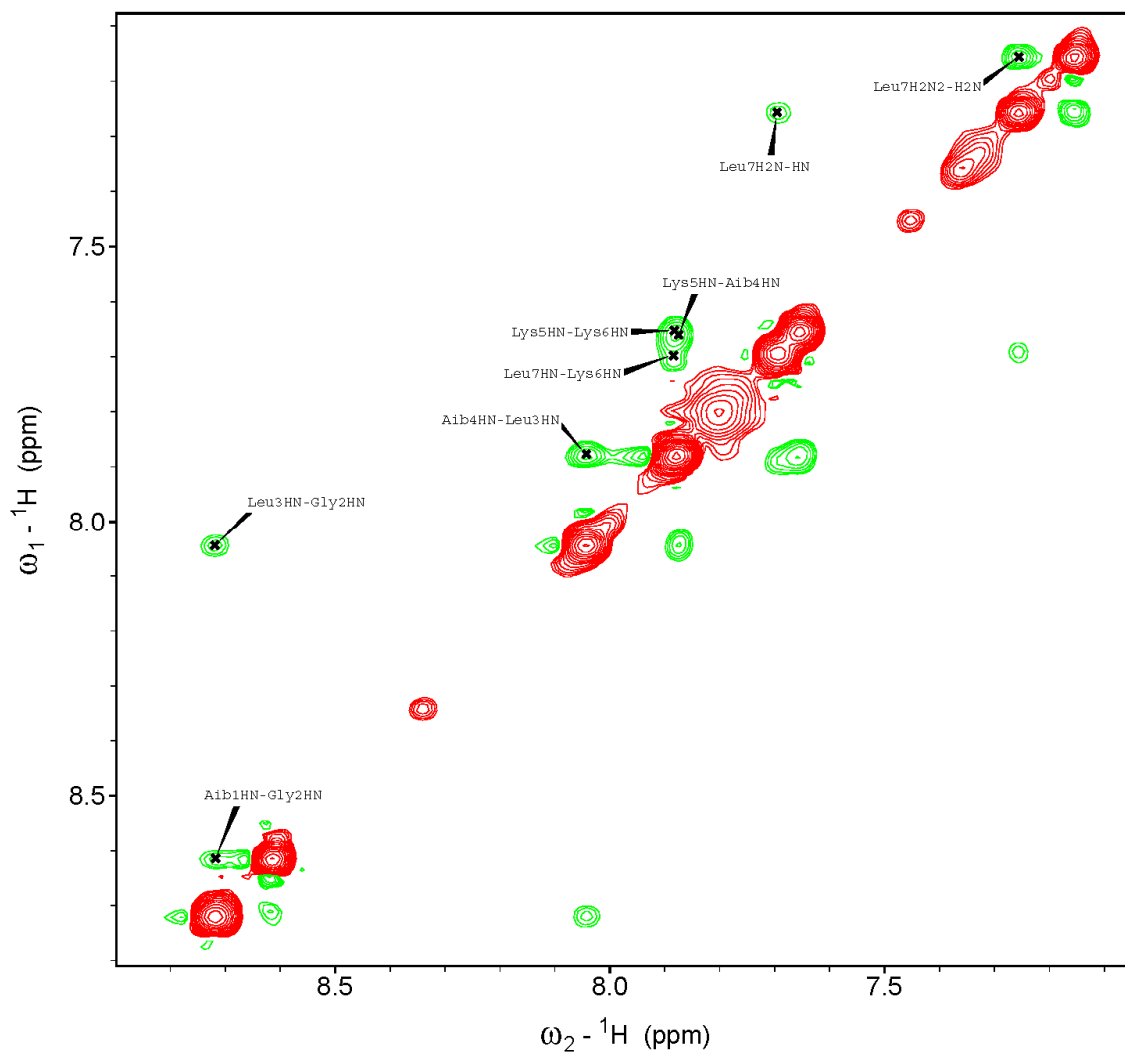
2D ${}^1\text{H}$ NMR for peptide **4c** in $\text{H}_2\text{O}/\text{D}_2\text{O}$ 9:1 (+3 μl HCl 3M in D_2O), 297K, 400MHz (peptide concentration 1 mM). NH-NH (amide) cross-peaks region of the ROESY spectrum. All sequential $\text{NH}_i \rightarrow \text{NH}_{i+1}$ are detectable (apart from those falling under the diagonal).

2D-NMR peptide **4c**



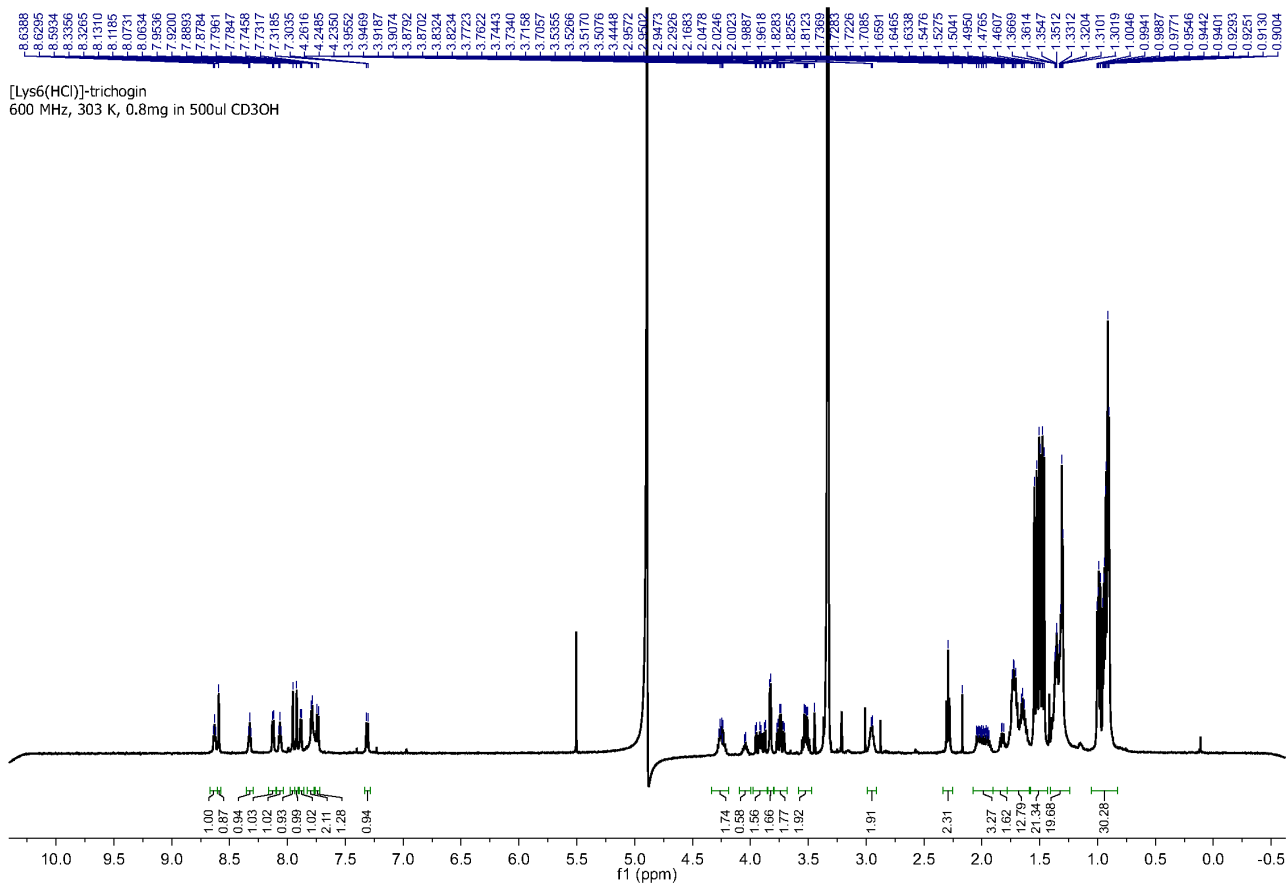
2D ${}^1\text{H}$ NMR for peptide **4c** in $\text{H}_2\text{O}/\text{D}_2\text{O}$ 9:1 (+3 μl HCl 3M in D_2O), 297K, 400MHz (peptide concentration 1 mM). αH -NH cross-peaks region (*fingerprint region*) of the ROESY spectrum. All sequential $\alpha\text{H}_i \rightarrow \text{NH}_{i+1}$ are detectable. There are no long-range correlations ($\alpha\text{H}_i \rightarrow \text{NH}_{i+n}$). This observation is (mainly) due to the extensive peak overlap, but it is usually associated with a somewhat flexible helical conformation, in agreement with CD findings.

2D-NMR peptide **4c1**



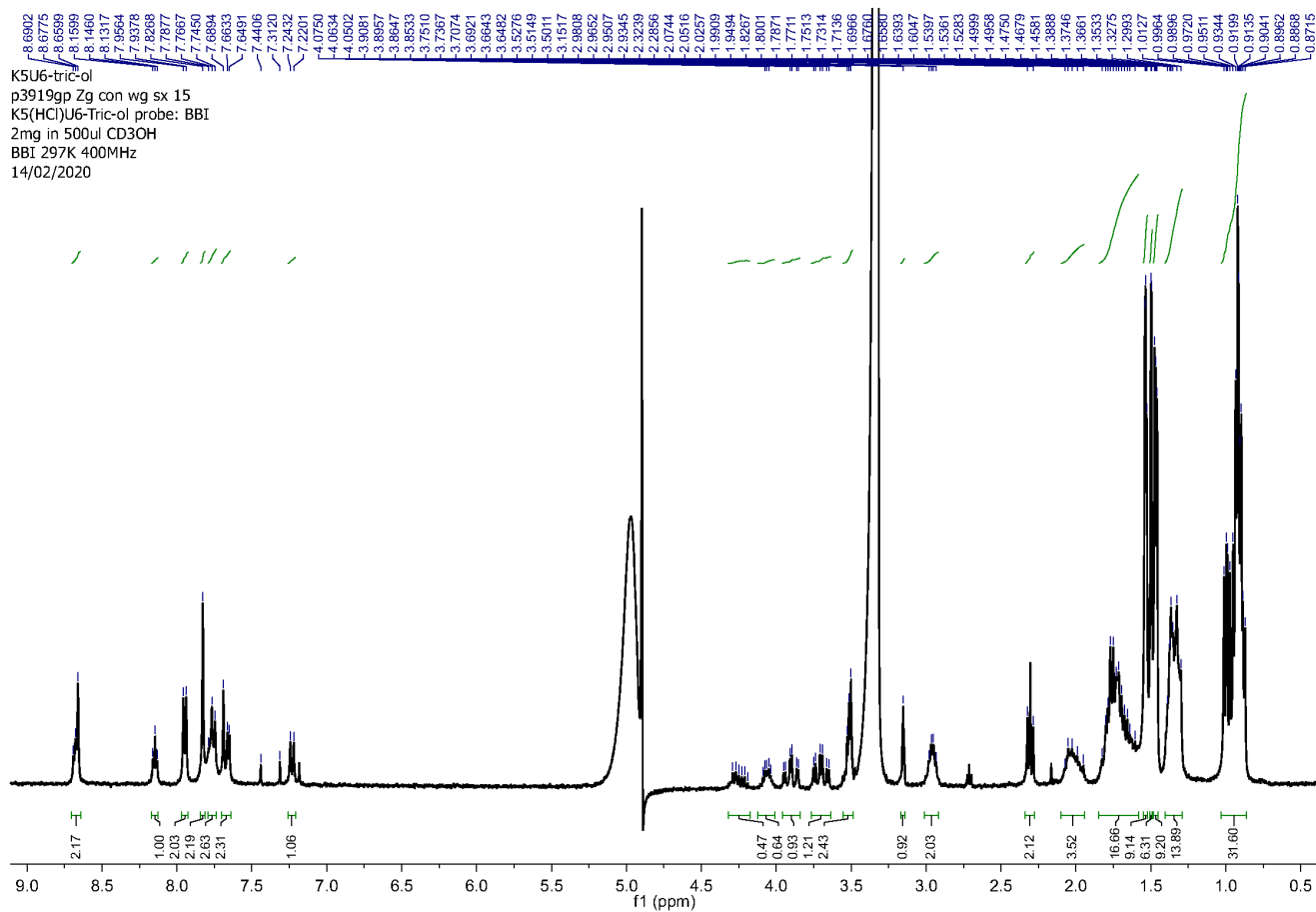
2D ${}^1\text{H}$ NMR for peptide **4c1** in CD_3OH , 297K, 400MHz (peptide concentration about 5 mM). The figure reports the NH-NH (amide) portion of the ROESY spectrum. All sequential $\text{NH}_i \rightarrow \text{NH}_{i+1}$ are detectable.

peptide 5 (K6)



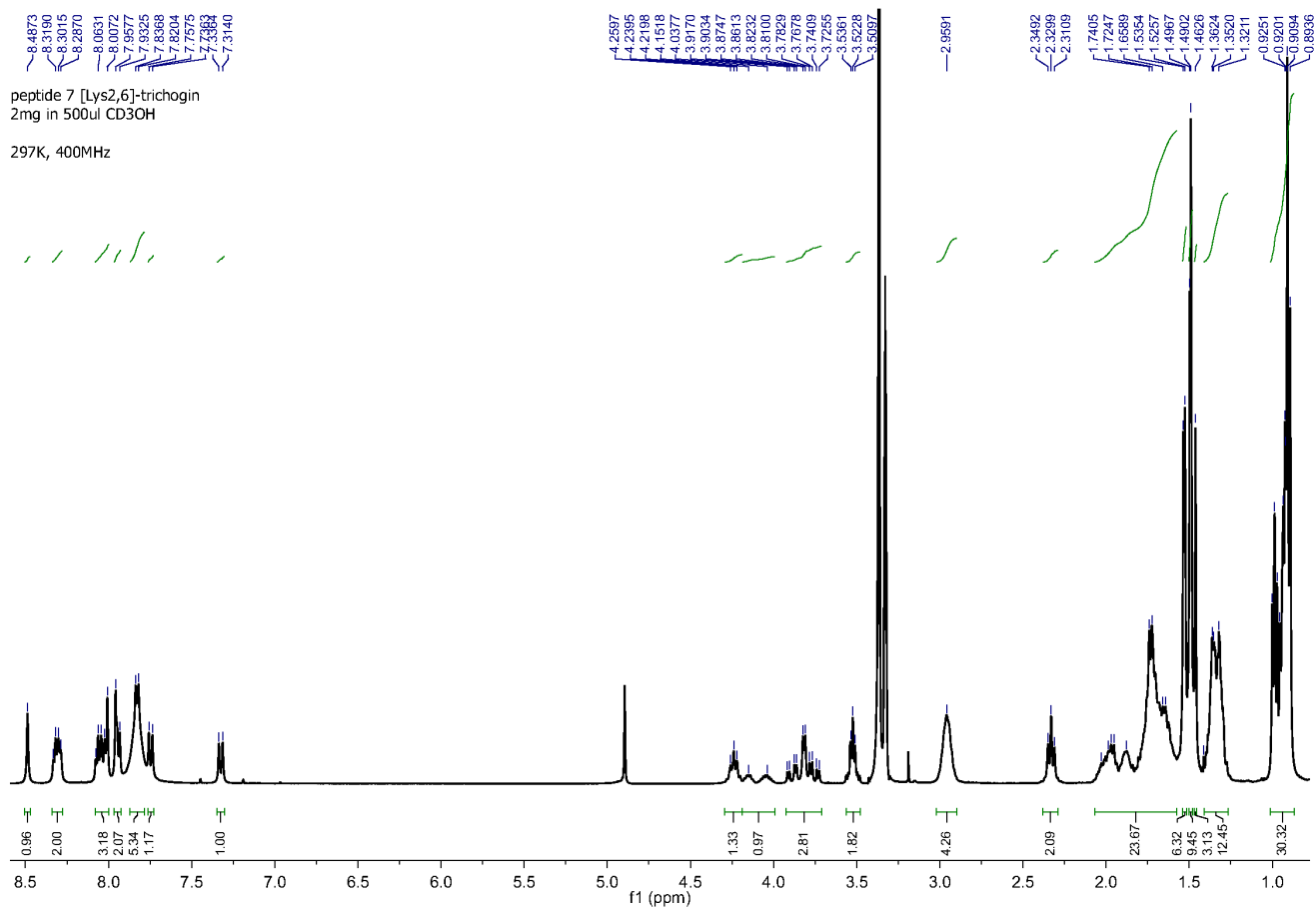
1D ^1H NMR spectrum for peptide 5 in CD_3OH , 298K, 600MHz (peptide concentration ca. 1.3 mM).

peptide 6 (K5U6)



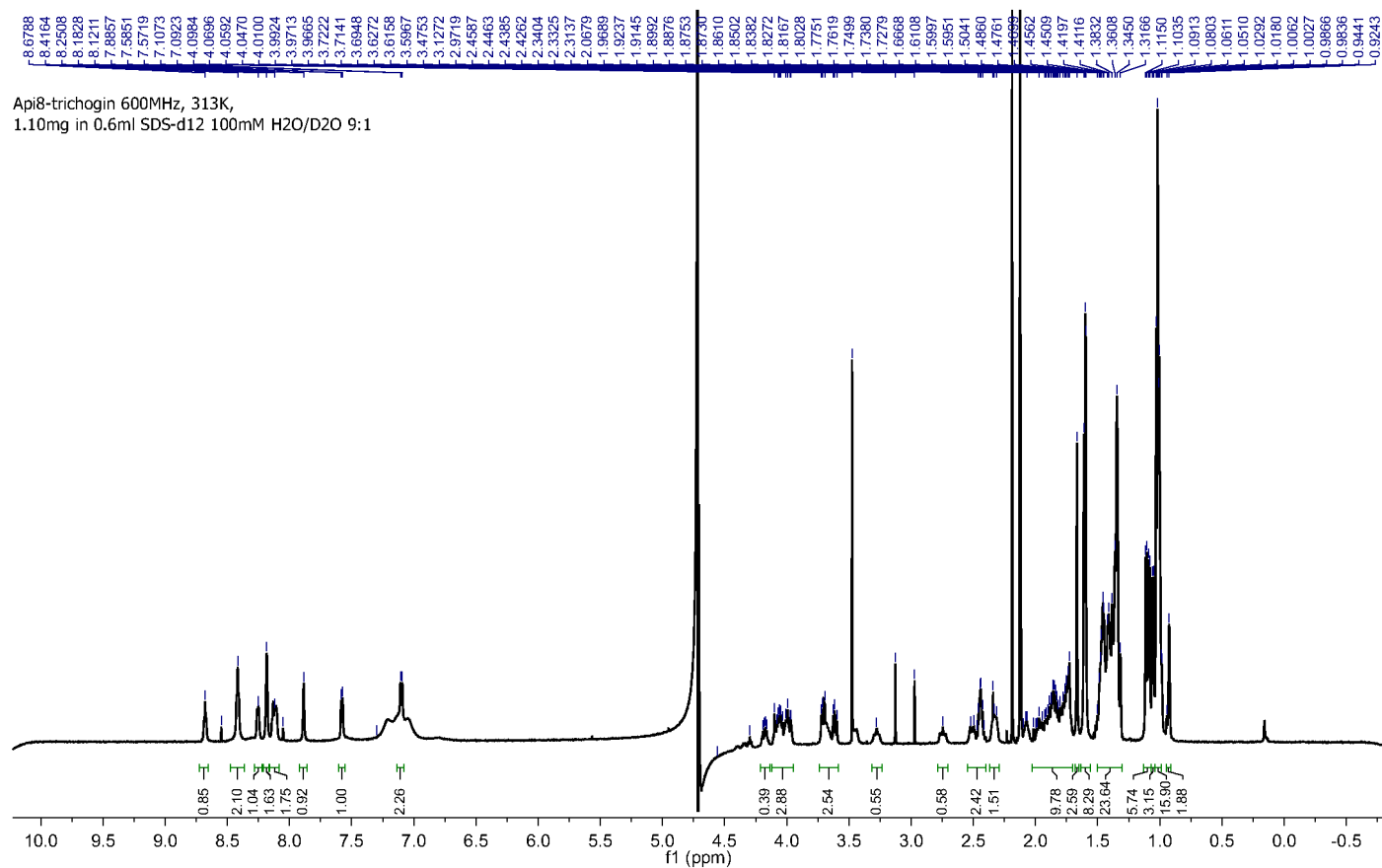
1D ¹H NMR spectrum for peptide 6 in CD₃OH, 297K, 400MHz (peptide concentration ca. 3 mM).

peptide 7 (K26)



1D ¹H NMR spectrum for peptide 7 in CD₃OH, 297K, 400MHz (peptide concentration ca. 3 mM).

peptide **8** (Api8)



1D ^1H NMR spectrum for peptide **8** in SDS-d₁₂ 100mM in H₂O/D₂O 9:1, 313K, 600MHz (peptide concentration ca. 2 mM).