Supplementary information

T cell-intrinsic role for Nod2 in protection against Th17-mediated uveitis

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Supplemental Figure 1. Anatomical and functional evaluation of eyes from Nod2^{-/-} mice. (a) Eyes of naive WT and Nod2^{-/-} mice were examined under magnification with a dissecting microscope (left), by fundoscopy (center), and by histology (right). The thickness of: (b) inner nuclear layer (INL) and (c) outer nuclear layer (ONL) of retinas of 8 week old WT and Nod2^{-/-} mice was assessed. Measurements were made from H&E-stained sections (6-8 sections obtained per mouse, n = 3 mice/genotype and are representative of 2 experiments, p=NS). (d, e) Electroretinography of 8 wk old WT and Nod2^{-/-} mice was performed to evaluate retinal function. Scotopic A-wave (d) and scotopic B-wave (e) amplitudes were used to assess functional cell responses dominated by photoreceptor and bipolar cell activity, respectively. Recordings were made from 6 mice/genotype and are representative of 2 independent experiments (p=NS). Data are mean \pm SEM. Statistical differences between groups were calculated using two-tailed Mann-Whitney U test (b,c) and unpaired Student's t-test (d, e). Source data are provided as a Source Data file.



Supplemental Figure 2. Examination of microbiota influence on uveitis. (a) To evaluate colony-associated differences, WT and Nod2⁴ mice, either bred in the VA Medical Unit (VMU) or newly arrived (within one day of experiment) from The Jackson Laboratory (JAX), were immunized with IRBP and evaluated by histopathology 20 days post-immunization. Data are combined from 2 independent experiments for n =10 (JAX-WT and VMU-Nod2+), 11 (JAX-Nod2⁺) and 18 (VMU-WT) mice. ***p<0.0001 (VMU comparison) and ***p = 0.0005 (JAX comparison). (b) To evaluate strain-specific microbiota, EAU was induced in WT and Nod2+ mice that were cohoused from the time of weaning (i.e. 4 weeks prior to immunization) and for the duration of the experiment. Uveitis was evaluated by histopathology 20 days postimmunization. Data are combined from 2 independent experiments where n = 16 mice/group. ***p<0.0001. (c) Mice were orally administered broad-spectrum antibiotics in their drinking water for 3 weeks prior to induction of EAU and uveitis was evaluated 20 days postimmunization by histopathology. Data are combined from 2 independent experiments where n =23 (WT-water), 21 (WT-Rx), 14 (Nod2⁴ water), 15 (Nod2⁴ Rx) mice. ***p<0.0001 (WT water vs. WT Rx, Nod2^{\pm} water vs. Nod2^{\pm} Rx, WT water vs. Nod2^{\pm} water) and ***p = 0.0002 (WT Rx vs. Nod2⁺ Rx). Data are box-whisker plots showing median, 25-75⁺ percentile, and min – max range. Statistical differences between groups were calculated using the two-tailed Mann-Whitney U test. Source data are provided as a Source Data file.



Supplemental Figure 3. *Nod2* expression in the eye. qPCR was used to evaluate mRNA expression of *Nod2* in distinct ocular tissues of naïve C57BL/6J mice, which are expressed as relative to mRNA expression of *Nod2* in spleen. Data are combined from 3 independent experiments (for each, dissected ocular tissues were pooled from n = 6 mice for RNA purification). Data are presented as floating bars with min – max range. C; cornea; I/CB, iris/ciliary body; R, retina; RPE/Ch, retinal pigment epithelium/choroid. Source data are provided as a Source Data file.



Supplemental Figure 4. The role of IFN γ in uveitis in WT and Nod2⁴ mice. The effect of IFN γ neutralization via administration of anti-IFN γ Ab or isotype control (IC) Ab on uveitis in WT and Nod2⁴ mice was assessed. (a) Clinical uveitis as evaluated by fundoscopy was scored repeatedly. Data are combined from 3 independent experiments where on day 14, n = 20 for all conditions. For day 21, n = 15 (WT IC), 19 (WT anti-IFN γ), 16 (Nod2⁴ IC), and 20 (Nod2⁴ anti-IFN γ). For 14 days; **p=0.0007 for Nod2⁴ IC vs. Nod2⁴ anti-IFN γ , **p=0.0189 for WT IC vs. Nod2⁴ IC. For 21 days; **p = 0.0006 for WT IC vs. WT anti-IFN γ , ***p<0.0001 for WT IC vs. Nod2⁴ IC. (b) Representative images of fundi and H&E-stained retinal sections 21 days post-immunization. Data are mean <u>+</u> SEM with dots representing individual values. Statistical differences between groups were calculated by unpaired Student's t-test. Source data are provided as a Source Data file.



Supplemental Figure 5. Increased expansion of Nod2⁴ CD4⁺ T cells in IRBP-immunized lymphopenic hosts. Splenocytes harvested from IRBP-immunized Rag1⁺ mice that had received either WT or Nod2⁴ CD4⁺ T cells were analyzed by flow cytometry for total numbers of indicated T cell subsets. Data are combined from 2 independent experiments where n = 6 (WT CD4⁺), 9 (Nod2⁴ CD4⁺, Nod2⁴ CD8⁺, Nod2⁴ CD4⁺ CD8⁺ "DN"), 8 (WT CD8⁺), 7 (WT DN) mice, **p=0.0008. Data are box-whisker plots with medians, 25-75^a percentile, and min-max range, and were compared using the two-tailed Mann-Whitney U test. Source data are provided as a Source Data file.



Supplemental Figure 6. Nod2-mediated protection against uveitis is independent of CFAcontaining immunization or MDP-stimulation. To evaluate a CFA-independent means of EAU induction, fungal-triggered uveitis was induced by immunization with IRBP emulsified in IFA containing heat-killed *S. cerevisiae* (HKSC) or heat-killed *C. albicans* (HKCA). Uveitis was evaluated by histopathology in WT and Nod2⁴ mice 20 days post-immunization using an expanded scale ranging 0 to 5. Data are combined from 2 independent experiments. For HKSC; n = 9 (WT) and 12 (Nod2⁴) and for HKCA; n = 10 mice/group. *p = 0.0062 (HKSC) and *p = 0.0151 (HKCA). (b) Splenocytes from naïve R161M or Nod2⁴ R161M mice were stimulated *in vitro* with MDP (10 µg/ml), versus PMA/ionomycin as a positive control. Culture supernatants were collected 72 hours post-stimulation and IL-17 production was evaluated by ELISA. Data is representative of 2 independent experiments where n = 5 mice/group. P = NS between WT and Nod2⁴ responses. Data are mean ± SEM with dots representing individual values (a), or boxwhisker plots with median, 25-75^a percentile, and min – max range (b). Statistical differences between groups were calculated using unpaired Student's t-test (a) or Mann-Whitney U test (b). Source data are provided as a Source Data file.