## Characterization of iRGD-ligand modified arginine-histidine-rich peptides for nucleic acid therapeutics delivery to $\alpha v\beta 3$ integrinexpressing cancer cells

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**Figure S1.** Detection of αvβ3 integrins on the surface of 293T (**a**,**b**), HeLa (**c**,**d**), PANC-1 (e,f) cell lines by flow cytometry analysis. (a,c,e) - unstained cells; (b,d,f) - CD51/CD61 antibody staining. The FACS data were processed by FlowJo software (FlowJo, LLC).



**Figure S2.** Cytotoxicity evaluation of free carriers in 293T, HeLa, PANC-1, and MDA-MB-231 cells by the Alamar blue assay. Values are the mean ± SD of the mean of triplicates. Amounts of carriers tested correspond to those applied for formation of NA/carriers complexes at N/P ratio 12/1.



**Figure S3.** Typical MALDI TOF mass spectra of RGD0 (a) and RGD1 (b) peptides.

Peptide analyzed	Calculated value of	Experimental value of
	molecular mass, g/mole	molecular mass, g/mole
RGD0	1972.28	1972.17
RGD1	2904.33	2903.54

Table S1. Experimental and theoretical molecular masses of RGD0 and RGD1 peptides.



**Figure S4.** SDS-PAGE results for RGD0 (1) and RGD1 (2) peptides; (3) - human protamine 1 protein; (4) - protein test mixture 6 for SDS PAGE (SERVA).

**Note 1:** RGD1 dimer cannot be fully denatured by SDS and possesses changed electrophoretic mobility.

**Note 2:** Tricine-sodium dodecyl sulphate-polyacrylamide gel electrophoresis 4%/16.5% (tricine-SDS-PAGE) was performed to separate low-molecular-weight peptides. Human protamine 1 protein (10.4 kDa) and protein test mixture 6 for SDS PAGE (SERVA) were used as molecular weight controls. Coomassie Brilliant Blue was used for the gel staining.



(b)

**Figure S5.**Typical HPLC results for RGD0 (a) and RGD1 (b) peptides.



**Figure S6.** Free thiol groups measurement by Ellman's assay after 24 h of RGD1 peptide cyclization.