

Expanded View Figures

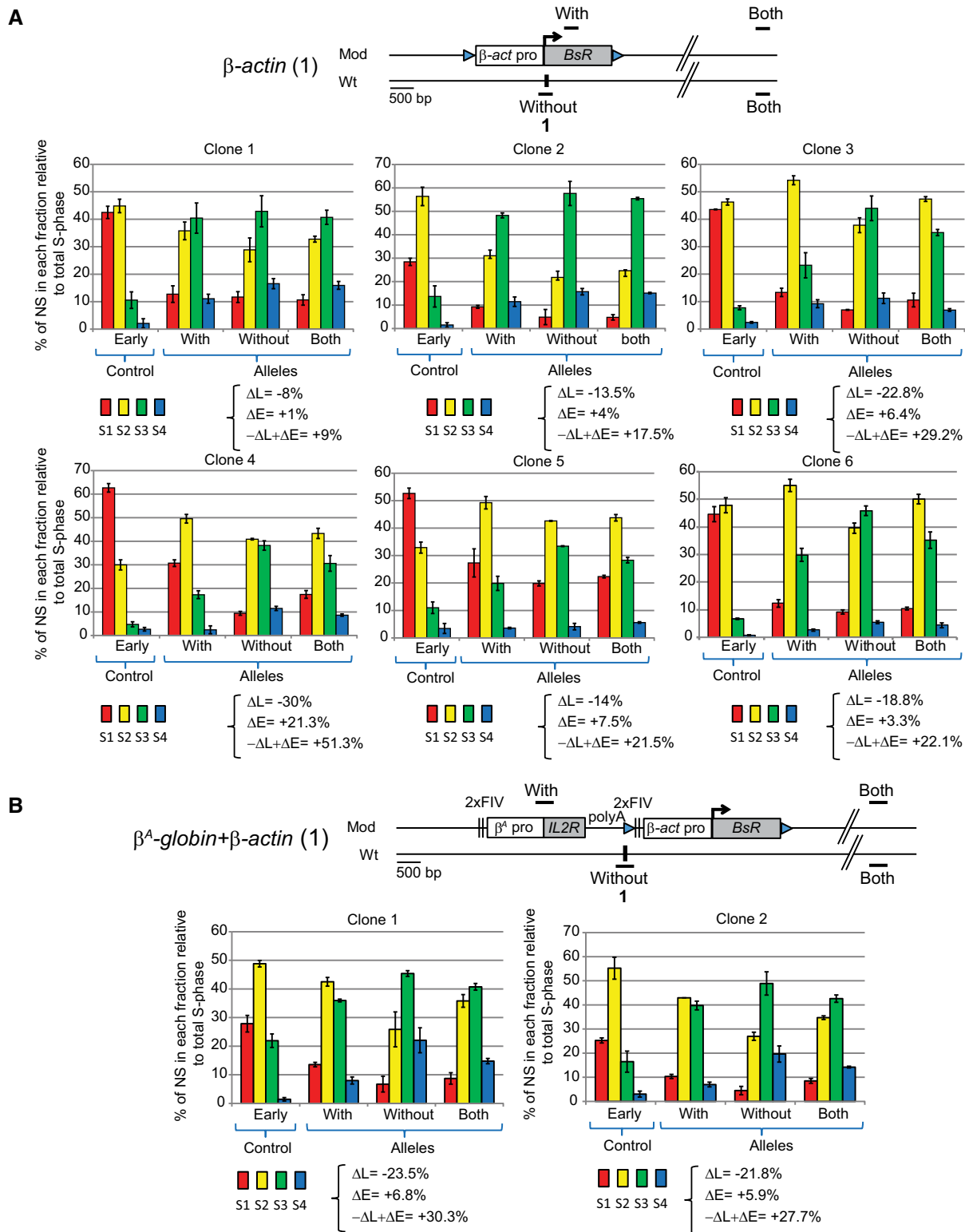


Figure EV1.

Figure EV1. Additional RT assays for transgenes shifting the timing of replication.

A, B RT profiles of each chromosomal allele are determined after targeted transgene integration using the allele-specific analysis of RT method by real-time PCR quantification described in Appendix Fig S1. Differences in $-\Delta L + \Delta E$ values calculated at the target site following transgene integration are indicated. Blue triangles represent reactive *loxP* sites. Error bars correspond to the standard deviation for qPCR duplicates. Analysis of six β^A -globin clonal cell lines (A) or two β^A -globin + β -actin clonal cell lines (B) described in Fig 1 is reported. Black vertical bars represent insertion sites.

Figure EV2. One advanced replicon inserted at site 1 or at site 3 directs a shift to earlier replication independently of the presence of a GFP reporter construct inserted at site 2.

RT profiles of each chromosomal allele are determined after targeted transgene integration using the allele-specific analysis of RT method by real-time PCR quantification described in Appendix Fig S1. Differences in $-\Delta L + \Delta E$ values calculated at the target site following transgene integration are indicated.

- A Analysis of one clonal cell line containing one *GFP* reporter construct composed of the *GFP* reporter gene under the control of the β^A -globin promoter (β^A pro) and linked to a 1.6 kb fragment of human chromosome 7 (h.K7) inserted at site 2.
- B, C Analysis of clonal cell lines containing one β^A -globin + β -actin construct described in Fig 1 inserted at site 1 and one *GFP* reporter construct inserted at site 2 on the same chromosome (B) or on the other chromosome (C).
- D, E Analysis of clonal cell lines containing one β^A -globin+ β -actin construct inserted at site 3 and one *GFP* reporter construct inserted at site 2 on the same chromosome (D) or on the other chromosome (E).

Data information: Blue and black triangles represent reactive *loxP* sites and recombined inactive *loxP* sites, respectively. Black vertical bars represent insertion sites. Error bars correspond to the standard deviation for qPCR duplicates.

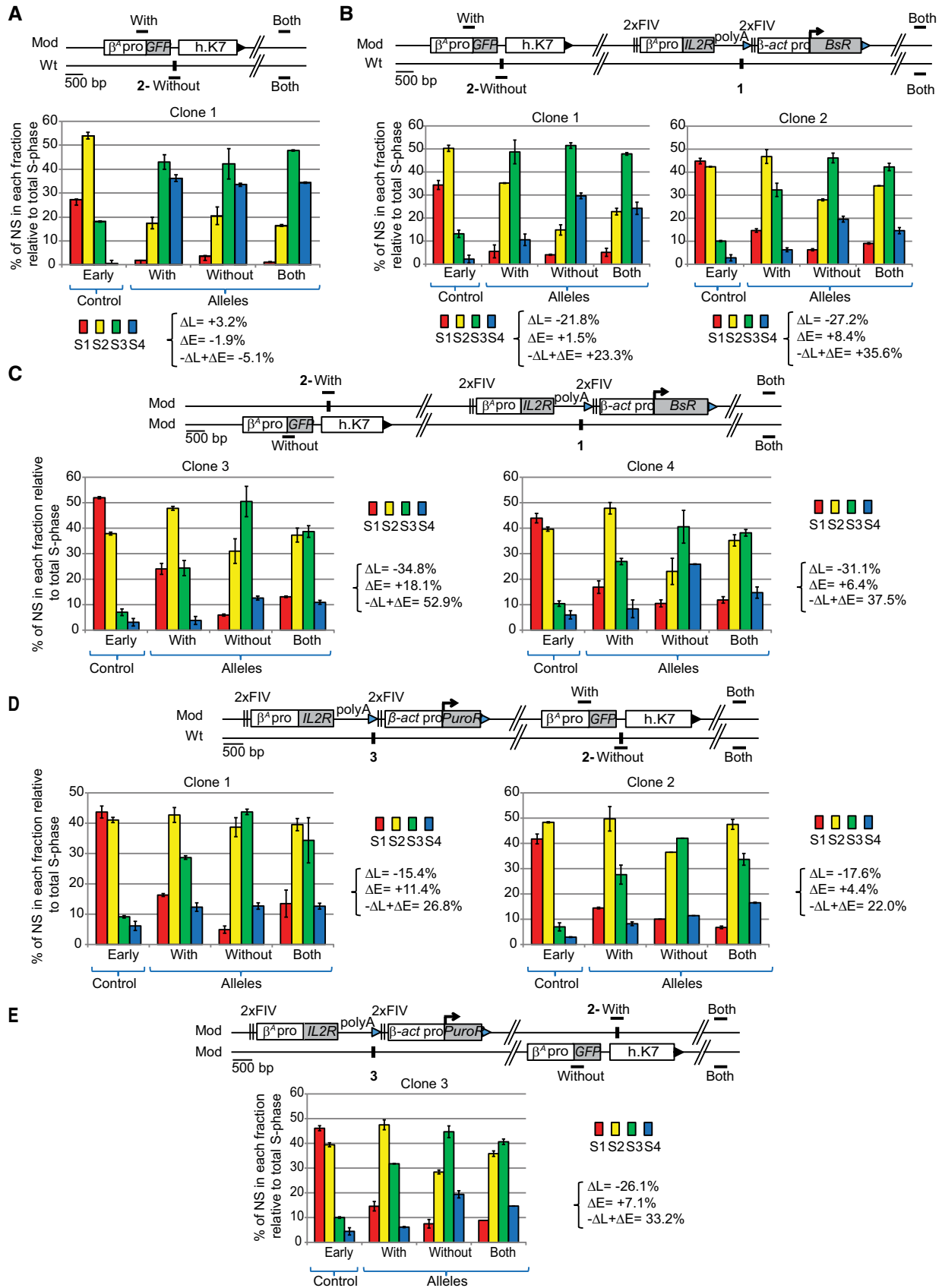


Figure EV2.

Figure EV3. Two β^A -globin + β -actin constructs as well as two β -actin constructs inserted at sites 1 and 3 form an early-replicated domain.

A–C RT profiles of each chromosomal allele are determined after targeted transgene integration using the allele-specific analysis of RT method by real-time PCR quantification described in Appendix Fig S1. Differences in $-\Delta L + \Delta E$ values calculated at the target site following transgene integration are indicated. Error bars correspond to the standard deviation for qPCR duplicates. (A, B) Analysis of two 1 + 2 + 3 (A) and three 1 + 3 (B) clonal cell lines described in Fig 3A. (C) Analysis of five clonal cell lines containing two β -actin constructs inserted at sites 1 and 3 on the same chromosome described in Fig 3B. Blue and black triangles represent reactive *loxP* sites and recombined inactive *loxP* sites, respectively. Black vertical bars represent insertion sites. Error bars correspond to the standard deviation for qPCR duplicates.

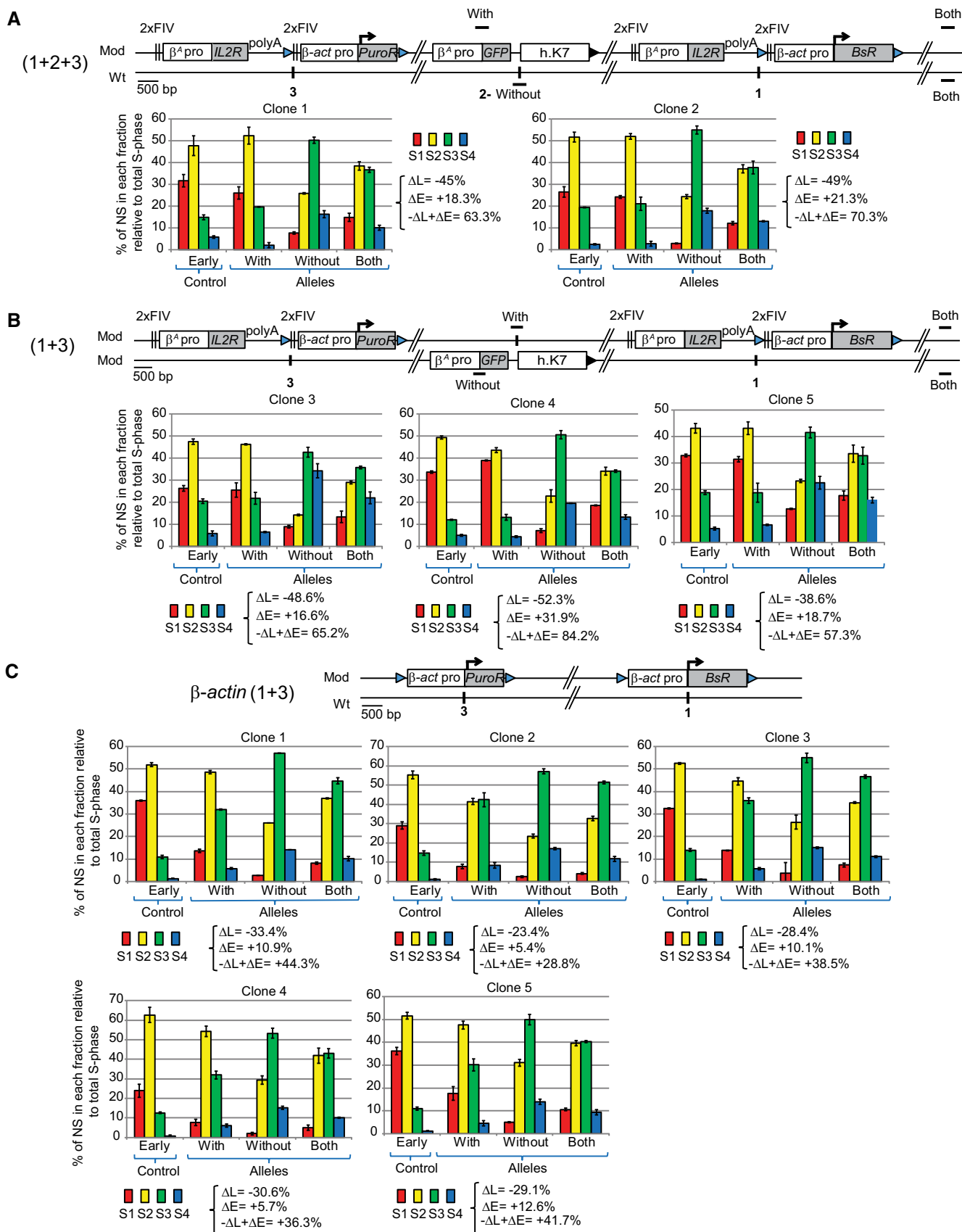
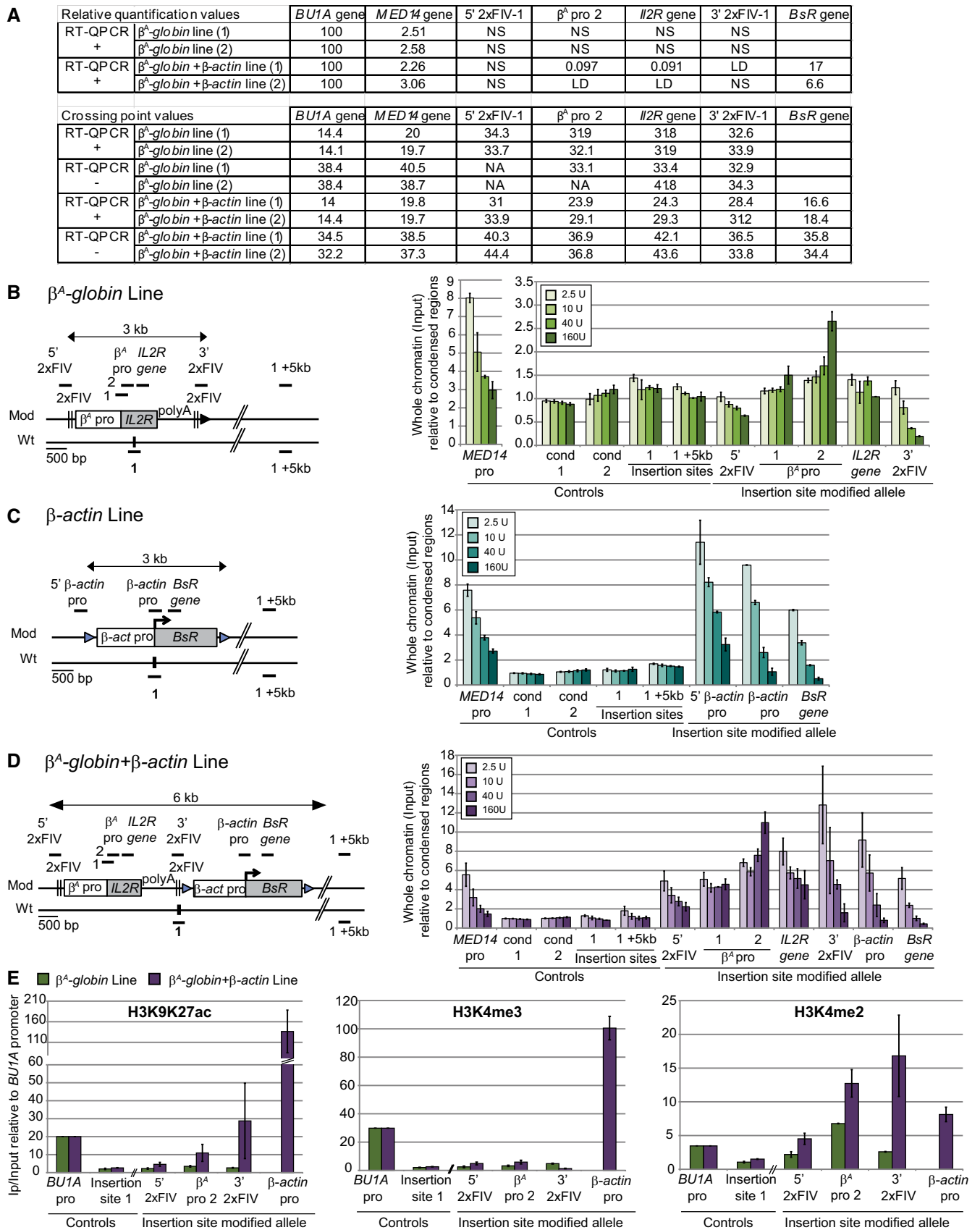


Figure EV3.

Figure EV4. The open chromatin structure of the whole advanced replicon is imposed by the active β -actin promoter.

- A Relative quantification, by real-time qPCR, of mRNA expression levels (RT-QPCR+) or background levels (RT-QPCR–), was performed in the β^A -globin and β^A -globin + β -actin clonal cell lines. For relative quantification, mRNA levels were normalized against *BU1A* mRNA levels arbitrarily set at 100 (first table). Crossing point (Cp) values for RT-QPCR+ and RT-QPCR– experiments are reported in the second table. NA corresponds to nonamplified samples, NS to nonspecific signals, and LD to the limit of detection.
- B–D The β^A -globin (B), β -actin (C), and β^A -globin + β -actin (D) transgenes are shown on the left, with the positions of the amplicons used for quantification (thick black lines with names indicated above). The endogenous active *MED14* promoter and two genomic regions located within the condensed region upstream from the β -globin locus (cond1 and cond2) were analyzed as controls. Quantification, by real-time qPCR, of total chromatin (input) extracted from the β^A -globin (B), β -actin (C), and β^A -globin + β -actin (D) clonal cell lines after digestion with increasing concentrations of micrococcal nuclease (MNAse, 2.5 U, 10 U, 40 U, 160 U/ml). Error bars indicate the standard deviation for qPCR triplicates made on two independent clones.
- E Immunoprecipitations of H3K9K27 acetylation (ac), H3K4 trimethylation (me3), or H3K4 dimethylation (me2) on formaldehyde-cross-linked chromatin extracted from the β^A -globin (green) and β^A -globin+ β -actin (purple) clonal cell lines. The endogenous active *BU1A* promoter (*BU1A pro*) was analyzed as a control. Data are presented as enrichments of immunoprecipitated material relative to input DNA and normalized against *BU1A* enrichment. Error bars indicate the standard deviation for at least qPCR duplicates made on two independent clones.



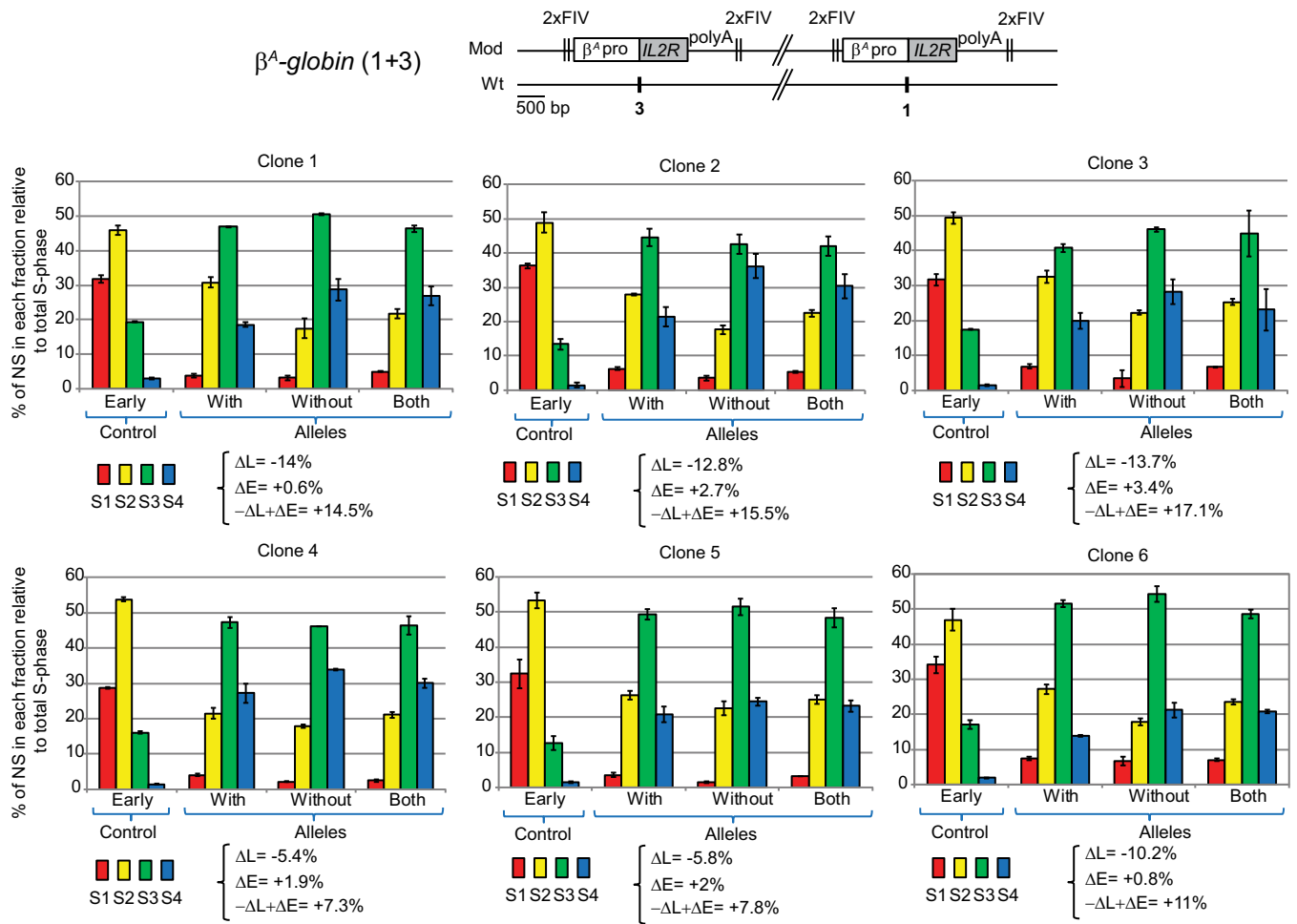


Figure EV5. Two β^A -globin constructs inserted at sites 1 and 3 are not sufficient to form an early-replicated domain.

RT profiles of each chromosomal allele are determined after targeted transgene integration using the allele-specific analysis of RT method by real-time PCR quantification described in Appendix Fig S1. Differences in $-\Delta L + \Delta E$ values calculated at the target site following transgene integration are indicated. Error bars correspond to the standard deviation for qPCR duplicates. Analysis of six β^A -globin (1 + 3) clonal cell lines described in Fig 4B.