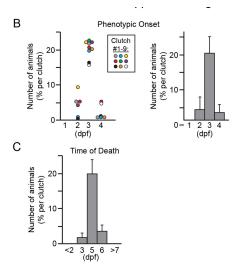
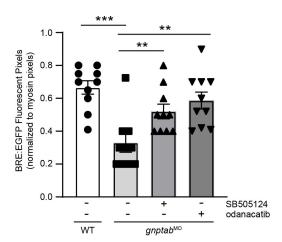
Α	Table o	of F0 foun	der ani	mals outcomes		
	Line	Female	Male	Concentration Tale mRNA (pg		outcross progeny with Mutation
	1	F		0.5 pg	5bp del	11/46
	3*	F		0.5 pg	2bp del	14/14
	5	F		0.5 pg	1bp del	1/20
	6	F		0.5 pg	25bp ins; 8bp del	18/20
	8	F		0.5 pg	5bp ins; 6bp del	15/20
	10	F		0.3 pg	8bp del	4/20
	14	F		0.3 pg	7bp del	2/20
	19	F		0.3 pg	4bp ins; 3bp ins	9/20
	1		М	0.5 pg	5bp del; 8bp del	2/11
	2*		M	0.5 pg	5bp del	17/20
	3*		M	0.5 pg	2bp del; 7bp del	13/13
	5		M	0.5 pg	4bp del; 5bp del	7/14
	6		M	0.5 pg	5bp del; 8bp del; 10bp in	s 6/15
	9		M	0.5 pg	1bp del	2/20
	10		M	0.5 pg	1bp ins	4/20
	13		M	0.3 pg	6bp del	10/21
	14		M	0.3 pg	5bp del;7bp del	5/20

D

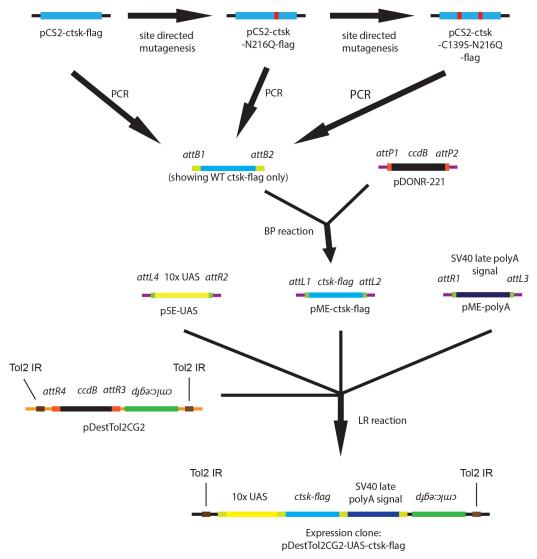


Primer	Primer	Gene	Experimenta
Name	Sequence	Location	Use
	GGCATATCAGACTTGA	Intron 7 to Intron 8	RFLP genotyping
gnptab TALE	ATTOAOAAAOATOOOAT		
For: AATGG	AGTCGAGATCAGTACCA CACTCACCTCGTCTG	Ex2 to Intron2	HRM genotyping
	GGCATATCAGACTTGA ATTCACAAACATCCCAT	Ex7 to Ex8	RT-PCR an Sequencing
	TCTAACAATTTGTCCGTG		RFLP genotyping
	AGCAGTGTGCCTACA TGCATGGCATCTATG	Ex4-5 to Ex5	Traditional Po
	GAAGCTCAATGGCACA ATTAGCAGCCTCTTG	Ex4-5 to Ex6	Quantitative a Non-Quantitat RT-PCR
For: GGAGG	GTCAGATGTTCAGGA CACCAGGTTCTGCTC	Ex5 to Ex5	Quantitative a Non-Quantitat RT-PCR
Rev: GCCCG	ACTGGCTTCAGGATT ATGTTAGCGACAG	Ex6 to Ex6	Quantitative a Non-Quantitat RT-PCR
	ACTCTCCTCACAGCA CACAAAGTTCTCTCC	Ex17-Ex18	Quantitative a Non-Quantitat RT-PCR
	CGACCGTTAATCTC GCTGGCATAACCACAT	Ex2 to Ex2	Quantitative a Non-Quantitat RT-PCR

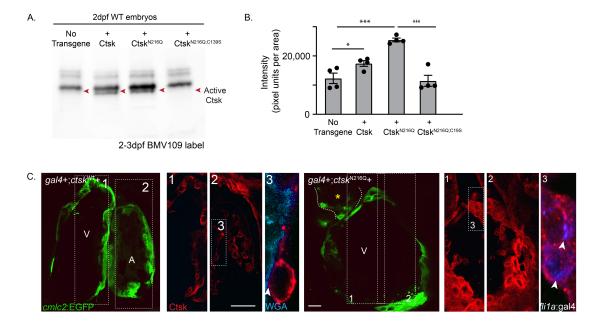
Supplemental Figure 1. A) Table illustrating the outcomes of F0 founder analyses. 20 males and 20 females were screened following injection of indicated TALEN concentrations. Shown are animals with interesting insertions and deletions following sequence analyses. The genetic mutations detected and the number of offspring from outcross matings with TLAB carrying the mutation listed. F1 outcrossed lines were generated from female 3 and males 2 and 3. B) Graph illustrates timing of phenotypic onset as scored from 9 distinct clutches. Circles represent individual clutches and indicate the number of embryos in the clutch (y axis) and time of phenotypic onset (x axis). This is also shown in the accompanying bar graph. C) Graph illustrates time of death for embryos in these phenotypically scored clutches. D) List of primers used in the study.



Supplemental Figure 2. BMP signaling is reduced in the hearts of *gnptab*-deficient MLII embryos. A) Graphs represent quantitation of BMP signaling, as assessed using BRE:EGFP transgenic zebrafish. EGFP fluorescence was calculated in the right half of the ventricle. EGFP fluorescence was normalized to fluorescent units for myosin staining within the same region.



Supplemental Figure 3. Schematic illustrating generation of TOL2 constructs to create transgenic zebrafish lines expressing WT, N216Q, and N216Q;C139S forms of cathepsin K under the control of UAS.



Supplemental Figure 4. A) BMV109 labeling of all transgenic conditions show Ctsk activity is increased in the CtskN216Q variant but completely lost in the Ctsk^{C139S;N216Q}, suggesting the variant is indeed catalytically dead. Representative gel from 4 independent experiments. B) Densitometric quantitation of Ctsk activity in 4 experiments. All measurements normalized to total protein load as assayed by Coomasie stain following gel imaging. C) Enlarged images from from Figure 7D showing the N216Q Ctsk variant is secreted from endothelial cells (see white arrows, panels 3). This is demonstrated by overlap with WGA (blue), which labels glycans at the cell surface and in the extracellular space. This is in contrast to WT Ctsk which is clearly distinct from extracellular WGA. Scale bars=20μm.