

Additional information for

Culicidae evolutionary history focusing on the Culicinae subfamily based on mitochondrial phylogenomics

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Supplementary table 1. Annotation of draft mitogenomes using MITOS.

Species	<i>nad2</i>	<i>COI</i>	<i>COII</i>	<i>atp8</i>	<i>atp6</i>	<i>COIII</i>	<i>nad3</i>	<i>nad5</i>	<i>nad4</i>	<i>nad4l</i>	<i>nad6</i>	<i>cytb</i>	<i>nad1</i>	<i>rrnL</i>	<i>rrnS</i>	<i>tRNA</i>	<i>PCG</i>	<i>rRNA</i>	Total of genes
<i>Ad. squamipennis</i>	1	1	1	-	-	1	1	1	1	-	-	1	1	1	1	16	9	2	27
<i>Ae. alboannulatus</i>	1	1	1	-	-	1	1	1	1	1	1	1	1	1	1	13	11	2	26
<i>Ae. camptorhynchus</i>	1	1	1	1	-	1	1	1	1	1	1	1	1	1	1	16	12	2	30
<i>Ae. detritus</i>	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	16	13	2	31
<i>Ae. fluviatilis</i>	1	1	1	1	1	1	-	1	1	1	1	1	1	1	-	4	12	1	17
<i>Ae. polynesiensis</i>	1	1	1	1	-	1	1	1	1	1	1	1	1	1	1	20	12	2	34
<i>Ae. riversi</i>	-	1	1	1	1	1	1	1	1	-	-	1	1	1	-	6	10	1	17
<i>Ae. scapularis</i>	1	1	1	1	1	1	-	1	1	-	1	1	1	-	1	13	11	1	25
<i>Ae. taeniorhynchus</i>	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	22	13	2	37
<i>An. albimanus</i>	1	1	1	1	-	1	1	1	1	1	1	1	1	1	1	20	12	2	34
<i>An. aquasalis</i>	1	1	1	1	-	1	-	1	1	-	-	1	1	1	1	3	9	2	14
<i>An. freeborni</i>	1	1	1	1	-	1	1	1	1	1	1	1	1	1	1	19	12	2	33
<i>An. nuneztovari</i>	1	1	1	1	-	1	1	1	1	1	1	1	1	1	1	9	12	2	23
<i>An. quadriannulatus</i>	1	1	1	1	-	1	1	1	1	1	1	1	1	1	1	20	12	2	34
<i>Cq. albicosta</i>	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	21	13	2	36
<i>Cq. chrysonotum</i>	1	1	1	-	1	1	1	1	1	1	1	1	1	1	1	20	12	2	34
<i>Cq. hermanoi</i>	1	1	1	1	1	1	1	1	1	-	-	1	-	1	1	14	10	2	26
<i>Cq. juxtamazonia</i>	1	1	1	-	1	1	-	1	1	-	-	1	-	1	1	8	8	2	18
<i>Cq. venezuelensis</i>	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	19	13	2	34
<i>Cx. amazonensis</i>	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	21	13	2	36
<i>Cx. australicus</i>	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	17	13	2	32
<i>Cx. corniger</i>	1	1	1	1	-	1	-	1	1	1	-	1	-	1	1	9	9	2	20
<i>Cx. globocoxitus</i>	1	1	1	1	-	1	1	1	1	1	1	1	1	1	1	17	12	2	31
<i>Cx. hortensis</i>	1	1	1	-	-	1	1	1	1	1	1	1	1	1	1	12	11	2	25
<i>Cx. pipiens molestus</i>	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	6	13	2	21
<i>Cx. nigripalpus</i>	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	22	13	2	37
<i>Cx. tarsalis</i>	1	1	1	1	-	1	1	1	1	1	1	1	1	1	1	19	12	2	33
<i>Cx. torrentium</i>	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	11	13	2	26
<i>Ma. wilsoni</i>	1	1	1	1	1	1	1	1	1	1	-	1	1	1	1	14	12	2	28
<i>Ma.humeralis</i>	-	1	1	-	1	-	1	1	-	-	-	1	1	1	1	5	7	2	14
<i>Ma.titillans</i>	1	1	1	-	1	1	1	1	1	-	1	1	1	1	1	14	11	2	27
<i>Ps. cingulata</i>	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	21	13	2	36
<i>Ps.albipes</i>	1	1	1	1	-	1	1	1	1	1	1	1	1	1	1	19	12	2	33
<i>Tp. aranoioides</i>	1	1	-	-	-	1	-	1	-	1	-	1	1	1	1	-	7	2	9
<i>Tr. digitatum</i>	1	1	1	-	1	1	1	1	1	1	-	1	-	1	1	16	10	2	28
<i>Tx. amboinensis</i>	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	19	13	2	34
<i>Ur. pulcherrima</i>	1	1	1	-	1	1	1	1	1	1	1	1	1	1	1	18	12	2	32

Legend: Bold names represent sequenced mitogenomes. Disrupted or duplicated gene are represented by bold numbers. Unidentified gene are represented by -.

Supplementary table 2. Predicted TMRCA values from Bayesian inference analysis for studied species clades.

Node*	HPD 95% ¹	Median dating ¹	PP of analysis ₁	PP of analysis ₂	UFboot of analysis ₂	PP of analysis ₃	PP of analysis ₄	PP of analysis ₅	UFboot of analysis ₅	PP of analysis ⁶	UFboot of analysis ⁷
A	243.79-332.41	273.74	1	1	-	1	1	1	-	1	-
B	145.88-232.95	182.75	1	1	-	1	1	1	-	1	-
F	128.09-204.91	160.86	1	-	-	-	-	-	-	-	-
I	124.77-200.69	157.02	0.49	0.81	-	0.49	-	0.96	69	0.99	-
H	117.92-195.06	151.36	0.5	-	-	-	-	-	-	-	-
C	114.58-187.29	145.83	1	1	100	1	1	1	100	1	100
N	101.87-168.21	130.49	1	1	93	1	1	0.99	92	1	79
J	88.78-146.1	113.73	1	1	100	1	1	-	-	0.81	83
D	87.14-141.71	110.73	1	1	100	1	1	1	100	1	100
U	79.07-135.87	103.9	1	1	100	1	1	1	100	1	100
O	81.02-132.17	102.87	1	1	100	1	1	1	100	1	100
Q	74.02-121.02	94.04	1	1	100	1	1	1	100	1	-
K	66.37-115.22	88.32	1	1	100	1	1	1	100	1	100
L	66.31-109.73	85.29	1	1	100	1	1	1	100	1	100
S	53.53-88.54	68.94	1	-	100	1	1	-	-	-	-
P	45.84-84.94	63.25	1	1	100	1	1	1	100	1	100
T	43.26-73.19	56.37	0.92	-	65	0.95	-	-	-	-	-
R	24.82-45.09	33.9	1	1	100	1	1	1	100	1	100
V	16.73-28.91	22.09	1	1	100	1	-	-	100	0.85	89
Z	15.05-26.45	20.08	1	1	100	1	0.85	0.86	97	-	98
Y	9.82-17.37	13.15	1	1	-	1	1	-	-	0.96	99
X	7.78-16.85	11.85	1	1	100	1	1	1	100	1	99
M	3.13-8.92	5.71	1	1	100	1	1	1	100	1	100
E	2.35-4.42	3.27	1	1	100	1	1	1	100	1	100
G	1.43-2.81	2.04	1	1	100	1	1	1	-	1	-

The HPD95% values represent the confidence intervals for each node. PP represents the probability posterior. UFboot represents ultrafast bootstrapping. * Nodes present in Fig. 3 and Fig. 4. ¹ Analysis based on partitioned PCG with codon partition (1st + 2nd and 3rd separately). ² Analysis based on complete mitogenome sequences. ³ Analysis based on partitioned PCG without codon partition. ⁴ Analysis based on Partitioned PCG without 3rd codon positions. ⁵ Analysis based on concatenated predicted amino acid sequences. ⁶ Analysis based on partitioned predicted amino acid sequences. ⁷ Analysis based on concatenated PCG without 3rd codon position.

Supplementary table 3. Information of sequenced samples.

Taxon	Genus	Subgenus	Tribe	N° of specimens	Location	N° of reads (Mi)
<i>Ad. squamipennis</i>	Aedeomyia	Aedeomyia	Aedeomyiini	3	Aldeia-PE+REC ²	3.9
<i>Ae. scapularis</i>	Aedes	Ochlerotatus	Aedini	7	Juazeiro-BA	7
<i>Ae. taeniorhynchus</i>	Aedes	Ochlerotatus	Aedini	10	São Luís -MA	11.3
<i>Cq. hermanoi</i>	Coquillettidia	Rhynchoaenia	Mansoniini	1	Aldeia-PE	1.8
<i>Cq. chrysonotum</i>	Coquillettidia	Rhynchoaenia	Mansoniini	14	PDI ¹	6.2
<i>Cq. albicosta</i>	Coquillettidia	Rhynchoaenia	Mansoniini	3	PDI ¹	2.4
<i>Cq. juxtamansonia</i>	Coquillettidia	Rhynchoaenia	Mansoniini	2	REC ²	5.5
<i>Cq. venezuelensis</i>	Coquillettidia	Rhynchoaenia	Mansoniini	4	PDI ¹ +REC ²	4.3
<i>Cx. amazonensis</i>	Culex	Aedinus	Culicini	1	REC ²	2.5
<i>Cx. corniger</i>	Culex	Phenacomyia	Culicini	5	Mato Grosso	3.9
<i>Cx. nigripalpus</i>	Culex	Culex	Culicini	6	PDI ¹	4.8
<i>Ma. titillans</i>	Mansonia	Mansonia	Mansoniini	3	REC ²	6.6
<i>Ma. wilsoni</i>	Mansonia	Mansonia	Mansoniini	2	REC ²	8.2
<i>Ma. humeralis</i>	Mansonia	Mansonia	Mansoniini	1	Aldeia-PE	2.2
<i>Ps. cingulata</i>	Psorophora	Grabhamia	Aedini	3	Mato Grosso	4.9
<i>Tr. digitatum</i>	Trichoprosopon	Trichoprosopon	Sabethini	1	Mato Grosso	7.6
<i>Ur. pulcherrima</i>	Uranotaenia	Uranotaenia	Uranotaeniini	1	REC ²	1.1

PE refers to state of Pernambuco, Brazil. MA refers to state of Maranhão, Brazil. BA refers to state of Bahia, Brazil. Mi refers to million of reads. ¹ Parque Estadual Dois Irmãos-PE . ² Reserva Ecológica de Carnijó-PE

Supplementary table 4. NCBI accession number of mitochondrial genomes retrieved from NCBI.

Species	Genus	Subgenus	Tribe	Access	Length (bp)	References *
<i>Aedes aegypti</i>	Aedes	Stegomyia	Aedini	NC_010241.1	16,655	1
<i>Aedes albopictus</i>	Aedes	Stegomyia	Aedini	NC_006817.1	16,665	-
<i>Aedes notoscriptus</i>	Aedes	Finlaya	Aedini	KM676218.1	15,846	2
<i>Aedes vigilax</i>	Aedes	Ochlerotatus	Aedini	KP995260.1	15,877	3
<i>Anopheles albitarsis</i>	Anopheles	Nyssorhynchus	-	HQ335344.1	15,413	4
<i>Anopheles bellator</i>	Anopheles	Kerteszia	-	KU551287.1	15,668	5
<i>Anopheles christyi</i>	Anopheles	Cellia	-	NC_028214.1	14,967	6
<i>Anopheles cruzii</i>	Anopheles	Kerteszia	-	KJ701506.1	15,449	-
<i>Anopheles culicifacies</i>	Anopheles	Cellia	-	NC_027502.1	15,330	7
<i>Anopheles darlingi</i>	Anopheles	Nyssorhynchus	-	NC_014275.1	15,386	8
<i>Anopheles deaneorum</i>	Anopheles	Nyssorhynchus	-	HQ335347.1	15,424	4
<i>Anopheles dirus</i>	Anopheles	Cellia	-	JX219731.1	15,404	9
<i>Anopheles epiroticus</i>	Anopheles	Cellia	-	NC_028217.1	15,379	6
<i>Anopheles farauti</i>	Anopheles	Cellia	-	NC_020770.1	15,412	9
<i>Anopheles funestus</i>	Anopheles	Cellia	-	NC_008070.1	15,354	10
<i>Anopheles gambiae</i>	Anopheles	Cellia	-	NC_002084.1	15,363	11
<i>Anopheles homunculus</i>	Anopheles	Kerteszia	-	KU551283.1	15,739	5
<i>Anopheles janconnae</i>	Anopheles	Nyssorhynchus	-	HQ335348.1	15,425	4
<i>Anopheles koliensis</i>	Anopheles	Cellia	-	JX219743.1	15,412	9
<i>Anopheles melas</i>	Anopheles	Cellia	-	NC_028219.1	15,366	6
<i>Anopheles merus</i>	Anopheles	Cellia	-	NC_028220.1	15,365	6
<i>Anopheles punctulatus</i>	Anopheles	Cellia	-	NC_028222.1	15,322	6
<i>Anopheles quadrimaculatus</i>	Anopheles	Anopheles	-	NC_000875.1	15,455	12
<i>Anopheles sinensis</i>	Anopheles	Anopheles	-	NC_028016.1	15,076	-
<i>Anopheles stephensi</i>	Anopheles	Cellia	-	NC_028223.1	15,387	6
<i>Bironella hollandi</i>	Bironella	Brugella	-	NC_037796.1	15,772	13
<i>Chagasia sp.</i>	Chagasia	-	-	MF381717.1	15,717	13
<i>Culex bidens</i>	Culex	Culex	Culicini	NC_037809.1	15,583	13
<i>Culex gelidus</i>	Culex	Culex	Culicini	KX753344.1	15,600	14
<i>Culex bilineatus</i>	Culex	Culex	Culicini	NC_037819.1	15,599	13
<i>Culex brami</i>	Culex	Culex	Culicini	NC_037828.1	15,586	13
<i>Culex camposi</i>	Culex	Culex	Culicini	NC_036008.1	15,570	15

<i>Culex coronator</i>	Culex	Culex	Culicini	NC_036006.1	15,576	15
<i>Culex declarator</i>	Culex	Culex	Culicini	NC_037822.1	15,575	13
<i>Culex dolosus</i>	Culex	Culex	Culicini	MF381620.1	15,851	13
<i>Culex chidesteri</i>	Culex	Culex	Culicini	NC_037826.1	16,052	13
<i>Culex lygrus</i>	Culex	Culex	Culicini	MF381718.1	15,572	13
<i>Culex mollis</i>	Culex	Culex	Culicini	MF381705.1	15,576	13
<i>Culex pipiens pallens</i>	Culex	Culex	Culicini	KT851543.1	15,617	16
<i>Culex pipiens pipiens</i>	Culex	Culex	Culicini	NC_015079.1	14,856	-
<i>Culex quinquefasciatus</i>	Culex	Culex	Culicini	NC_014574.1	15,587	1
<i>Culex surinamensis</i>	Culex	Culex	Culicini	MF381615.1	15,568	13
<i>Culex tritaeniorhynchus</i>	Culex	Culex	Culicini	KT851544.1	14,844	16
<i>Culex usquatissimus</i>	Culex	Culex	Culicini	NC_036007.1	15,574	15
<i>Culex usquatus</i>	Culex	Culex	Culicini	NC_036005.1	15,573	15
<i>Drosophila melanogaster</i>	Drosophila	Drosophilinae	Drosophilini	U37541.1	19,517	17
<i>Haemagogus janthinomys</i>	Haemagogus	Haemagogus	Aedini	NC_028025.1	15,698	18
<i>Sabethes belisarioi</i>	Sabethes	Sabethes	Sabethini	MF957171.1	15,911	19
<i>Sabethes chloropterus</i>	Sabethes	Sabethoides	Sabethini	NC_037499.1	15,609	19
<i>Sabethes glaucodaemon</i>	Sabethes	Sabethoides	Sabethini	NC_037500.1	15,620	19

* References listed at the end of the document.

Supplementary table 5. Information about retrieved raw sequencing datasets from NCBI SRA database.

Species	Genus	Subgenus	Tribe	SRA accession number	Original study
<i>Ae. alboannulatus</i>	Aedes	Finlaya	Aedini	SRR5665565	RNA-Seq (Species virome)
<i>Ae. camptorhynchus</i>	Aedes	Ochlerotatus	Aedini	SRR5665568	RNA-Seq (Species virome)
<i>Ae. detritus</i>	Aedes	Ochlerotatus	Aedini	SRR6489869	RNA-Seq (Differential gene expression)
<i>Ae. fluviatilis</i>	Aedes	Ochlerotatus	Aedini	SRR3574348	RNA-Seq (Differential gene expression)
<i>Ae. polynesiensis</i>	Aedes	Stegomyia	Aedini	ERR1706659	RNA-Seq (Comparative transcriptomes)
<i>Ae. riversi</i>	Aedes	Stegomyia	Aedini	ERR1706661	RNA-Seq (Comparative transcriptomes)
<i>An. albimanus</i>	Anopheles	Nyssorhynchus	-	SRR314655	Genome sequencing
<i>An. aquasalis</i>	Anopheles	Nyssorhynchus	-	SRR927456	RNA-Seq (Transcriptome)
<i>An. freeborni</i>	Anopheles	Anopheles	-	SRR908289	RNA-Seq (Saliva transcriptome)
<i>An. nuneztovari</i>	Anopheles	Nyssorhynchus	-	SRR6471062	RNA-Seq (Saliva transcriptome)
<i>An. quadriannulatus</i>	Anopheles	Cellia	-	SRR529986	Genome sequencing
<i>Cx. australicus</i>	Culex	Culex	Culicini	SRR5665566	RNA-Seq (Species virome)
<i>Cx. globocoxitus</i>	Culex	Culex	Culicini	SRR5665567	RNA-Seq (Species virome)
<i>Cx. hortensis</i>	Culex	Maillotia	Culicini	SRR1324883	RNA-Seq (Comparative transcriptomes)
<i>Cx. pipiens molestus</i>	Culex	Culex	Culicini	SRR1462325	RNA-Seq (Comparative transcriptomes)
<i>Cx. tarsalis</i>	Culex	Culex	Culicini	SRR5149179	RNA-Seq (Transcriptome)
<i>Cx. torrentium</i>	Culex	Culex	Culicini	SRR1324895	RNA-Seq

<i>Ps. albipes</i>	Psorophora	Janthinossoma	Aedini	SRR908278	(Comparative transcriptomes) RNA-Seq (Saliva transcriptome)
<i>Tp. aranoides</i>	Tripteroides	Rachionotomyia	Sabethini	SRR6155938	RNA-Seq (Transcriptome)
<i>Tx. amboinensis</i>	Toxorhynchites	Toxorhynchites	Toxorhynchitini	SRR2061845	RNA-Seq (Transcriptome)

Supplementary table 6. Calibration points used in BEAST 1.8.4

Clade	Represented taxa	Minimum bound	Maximum bound	References
<i>Diptera</i>	<i>Culicidae</i> and <i>D. melanogaster</i>	240	241	KRZEMIŃSKI, KRZEMIŃSKA & PAPIER, 1994 ²⁰
<i>Culicidae</i>	<i>Culicinae</i> e <i>Anophelinae</i>	90	99	BORKENT & GRIMALDI, 2004 ²¹
<i>Culicinae</i>	<i>Culicinae</i>	76.5	79	POINAR, ZAVORTINK., PIKE & JOHNSTON, 2000 ²²
<i>Anophelinae</i>	<i>Anophelinae</i>	15	34	ZAVORTINK & POINAR, 2000 ²³

Supplementary table 7. Evolutionary models used in BEAST analysis.

Partition	Best nucleotide model ¹	Best protein model ²
Nucleotide mitogenome alignment	GTR+G+I	-
Concatenated protein alignment	-	mtREV+G+I
Atp6	GTR+G+I	mtREV+G+I
Atp8	GTR+G	JTT+G+I
COIII	GTR+G+I	mtREV+G+I
COII	GTR+G+I	mtREV+G
COI	GTR+G+I	JTT+G+I
CytB	GTR +G+I	mtREV+G+I
Nad1	GTR +G+I	JTT+G+I
Nad2	GTR +G+I	JTT+G
Nad3	GTR +G+I	mtREV+G+I
Nad4L	GTR +G+I	mtREV+G
Nad4	GTR +G+I	JTT+G+I
Nad5	GTR +G+I	JTT+G
Nad6	GTR +G+I	mtREV+G
rrnL	GTR +G+I	-
rrnS	GTR +G+I	-

¹Best model as suggested by SMS. ² More likely model suggested by Prottest and present in BEAST 1.8.4.

Supplementary file 1. Test of substitution saturation of mitochondrial PCG.

Analysis performed on all sites.

Testing whether the observed Iss is significantly lower than Iss.c.

IssSym is Iss.c assuming a symmetrical topology.

IssAsym is Iss.c assuming an asymmetrical topology.

Note: two-tailed tests are used.

COI 1st+2nd codon positions:

NumOTU	Iss	Iss.cSym	T	DF	P	Iss.cAsym	T	DF	P
4	0,119	0,821	72,061	1023	0,0000	0,789	68,777	1023	0,0000
8	0,120	0,789	64,934	1023	0,0000	0,684	54,715	1023	0,0000
16	0,124	0,773	64,019	1023	0,0000	0,576	44,550	1023	0,0000
32	0,147	0,750	59,214	1023	0,0000	0,445	29,334	1023	0,0000

COI 3rd codon positions:

NumOTU	Iss	Iss.cSym	T	DF	P	Iss.cAsym	T	DF	P
4	0,537	0,796	11,883	510	0,0000	0,762	10,330	510	0,0000
8	0,580	0,753	7,928	510	0,0000	0,641	2,812	510	0,0051
16	0,600	0,722	5,837	510	0,0000	0,513	4,163	510	0,0000
32	0,608	0,703	4,751	510	0,0000	0,378	11,490	510	0,0000

COII 1st+2nd codon positions:

NumOTU	Iss	Iss.cSym	T	DF	P	Iss.cAsym	T	DF	P
4	0,258	0,793	31,684	456	0,0000	0,759	29,701	456	0,0000
8	0,224	0,747	30,309	456	0,0000	0,636	23,846	456	0,0000
16	0,274	0,713	25,787	456	0,0000	0,504	13,505	456	0,0000
32	0,285	0,697	25,475	456	0,0000	0,371	5,276	456	0,0000

COII 3rd codon positions:

NumOTU	Iss	Iss.cSym	T	DF	P	Iss.cAsym	T	DF	P
4	0,630	0,777	4,490	227	0,0000	0,762	4,033	227	0,0001
8	0,673	0,732	1,878	227	0,0617	0,631	1,319	227	0,1885
16	0,718	0,654	2,087	227	0,0380	0,459	8,498	227	0,0000
32	0,750	0,685	2,250	227	0,0254	0,363	13,387	227	0,0000

COIII 1st+2nd codon positions:

NumOTU	Iss	Iss.cSym	T	DF	P	Iss.cAsym	T	DF	P
4	0,230	0,797	34,999	525	0,0000	0,764	32,946	525	0,0000
8	0,236	0,754	32,263	525	0,0000	0,643	25,328	525	0,0000
16	0,261	0,725	29,985	525	0,0000	0,515	16,431	525	0,0000
32	0,281	0,705	29,101	525	0,0000	0,378	6,659	525	0,0000

COIII 3rd codon positions:

NumOTU	Iss	Iss.cSym	T	DF	P	Iss.cAsym	T	DF	P
4	0,651	0,778	4,004	262	0,0001	0,758	3,356	262	0,0009

8	0,716	0,732	0,487	262	0,6265	0,627	2,761	262	0,0062
16	0,748	0,666	2,525	262	0,0121	0,465	8,773	262	0,0000
32	0,796	0,682	3,501	262	0,0005	0,356	13,527	262	0,0000

ND1 1st+2nd codon positions:

NumOTU	Iss	Iss.cSym	T	DF	P	Iss.cAsym	T	DF	P
4	0,232	0,803	41,973	627	0,0000	0,772	39,681	627	0,0000
8	0,270	0,763	31,688	627	0,0000	0,653	24,621	627	0,0000
16	0,257	0,741	33,136	627	0,0000	0,530	18,739	627	0,0000
32	0,307	0,715	27,312	627	0,0000	0,388	5,436	627	0,0000

ND1 3rd codon positions:

NumOTU	Iss	Iss.cSym	T	DF	P	Iss.cAsym	T	DF	P
4	0,616	0,782	5,860	313	0,0000	0,756	4,938	313	0,0000
8	0,773	0,735	1,302	313	0,1939	0,626	5,017	313	0,0000
16	0,787	0,682	3,693	313	0,0003	0,475	10,927	313	0,0000
32	0,776	0,683	3,322	313	0,0010	0,354	15,165	313	0,0000

ND2 1st+2nd codon positions:

NumOTU	Iss	Iss.cSym	T	DF	P	Iss.cAsym	T	DF	P
4	0,462	0,806	19,700	683	0,0000	0,775	17,927	683	0,0000
8	0,505	0,768	14,386	683	0,0000	0,658	8,401	683	0,0000
16	0,505	0,747	13,360	683	0,0000	0,538	1,812	683	0,0704
32	0,598	0,721	6,691	683	0,0000	0,396	10,982	683	0,0000

ND2 3rd codon positions:

NumOTU	Iss	Iss.cSym	T	DF	P	Iss.cAsym	T	DF	P
4	0,778	0,784	0,226	341	0,8212	0,756	0,810	341	0,4185
8	0,828	0,737	3,275	341	0,0012	0,627	7,222	341	0,0000
16	1,031	0,689	12,631	341	0,0000	0,481	20,299	341	0,0000
32	1,037	0,685	13,325	341	0,0000	0,356	25,823	341	0,0000

ND3 1st+2nd codon positions:

NumOTU	Iss	Iss.cSym	T	DF	P	Iss.cAsym	T	DF	P
4	0,367	0,777	16,360	233	0,0000	0,761	15,719	233	0,0000
8	0,332	0,732	15,880	233	0,0000	0,630	11,819	233	0,0000
16	0,379	0,656	10,371	233	0,0000	0,460	3,005	233	0,0029
32	0,415	0,684	10,433	233	0,0000	0,362	2,066	233	0,0400

ND3 3rd codon positions:

NumOTU	Iss	Iss.cSym	T	DF	P	Iss.cAsym	T	DF	P
4	0,669	0,793	2,773	116	0,0065	0,821	3,396	116	0,0009
8	0,770	0,773	0,069	116	0,9452	0,707	1,383	116	0,1694
16	0,826	0,599	4,990	116	0,0000	0,469	7,845	116	0,0000
32	0,950	0,763	4,278	116	0,0000	0,512	10,022	116	0,0000

ND4L 1st+2nd codon positions:

NumOTU	Iss	Iss.cSym	T	DF	P	Iss.cAsym	T	DF	P
4	0,330	0,780	17,564	157	0,0000	0,784	17,747	157	0,0000
8	0,334	0,745	14,305	157	0,0000	0,658	11,299	157	0,0000
16	0,349	0,623	10,092	157	0,0000	0,454	3,879	157	0,0002
32	0,432	0,712	10,365	157	0,0000	0,417	0,545	157	0,5863

ND4L 3rd codon positions:

NumOTU	Iss	Iss.cSym	T	DF	P	Iss.cAsym	T	DF	P
4	0,659	0,833	3,248	78	0,0017	0,903	4,560	78	0,0000
8	0,707	0,848	2,665	78	0,0093	0,823	2,189	78	0,0316
16	0,793	0,573	4,465	78	0,0000	0,524	5,454	78	0,0000
32	0,839	0,891	1,139	78	0,2580	0,739	2,217	78	0,0296

ND4 1st+2nd codon positions:

NumOTU	Iss	Iss.cSym	T	DF	P	Iss.cAsym	T	DF	P
4	0,344	0,816	31,298	895	0,0000	0,785	29,198	895	0,0000
8	0,357	0,782	24,785	895	0,0000	0,675	18,555	895	0,0000
16	0,402	0,765	18,447	895	0,0000	0,563	8,178	895	0,0000
32	0,407	0,740	15,626	895	0,0000	0,428	0,971	895	0,3318

ND4 3rd codon positions:

NumOTU	Iss	Iss.cSym	T	DF	P	Iss.cAsym	T	DF	P
4	0,682	0,792	4,404	447	0,0000	0,759	3,068	447	0,0023
8	0,750	0,747	0,111	447	0,9118	0,635	4,355	447	0,0000
16	0,811	0,711	3,659	447	0,0003	0,502	11,350	447	0,0000
32	0,834	0,696	4,726	447	0,0000	0,369	15,958	447	0,0000

ND5 1st+2nd codon positions:

NumOTU	Iss	Iss.cSym	T	DF	P	Iss.cAsym	T	DF	P
4	0,390	0,826	28,458	1161	0,0000	0,794	26,372	1161	0,0000
8	0,375	0,796	23,303	1161	0,0000	0,692	17,559	1161	0,0000
16	0,413	0,780	17,244	1161	0,0000	0,587	8,194	1161	0,0000
32	0,473	0,758	11,503	1161	0,0000	0,461	0,452	1161	0,6514

ND5 3rd codon positions:

NumOTU	Iss	Iss.cSym	T	DF	P	Iss.cAsym	T	DF	P
4	0,723	0,801	3,337	580	0,0009	0,769	1,964	580	0,0501
8	0,823	0,759	2,356	580	0,0188	0,648	6,451	580	0,0000
16	0,885	0,734	4,978	580	0,0000	0,524	11,951	580	0,0000
32	0,973	0,710	7,593	580	0,0000	0,382	17,075	580	0,0000

ND6 1st+2nd codon positions:

NumOTU	Iss	Iss.cSym	T	DF	P	Iss.cAsym	T	DF	P
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4	0,463	0,781	12,219	307	0,0000	0,756	11,237	307	0,0000
8	0,528	0,734	7,426	307	111,3362	0,626	3,521	307	0,0005
16	0,564	0,680	4,021	307	728760384	0,474	3,118	307	0,0020
32	0,632	0,683	1,713	307	0,0877	0,354	9,373	307	0,0002

ND6 3rd codon positions:

NumOTU	Iss	Iss.cSym	T	DF	P	Iss.cAsym	T	DF	P
4	0,847	0,781	1,620	152	0,1073	0,788	1,454	152	0,1479
8	0,986	0,747	5,532	152	13484792,	0,662	7,495	152	510,3412
16	1,049	0,620	10,045	152	0,0002	0,455	13,912	152	0,0000
32	1,118	0,716	9,303	152	0,0014	0,425	16,021	152	0,0000

Interpretation of results:

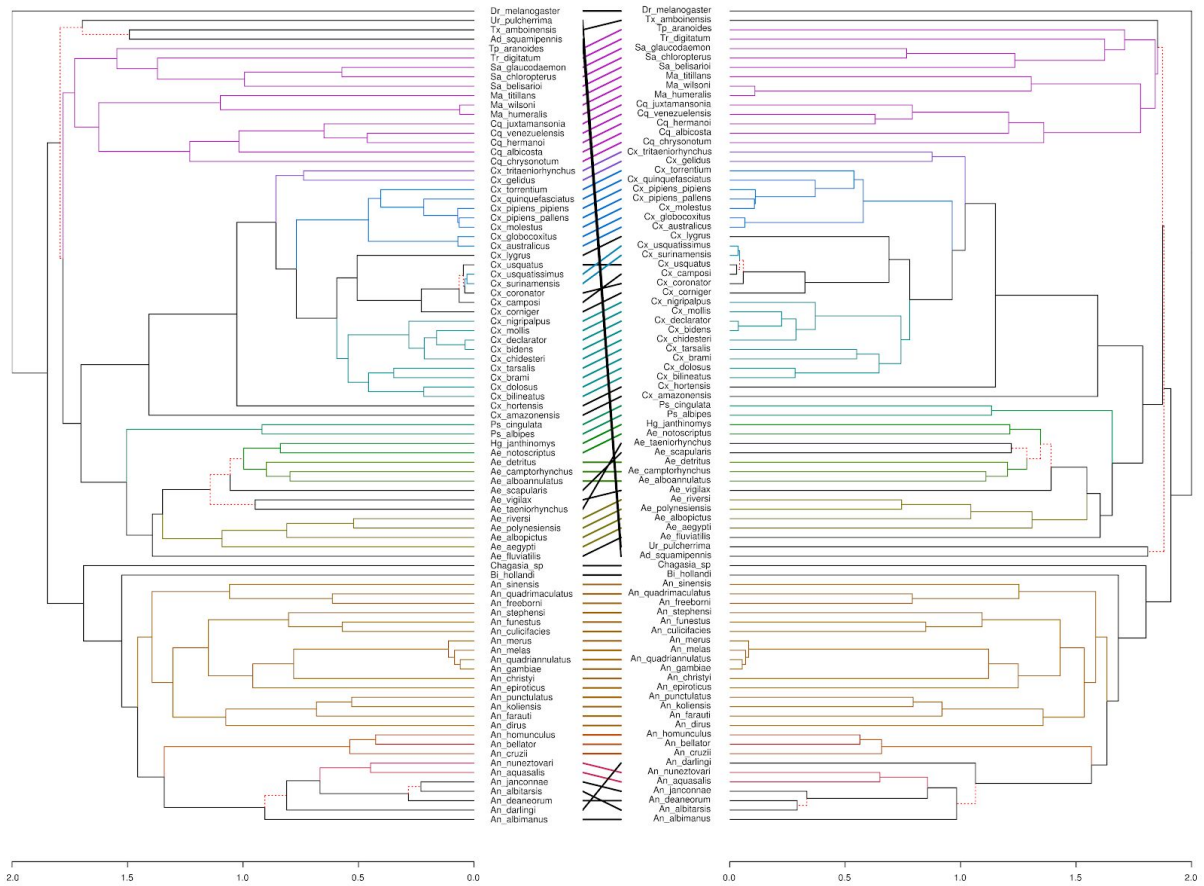
Significant Difference

 Yes No

 Iss < Iss.c Little Substantial
 saturation saturation

Complete nucleotide mitogenome alignment sequences

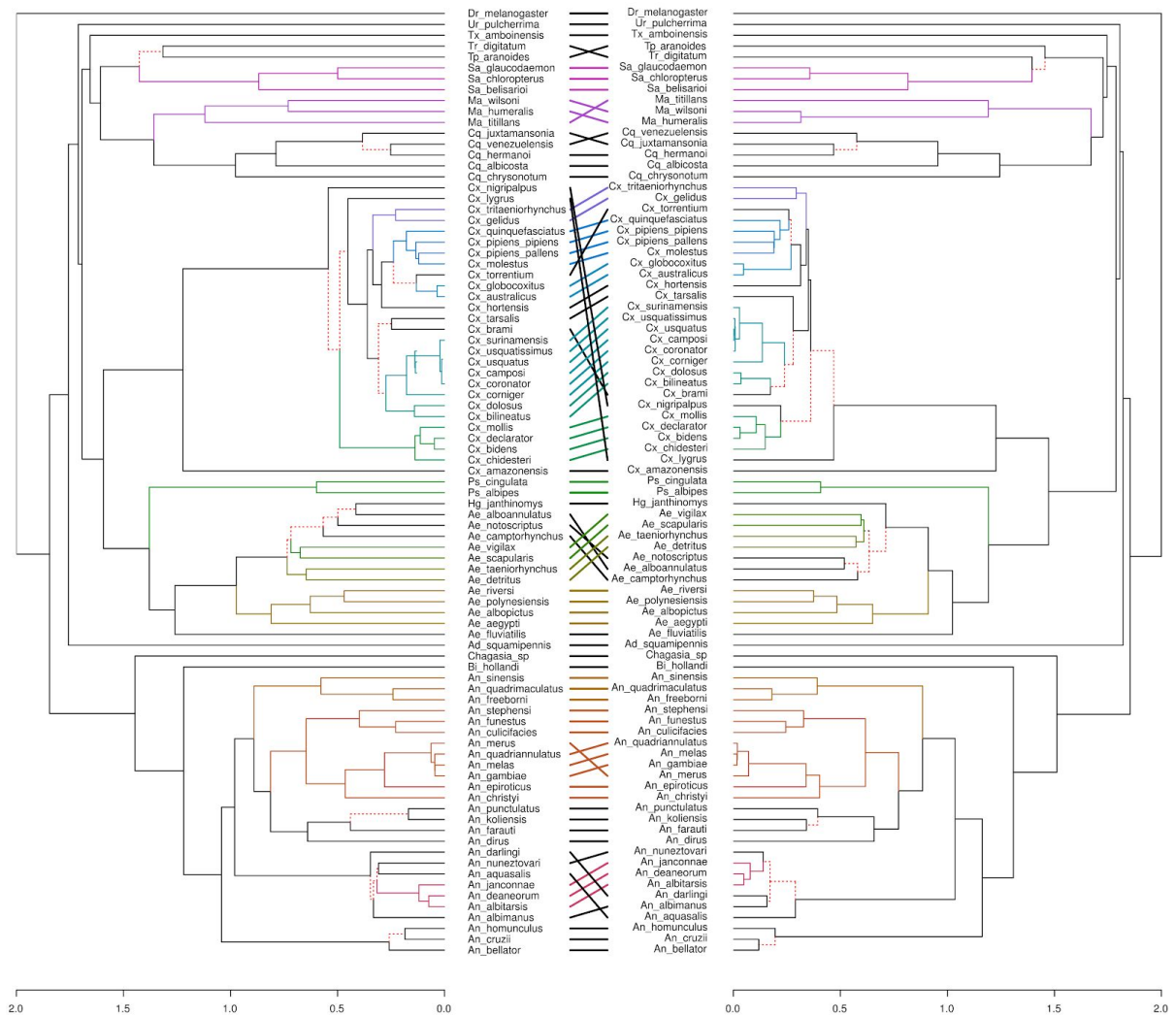
Partitioned PCG with codon partition



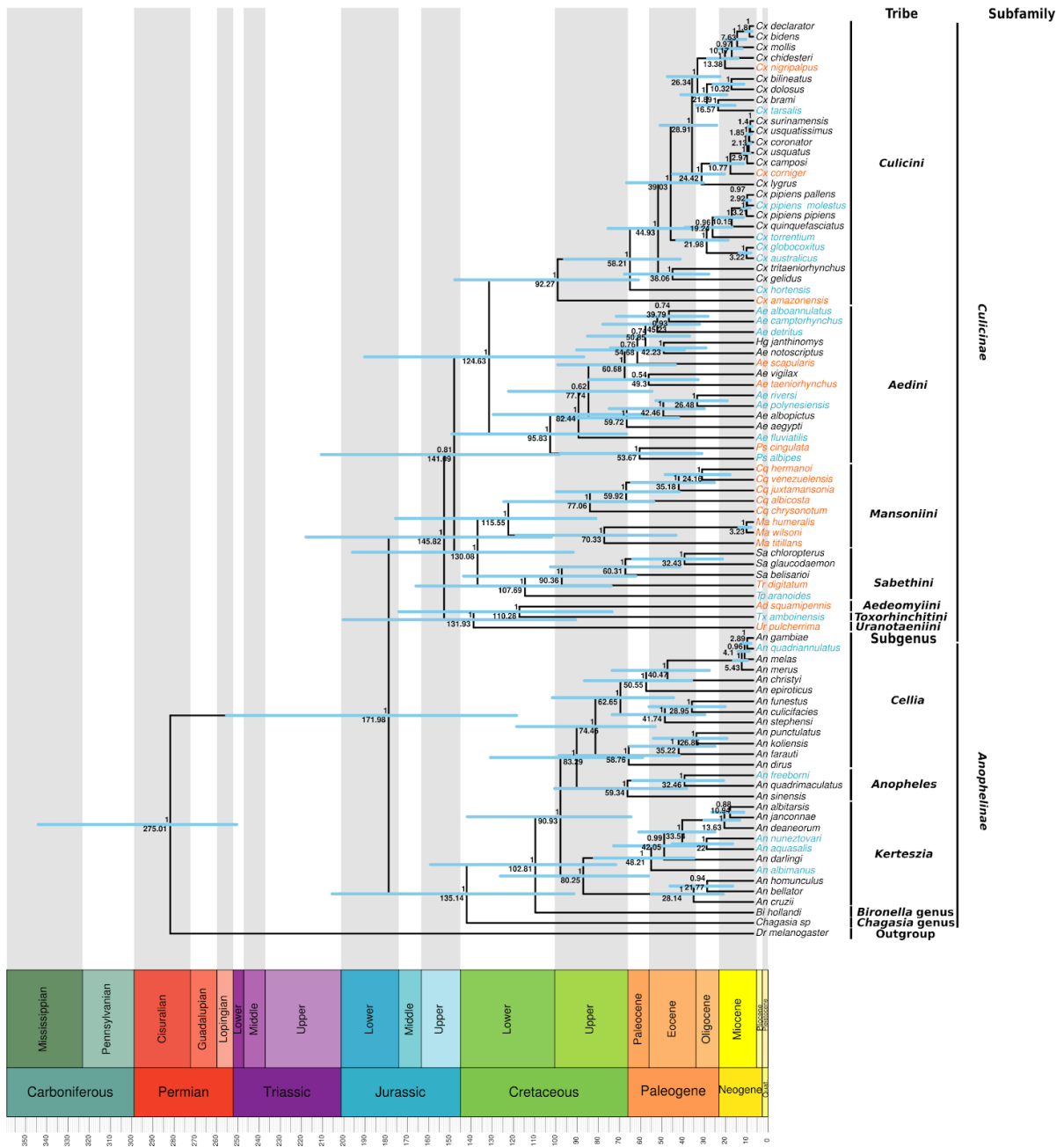
Supplementary figure 1. Comparative dendrogram topologies of trees obtained from different approaches using complete nucleotide mitogenomes and PCG with codon partition in the BEAST analysis. Dashed red lines show the branches that were different between the approaches used.

Concatenated predicted amino acid sequences

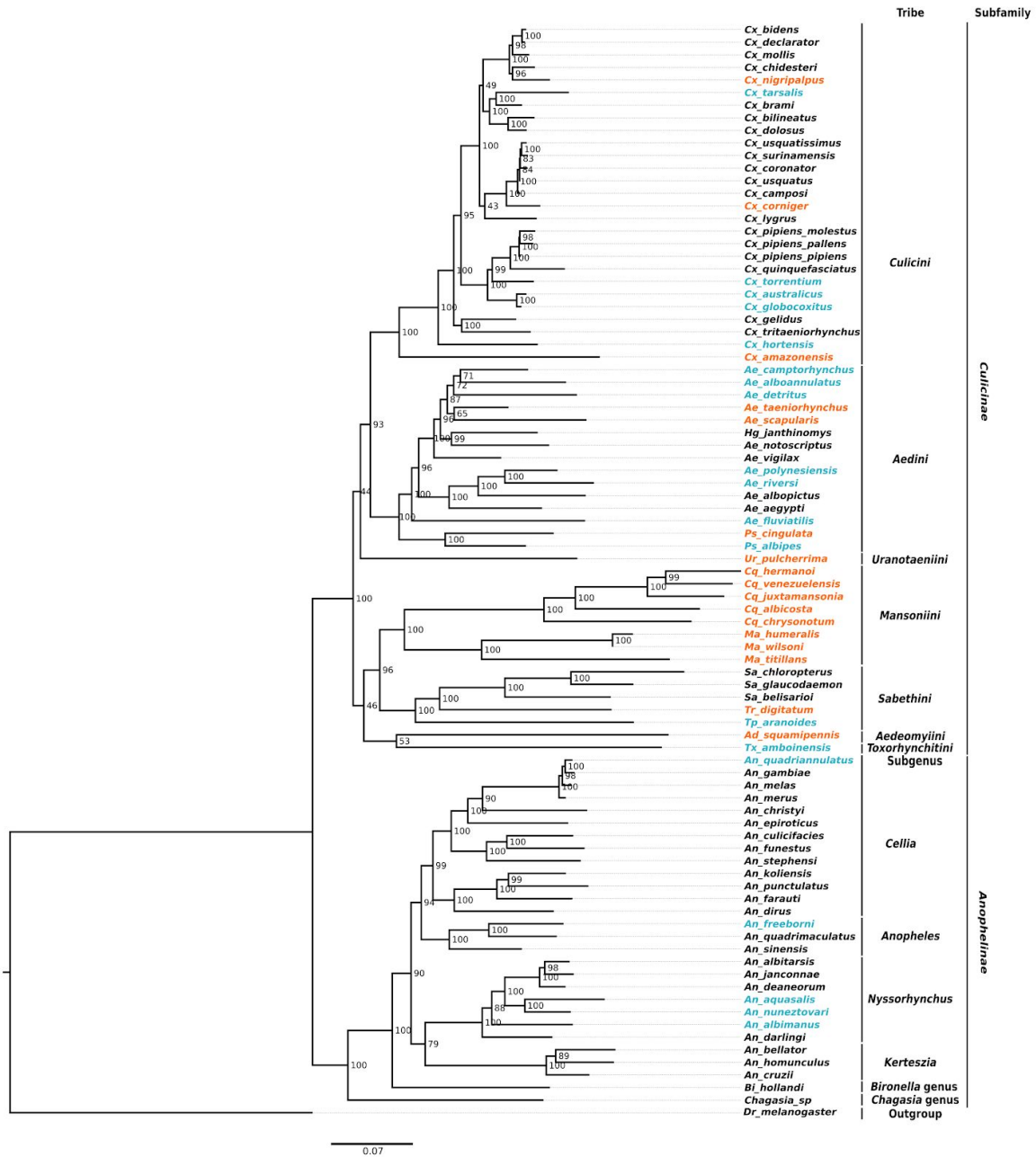
Partitioned predicted amino acid sequences



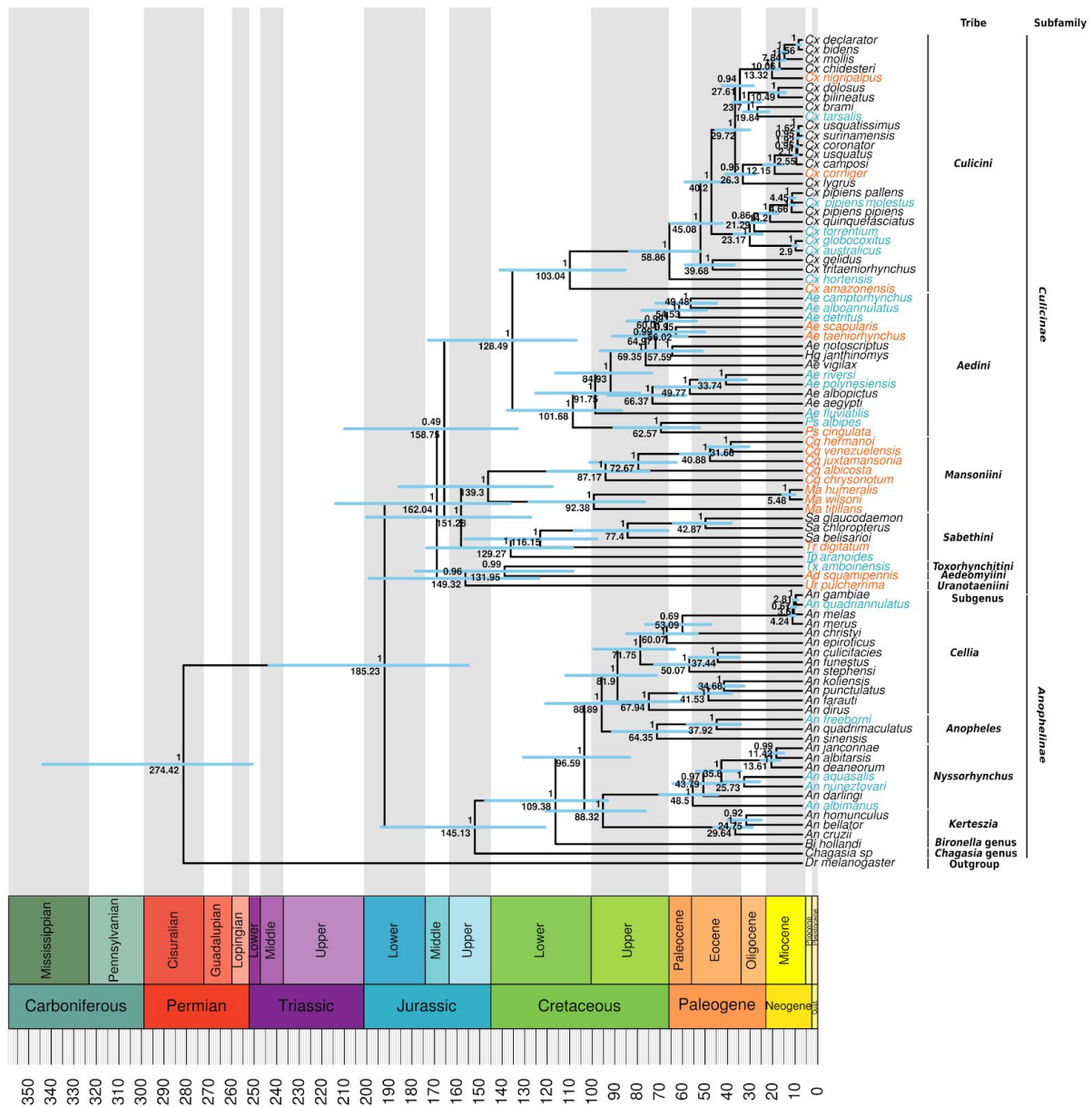
Supplementary figure 2. Comparative dendrogram topologies of trees obtained from different approaches using concatenated predicted amino acid sequences and partitioned predicted amino acid sequences in the BEAST analysis. Dashed red lines show the branches that were different between the approaches used.



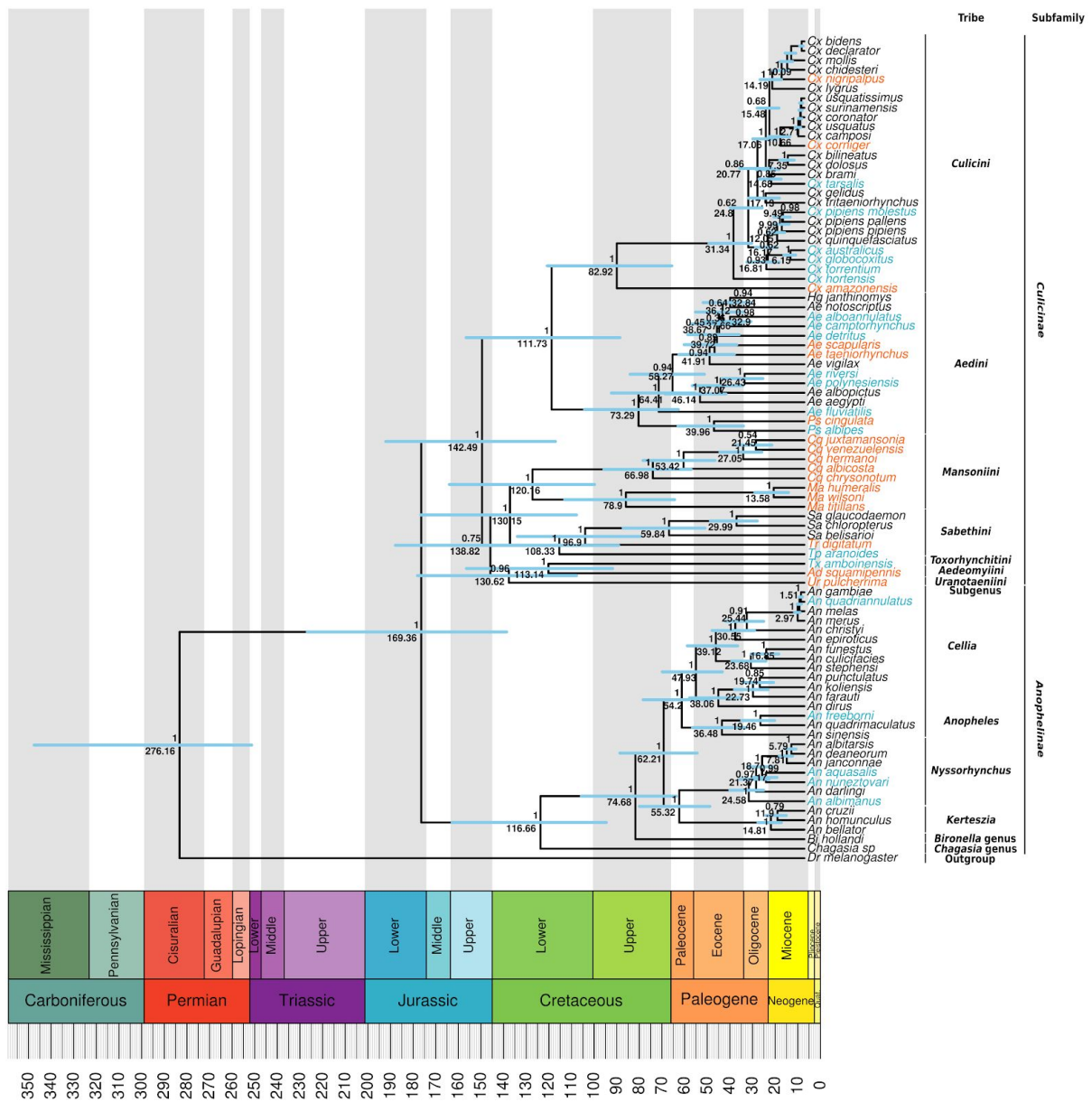
Supplementary figure 3. Evolutionary timescale of the Culicidae family based on BEAST analysis using complete mitogenome sequences. Blue bars represent the HPD95%. The numbers above and below the bars show the posterior probability and the predicted median dating respectively for each node. Light blue tip names represent mitogenomes characterized from SRA data. Orange tip names represent sequenced mitogenomes from this study.



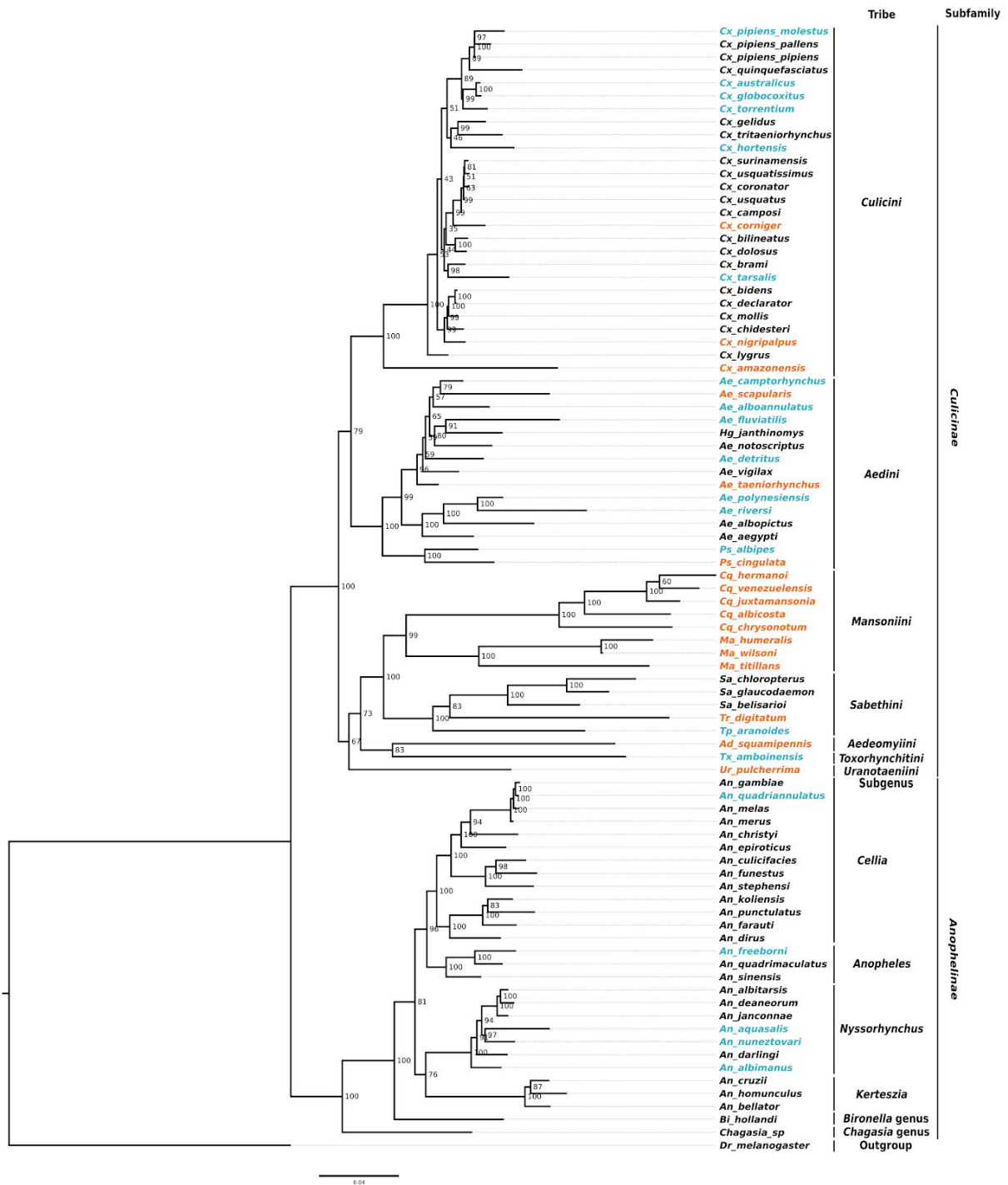
Supplementary figure 4. Maximum likelihood phylogenetic tree reconstructed with IQ-TREE version 1.6.12 based on complete mitogenome sequences using the GTR+F+I+G4 evolutionary model and the ultrafast bootstrapping with 1000 replicates. Light blue tip names represent mitogenomes characterized from SRA data. Orange tip names represent sequenced mitogenomes from this study.



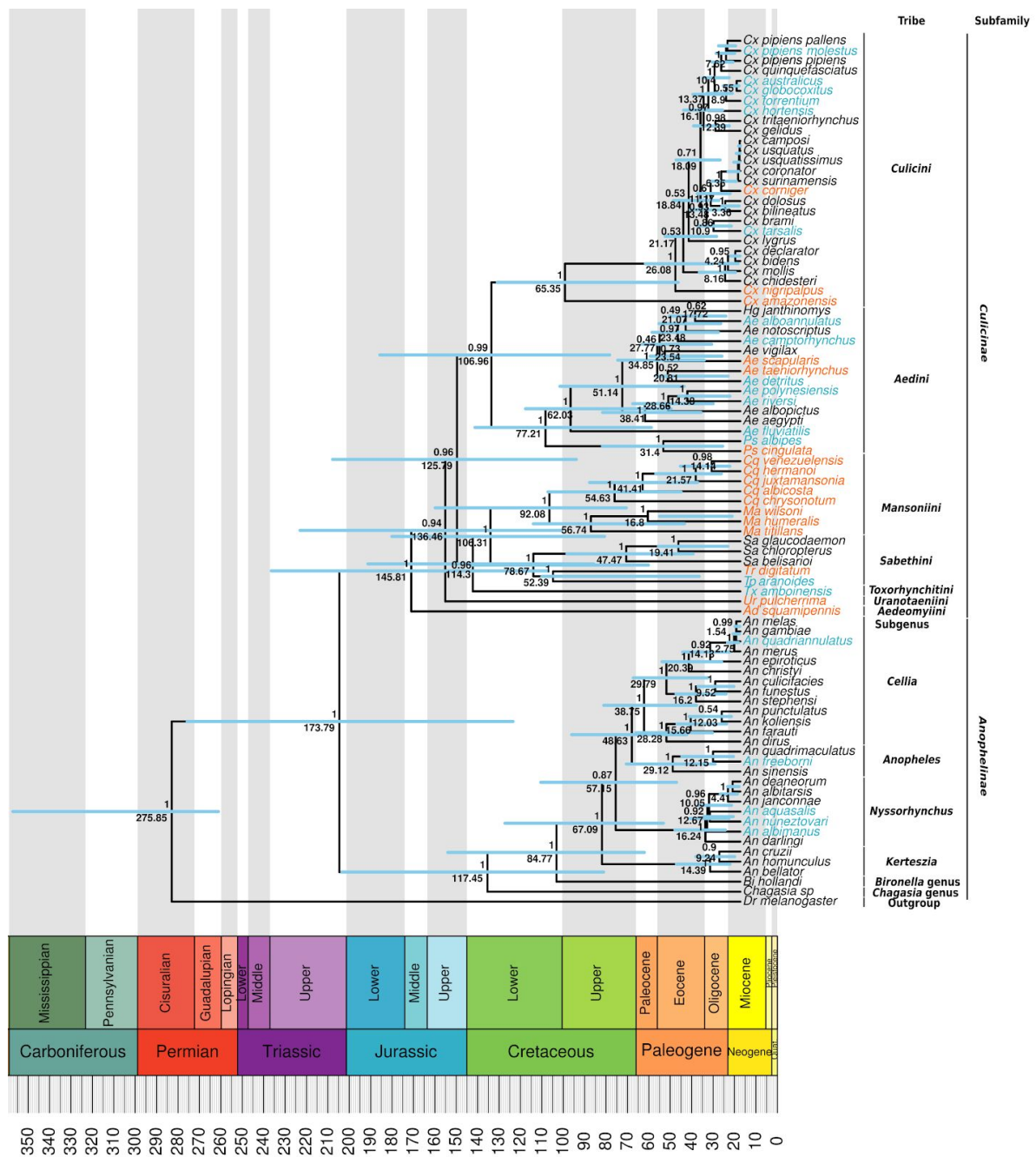
Supplementary figure 5. Evolutionary timescale of Culicidae family based on BEAST analysis using a partitioned scheme of PCG without codon partition. Blue bars represent the HPD95%. The numbers above and below the bars show the posterior probability and the predicted median dating respectively for each node. Light blue tip names represent mitogenomes characterized from SRA data. Orange tip names represent sequenced mitogenomes from this study.



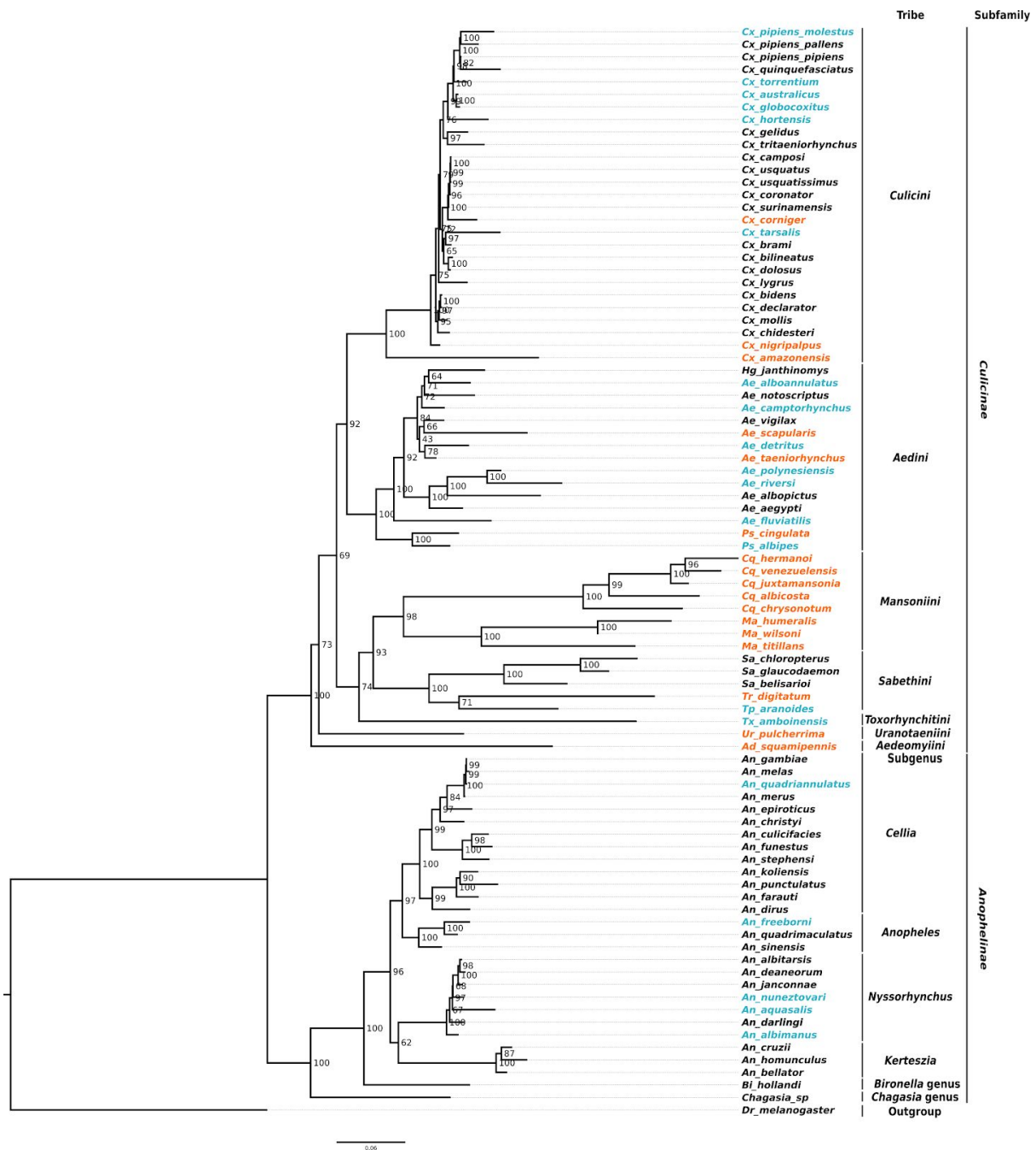
Supplementary figure 6. Evolutionary timescale of Culicidae family based on BEAST analysis using a partitioned scheme of PCG without 3rd codon positions. Blue bars represent the HPD95%. The numbers above and below the bars show the posterior probability and the predicted median dating respectively for each node. Light blue tip names represent mitogenomes characterized from SRA data. Orange tip names represent sequenced mitogenomes from this study.



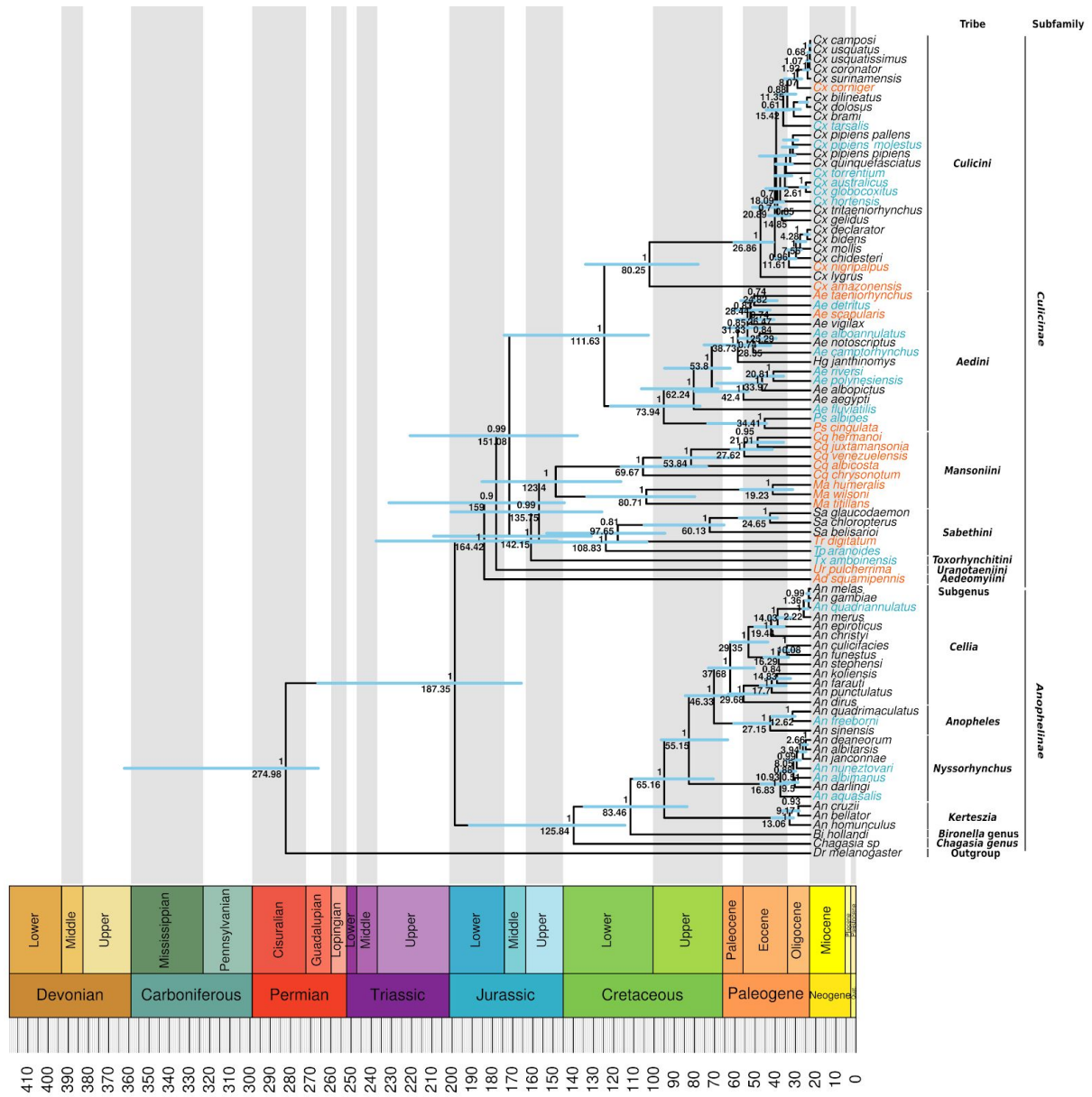
Supplementary figure 7. Maximum likelihood phylogenetic tree reconstructed with IQ-TREE version 1.6.12 based on concatenated PCG without 3rd codon positions using the GTR+F+I+G4 evolutionary model and the ultrafast bootstrapping with 1000 replicates. Light blue tip names represent mitogenomes characterized from SRA data. Orange tip names represent sequenced mitogenomes from this study.



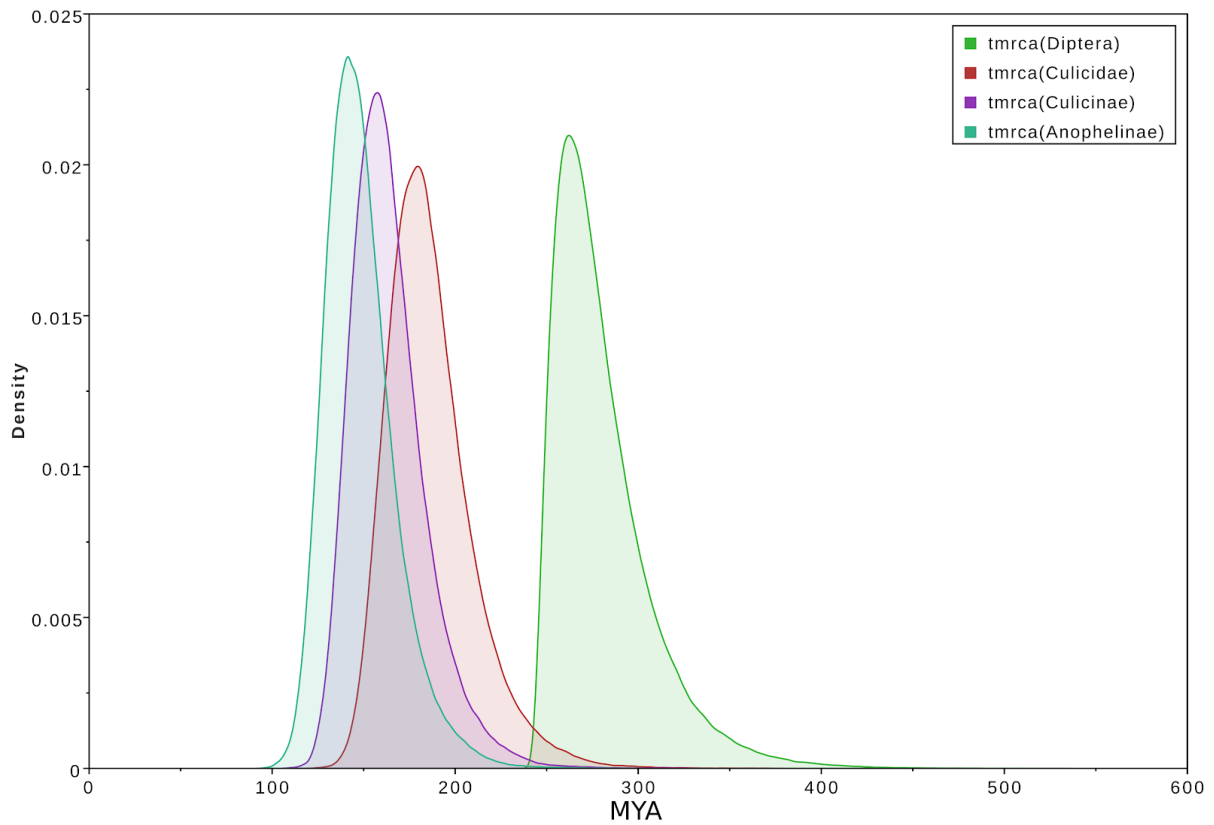
Supplementary figure 8. Evolutionary timescale of Culicidae family based on BEAST analysis using a concatenated predicted amino acid sequence. Blue bars represent the HPD95%. The numbers above and below the bars show the posterior probability and the predicted median dating respectively for each node. Light blue tip names represent mitogenomes characterized from SRA data. Orange tip names represent sequenced mitogenomes from this study.



Supplementary figure 9. Phylogenetic tree of Maximum likelihood reconstructed with IQ-TREE version 1.6.12 based on concatenated predicted amino acid sequences using the mtInv+I+G4 evolutionary model and the ultrafast bootstrapping with 1000 replicates. Light blue tip names represent mitogenomes characterized from SRA data. Orange tip names represent sequenced mitogenomes from this study.



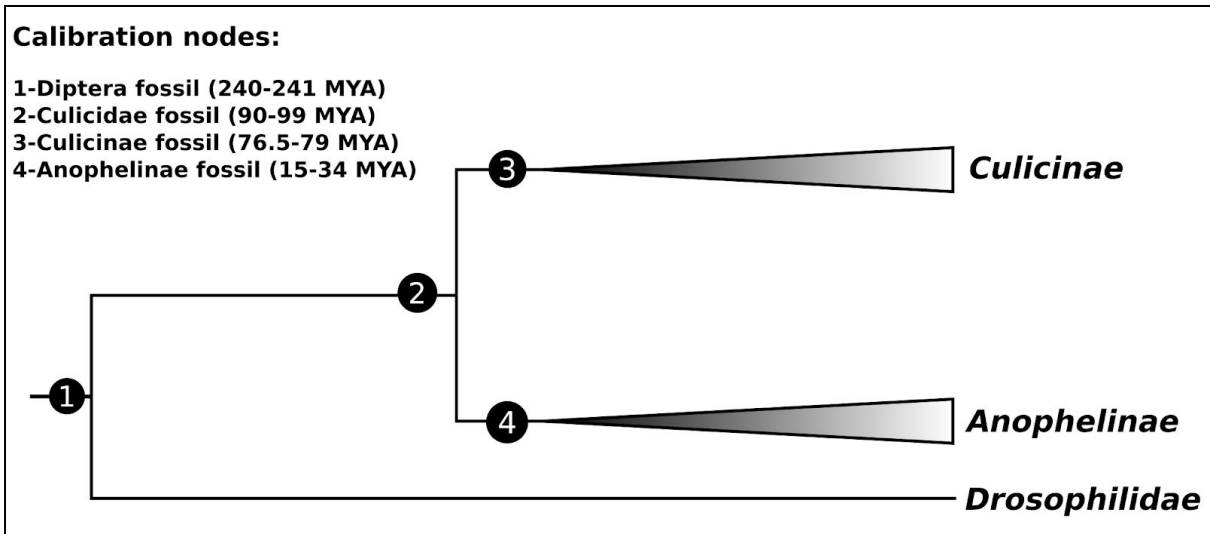
Supplementary figure 10. Evolutionary timescale of Culicidae family based on BEAST analysis using a partitioned scheme of predicted amino acid sequences. Blue bars represent the HPD95%. The numbers above and below the bars show the posterior probability and the predicted median dating respectively for each node. Light blue tip names represent mitogenomes characterized from SRA data. Orange tip names represent sequenced mitogenomes from this study.



Supplementary figure 11. Density plot showing the 95% posterior distribution of ancestors time estimate based on the alignment of partitioned mitochondrial PCG with codon partition.



Supplementary figure 12. Quality of raw reads generated from mosquito samples. Figure generated from FasqQC results and summarized by MultiQC tool. The yellow line represents results from *Cq. albicosta* reads and green lines represent the remaining species.



Supplementary figure 13. Schematic tree representing fossil dating used as calibration points in the molecular clock bayesian analyses. Circles with numbers inside represent each calibration point used based on the literature shown in Supplementary table 6.

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